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Convergent evolution of bird-mammal shared characteristics for adapting to nocturnality

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The diapsid lineage (birds) and synapsid lineage (mammals), share a suite of functionally similar characteristics (e.g. endothermy) that are considered to be a result of their convergent evolution, but the candidate selections leading to this convergent evolution are still under debate. Here, we used a newly developed molecular phyloecological approach to reconstruct the diel activity pattern of the common ancestors of living birds. Our results strongly suggest that they had adaptations to nocturnality during their early evolution, which is remarkably similar to that of ancestral mammals. Given their similar adaptation to nocturnality, we propose that the shared traits in birds and mammals may have partly evolved as a result of the convergent evolution of their early ancestors adapting to ecological factors (e.g. low ambient temperature) associated with nocturnality. Finally, a conceptually unifying ecological model on the evolution of endothermy in diverse organisms with an emphasis on low ambient temperature is proposed. We reason that endothermy may evolve as an adaptive strategy to enable organisms to effectively implement various life-cycle activities under relatively low-temperature environments. In particular, a habitat shift from hightemperature to relatively low-temperature environments is identified as a common factor underlying the evolution of endothermy.

1. Introduction

Extant birds and mammals share a number of highly similar characteristics, including but not limited to, enhanced hearing, vocal communication, endothermy, insulation, shivering, respiratory turbinates, high basal metabolism, grinding, sustained activity, four-chambered heart, high blood pressure, and intensive parental care [1-8]. These bird-mammal shared characteristics (BMSC) are considered to have evolved convergently in the two groups [4], while the selection pressures that underlie their convergent evolution are still under debate. To date, many candidate selections, which are not necessarily mutually exclusive, have been proposed [9-11] and popular views suggest that BMSC (e.g. endothermy) evolved primarily for increased sustained activity [1] and enhanced parental care [4,5]. On the other hand, the traditional view, the thermal niche expansion model, proposes that nocturnality may have facilitated the evolution of endothermy in ancestral mammals that would not necessarily feature a high resting metabolic rate typical of most living mammals [2,8]. Indeed, early mammals are widely accepted as having been nocturnal [12,13], and it has been hypothesized that the evolution of endothermy may have allowed them to overcome ambient low-temperature constraints to be active at night, while diurnal ectothermic reptiles essentially depend on sunlight to increase their body temperature to operational levels [12,14]. The nocturnality hypothesis is criticized as being suitable only to mammals [1], partly because

so far only the nocturnality of the synapsid lineages (e.g. early mammals) is well known, while the possible nocturnal activity of the diapsid lineages (e.g. birds), especially during their early evolution, is unknown.

The diel activity pattern of ancestral birds has received less attention. To date, there are only two relevant studies based on behaviour or morphology analyses. A maximumlikelihood reconstruction based on behavioural data suggests the diurnality of the common ancestor of living birds (CALB) [15]. This study classifies each species as either diurnal or nocturnal without consideration of other behavioural possibilities (e.g. crepuscular or cathemeral) or behavioural complexity, for instance, the widespread existence of the occasional nocturnality of typically diurnal birds [16], and hence this result may be less accurate. Another study uses a phylogenetic discriminant analysis to reconstruct the diel activity patterns of four fossil basal birds (Archaeopteryx lithographica, Confuciusornis sanctus, Sapeornis chaoyangensis, Yixianornis grabaui) based on eye morphology data, and the result demonstrates their diurnality [17]. The phylogenetic discriminant analysis approach is criticized for its inability to distinguish diurnality from cathemerality (i.e. active both day and night) [18,19], and hence uncertainty remains concerning the diel activity patterns of these basal birds. Given the uncertainties of the results, the diel activity pattern of ancestral birds remains unclear.

The recent development of a molecular phyloecological (MPE) approach allows us to reconstruct the diel activity pattern of ancestral taxa using molecular data [13,20-22]. The MPE approach has been applied to analyse the diel activity patterns of birds and mammals, and the results demonstrate its sensitivity to distinct animals with different diel activity patterns, showing that nocturnality, diurnality, and cathemerality are featured by enhanced positive selection of dimlight vision genes (rod-expressed genes), bright-light vision genes (cone-expressed genes), and both, respectively [13,20,22]. In this study, we employed the MPE approach to reconstruct the early evolution of the diel activity patterns in diapsid lineages. Our results reveal that diapsid lineages exhibit similar nocturnality adaption to that of synapsid lineages. Our findings provide insights into the candidate selection that underlies the convergent evolution of the shared characteristics, especially endothermy, in birds and mammals.

2. Results and discussion

(a) Evidence for the nocturnality of diapsid lineages

Following the MPE approach for studying the diel activity pattern of ancestral taxa [13,20–22], we reconstructed the diel activity patterns of our focal taxa by analysing the adaptive evolution of 33 phototransduction genes (electronic supplementary material, table S1) involved in the rod and cone phototransduction pathway [23,24] in the context of sauropsid phylogeny (figure 1). Positively selected genes were identified based on the branch-site model (electronic supplementary material, table S2), and the positive selection remained robust when phylogenetic uncertainties [25,37,38] (electronic supplementary material, tables S3 and S4) and initial value variations of kappa and ω were considered.

We initially examined the adaptive evolution of the phototransduction genes along the common ancestor branch of living birds. One rod-expressed gene, *GRK1*, showed the most significant positive selection signal ($\omega = 13.983$, d.f. = 1, p = 0.002) (electronic supplementary material, table S2 and figure S1), and another rod-expressed gene, SLC24A1, showed a positive selection signal with marginal significance $(\omega = 25.293, d.f. = 1, p = 0.053)$. GRK1 is rhodopsin kinase, involved in the inactivation of activated rhodopsin in rods. *SLC24A1* encodes the Na⁺/Ca²⁺-K⁺ exchanger and helps to extrude free calcium in the outer segment of rods to allow restoration of the cyclic guanosine monophosphate (cGMP) concentration. Both genes are involved in photoresponse recovery and aid in motion detection [39]. The finding of positive selection signals in the two rod-expressed genes suggests a particularly enhanced capability of the CALB for motion detection in dim-light conditions (e.g. nocturnality). Moreover, we found one cone-expressed gene, PDE6C, to be under positive selection ($\omega = 26.734$, d.f. = 1, p = 0.014). *PDE6C* encodes the hydrolytic subunits of cGMP phosphodiesterase (PDE6) in cones and is known to play a critical role in amplifying signals in the phototransduction cascade. The positive selection of the cone-expressed gene suggests intensified selection for increased visual acuity in bright-light environments (e.g. diurnality). Our findings of the positive selections of both the rod-expressed genes and the cone-expressed gene suggest that the CALB may have been active at both night-time and daytime, and that they may have evolved an enhanced capability for motion detection at night.

