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A rare missense variant in *CASP7* is associated with familial late-onset Alzheimer disease

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Abstract

INTRODUCTION: The genetic architecture of Alzheimer disease (AD) is only partially understood.

METHODS: We conducted an association study for AD using whole sequence data from 507 genetically enriched AD cases (i.e., cases having close relatives affected by AD) and 4,917

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cognitively healthy controls of European Ancestry (EA) and 172 enriched cases and 179 controls of Caribbean Hispanic (CH) ancestry. Confirmation of top findings from Stage 1 was sought in two family-based GWAS datasets and in a whole genome sequencing dataset comprising members from 42 EA and 115 CH families.

RESULTS: We identified associations in EAs with variants in 12 novel loci. The most robust finding is a rare *CASP7* missense variant (rs116437863; $p=2.44 \times 10^{-10}$) which improved when combined with results from the Stage 2 datasets ($p=1.92 \times 10^{-10}$).

DISCUSSION: Our study demonstrated that an enriched cases design can strengthen genetic signals, thus allowing detection of associations that would otherwise be missed in a traditional case-control study.

Keywords

enriched case-control; whole exome sequencing; association study; genome-wide association studies; gene-based analyses

1. Introduction

Late-onset Alzheimer disease (AD) is a progressive neurodegenerative disorder in persons ages 65 years and older characterized by memory loss and dementia. AD risk increases exponentially with age with a prevalence of 30–40% among 85–89 year-olds [1]. As average life expectancy has increased, the number of AD cases will increase to 11–16 million by 2050 with nearly 1 million new cases per year unless measures are identified to delay or prevent the disease [2, 3].

Risk of AD is modulated by variants in multiple genes, most notably the *APOE* $\epsilon 2$ and $\epsilon 4$ alleles, in combination with lifestyle and environmental factors. AD has a substantial genetic component with an estimated heritability of 58–79% [4]. Genome-wide association studies (GWAS) have identified >20 common susceptibility variant loci showing robust evidence for association with AD [5–7]. Recently, studies that performed whole exome sequencing (WES), targeting gene sequencing and rapid throughput genotyping using SNP microarray chips with high exome content have reported associations with rare risk variants in multiple novel loci including *TREM2* (R47H) [8–10], *PLD3* [11], *AKAP9* [12], *UNC5C* [13], *PLCG2* [14], *ABI3* [14] as well as with rare risk and protective variants in several previously known AD genes (*APP* [15, 16], *APOE* (*p.V236E*) [17], *SORL1* [18] and *ABCA7* [19, 20]).

The Alzheimer’s Disease Sequencing Project (ADSP) is an NIH-funded initiative to identify novel genes and rare risk and protective variants using WES and whole genome sequencing (WGS) approaches. In this study, we analyzed a subset of the ADSP WES cohort including “enriched” AD cases (i.e., cases who have close relatives also affected by AD and thus more likely to have a high burden of AD risk alleles compared to cases not ascertained on the basis of a positive family history) and all controls to identify novel associations with single nucleotide variants (SNVs) and short insertions and deletions (indels).

2. Methods

2.1 Participants, Sequencing and Data Processing

The Alzheimer's Disease Sequencing Project (ADSP) performed WES of DNA specimens from 5,778 AD cases and 5,136 controls at three NHGRI Genome Centers (Broad Institute, Baylor College of Medicine, and McDonnell Genome Institute at the Washington University). Detailed description of the ADSP WES study design has been published elsewhere [21]. In brief, subjects for this study were selected from datasets assembled by the Alzheimer's Disease Genetics Consortium (ADGC), Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium, and the Rotterdam Study [5, 22]. In the ADSP WES EA cohort, ~5,000 AD cases that were not ascertained on the basis of family history of AD were selected because they have the lowest risk explained by age and APOE genotypes (young onset, APOE $\epsilon 2/\epsilon 2$, $\epsilon 2/\epsilon 3$, or $\epsilon 3/\epsilon 3$). ~5,000 cognitively normal controls were selected as controls least likely to convert to a case based on age, APOE, and autopsy data (old, APOE $\epsilon 2/\epsilon 2$, $\epsilon 2/\epsilon 3$, or $\epsilon 3/\epsilon 3$, and little or no AD neuropathology). In addition, ~700 unrelated cases were selected from additional multiplex families with >2 close relatives affected by AD in each family, but only one case was selected from each family (i.e., "enriched-cases"). To enhance discovery of novel AD-related variants, enriched cases which could be explained by cosegregation of *APOE* $\epsilon 4$ were excluded.

After performing a series of filtering steps to identify duplicate samples and subjects with low genotype call rates, there remained a sample containing 10,441 individuals of European ancestry (EA) and 395 Caribbean Hispanics (CH). Subjects for this study included 679 unrelated AD cases (507 EA and 172 CH) from families containing at least three members affected by AD and 5,094 unrelated controls (4,917 EA and 177 CH). Characteristics of the 5,773 subjects included in this study are shown in Supplementary Table 1. Compared to the overall ADSP case-control study design, EA enriched cases are older (age at onset = 83.6 years) and similar in age to the controls (age at last examination = 86.5 years). The mean age at onset of CH enriched cases (75.4 years) is similar to the mean age at last examination (73.5 years) of the CH controls.

2.2 Whole Exome Sequencing and Quality Control

Details of library preparation, sequencing protocols, and variant calling pipelines are described in Supplementary Methods. After sequencing, 100 bp paired-end reads were mapped to human reference genome (GRCh37/hg19) using the Burrows-Wheeler Aligner (BWA) [23]. The ADSP Quality Control (QC) Working Group applied QC protocols to autosomal bi-allelic single nucleotide variants (SNVs) and short insertions and deletions (indels) to generate a high-quality variant call set. After QC, there remained 1,454,483 SNVs and 69,931 indels for association analyses.

2.3 Single-variant Association Analyses

In Stage 1, association of AD with each variant having a minor allele count (MAC) ≥ 10 (100,338 variants for EA and 64,691 for CH; Figure 1 and Supplementary Table 2) was tested in each population using score tests in seqMeta (<https://github.com/DavisBrian/seqMeta>) with three additive logistic regression models. A MAC cutoff of 10 is the

minimum number of alleles to achieve statistical significance in this sample. A minimal adjustment model (Model 0) included covariates for sequencing center and principal components (PCs) of ancestry (the first 10 PCs for EAs and the first 3 PCs for CHs) in order to identify variants whose effects on AD risk are confounded by age and sex. This model was previously shown to increase detection of associations in this sample in which the mean age is substantially different between cases and controls [24]. A second model (Model 1) also included terms for age and sex, and a third model (Model 2) included all covariates from model 2 plus terms for the number of APOE ϵ 4 and ϵ 2 alleles. Results from analyses of 46,425 variants that were successfully called and passed criteria for single-variant analysis in the EA and CH data sets were combined using an inverse variance-weighted meta-analysis approach implemented in seqMeta after applying genomic control. A Bonferroni correction was applied to define study-wide significance (SWS) in each group (EA: $p < 4.98 \times 10^{-7}$, CH: $p < 7.73 \times 10^{-7}$, and meta $p < 1.08 \times 10^{-6}$; Supplementary Table 2).