The combined positive selection of the rod- and coneexpressed genes may also be selected for mesopic vision in crepuscular conditions, in which both rods and cones are functionally active. To test if there are shifts in their wavelengths of maximal absorption (λ_{max}) towards the relatively abundant short-wavelength light in twilight, which characterizes crepuscularly active animals [13,20], we subsequently examined the possible spectral tuning of four cone opsin genes, LWS, RH2, SWS2, and SWS1, in the CALB. Among examined critical amino acid sites (electronic supplementary material, table S5) that are associated with spectral tuning of cone opsins [40-43], no critical amino acid replacement was found along the branch of the CALB, suggesting that change of the spectral tuning of the cone opsins is less likely. This may suggest that the CALB may have been mainly active at both night-time and daytime (e.g. cathemeral) rather than being mainly active crepuscularly. This is consistent with a previous morphological study showing that the early stem bird species, Archaeopteryx lithographica, is somewhat intermediate in orbital characteristics between nocturnality and diurnality [44].

We also reconstructed the diel activity pattern of an ancestral archosaur, one of the relatively distant CALB. Intriguingly, we found four rod-expressed genes, CNGA1, GNAT1, PDE6B, and SLC24A1, and two genes that are rodand cone-expressed, GUCY2D and RCVRN, which are involved in phototransduction activation or photoresponse recovery, to be under positive selection along the ancestral archosaur branch (electronic supplementary material, table S2). The positive selection of the mainly rod-expressed genes suggests enhanced visual acuity and enhanced motion detection in dim-light conditions, and hence strongly supports that the ancestral archosaur was probably nocturnal, consistent with previous studies [13,15,17]. Furthermore, these positively selected genes incorporate almost all of the principle components of the phototransduction pathway (electronic supplementary material, figure S2), suggesting that the



Figure 1. Diel activity pattern reconstruction. The phylogenetic relationships of species used in this study follows published literature [25–36] and the Tree of Life Web Project (http://tolweb.org/Passeriformes). The diel activity classification of taxonomic bird orders follows one published study [16]. Red shows bird orders involved in nocturnal and/or crepuscular activities. Except for five regularly nocturnal bird groups, which harbour mainly nocturnally active species, all other bird groups are considered as typically diurnal, but widely show occasional nocturnality or engage in partial behaviours during crepuscular periods [16].

adaptation to nocturnality leads to a substantial modification of the visual system of the ancestral archosaur. This finding is remarkably similar to that in ancestral mammals [13].

Altogether, our molecular reconstruction based on the phylogenetic analysis by maximum likelihood (PAML) results suggests that while the CALB may have been active both night and day, the ancestral archosaur was more likely nocturnal (figure 1). When the branch site-unrestricted statistical test for episodic diversification (BUSTED) was used to confirm the selection signals of these positively selected genes identified by PAML, one rod-expressed gene (*SLC24A1*) and one coneexpressed gene (*PDE6C*) along the ancestral bird branch and three rod-expressed genes (*CNGA1*, *PDE6B*, and *SLC24A1*) along the ancestral archosaur branch were found to be under positive selection (electronic supplementary material, table S6), providing further support for our reconstructed diel activity results of our focal taxa. In light of these robust reconstructions, our molecular results and previous studies [13,17] consistently suggest that the diapsid lineages may have experienced strong adaptation to nocturnality, in parallel with the synapsid lineages (e.g. early mammals) [13]. The similar evolution of nocturnality in both diapsid and synapsid lineages should be considered as a derived result through their independent and convergent evolution, given that one previous study shows that the ancestral amniote and ancestral reptile were predominately diurnal (figure 2) [13].

(b) Nocturnality and endothermy evolution in diapsid and synapsid lineages

Our molecular results suggest the CALB may have experienced nocturnal periods to some extent. This nocturnality is



Figure 2. Reconstructed diel activity patterns of main groups of amniotes based on the results of this study and previous studies [13,45]. Living endothermic groups and their possible early evolution of endothermy are shown in red based on previous studies [7,46–51].

also supported by multiple lines of morphological and behavioural evidence. (i) Theropod dinosaurs, representing one of the CALB, are reconstructed as mainly scotopic/ cathemeral based on eye morphology, suggesting that the dinosaur ancestors of birds were mainly active in dim-light conditions [17]. The discoveries of feathers (insulation) and endothermy or mesothermy in many theropod dinosaurs [46,52,53] would enable them to be active at low temperature during the night, and thus are consistent with their nocturnality. (ii) Though considerable controversy remains about its taxonomy, Protoavis texensis, which was regarded as the first bird, is characterized by owl-like large and forward-facing orbits, indicating its nocturnality [54]. (iii) Though most living birds are regarded as typically diurnal, they widely exhibit potential capability for nocturnal activities (e.g. night migration) (figure 1) [16,55], consistent with the possible nocturnal activity of the CALB. (iv) Birds have the largest eyes of any of the terrestrial vertebrates [56], and large eyes are normally linked to increased visual acuity and are generally a character of nocturnal taxa [16,57,58]. (v) Birds have evolved vocal communication and much more sensitive hearing than reptiles [3], suggesting their greater dependence on acoustic communication, which normally characterizes nocturnal species [57,58]. Altogether, our molecular results and the behavioural and morphological evidence strongly support the nocturnality of the CALB, which challenges its pure diurnality perspectives [15,17]. The underlying causes of the nocturnality of the CALB remain unclear, while for ancestral mammals, their nocturnality is generally considered to be a result of the predation and/or competition from diurnal reptiles (e.g. dinosaurs) [59-61].

Crompton *et al.* proposed that the evolution of endothermy in synapsid lineages such as mammals may be facilitated by their nocturnality [2]. Similarly, we propose that nocturnality may have facilitated the evolution of endothermy in diapsid lineages to some extent as well, since ancestral archosaurs, theropod dinosaurs [17], and ancestral birds were found to be subjected to a similar nocturnal adaptation (figure 1). In support of this, previous studies demonstrate that the evolution of endothermy in diapsids may be traced back to non-avian dinosaurs [7,46,62,63] or even earlier to basal archosaurs [46-48]. In particular, the existence of a four-chambered heart in both crocodiles and birds suggests that basal archosaurs may have evolved a four-chambered heart, which may strengthen their possible endothermy [48]. Similarly, for synapsid lineages, their nocturnality has been determined to have occurred much earlier (e.g. the Late Carboniferous almost 300 Ma) than true mammals [45], and their endothermy has been demonstrated to have evolved much earlier than true mammals as well, and may be traced back to the common ancestor of Neotherapsida, more than 260 Ma [7,49-51]. In light of this, it therefore appears that the timing of the evolution of endothermy in both the diapsid and synapsid lineages coincides with the appearance of their nocturnality, supporting the nocturnality hypothesis as a candidate explanation for their evolution of endothermy. In addition to endothermy, birds and mammals share many other characteristics, and the time for their independent evolution in the two lineages remains largely unknown. Future palaeobiological findings may help to determine the timing of their origins and help to understand the possible effects of nocturnality on their evolution.

(c) Nocturnality and the integrated evolution of birdmammal shared characteristics

The diapsid (e.g. birds) and synapsid (e.g. mammals) lineages share many functionally similar characteristics, such as enhanced hearing, vocal communication, endothermy, insulation, shivering, respiratory turbinates, high basal metabolism, grinding, sustained activity, four-chambered heart, high blood pressure, and intensive parental care [1-8]. The possible integrated and correlated evolution of these BMSC has long been recognized [1,4,8,14]. Here, we attempt to explain their possible integrated evolution in the context of the adaptation to nocturnality (figure 3).

During the shift towards nocturnality of a diurnal amniote (reptile) ancestor (figure 2), two basal pressures: visual function constraint and low ambient temperature, may have exerted strong selection pressure for integrated evolution of the BMSC. The visual functional constraints

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Figure 3. Integrated evolution of bird-mammal shared characteristics for adapting to nocturnality. Arrows show possible cause and effect relationships between two nocturnality-associated selection pressures (visual constraints and low temperature) and the evolution of BMSC. Please see text for details. The black background represents dim-light environments (e.g. night) and red represents the evolution of endothermy-related characteristics under low-temperature selection pressure. BMR, basal metabolic rate.

may lead to the evolution of non-visual senses, such as enhanced hearing and vocal communication, as a result of compensatory selection pressure under dim-light conditions. In support of this, nocturnal animals generally have highly developed non-visual senses [57,58,64]. For the lowtemperature selection pressure, previous studies suggest that it may lead to the evolution of endothermy, insulation (fur and feathers), respiratory turbinates, or even shivering. Endothermy has long been considered as an adaptive strategy to cold temperature [2,8,65-68] given the temperature-dependence of many life-cycle activities [14]. Insulation (fur and feathers) and respiratory turbinates are believed to reduce heat loss [4,7,8], and shivering may then evolve to generate additional heat for rapid elevation of core temperature. Though its underlying thermogenesis mechanism is not fully understood, endothermy is considered to be partly linked to high basal metabolic rate (BMR) [1], which may then require not only accelerated assimilation through, for instance, grinding by mastication in mammals and in the gizzard in birds but also sufficient food supply [8]. The increased food requirement may subsequently exert selection

for enhanced sustained activity for efficient food acquisition. Both high BMR and sustained activity depend on an adequate oxygen supply, which may subsequently have led to the evolution of the four-chambered heart, high blood pressure, and other related anatomical structures [4].

The two selections, namely the visual function constraint and low ambient temperature associated with nocturnality, may have favoured the evolution of intensive parental care as well. A juvenile of small body size has apparent disadvantages during cold nights due to its higher surface area to volume ratio, and intensive parental care would necessarily help to reduce offspring mortality [8]. With the benefits of parental care, endothermy, hearing, and vocal communication would be further strengthened to promote the offspring's survival rate [4] at night. Despite this, parental care is believed to be accompanied by high costs to provision young, and this may exert additional selection pressure on sustained activity to obtain extra food [4,5]. In addition to supporting foraging, sustained activity inevitably generates heat continuously (activity thermogenesis) [1,69] and may work as an important source of thermogenesis to contribute to endothermy. Activity thermogenesis, along with thermogenesis from BMR and shivering, may have worked as three principle thermogenesis sources contributing to endothermy, thereby guaranteeing the implementation of various life-cycle activities under low-temperature conditions. Taken together, the evolution of BMSC is consistent with the adaptation to nocturnality (figure 3), and the evolution of the BMSC may be a consequence of the convergent adaption of birds and mammals to specific selections associated with nocturnality.

(d) Low temperature and endothermy evolution

Our results suggest that the low-temperature selection pressure associated with nocturnality may have facilitated the evolution of endothermy in both diapsid and synapsid lineages. Intriguingly, although there is some controversy, low temperature is also believed to underlie the evolution of regional endothermy found in fishes (e.g. lamnid sharks, billfishes, and tunas). These warm water-adapted fishes are found to frequently dive into cool water, and the origin of their endothermy was proposed to be linked to multiple oceanic cooling events since the Eocene [65-67]. Likewise, the endogenous thermogenesis of the endothermic leatherback turtle (Dermochelys coriacea) is found to be low-temperature dependent [70]. It is possible that low temperature also plays a role in the evolution of facultative endothermy, which was discovered in at least two reptiles during the reproductive period, the Indian python and the tegu lizard [71,72]. The Indian python only starts to generate heat for incubation when the ambient temperature is below a critical low temperature (33°C) [71], which likely suggests a possible association between the appearance of facultative endothermy and the occurrence of relatively low ambient temperature. In addition, even for endothermic insects and plants, some explanations of the appearance of their endothermy have incorporated the effects of low ambient temperature [73-75]. It is conceivable that all of these endotherms would not necessarily have evolved to generate heat themselves if their external environment constantly were similar to their elevated body temperature.