It is well-known that the standard maximum likelihood estimation of the logistic model can suffer from small-sample bias [25]. We applied a penalized likelihood method (i.e., the Firth logistic regression test [25–27]) to evaluate association of the odds ratio (OR) and confidence intervals for all top single variants using Heinze's "logistf" package in R (<http://cemsis.meduniwien.ac.at/en/kb/science-research/software/statistical-software/fllogistf/>).

2.4 Gene-based Association Analysis

In an attempt to improve power by removing non-functional variants, we selected variants on the basis of annotated function using the Ensembl Variant Effect Predictor (VEP) [28] and SnpEff [29] software as follows: a) HIGH IMPACT: variants classified as splice acceptor, splice donor, stop gained, frameshift, stop lost, start lost, or transcript amplification; b) HIGH or MODERATE IMPACT: included the categories above plus variants annotated as in-frame insertion, in-frame deletion, missense variant, or protein altering. Association was tested for genes with ≥ 2 variants and a cumulative MAC (cMAC)

≥ 10 after excluding variants with a minor allele frequency (MAF) ≥ 0.05 using the same three models tested in the individual variant analyses and the *SKAT-O* program in seqMeta [30]. Separate analyses were performed for high impact variants only (2,298 genes in EAs and 314 genes in CHs) and for high and moderate impact variants (16,026 genes in EAs and 11,743 genes in CHs). Analyses of the combined populations included 1,941 genes with high-impact variants and 14,960 genes with high/moderate impact variants. The ethnic-specific gene-based results were combined by meta-analysis of Z-scores weighted by the number of subjects using seqMeta, assuming the same direction of effect on a gene in both populations. Significance thresholds for each analysis were determined based on the number of genes tested (Supplementary Table 3).

2.5 Stage 2 Analyses in GWAS datasets

We attempted to confirm the top-ranked discovery stage results from single variant analysis ($p < 1.0 \times 10^{-5}$) and gene-based tests ($p < 1.0 \times 10^{-4}$) obtained from the best-model (i.e. the smallest P-value among the three models tested for each individual variant or gene) using ADGC GWAS datasets in which genotypes for ~ 39 M variants as rare as MAF=0.0004 were imputed with the Haplotype Reference Consortium (HRC) r1.1 reference panel [31] using

MiniMac3 (see Supplementary Methods for additional details of imputation procedures). In order to be consistent with enriched cases design of the discovery analyses, we evaluated the two ADGC family-based cohorts, MIRAGE (449 AD cases and 704 controls) and NIA-LOAD (1,568 cases and 1,457 controls), after excluding subjects who were included in the ADSP WES dataset (Table 1). Models 1 and 2 only were evaluated in each data set using imputed allele dosages for each variant, generalized estimating equations (GEE) implemented in *geepack* R package for single variant tests, and ‘F-SKAT’ [32] for gene-based tests to account for the family structure. Model 0 was not evaluated in the Stage 2 datasets because these samples did not have unique ascertainment schemes for AD cases and controls on the basis of age and sex. Results from the Stage 2 datasets were combined using a fixed-effects inverse variance-weighted method in METAL [33] applied to single variant results and the sample-size weighted Z-score method applied to gene-based results. Successful replication was determined using a nominal significance threshold ($p < 0.05$) and, for single variants, if the effect direction was the same in the Stage 1 and Stage 2 datasets. Results from Stages 1 and 2 were combined using the same meta-analysis approach. Results for Models 0 and 1 in the Stage 1 dataset were each meta-analyzed with those obtained from Model 1 in the Stage 2 datasets.

2.6 Stage 2 Analyses in the ADSP Family-based WGS Dataset

We further examined the top-ranked discovery stage results (52 individual variants and eight genes) in the ADSP WGS family-based dataset [21, 34]. This dataset includes 197 individuals sequenced in 42 EA families and 501 individuals in 115 CH families. Association of individual variants was evaluated by inspecting their segregation within families. Gene-based tests were conducted separately for EA and CH families using F-SKAT.

3. Results

There was little evidence for genomic inflation in single variant based exome-wide results in the EA ($\lambda = 0.92$), CH ($\lambda = 1.05$), or combined populations ($\lambda = 1.07$; Supplementary Fig. 1).

3.1 Single-variant Results in EAs

In Model 0, rare variants in *TREM2* ($p = 4.56 \times 10^{-12}$), *NPC1* ($p = 5.78 \times 10^{-9}$), *CASP7* ($p = 2.44 \times 10^{-10}$) and *KCNK13* ($p = 1.55 \times 10^{-7}$) were significantly associated with AD (Table 2, Supplementary Fig. 2). After adjusting for age and sex (Model 1), the evidence for association with all four genes was reduced but the *TREM2* and *NPC1* variants were still significant ($P < 5.0 \times 10^{-7}$). Additional adjustment for *APOE* genotype (Model 2) further diminished associations with all genes but *NPC1* ($p < 9.41 \times 10^{-5}$; Supplementary Table 4). Variants in *HOXB-AS1/HOXB2* ($p = 7.63 \times 10^{-8}$), *HTR3A* ($p = 1.28 \times 10^{-7}$), *ZNF333* ($p = 1.28 \times 10^{-7}$) and *STAB1* ($p = 3.58 \times 10^{-7}$) surpassed the SWS threshold for Model 1, and the association with the *HOXB-AS1/HOXB2* variant was also significant in Model 2 ($p = 1.30 \times 10^{-7}$). For Model 2, SWS associations were identified with rare variants in six additional novel gene regions including *SCN4A* ($p = 6.30 \times 10^{-14}$), *MUC17* ($p = 1.63 \times 10^{-9}$), *AKNAD1* (two variants in complete LD, $p = 2.71 \times 10^{-8}$ for both), *KANSL3* ($p = 6.40 \times 10^{-8}$), *TMEM87A* ($p = 2.79 \times 10^{-7}$) and *OTOG* ($p = 4.16 \times 10^{-7}$). Suggestive evidence for association

($p < 5.0 \times 10^{-6}$) was obtained for variants in seven additional genes (Supplementary Table 4). In addition to the previously known *TREM2* R47H variant [8–10], 52 variants including two indels from 50 novel loci met criteria for follow up in the Stage 2 datasets. Analyses of these variants yielded nominally significant ($P < 0.05$) results, however only the *CASP7* variant showed the same effect direction, and this association was slightly more significant ($p = 1.92 \times 10^{-10}$) in the combined Stage 1+2 sample (Table 2, Supplementary Table 5). This *CASP7* variant is a previously identified missense mutation (rs116437863) that results in an amino acid substitution of glycine for arginine and is predicted to be deleterious and probably damaging by SIFT [35] (score = 0) and PolyPhen [36] (score = 0.99).