Given the possibility of the evolution of endothermy due to low temperatures (figure 3), parental care and sustained activity, which are popularly regarded as the initial selection pressures for the evolution of endothermy [1,4,5], may be only partial consequences of selection for various life-cycle activities under low-temperature conditions, instead of the sole or even the initial selective target. Regarding the two popular views, criticism can be found elsewhere [4,5,76], while one additional issue is that no explanation is provided to interpret why only avian and mammalian lineages were selected for sustained activity or intensive parental care but other living reptiles have not done likewise. In addition to parental care and sustained activity, the body-size miniaturization, which occurs in parallel along the evolutions of both avian and mammalian lineages, is also proposed to contribute to the arising of endothermy [68,77]. However, the miniaturization model is criticized elsewhere [9]. In fact, the model was originally used to interpret how endothermy may evolve in mammals rather than explaining the causes, and the underlying cause may be partly linked to the nocturnality of ancestral mammals [68]. Moreover, previous studies indicate the evolution of endothermy predates the occurrences of their miniaturization in both avian and mammalian lineages [68,77,78], suggesting that body-size miniaturization may not be the initial selection driver for their evolution of endothermy. With respect to these different models on the evolution of endothermy, even for the biological functions of endothermy, it would be biased to only emphasize one of its many potential biological functions, leading to many theoretical models applicable only to specific groups [9,11]. For instance, neither the aerobic capacity model [1] nor the parental care model [4] explain the evolution of regional endothermy found in fishes, which are known to have evolved for other possible functions rather than for sustained activity or parental care [65,67]. In fact, for all different endothermic organisms found to date, endothermy is involved in diverse functions, spanning all possible life-cycle activities related to locomotion, sensory communication, foraging, digestion, reproduction, breeding, antipredation, and parental care [1,4,14,65,67,73]. A conceptually unifying model accounting for all of these diverse functions of endothermy in the context of low-temperature environments would be more reasonable to explain the appearance of endothermy in diverse organisms. In light of this, we propose that endothermy may evolve as an adaptive strategy to enable organisms to efficiently implement various life-cycle activities (e.g. parental care and sustained activity) under relatively low-temperature environments.

(e) Endothermy evolution and the habitat shift from high-temperature environments to low-temperature environments

The diel activity changes of both diapsid and synapsid lineages indicate that they may have experienced a habitat shift from high-temperature environments (day) to relatively low-temperature environments (night) (figure 2), which may have facilitated the evolution of their endothermy. Similarly, it is suggested that the evolution of endothermy in diverse fishes may be linked to their habitat change from warmwater habitat to cold-water habitat [65-67]. Likewise, for leatherback turtles, the appearance of their endothermy is associated with their habitat change from tropical nesting sites to cold northern foraging grounds [70]. For the facultative endothermic reptile, Indian python (Python molurus) [71], one molecular phylogeographic study reveals a possible origin of the genus Python in West Africa followed by dispersal to Europe and Asia [79], suggesting that the evolutionary origin of the Indian python may have experienced a habitat shift from high-temperature areas (tropical climate) to relatively low-temperature areas (subtropical/temperate climate). The other facultative endothermic reptile, the tegu lizard (Salvator merianae) [72], might have experienced a similar high-to-low temperature habitat shift, as it extends geographical distribution from tropical, subtropical to relatively low-temperature, temperate climates compared with its closest relatives (Salvator duseni, Salvator rufescens) [80].

In light of these findings, a habitat shift from hightemperature to relatively low-temperature environments seems to be a common factor that underlies the evolution of endothermy in these endothermic vertebrates. This specific habitat shift may be an important means to stimulate the evolution of endothermy in vertebrates. Endothermy would be selectively favoured in low-temperature environments

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for the optimum activity of enzymes and other molecules that had been adapted to previous relatively high-temperature conditions. If this is true, we may expect that endothermy would be more likely to be found in taxa with their biogeographical histories involving a habitat shift from high-temperature environments to relatively low-temperature environments. This pattern remains to be tested in relevant endothermic vertebrates, insects, and plants [73,75].

Endothermy may not be the sole strategy to adapt to lowtemperature environments. Apparently, some ectothermic animals, for instance, geckos and cold-water fish, are capable of being active in cold-temperature environments, suggesting the existence of alternative physiological or behavioural strategies to respond to cold temperatures. In addition to low temperature, many other possible factors such as biogeographic history, food availability, body size, and selection intensity may also affect the evolution of endothermy. Future study through a comparative and integrated analysis of possible factors between endotherms and their phylogenetically closest ectotherms should help to explain why endothermy is only restricted to some specific groups.

3. Conclusion

This study provides strong evidence for the nocturnality of the diapsid lineages, paralleling that of the synapsid lineages. Given their nocturnality, an integrated perspective on the evolution of BMSC as a convergent adaptation to nocturnality is proposed. Moreover, after summarizing our findings and relevant empirical studies on the evolution of endothermy, low temperature is suggested as a possible common factor underlying endothermy evolution in vertebrates. Given the significance of low temperature in endothermy evolution, a conceptually unifying ecological model of endothermy evolution with an emphasis on low temperature is proposed. We reason that endothermy may evolve as an adaptive strategy to enable organisms to effectively implement various life-cycle activities under relatively low-temperature environments, which happens during a habitat shift from a high-temperature environment to a relatively low-temperature environment.

4. Material and methods

(a) Taxa covered

The retinal transcriptome sequencing data of 17 bird species of which 15 were published previously by our laboratory [20] and two newly generated in this study, along with all of the available genome data of birds and reptiles in GenBank were used in this study (electronic supplementary material, table S1). In total, 95 species were used, of which 80 bird species from 34 orders, representing the majority of living bird orders (34/39) [81], were covered. During our initial sequence analyses, we found several of our focal phototransduction genes missing from palaeognath genome data, and thus two species, the ostrich (*Struthio camelus*) and the emu (*Dromaius novaehollandiae*), were selected to be subjected to retinal transcriptome sequencing.

(b) Sampling, RNA isolation, and cDNA library

construction

One ostrich (three months old) and one emu (six months old) were obtained with permission from an artificial breeding

company (Quanxin, Daqing). Active individuals were transported to the laboratory on 2 December 2017 and maintained under natural light:dark conditions with food and water provided. The ostrich and emu were sacrificed after 24 h and their retinal tissues were dissected during the daytime (15.00 to 16.00) and were preserved in RNA-locker (Sangon Biotech, Shanghai), flash frozen in liquid nitrogen, and then stored at -80 °C until processed for RNA isolation one week later. The experimental procedures were performed in accordance with an animal ethics approval granted by Northeast Normal University. All experimental procedures carried out in this study were approved by the National Animal Research Authority of Northeast Normal University, China (approval number: NENU-20080416) and the Forestry Bureau of Jilin Province of China (approval number: [2006]178).