3.2 .Single-variant results in Caribbean Hispanics

No variants reached SWS ($p < 7.73 \times 10^{-7}$) in the CH group. However, notable novel association signals were observed with a SNV in *LDB3* ($p = 5.11 \times 10^{-6}$), a previously known 6 bp frameshift deletion (rs782084513) in *ORAI1* ($p = 5.34 \times 10^{-5}$), and a 3 bp deletion in *KLHL40* ($p = 7.98 \times 10^{-5}$). The strength of these associations was similar in all three models (Supplementary Table 6) indicating they are independent of age, sex and *APOE* genotype. The *LDB3* and *ORAI1* variants were also observed in EAs but were not associated with AD risk ($p > 0.25$). Conversely, none of the top-ranked rare variants in EAs had a MAC 10 in the CH group, except for the variants in *ZNF333* (MAC=26) and *SCN4A* (MAC=19) which were not associated with AD risk in the CH dataset ($p > 0.20$) (Supplementary Table 7). However, a rare variant *SLAIN1* showed mild evidence of association in both EA ($p = 0.00015$) and CH ($p = 0.0023$) groups, and the Model 0 result from the two groups combined approached SWS (meta $p = 4.68 \times 10^{-6}$; Supplementary Table 8).

3.3 .Gene-based association results

In analyses focused on high-impact variants, associations with six novel loci were SWS ($p < 2.18 \times 10^{-5}$) including *JMJD4*, *C1orf173*, *ANXA5*, *AARD*, *ASCC1*, and *ASB13* (Table 3). Results for two additional novel genes (*DTYMK* and *IGHJ6*) were SWS ($p < 3.12 \times 10^{-6}$) in analyses that included high and moderate-impact variants. Among these findings, only *AARD* showed evidence for association in the Stage 2 datasets ($p = 6.16 \times 10^{-3}$). However, associations were strengthened for *C1orf173* ($p = 1.50 \times 10^{-5}$), *ANXA5* ($p = 2.27 \times 10^{-5}$) and *AARD* ($p = 1.01 \times 10^{-6}$) in the combined Stage 1+2 dataset. Although none of the gene-based tests were SWS in the CH group, analyses of high and moderate impact variants revealed a significant association with *KLHL40* ($p = 9.98 \times 10^{-5}$ in model 0 and $p = 1.01 \times 10^{-4}$ in models 1 and 2).

3.4 .Stage 2 results in the ADSP Family-based WGS data

SWS and suggestive associations for 52 individual variants (Supplementary Table 4) and eight genes (Table 3) were further examined in the ADSP WGS family-based dataset. Low-frequency missense variants in *AIMIL* (rs80177817, MAF=0.02, Discovery $p = 6.14 \times 10^{-7}$) were observed and segregated nearly perfectly with disease in two EA families and one CH family. A previously identified rare frameshift mutation in *DHX37* (rs779974893, MAF=0.001, Stage 1 $p = 7.68 \times 10^{-6}$), and the known AD-associated rare missense mutation in *TREM2* (R47H, MAF=0.003) each occurred and segregated with disease in one EA family (Supplementary Table 9). A rare missense variant in *ENGASE* (rs11871357,

MAF=0.001, Stage 1 $p=7.47\times 10^{-6}$) and a rare synonymous variant in *ZNF333* (rs79724046, MAF=0.003, Stage 1 $p=1.28\times 10^{-7}$) each occurred and were observed predominantly among affected members with disease in three CH families. A rare variant in *KANSL3* (rs34406082, Stage 1 $p=6.40\times 10^{-8}$) perfectly co-segregated with disease in two CH families. Low-frequency variants in *SCN4A* (rs73992419, Stage 1 $p=6.30\times 10^{-14}$) and *PTGIS* (rs61322884, Stage 1 $p=4.55\times 10^{-6}$) each showed a high degree of co-segregation with disease in two CH families. None of the gene-based tests in EA or CH families were nominally significant ($P>0.20$; Supplementary Table 10), noting that none of the rare variants that primarily accounted for the gene associations in the Stage 1 sample were observed in the WGS families (Supplementary Table 11).

3.5 Findings for previously reported AD-associated rare variants

Of 15 rare variants previously reported to be associated with AD, two non-exonic *ABCA7* variants not included in the WES capture. Three of the tested seven variants (i.e., those with MAC = 10) were significantly associated with AD after correcting for the number of tests ($P<0.007$) in a model adjusting for age, sex and APOE dosage: *PLD3*-V232M, *ABI3*-S209F, and *SORL1*-A528T (Supplementary Table 12).

4. Discussion

We identified novel associations for AD with a single rare variant in *CASP7* and gene-based tests of aggregated rare variants in *C1orf173*, *ANXA5*, and *AARD* in 5,094 controls and a subset of 679 unrelated familial AD cases from the ADSP. These findings were study-wide significant and improved when combined with results obtained from HRC-imputed data from 2,161 AD cases and 2,017 controls in two family-based ADGC GWAS datasets. Study-wide significant findings at the variant or gene level were observed for several other loci (*NPC1*, *KCNK13*, *HOXB2*, *HTR3A*, *ZNF333*, *STAB1*, *SCN4A*, *MUC17*, *AKNAD1*, *KANSL3*, *TMEM87A*, *OTOG*, *DTYMK*, and *IGHJ6*), but these findings were not bolstered by the Stage 2 datasets. A previous study by the ADSP of the entire WES dataset including 5,740 AD cases reported associations with one rare variant in *AC099552.4* and nine high-impact aggregated variants in *ZNF655* [35]. The greatly improved detection of associations with rare variants in this smaller sample of AD cases is likely due to the enriched case design. For example, the result for the established rare *TREM2* R47H variant ($p=4.56\times 10^{-12}$) is virtually identical to the result for this variant in the prior study [35] which is consistent with the observation of the R47H variant only among enriched AD cases in the entire dataset. None of the other top association findings in this study were remarkable in the analysis of the enlarged ADSP sample including cases that were not ascertained on the basis of family history of AD; the strongest signals were observed for *KCNK13* ($P=0.0063$) and *HOXB2* ($P=0.0019$) (Supplementary Table 13).

CASP7 encodes a member of the cysteine-aspartic acid protease (caspase) family. Sequential activation of caspases plays a central role in the execution-phase of cell apoptosis. Caspase-7 is a protease involved in apoptosis and inflammation [37]. Activation of caspase apoptotic pathways involves apoptosome assembly [38] and multiple recent studies link this process to aging and AD neuropathology, including caspase cleavage of amyloid precursor protein

[39–41] and tau [40, 42] (Figure 2). Roles for other caspases in AD pathogenesis have been described, including caspase-6 in cognitive impairment [43] and tau cleavage [44], caspase-8 in amyloid processing, synaptic plasticity, learning, memory and control of microglia pro-inflammatory activation and associated neurotoxicity [45, 46], and caspase-9 activation in tau cleavage [47]. Su *et al.* reported that activated caspase-3 expression correlates with Alzheimer pathology [48]. Caspase 3 is processed by caspases 8, 9, and 10, and is the predominant caspase involved in the cleavage of amyloid precursor protein, which is associated with neuronal death and plaque formation in AD brain [49]. A recent targeted sequencing study of genes involved in amyloid metabolism found association of AD with two *CASP8* rare variants [46]. Our finding of a rare missense variant in *CASP7* provides additional evidence for the role of caspase apoptotic pathways in AD. Further functional studies of *CASP7*, and the rs116437863 missense variant in particular, are needed to define its role in AD pathogenesis and evaluate the potential of caspase 7 inhibition as an AD treatment strategy.