Total RNA isolation was performed using TRIzol Reagent (Invitrogen Life Technologies), following the manufacturer's protocol and instructions. RNA degradation and contamination was monitored on 1% agarose gels. RNA purity was checked using the NanoPhotometer® spectrophotometer (IMPLEN, CA, USA). RNA concentration was measured using the Qubit® RNA Assay Kit in a Qubit[®]2.0 Flurometer (Life Technologies, CA, USA). RNA integrity was assessed using the RNA Nano 6000 Assay Kit of the Agilent Bioanalyzer 2100 system (Agilent Technologies, CA, USA). For cDNA library construction, a total amount of 3 µg RNA per sample was used as input material for the RNA sample preparations. The cDNA library was generated using NEBNext®UltraTM RNA Library Prep Kit for Illumina[®] (NEB, USA) according to the manufacturer's protocol. Specifically, mRNA was purified from total RNA using poly-T oligo-attached magnetic beads. The enriched mRNA was fragmented into small pieces and was then used for the syntheses of the first and the second strands of cDNA. The cDNA was purified with the AMPure XP system (Beckman Coulter, Beverly, MA, USA), and then 3 µl USER Enzyme (NEB, USA) was used with size-selected, adaptor-ligated cDNA at 37°C for 15 min followed by 5 min at 95°C before PCR. Then, PCR was performed, PCR products were purified (AMPure XP system) and library quality was assessed on the Agilent Bioanalyzer 2100 system. Paired-ending sequencing of the cDNA library was performed using Illumina HiSeq X Ten (Biomarker Technology Co., Beijing).

(c) Data filtering and de novo assembly

In total, 10.48 G bases and 12.05 G bases were generated for the ostrich and emu, respectively. Raw data were filtered by removing reads containing adapter, reads containing ploy-N and low-quality reads, and clean reads with high quality were obtained and used for de novo assembly. De novo assembly was performed using Trinity [82] with default parameters except that min_kmer_cov = 2 was used, ensuring that only k-mers with at least $2 \times$ coverage were used. The longest transcript was used to yield unigenes, and the unigenes longer than 200 bp were retained for further analyses. The transcriptome sequencing data of the two species have been deposited into the National Center for Biotechnology Information Sequence Read Archive database under accession number (SRP134220).

(d) Sequence abstraction and alignment

The 33 phototransduction genes involved in both the rod phototransduction pathway and the cone phototransduction pathway were used in this study [23]. To extract the coding sequences from the ostrich and the emu unigene pools, the coding sequences of *Gallus gallus* downloaded from GenBank were used as query sequences to blast against the unigene pools of the two species by using Blastn. The unigene sequences obtained were then annotated by blasting against the National Center for Biotechnology Information (NCBI) nr/nt database using the online Blastn, and only the unigene sequences with the same gene annotation as that of the query sequences were used for subsequent analyses. In addition to these two species, the coding sequences of the 33 genes from our previously published 15 bird species were also included [20]. Additionally, we downloaded the longest transcript sequences of our target genes from all other birds and reptiles with genome data available in Gen-Bank. Gene sequences were aligned using webPRANK (http:// www.ebi.ac.uk/goldman-srv/webprank/), and individual sequences with low identities, long indels, multiple ambiguous bases Ns, and/or too short a length were removed or replaced by other relevant transcript variants. After this pruning, highquality alignments were constructed, and their translated protein sequences were blasted against the non-redundant protein sequence database to confirm the correctness of sequence cutting.

(e) Positive selection analyses

We used the branch model and branch-site model implemented in the Codeml program of PAML [83] for positive selection analyses. Upon analysis, we initially constructed an unrooted species tree following published studies [25–36] and the Tree of Life Web Project (http://tolweb.org/Passeriformes). The phylogenetic relationships among bird orders followed one genome-level study [25]. We used a codon-based maximumlikelihood method to estimate the ratio of non-synonymous to synonymous substitutions per site (dN/dS or ω), and employed likelihood ratio tests (LRT) to determine statistical significance. A statistically significant value of $\omega > 1$ suggests positive selection.

(i) Branch model

We used a two-rate branch model to detect possible positive selection signals along the ancestral bird branch and the ancestral archosaur branch. For analyses, the two branches were respectively labelled foreground branches and others were treated as background branches. In the two-rate branch model, ω is assumed to be different between foreground branches and background branches, and its goodness of fit was analysed using the LRT, in which the two-rate branch model was compared with the one-rate branch model that assumes a single ω value across the tree. If a statistically significant value of $\omega > 1$ in a foreground branch was detected, the two-ratio branch model was then compared with the two-ratio branch model with a constraint of $\omega = 1$ to further determine whether the $\omega > 1$ of the foreground branch was statistically significant.

(ii) Branch-site model

We used a branch-site model (Test 2) to detect positively selected sites for our focal branches (foreground branches). In this model, four classes of sites are assumed. Site class 0 ($0 < \omega_0 < 1$) and site class 1 ($\omega_1 = 1$) represent evolutionarily conserved and neutral codons, respectively, for both background branches and foreground branches. Site classes 2a and 2b represent evolutionarily conserved ($0 < \omega_0 < 1$) and neutral ($\omega_1 = 1$) codons, respectively, for background branches but are allowed to be under positive selection ($\omega_2 > 1$) for the foreground branches. Test 2 compares a modified model A with its corresponding null model with $\omega = 1$ constrained to determine the statistical significance. The empirical Bayes method was used to detect positively selected sites.

(f) Robustness test of positive selection genes

The positively selected genes detected were further subjected to a robustness test. First, to detect the dependency of our positive

selection results on the initial value variations of kappa and omega, we used two different initial values of kappa (kappa = 0.5, 3.0) and of omega ($\omega = 0.5, 2.0$) for our positive selection analyses. In total, four independent runs were conducted for each of the positively selected genes found. Moreover, we examined the effects of phylogenetic uncertainties on our positive selection results. So far, considerable phylogenetic inconsistencies among different bird orders have been found. To examine the phylogenetic uncertainty effects, we used three different phylogenetic relationships of bird orders [25,37,38] for our positive selection analyses. For convenience, the three different phylogenetics were referred to as Hackett's tree, Jarvis's tree, and Prum's tree in our study.

To ensure the robustness of our results, we also employed BUSTED, implemented in the HyPhy software (v.2.2.4) [84,85], to confirm the positive selection signals of those positively selected genes identified by PAML. Unlike PAML, in which only positive selection in foreground branches is allowed, BUSTED allows the occurrence of positive selection in both foreground and background branches (unconstrained branch-site model). The unconstrained model fit is compared with the branch-site model that disallows $\omega > 1$ among foreground branches (null model), and then LRT was used to determine statistical significance.