The association with a rare 3 bp in-frame deletion (rs550307753) in *TMEM87A* reached the study-wide significance in EAs ($p=2.79\times 10^{-7}$), but this finding could not be replicated because this variant was not genotyped or imputed well in the ADGC GWAS datasets.

Gene-based tests considering only highly deleterious SNVs and indels yielded highly significant associations with three novel genes (*Corf173*, *ANXA5*, and *AARD*) which were strengthened by meta analysis with results from the Stage 2 datasets. *ANXA5* is phospholipase A2 kinase C inhibitory protein that has been implicated in membrane-related events along exocytotic and endocytotic pathways, and in AD [50]. The function of *Corf173* (alias *ERICH3* - glutamate rich 3) is largely unknown. *AARD* has no obvious connection to AD or brain. *IGHJ6* was another SWS gene-based finding that did not replicate because none of the high or moderate impact rare variants each occurring only once or twice in the Stage 1 sample were observed in the Stage 2 datasets. This gene encodes one of the immunoglobulin heavy gamma variable chains and is a very good candidate given its functional similarity to *IGHG3*, one of the top associations in the entire ADSP WES sample (unpublished result) and evidence that antibodies to IgG cross-react with fibril and oligomer amyloid- β aggregates [51].

In comparison with the EA cohort, the CH cohort is very small with only 172 enriched cases and 177 controls. Nonetheless, we observed suggestive CH-specific association signals with three infrequent ($2\% < \text{MAF} < 5\%$) previously known variants including a SNV (rs76615432) in *LDB3* ($p=5.11\times 10^{-6}$), a small deletion causing a frameshift (rs782084513) in *ORAI1* ($p=5.34\times 10^{-5}$), and a small deletion in *KLHL40* ($p=7.98\times 10^{-5}$). Notably, *KLHL40* is the only gene which yielded top-ranked results from individual variant and gene-based tests, due largely to the fact that among nine distinct *KLHL40* variants that were observed one SWS variant (rs34020089) accounted for 21 of the 36 aggregated rare variants (i.e., $\text{MAC}=36$) in the gene-based test (Supplementary Table 6, Supplementary Table 14).

Several strengths and limitations of our study warrant discussion. One of the major strengths lies in the careful clinical and genetic characterization of all individuals enrolled in the ADSP. Another strength of the study is the enriched cases design which strengthened the

genetic signals thus allowing detection of associations that would otherwise be missed in a larger traditional case-control study.

A major limitation of the study is that the sample sizes are relatively small especially for enriched AD cases and the CH group overall. In the Stage 1 EA group, the enriched cases represent approximately 10% of the total sample which had sufficient power to detect association with moderate effect variants [24]. It has been shown that one can reduce the sample size of cases approximately four-fold using cases who have at least two affected relatives to have the same power as a sample of cases that are not ascertained on the basis of family history [52]. Although our case sample was not large enough to benefit fully from the enriched cases design, it was sufficient to detect association with highly penetrant variants whose effects would be diluted in a sample not ascertained on the basis of family history [52]. This idea is exemplified by the *TREM2* R47H variant for which we observed similar p values but a higher odds ratio in the current study ($p=4.56\times 10^{-12}$, OR=11.82, Supplementary Table 13) compared to the finding for this variant in the total group of ADSP cases ($p=4.8\times 10^{-12}$, OR=3.61) [35].

Another limitation is that coding variants were identified directly in the Stage 1 dataset, but imputed in the Stage 2 GWAS datasets with varying degrees of confidence for those with $MAF<0.5\%–1.0\%$. In order to be consistent with the study design of the discovery stage analysis, we included only family-based cohorts from the ADGC HRC-imputed GWAS dataset, MIRAGE and NIA-LOAD, in Stage 2 and thus had little power to confirm our findings. The relatively small size and unreliable imputation of very rare variants in the Stage 2 sample particularly limited our ability to replicate findings from gene-based tests. Also, the ADSP WES study design, for which AD cases were selected to have relatively early onset and a lower frequency of the *APOE* $\epsilon 4$ allele and controls were selected to be as old as possible with preference given to those having at least one *APOE* $\epsilon 4$ allele to enrich this group for protective variants, introduced confounding between age and AD status which reduced power for detecting associations. To overcome this limitation, we included a model without age adjustment. These limitations underscore the need to replicate our findings in other datasets containing enriched cases. Finally, our primary analysis relied on the score test which is prone to increased type-I errors for rare variants in unbalanced samples [53]. For this reason, we re-evaluated our top results using the Firth test. This analysis showed that the finding for the rare *CASP7* variant is attenuated ($p=2.21\times 10^{-5}$). Recognizing that the Firth test over-corrects for bias in very sparse data [24], the true p-value may be between those obtained using the score and Firth tests.

In summary, we identified multiple novel associations for AD with individual and aggregated rare variants using an enriched case-control study design. A better understanding of the molecular mechanisms underlying these associations will require functional experiments and *in silico* studies of the connections of genetic variants to gene expression and processing of AD-related proteins.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Appendix

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Abbreviations

AD	Alzheimer disease
ADGC	Alzheimer's Disease Genetics Consortium
ADSP	Alzheimer's Disease Sequencing Project
CH	Caribbean Hispanic
cMAC	cumulative minor allele count
EA	European ancestry

GWAS	genome-wide association study
HRC	Haplotype reference Consortium
MAC	minor allele count
MAF	minor allele frequency
OR	odds ratio
PC	principle component (of ancestry)
QC	quality control
SNV	single nucleotide variant
SWS	study-wide significant
VEP	Ensembl Variant Effect Predictor
WES	whole exome sequencing
WGS	whole genome sequencing