(g) Spectral tuning analyses

We analysed possible spectral tuning (i.e. the shifts of wavelengths of maximal absorption) of four cone-expressed opsins, LWS, RH2, SWS2, and SWS1, along the ancestral bird branch and the ancestral archosaur branch. For this, we reconstructed ancestral amino acid sequences using amino acid-based marginal reconstruction implemented by the empirical Bayes approach in PAML 4.8a to determine the amino acid replacements along the branches, considering that the spectral tuning of these cone opsins is controlled by specific critical amino acid replacements [40-43]. To test for result consistency, two different amino acid substitution models, JTT and Poisson, were used. The two models have different assumptions about amino acid substitution rates. The model ITT assumes different substitution rates of different amino acids, while the Poisson model assumes the same substitution rate of all amino acids. With the reconstruction of ancestral sequences for internal nodes, the critical amino acid replacements that are associated with the spectral tuning of cone opsins along branches can be inferred. To determine the numbering of the critical amino acid replacements in our sequences, we employed webPRANK to align our sequences against the bovine rhodopsin sequence (NP_001014890). The effects of the critical amino acid replacements on the shifts of the wavelengths of maximal absorption $(\Delta\lambda)$ of the four cone opsins were analysed following published studies [40-43].

Data accessibility. All data needed to evaluate the conclusions in the paper are present in the paper and/or the electronic supplementary material. Additional data are available from the authors upon request.

Authors' contributions. Y.W. designed research, performed analyses, and wrote the paper. H.W. revised the manuscript. All authors gave final approval for publication.

Competing interests. We have no competing interests.

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References

- Bennett AF, Ruben JA. 1979 Endothermy and activity in vertebrates. *Science* 206, 649–654. (doi:10.1126/science.493968)
- Crompton A, Taylor CR, Jagger JA. 1978 Evolution of homeothermy in mammals. *Nature* 272, 333–336. (doi:10.1038/272333a0)
- Dooling RJ, Fay RR, Popper AN. 2000 *Comparative hearing: birds and reptiles*, pp. 308–359. New York, NY: Springer.
- Farmer C. 2000 Parental care: the key to understanding endothermy and other convergent features in birds and mammals. *Am. Nat.* 155, 326–334. (doi:10.1086/303323)
- Koteja P. 2000 Energy assimilation, parental care and the evolution of endothermy. *Proc. R. Soc. Lond. B* 267, 479–484. (doi:10.1098/rspb.2000. 1025)
- Hohtola E. 2004 Shivering thermogenesis in birds and mammals. In *Life in the cold: evolution, mechanisms, adaptation, and application twelfth international hibernation symposium* (eds BM Barnes, HV Carey), pp. 241–252. Fairbanks, Alaska: Institute of Arctic Biology, University of Alaska Fairbanks.
- Hillenius WJ, Ruben JA. 2004 The evolution of endothermy in terrestrial vertebrates: Who? When? Why? *Physiol. Biochem. Zool.* 77, 1019–1042. (doi:10.1086/425185)
- Kemp TS. 2005 The origin and evolution of mammals. New York, NY: Oxford University Press.
- Grigg GC, Beard LA, Augee ML. 2004 The evolution of endothermy and its diversity in mammals and birds. *Physiol. Biochem. Zool.* **77**, 982–997. (doi:10. 1086/425188)
- Hayes JP, Garland T. 1995 The evolution of endothermy: testing the aerobic capacity model. *Evolution* 49, 836–847. (doi:10.1111/j.1558-5646. 1995.tb02320.x)
- Koteja P. 2004 The evolution of concepts on the evolution of endothermy in birds and mammals. *Physiol. Biochem. Zool.* **77**, 1043–1050. (doi:10. 1086/423741)
- Gerkema MP, Davies WI, Foster RG, Menaker M, Hut RA. 2013 The nocturnal bottleneck and the evolution of activity patterns in mammals. *Proc. R. Soc. B* 280, 20130508. (doi:10.1098/rspb. 2013.0508)
- Wu Y, Wang H, Hadly EA. 2017 Invasion of ancestral mammals into dim-light environments inferred from adaptive evolution of the phototransduction genes. *Sci. Rep.* 7, 46542. (doi:10.1038/srep46542)
- Clarke A, Portner H. 2010 Temperature, metabolic power and the evolution of endothermy. *Biol. Rev.* 85, 703–727. (doi:10.1111/j.1469-185X.2010.00122.x)
- Anderson SR, Wiens JJ. 2017 Out of the dark: 350 million years of conservatism and evolution in diel activity patterns in vertebrates. *Evolution* **71**, 1944–1959. (doi:10.1111/evo.13284)
- 16. Maitin G. 1990 *Birds by night*. London, UK: T & A D Poyser.

- Schmitz L, Motani R. 2011 Nocturnality in dinosaurs inferred from scleral ring and orbit morphology. *Science* 332, 705–708. (doi:10.1126/science.1200043)
- Hall MI, Kamilar JM, Kirk EC. 2012 Eye shape and the nocturnal bottleneck of mammals. *Proc. R. Soc. B* 279, 4962–4968. (doi:10.1098/rspb.2012.2258)
- Hall MI, Kirk EC, Kamilar JM, Carrano MT. 2011 Comment on 'Nocturnality in dinosaurs inferred from scleral ring and orbit morphology'. *Science* 334, 1641. (doi:10.1126/science.1208442)
- Wu Y, Hadly EA, Teng W, Hao Y, Liang W, Liu Y, Wang H. 2016 Retinal transcriptome sequencing sheds light on the adaptation to nocturnal and diurnal lifestyles in raptors. *Sci. Rep.* 6, 33578. (doi:10.1038/srep33578)
- Wu Y, Wang H, Wang H, Feng J. 2018 Arms race of temporal partitioning between carnivorous and herbivorous mammals. *Sci. Rep.* 8, 1713. (doi:10. 1038/s41598-018-20098-6)
- Wu Y, Wang H, Wang H, Hadly EA. 2017 Rethinking the origin of primates by reconstructing their diel activity patterns using genetics and morphology. *Sci. Rep.* 7, 11837. (doi:10.1038/s41598-017-12090-3)
- Lamb TD. 2013 Evolution of phototransduction, vertebrate photoreceptors and retina. *Prog. Retin. Eye Res.* 36, 52–119. (doi:10.1016/j.preteyeres. 2013.06.001)
- Larhammar D, Nordström K, Larsson TA. 2009 Evolution of vertebrate rod and cone phototransduction genes. *Phil. Trans. R Soc. B* 364, 2867–2880. (doi:10.1098/rstb.2009.0077)
- Jarvis ED *et al.* 2014 Whole-genome analyses resolve early branches in the tree of life of modern birds. *Science* **346**, 1320–1331. (doi:10.1126/ science.1253451)
- Crawford NG, Faircloth BC, McCormack JE, Brumfield RT, Winker K, Glenn TC. 2012 More than 1000 ultraconserved elements provide evidence that turtles are the sister group of archosaurs. *Biol. Lett.* 8, 783–786. (doi:10.1098/rsbl.2012.0331)
- Crawford NG, Parham JF, Sellas AB, Faircloth BC, Glenn TC, Papenfuss TJ, Henderson JB, Hansen MH, Simison WB. 2015 A phylogenomic analysis of turtles. *Mol. Phylogenet. Evol.* 83, 250–257. (doi:10.1016/j.ympev.2014.10.021)
- Fuchs J, Johnson JA, Mindell DP. 2015 Rapid diversification of falcons (Aves: Falconidae) due to expansion of open habitats in the Late Miocene. *Mol. Phylogenet. Evol.* 82, 166–182. (doi:10.1016/j. ympev.2014.08.010)
- Green RE *et al.* 2014 Three crocodilian genomes reveal ancestral patterns of evolution among archosaurs. *Science* **346**, 1254449. (doi:10.1126/ science.1254449)
- Jønsson KA, Fjeldså J. 2006 A phylogenetic supertree of oscine passerine birds (Aves: Passeri). *Zoolog. Scripta* 35, 149–186. (doi:10.1111/j.1463-6409.2006.00221.x)
- Kriegs JO, Matzke A, Churakov G, Kuritzin A, Mayr G, Brosius J, Schmitz J. 2007 Waves of genomic

hitchhikers shed light on the evolution of gamebirds (Aves: Galliformes). *BMC Evol. Biol.* **7**, 190. (doi:10.1186/1471-2148-7-190)

- Lerner HR, Mindell DP. 2005 Phylogeny of eagles, Old World vultures, and other Accipitridae based on nuclear and mitochondrial DNA. *Mol. Phylogenet. Evol.* 37, 327–346. (doi:10.1016/j.ympev.2005.04.010)
- Pyron RA, Burbrink FT, Wiens JJ. 2013 A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. *BMC Evol. Biol.* 13, 93. (doi:10.1186/1471-2148-13-93)
- Wink M, El-Sayed A-A, Sauer-Gürth H, Gonzalez J. 2009 Molecular phylogeny of owls (Strigiformes) inferred from DNA sequences of the mitochondrial cytochrome b and the nuclear RAG-1 gene. *Ardea* 97, 581–591. (doi:10.5253/078.097.0425)
- Wright TF *et al.* 2008 A multilocus molecular phylogeny of the parrots (Psittaciformes): support for a Gondwanan origin during the Cretaceous. *Mol. Biol. Evol.* 25, 2141–2156. (doi:10.1093/molbev/ msn160)
- Le Duc D *et al.* 2015 Kiwi genome provides insights into evolution of a nocturnal lifestyle. *Genome Biol.* 16, 147. (doi:10.1186/s13059-015-0711-4)
- Hackett SJ *et al.* 2008 A phylogenomic study of birds reveals their evolutionary history. *Science* **320**, 1763–1768. (doi:10.1126/science.1157704)
- Prum RO, Berv JS, Dornburg A, Field DJ, Townsend JP, Lemmon EM, Lemmon AR. 2015 A comprehensive phylogeny of birds (Aves) using targeted next-generation DNA sequencing. *Nature* 526, 569–573. (doi:10.1038/nature15697)
- Umino Y, Herrmann R, Chen C-K, Barlow RB, Arshavsky VY, Solessio E. 2012 The relationship between slow photoresponse recovery rate and temporal resolution of vision. *J. Neurosci.* 32, 14 364–14 373. (doi:10. 1523/JNEUROSCI.1296-12.2012)
- Takenaka N, Yokoyama S. 2007 Mechanisms of spectral tuning in the RH2 pigments of Tokay gecko and American chameleon. *Gene* **399**, 26–32. (doi:10.1016/j.gene.2007.04.036)
- Yokoyama S. 2002 Molecular evolution of color vision in vertebrates. *Gene* **300**, 69–78. (doi:10. 1016/S0378-1119(02)00845-4)
- Yokoyama S. 2008 Evolution of dim-light and color vision pigments. *Annu. Rev. Genom. Hum. Genet.* 9, 259–282. (doi:10.1146/annurev.genom.9.081307. 164228)
- Yokoyama S, Tada T. 2003 The spectral tuning in the short wavelength-sensitive type 2 pigments. *Gene* **306**, 91–98. (doi:10.1016/S0378-1119(03)00424-4)
- Hall MI. 2008 The anatomical relationships between the avian eye, orbit and sclerotic ring: implications for inferring activity patterns in extinct birds. *J. Anat.* 212, 781–794. (doi:10.1111/j.1469-7580.2008.00897.x)
- Angielczyk K, Schmitz L. 2014 Nocturnality in synapsids predates the origin of mammals by over 100 million years. *Proc. R. Soc. B* 281, 20141642. (doi:10.1098/rspb.2014.1642)

royalsocietypublishing.org/journal/rspb Proc. R. Soc. B 286: 20182185

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- Legendre LJ, Guénard G, Botha-Brink J, Cubo J.
 2016 Palaeohistological evidence for ancestral high metabolic rate in Archosaurs. *Syst. Biol.* 65, 989–996. (doi:10.1093/sysbio/syw033)
- 47. Bakker RT. 1986 *The dinosaur heresies*. Essex: Longman Scientific & Technical.
- Seymour RS, Bennett-Stamper CL, Johnston SD, Carrier DR, Grigg GC. 2004 Evidence for endothermic ancestors of crocodiles at the stem of archosaur evolution. *Physiol. Biochem. Zool.* **77**, 1051–1067. (doi:10.1086/422766)
- Olivier C, Houssaye A, Jalil N-E, Cubo J. 2017 First palaeohistological inference of resting metabolic rate in an extinct synapsid, *Moghreberia nmachouensis* (Therapsida: Anomodontia). *Biol. J. Linnean. Soc.* **121**, 409–419. (doi:10.1093/biolinnean/blw044)
- Rey K *et al.* 2017 Oxygen isotopes suggest elevated thermometabolism within multiple Permo-Triassic therapsid clades. *Elife* 6, e28589. (doi:10.7554/eLife. 28589)
- Ruben J. 1995 The evolution of endothermy in mammals and birds: from physiology to fossils. *Annu. Rev. Physiol.* 57, 69–95. (doi:10.1146/ annurev.ph.57.030195.000441)
- Seebacher F. 2003 Dinosaur body temperatures: the occurrence of endothermy and ectothermy. *Paleobiology* 29, 105–122. (doi:10.1666/0094-8373(2003)029<0105:DBTT00>2.0.C0;2)
- Xu X, Zhou Z, Dudley R, Mackem S, Chuong C-M, Erickson GM, Varricchio DJ. 2014 An integrative approach to understanding bird origins. *Science* 346, 1253293. (doi:10.1126/science.1253293)
- Chatterjee S. 2015 *The rise of birds: 225 million years of evolution*, pp. 72–105. Baltimore: Johns Hopkins University Press.
- La VT. 2012 Diurnal and nocturnal birds vocalize at night: a review. *The Condor* **114**, 245–257. (doi:10. 1525/cond.2012.100193)
- Hall M, Heesy C. 2011 Eye size, flight speed and Leuckart's Law in birds. *J. Zool.* 283, 291–297. (doi:10.1111/j.1469-7998.2010.00771.x)
- 57. Roots C. 2006 *Nocturnal animals*. London: Greenwood Press.
- Walls GL. 1942 The vertebrate eye and its adaptive radiation. New York, NY: The Cranbrook Institute of Science.