References

- [1]. Hebert LE, Weuve J, Scherr PA, Evans DA. Alzheimer disease in the United States (2010–2050) estimated using the 2010 census. *Neurology* 2013;80:1778–83. [PubMed: 23390181]
- [2]. Alzheimer's A 2015 Alzheimer's disease facts and figures. *Alzheimers Dement* 2015;11:332–84. [PubMed: 25984581]
- [3]. Brookmeyer R, Johnson E, Ziegler-Graham K, Arrighi HM. Forecasting the global burden of Alzheimer's disease. *Alzheimers Dement* 2007;3:186–91. [PubMed: 19595937]
- [4]. Gatz M, Reynolds CA, Fratiglioni L, Johansson B, Mortimer JA, Berg S, et al. Role of genes and environments for explaining Alzheimer disease. *Arch Gen Psychiatry* 2006;63:168–74. [PubMed: 16461860]
- [5]. Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet* 2013;45:1452–8. [PubMed: 24162737]
- [6]. Jun G, Ibrahim-Verbaas CA, Vronskaya M, Lambert JC, Chung J, Naj AC, et al. A novel Alzheimer disease locus located near the gene encoding tau protein. *Mol Psychiatry* 2016;21:108–17. [PubMed: 25778476]
- [7]. Jun G, Asai H, Zeldich E, Drapeau E, Chen C, Chung J, et al. PLXNA4 is associated with Alzheimer disease and modulates tau phosphorylation. *Ann Neurol* 2014;76:379–92. [PubMed: 25043464]
- [8]. Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, et al. TREM2 variants in Alzheimer's disease. *N Engl J Med* 2013;368:117–27. [PubMed: 23150934]
- [9]. Jonsson T, Stefansson H, Steinberg S, Jonsdottir I, Jonsson PV, Snaedal J, et al. Variant of TREM2 associated with the risk of Alzheimer's disease. *N Engl J Med* 2013;368:107–16. [PubMed: 23150908]
- [10]. Reitz C, Mayeux R, Alzheimer's Disease Genetics C. TREM2 and neurodegenerative disease. *N Engl J Med* 2013;369:1564–5. [PubMed: 24131184]
- [11]. Cruchaga C, Karch CM, Jin SC, Benitez BA, Cai Y, Guerreiro R, et al. Rare coding variants in the phospholipase D3 gene confer risk for Alzheimer's disease. *Nature* 2014;505:550–4. [PubMed: 24336208]

- [12]. Logue MW, Schu M, Vardarajan BN, Farrell J, Bennett DA, Buxbaum JD, et al. Two rare AKAP9 variants are associated with Alzheimer's disease in African Americans. *Alzheimers Dement* 2014;10:609–18 e11. [PubMed: 25172201]
- [13]. Wetzel-Smith MK, Hunkapiller J, Bhangale TR, Srinivasan K, Maloney JA, Atwal JK, et al. A rare mutation in UNC5C predisposes to late-onset Alzheimer's disease and increases neuronal cell death. *Nat Med* 2014;20:1452–7. [PubMed: 25419706]
- [14]. Sims R, van der Lee SJ, Naj AC, Bellenguez C, Badarinarayan N, Jakobsdottir J, et al. Rare coding variants in PLCG2, ABI3, and TREM2 implicate microglial-mediated innate immunity in Alzheimer's disease. *Nat Genet* 2017;49:1373–84. [PubMed: 28714976]
- [15]. Jonsson T, Atwal JK, Steinberg S, Snaedal J, Jonsson PV, Bjornsson S, et al. A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. *Nature* 2012;488:96–9. [PubMed: 22801501]
- [16]. Wang LS, Naj AC, Graham RR, Crane PK, Kunkle BW, Cruchaga C, et al. Rarity of the Alzheimer disease-protective APP A673T variant in the United States. *JAMA Neurol* 2015;72:209–16. [PubMed: 25531812]
- [17]. Medway CW, Abdul-Hay S, Mims T, Ma L, Bisceglia G, Zou F, et al. ApoE variant p.V236E is associated with markedly reduced risk of Alzheimer's disease. *Mol Neurodegener* 2014;9:11. [PubMed: 24607147]
- [18]. Vardarajan BN, Zhang Y, Lee JH, Cheng R, Bohm C, Ghani M, et al. Coding mutations in SORL1 and Alzheimer disease. *Ann Neurol* 2015;77:215–27. [PubMed: 25382023]
- [19]. Steinberg S, Stefansson H, Jonsson T, Johannsdottir H, Ingason A, Helgason H, et al. Loss-of-function variants in ABCA7 confer risk of Alzheimer's disease. *Nat Genet* 2015;47:445–7. [PubMed: 25807283]
- [20]. Sassi C, Nalls MA, Ridge PG, Gibbs JR, Ding J, Lupton MK, et al. ABCA7 p.G215S as potential protective factor for Alzheimer's disease. *Neurobiol Aging* 2016;46:235 e1–9.
- [21]. Beecham GW, Bis JC, Martin ER, Choi SH, DeStefano AL, van Duijn CM, et al. The Alzheimer's Disease Sequencing Project: Study design and sample selection. *Neurol Genet* 2017;3:e194. [PubMed: 29184913]
- [22]. Ahmad S, Bannister C, van der Lee SJ, Vojinovic D, Adams HHH, Ramirez A, et al. Disentangling the biological pathways involved in early features of Alzheimer's disease in the Rotterdam Study. *Alzheimers Dement* 2018;14:848–57. [PubMed: 29494809]
- [23]. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 2009;25:1754–60. [PubMed: 19451168]
- [24]. Bis JC, Jian X, Kunkle BW, Chen Y, Hamilton-Nelson KL, Bush WS, et al. Whole exome sequencing study identifies novel rare and common Alzheimer's-Associated variants involved in immune response and transcriptional regulation. *Mol Psychiatry* 2018.
- [25]. Firth D Bias reduction of maximum likelihood estimates. *Biometrika* 1993;80:27–38.
- [26]. Heinze G, Schemper M. A solution to the problem of separation in logistic regression. *Stat Med* 2002;21:2409–19. [PubMed: 12210625]
- [27]. Heinze G A comparative investigation of methods for logistic regression with separated or nearly separated data. *Stat Med* 2006;25:4216–26. [PubMed: 16955543]
- [28]. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GR, Thormann A, et al. The Ensembl Variant Effect Predictor. *Genome Biol* 2016;17:122. [PubMed: 27268795]
- [29]. Cingolani P, Platts A, Wang le L, Coon M, Nguyen T, Wang L, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of *Drosophila melanogaster* strain w1118; iso-2; iso-3. *Fly (Austin)* 2012;6:80–92. [PubMed: 22728672]
- [30]. Lee S, Teslovich TM, Boehnke M, Lin X. General framework for meta-analysis of rare variants in sequencing association studies. *Am J Hum Genet* 2013;93:42–53. [PubMed: 23768515]
- [31]. McCarthy S, Das S, Kretzschmar W, Delaneau O, Wood AR, Teumer A, et al. A reference panel of 64,976 haplotypes for genotype imputation. *Nat Genet* 2016;48:1279–83. [PubMed: 27548312]

- [32]. Yan Q, Tiwari HK, Yi N, Gao G, Zhang K, Lin WY, et al. A Sequence Kernel Association Test for Dichotomous Traits in Family Samples under a Generalized Linear Mixed Model. *Hum Hered* 2015;79:60–8. [PubMed: 25791389]
- [33]. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* 2010;26:2190–1. [PubMed: 20616382]
- [34]. Blue EE BJ, Dorschner MO, Tsuang D, Barral SM, Beecham G, Below JE, Bush WS, Butkiewicz M, Cruchaga C, DeStefano A, Farrer LA, Goate A, Haines J, Jaworski J, Jun G, Kunkle B, Kuzma A, Lee JJ, Lunetta K, Ma Y, Martin E, Naj A, Nato AG, Jr, Navas P, Nguyen H, Reitz C, Reyes D, Salerno W, Schellenberg G, Seshadri S, Sohi H, Thornton TA, Valladares O, van Duijn C, Vardarajan BN, Wang L- S, Boerwinkle E, Dupuis J, Pericak-Vance MA, Mayeux R, Wijsman EM, on behalf of the Alzheimer’s Disease Sequencing Project. Genetic variation in genes underlying diverse dementias may explain a small proportion of cases in the Alzheimer’s Disease Sequencing Project. *Dement Ger Cog Disorders* 2018.
- [35]. Vaser R, Adusumalli S, Leng SN, Sikic M, Ng PC. SIFT missense predictions for genomes. *Nat Protoc* 2016;11:1–9. [PubMed: 26633127]
- [36]. Adzhubei IA, Schmidt S, Peshkin L, Ramensky VE, Gerasimova A, Bork P, et al. A method and server for predicting damaging missense mutations. *Nat Methods* 2010;7:248–9. [PubMed: 20354512]
- [37]. Lamkanfi M, Kanneganti TD. Caspase-7: a protease involved in apoptosis and inflammation. *Int J Biochem Cell Biol* 2010;42:21–4. [PubMed: 19782763]
- [38]. Cullen SP, Martin SJ. Caspase activation pathways: some recent progress. *Cell Death Differ* 2009;16:935–8. [PubMed: 19528949]
- [39]. Zhao M, Su J, Head E, Cotman CW. Accumulation of caspase cleaved amyloid precursor protein represents an early neurodegenerative event in aging and in Alzheimer’s disease. *Neurobiol Dis* 2003;14:391–403. [PubMed: 14678756]
- [40]. Rohn TT, Kokoulina P, Eaton CR, Poon WW. Caspase activation in transgenic mice with Alzheimer-like pathology: results from a pilot study utilizing the caspase inhibitor, Q-VD-OPH. *Int J Clin Exp Med* 2009;2:300–8. [PubMed: 20057974]
- [41]. Fiorelli T, Kirouac L, Padmanabhan J. Altered processing of amyloid precursor protein in cells undergoing apoptosis. *PLoS One* 2013;8:e57979. [PubMed: 23469123]
- [42]. Cotman CW, Poon WW, Rissman RA, Blurton-Jones M. The role of caspase cleavage of tau in Alzheimer disease neuropathology. *J Neuropathol Exp Neurol* 2005;64:104–12. [PubMed: 15751224]
- [43]. Albrecht S, Bourdeau M, Bennett D, Mufson EJ, Bhattacharjee M, LeBlanc AC. Activation of caspase-6 in aging and mild cognitive impairment. *Am J Pathol* 2007;170:1200–9. [PubMed: 17392160]
- [44]. Ramcharitar J, Albrecht S, Afonso VM, Kaushal V, Bennett DA, Leblanc AC. Cerebrospinal fluid tau cleaved by caspase-6 reflects brain levels and cognition in aging and Alzheimer disease. *J Neuropathol Exp Neurol* 2013;72:824–32. [PubMed: 23965742]
- [45]. Rohn TT, Head E, Nesse WH, Cotman CW, Cribbs DH. Activation of caspase-8 in the Alzheimer’s disease brain. *Neurobiol Dis* 2001;8:1006–16. [PubMed: 11741396]
- [46]. Rehker J, Rodhe J, Nesbitt RR, Boyle EA, Martin BK, Lord J, et al. Caspase-8, association with Alzheimer’s Disease and functional analysis of rare variants. *PLoS One* 2017;12:e0185777. [PubMed: 28985224]
- [47]. Rohn TT, Rissman RA, Davis MC, Kim YE, Cotman CW, Head E. Caspase-9 activation and caspase cleavage of tau in the Alzheimer’s disease brain. *Neurobiol Dis* 2002;11:341–54. [PubMed: 12505426]
- [48]. Su JH, Zhao M, Anderson AJ, Srinivasan A, Cotman CW. Activated caspase-3 expression in Alzheimer’s and aged control brain: correlation with Alzheimer pathology. *Brain Res* 2001;898:350–7. [PubMed: 11306022]
- [49]. Gervais FG, Xu D, Robertson GS, Vaillancourt JP, Zhu Y, Huang J, et al. Involvement of caspases in proteolytic cleavage of Alzheimer’s amyloid-beta precursor protein and amyloidogenic A beta peptide formation. *Cell* 1999;97:395–406. [PubMed: 10319819]

- [50]. Arispe N, Diaz JC, Simakova O. Abeta ion channels. Prospects for treating Alzheimer's disease with Abeta channel blockers. *Biochim Biophys Acta* 2007;1768:1952–65. [PubMed: 17490607]
- [51]. O'Nuallain B, Acero L, Williams AD, Koeppen HP, Weber A, Schwarz HP, et al. Human plasma contains cross-reactive Abeta conformer-specific IgG antibodies. *Biochemistry* 2008;47:12254–6. [PubMed: 18956886]
- [52]. Antoniou AC, Easton DF. Polygenic inheritance of breast cancer: Implications for design of association studies. *Genet Epidemiol* 2003;25:190–202. [PubMed: 14557987]
- [53]. Ma C, Blackwell T, Boehnke M, Scott LJ, Go TDi. Recommended joint and meta- analysis strategies for case-control association testing of single low-count variants. *Genet Epidemiol* 2013;37:539–50. [PubMed: 23788246]
- [54]. Nhan HS, Chiang K, Koo EH. The multifaceted nature of amyloid precursor protein and its proteolytic fragments: friends and foes. *Acta Neuropathol* 2015;129:1–19. [PubMed: 25287911]

RESEARCH IN CONTEXT

1. Systematic review: The authors are members of the Alzheimer's Disease Sequencing Project and therefore are familiar with emerging pertinent literature. PubMed searches were conducted to identify other relevant publications. References that support the significance of the identified risk loci are cited.
2. Interpretation: Although both common and rare variants in >30 late-onset Alzheimer's disease (LOAD) risk genes have been identified from genome-wide association and whole exome sequencing studies, this report identifies associations with rare variants in several novel loci for LOAD using a design focused on LOAD cases that are likely genetically enriched because they are members of families with multiple affected members. *CASP7* provides further evidence for the role of caspase apoptotic pathways in AD.
3. Future directions: A better understanding of the molecular mechanisms underlying these associations will require functional experiments and *in silico* studies of the connections of genetic variants to gene expression and processing of AD-related proteins. Further studies are also needed to determine whether *CASP7* is a suitable target for development of novel therapies.