- Crompton AW. 1980 Biology of the earliest mammals. In *Comparative physiology: primitive mammals* (eds K Schmidt-Nielsen, L Bolis, CR Taylor), pp. 1–12. Cambridge: Cambridge University Press.
- 60. Cowen R. 2005 *History of life*. Malden, MA: Blackwell Publishing.
- Kermack DM, Kermack K. 1984 The evolution of mammalian characters. Washington DC: Kapitan Szabo Publishers.
- Eagle RA *et al.* 2015 Isotopic ordering in eggshells reflects body temperatures and suggests differing thermophysiology in two Cretaceous dinosaurs. *Nat. Commun.* 6, 8296. (doi:10.1038/ncomms9296)
- Grady JM, Enquist BJ, Dettweiler-Robinson E, Wright NA, Smith FA. 2014 Evidence for mesothermy in dinosaurs. *Science* 344, 1268–1272. (doi:10.1126/ science.1253143)
- Healy S, Guilford T. 1990 Olfactory-bulb size and nocturnality in birds. *Evolution* 44, 339–346. (doi:10.1111/j.1558-5646.1990.tb05203.x)
- Block B. 2011 Endothermy in tunas, billfishes, and sharks. In *Encyclopedia of fish physiology: from genome to environment*, vol. 3 (ed. A.P Farrell.), pp. 1914–1920. San Diego: Academic Press.
- Block BA, Finnerty JR, Stewart AFR, Kidd J. 1993 Evolution of endothermy in fish: mapping physiological traits on a molecular phylogeny. *Science* 260, 210–214. (doi:10.1126/science.8469974)
- Dickson KA, Graham JB. 2004 Evolution and consequences of endothermy in fishes. *Physiol. Biochem. Zool.* 77, 998–1018. (doi:10.1086/ 423743)
- McNab BK. 1978 The evolution of endothermy in the phylogeny of mammals. *Am. Nat.* 112, 1–21. (doi:10.1086/283249)
- Ramsay JA. 2017 Endothermy. In *Principles of thermal* ecology: temperature, energy and life (ed. A Clarke), pp. 214–244. Oxford, UK: Oxford University Press.
- Bostrom BL, Jones TT, Hastings M, Jones DR. 2010 Behaviour and physiology: the thermal strategy of leatherback turtles. *PLoS ONE* 5, e13925. (doi:10. 1371/journal.pone.0013925)
- Hutchison VH, Dowling HG, Vinegar A. 1966 Thermoregulation in a brooding female Indian python, *Python molurus bivittatus. Science* **151**, 694–695. (doi:10.1126/science.151.3711.694)

- Tattersall GJ, Leite CAC, Sanders CE, Cadena V, Andrade DV, Abe AS, Milsom WK. 2016 Seasonal reproductive endothermy in tegu lizards. *Sci. Adv.* 2, e1500951. (doi:10.1126/sciadv.1500951)
- Heinrich B. 1974 Thermoregulation in endothermic insects. *Science* 185, 747–756. (doi:10.1126/ science.185.4153.747)
- May ML. 1979 Insect thermoregulation. *Annu. Rev. Entomol.* 24, 313 – 349. (doi:10.1146/annurev.en.24. 010179.001525)
- Patiño S, Grace J, Bänziger H. 2000 Endothermy by flowers of *Rhizanthes lowii* (Rafflesiaceae). *Oecologia* 124, 149–155. (doi:10.1007/s004420050001)
- Angilletta J, Michael J, Sears MW. 2003 Is parental care the key to understanding endothermy in birds and mammals? *Am. Nat.* 162, 821–825. (doi:10. 1086/380921)
- Lovegrove BG. 2017 A phenology of the evolution of endothermy in birds and mammals. *Biol. Rev.* 92, 1213–1240. (doi:10.1111/brv.12280)
- Lee MS, Cau A, Naish D, Dyke GJ. 2014 Sustained miniaturization and anatomical innovation in the dinosaurian ancestors of birds. *Science* 345, 562–566. (doi:10.1126/science.1252243)
- Reynolds RG, Niemiller ML, Revell LJ. 2014 Toward a Tree-of-Life for the boas and pythons: multilocus species-level phylogeny with unprecedented taxon sampling. *Mol. Phylogenet. Evol.* **71**, 201–213. (doi:10.1016/j.ympev.2013.11.011)
- 80. Uetz P. 2006 The Reptile Database. http://www.reptile-database.org.
- Gill F, Donsker D. 2018 IOC World Bird List (v8.2). (doi:10.14344/IOC.ML.8.2)
- Grabherr MG *et al.* 2011 Full-length transcriptome assembly from RNA-seq data without a reference genome. *Nat. Biotechnol.* 29, 644–652. (doi:10. 1038/nbt.1883)
- Yang Z. 2007 PAML 4: phylogenetic analysis by maximum likelihood. *Mol. Biol. Evol.* 24, 1586–1591. (doi:10.1093/molbev/msm088)
- Murrell B *et al.* 2015 Gene-wide identification of episodic selection. *Mol. Biol. Evol.* **32**, 1365–1371. (doi:10.1093/molbev/msv035)
- Pond SL, Frost SD, Muse SV. 2005 HyPhy: hypothesis testing using phylogenies. *Bioinformatics* 21, 676–679. (doi:10.1093/bioinformatics/bti079)