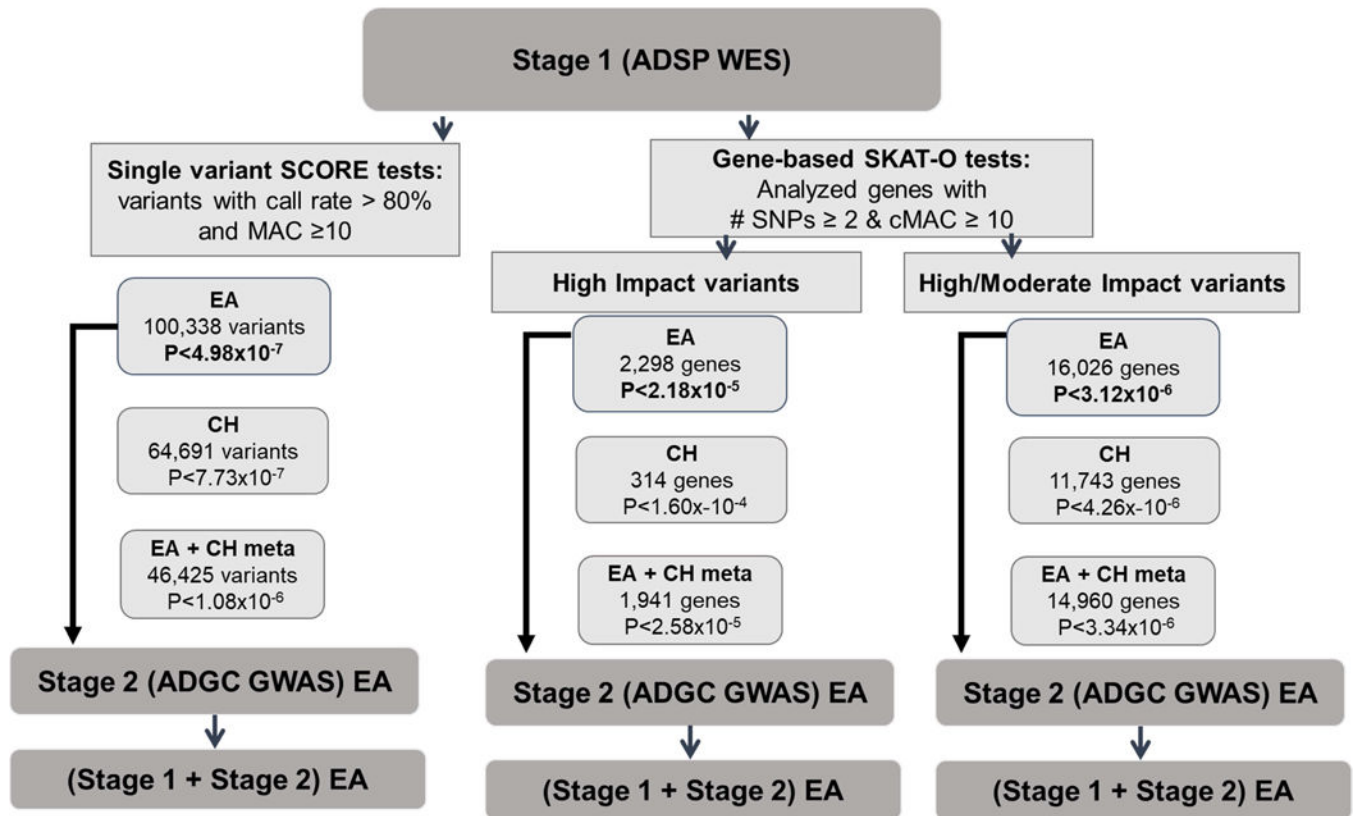


Figure 1.

Analysis design. MAC = minor allele account; cMAC = cumulative minor allele count; EA = European ancestry; CH = Caribbean Hispanic; WES = whole exome sequencing; ADSP = Alzheimer's Disease Sequencing Project; ADGC = Alzheimer's Disease Genetics Consortium; GWAS = genome-wide association study.

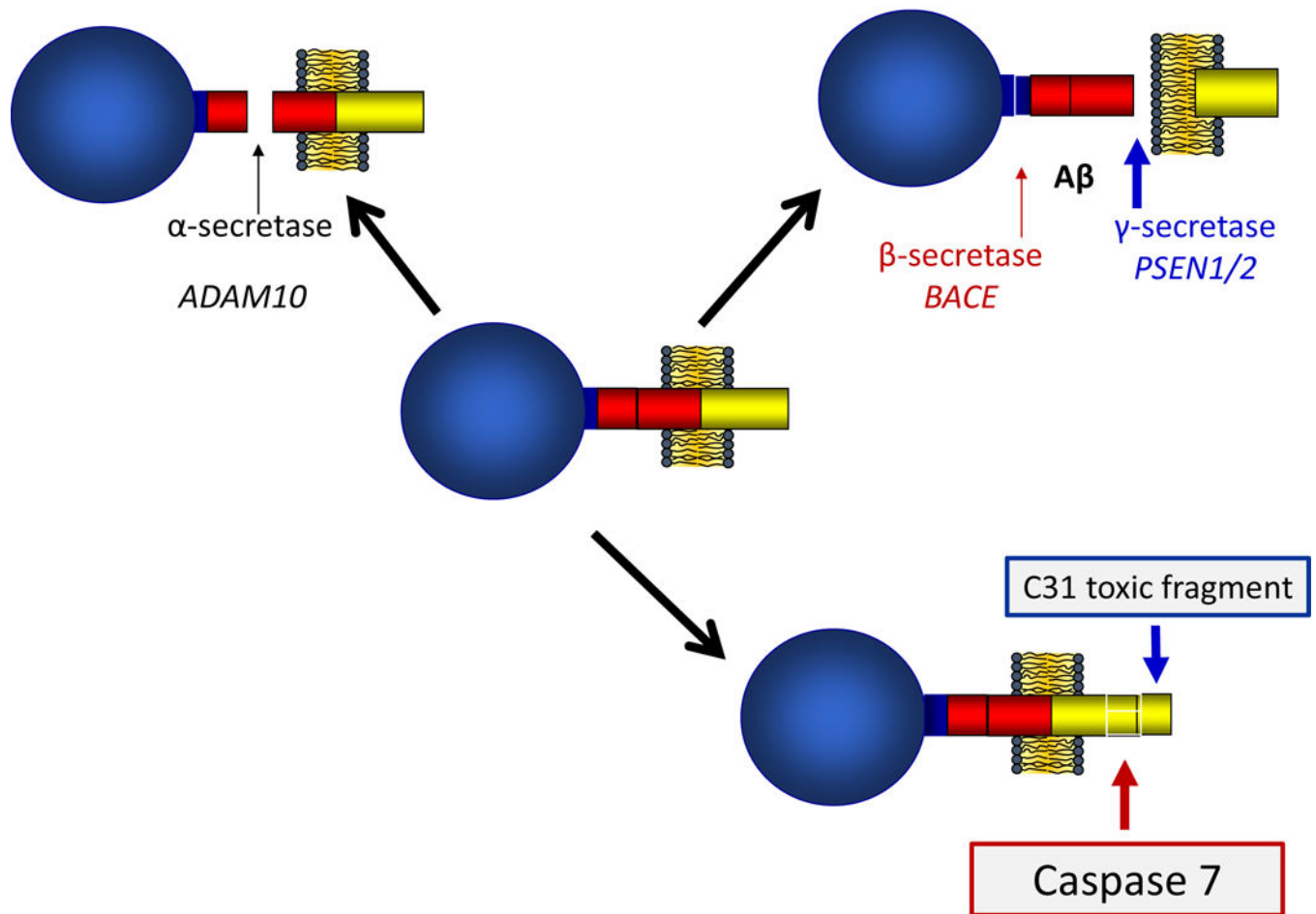


Figure 2. Processing of the amyloid precursor protein (APP) by α (*ADAM10*), β (*BACE*), and γ (*PSEN1/2*) secretases. Alternate processing of APP may result in cleavage of the C31 fragment by the protease encoded by *CASP7* [41]. C31 is one of several C-terminal fragments produced from APP and there is some evidence that it is toxic [54].

Table 1.

Number of AD cases and controls in the Stage 1 WES dataset and Stage 2 ADGC family-based GWAS datasets.

Stage	Cohort	Number controls	Number AD cases	Total
Stage 1 (ADSP WES)	EA	4,917	507	5,424
	CH	177	172	349
	Total	5,094	679	5,773
Stage 2 (ADGC GWAS)	MIRAGE (EA)	704	449	1,153
	NIA-LOAD (EA)	1,457	1,568	3,025
	Total	2,161	2,017	4,178
Combined		7,255	2,696	9,951

AD = Alzheimer's Disease; EA = European Ancestry; CH = Caribbean Hispanic; WES = whole exome sequencing; ADSP= Alzheimer's Disease Sequencing Project; ADGC = Alzheimer's Disease Genetics Consortium; GWAS = genome-wide association study; MIRAGE = Multi Institutional Research in Alzheimer's Genetic Epidemiology Study; NIA- LOAD = National Institute on Aging - Late Onset Alzheimer's Disease Family Study

Table 2.

Single-variant association results in European ancestry individuals excluding the *APOE* region *

Chr: Map Position : Effect Allele : Reference Allele	dbSNP ID	Function	Gene Symbol	Stage 1				Stage 2 Datasets P-value				Stage 1 + 2		
				MAC (EA)	MAC (CH)	MAC Total	OR** (95% CI)	Minimum P-value	Best Model	meta	MIRAGE [†]	NIA-LOAD [‡]	P-value	Direction
6:41129252:C:T	rs75932628 (R47H)	missense	<i>TREM2</i>	34	0	34	11.8(5.0–29.1)	4.56E-12	M0	1.49E-01	7.68E-01	1.06E-01	5.23E-05	+++
18:21118536:G:A	rs150334966	missense	<i>NPC1</i>	11	0	11	38.2(6.7–506.4)	5.78E-09	M0	8.94E-02	9.09E-02	5.83E-01	4.68E-08	---
10:115489177:A:G	rs116437863	missense	<i>CASP7</i>	10	0	10	19.0(5.6–68.2)	2.44E-10	M0	1.28E-01	3.51E-01	6.08E-02	1.92E-10	+++
14:90651236:C:T	rs112153420	synonymous	<i>KCNK13</i>	59	2	61	4.8(2.4–9.3)	1.55E-07	M0	5.86E-01	4.64E-01	3.56E-01	0.0024	++
17:46620725:T:A	rs183316427	missense	<i>HOXB2</i>	78	5	83	5.1(2.6–10.2)	7.63E-08	M1	7.92E-01	6.85E-01	5.05E-01	0.0026	++
11:113846023:C:T	rs62625041	5'UTR	<i>HTR3A</i>	12	0	12	114.9(5.1–24.6)	1.28E-07	M1	1.23E-01	1.33E-01	5.23E-01	0.012	---
19:14829543:C:T	rs79724046 #	synonymous	<i>ZNF333</i>	35	26	61	8.8(3.4–22.0)	1.28E-07	M1	1.23E-01	1.33E-01	5.23E-01	0.00033	+++
3:52552876:C:T	rs372601175	synonymous	<i>STAB1</i>	10	0	10	13.9(3.5–62.7)	3.58E-07	M1	2.01E-01	2.96E-01	2.02E-01	8.68E-05	+++
17:62026823:G:A	rs73992419 #	synonymous	<i>SCN4A</i>	10	19	29	48.2(9.1–242.1)	6.30E-14	M2	3.76E-01	6.84E-01	4.13E-01	0.10	---
7:100679240:G:C	rs144027969	missense	<i>MUC17</i>	35	0	35	191.2(11.4–28.0)	1.63E-09	M2	NA	NA	NA	NA	NA
1:109394718:G:T	rs141968711	missense	<i>AKNAD1</i>	11	0	11	22.8(4.6–113.1)	2.71E-08	M2	1.26E-01	2.73E-02	2.22E-02	0.30	++
1:109394720:C:G	rs747976210	synonymous	<i>AKNAD1</i>	11	0	11	22.8(4.6–113.1)	2.71E-08	M2	1.26E-01	2.73E-02	2.22E-02	0.30	++
2:97270095:C:T	rs34406082	missense	<i>KANSL3</i>	12	7	19	47.7(6.6–366.3)	6.40E-08	M2	9.33E-01	6.83E-01	7.61E-01	0.21	+++
15:42565552:CAGA:C	rs550307753	Inframe deletion	<i>TMEM87A</i>	11	2	13	36.9(7.4–232.6)	2.79E-07	M2	NA	NA	NA	NA	NA
11:17581152:C:T	rs199968574	missense	<i>OTOG</i>	11	1	12	29.3(4.4–161.4)	4.16E-07	M2	9.90E-02	1.51E-01	1.07E-01	0.78	---

* Table shows variants with $p < 5.0 \times 10^{-7}$, bonferroni significance threshold (i.e., SWS);

** OR was based on Firth logistic regression tests;

[†] Family-based ADGC GWAS dataset excluding subjects included in the Stage 1 sample.

[#] variant co-segregated with AD in 2 CH Stage 2 families;

MAC = minor allele account; EA = European ancestry; CH = Caribbean Hispanic;

OR = Odds Ratio for best model (OR and 95% CI for all three model are provided in Supplementary Table 4) M0 = Model 0; adjustment for PCs and sequencing center;

M1 = Model 1: same adjustments as Model 0 + age and sex;

M2 = Model 2: same adjustments as Model 1 + *APOE* $\epsilon 4$ status and *APOE* $\epsilon 2$ status Study-wide significant results highlighted in **bold**

Table 3.

Gene-based association results identified from EA individuals

Gene Symbol	Chr	Start	End	cMAC	# SNPs	cMAF	p-value	Best Model	# SNPs MIRAGE	# SNPs LOAD	Stage 1 + 2	
											Meta p-value	Meta p-value
High Impact Variants (SWS threshold - $p < 2.18 \times 10^{-5}$)												
<i>JMJD4</i>	1	227,920,244	227,922,975	16	10	0.00148	3.56E-05	M0	7	6	9.36E-01	0.0019
<i>ANXA5</i>	4	122,593,712	122,605,898	13	6	0.0012	4.66E-05	M0	6	2	1.24E-01	2.27E-05
<i>ASB13</i>	10	5,682,665	5,695,015	12	2	0.00111	4.32E-05	M1	3	0	1.83E-01	5.20E-05
<i>ASCC1</i>	10	73,857,161	73,970,584	122	5	0.01126	2.51E-06	M2	6	5	7.89E-01	0.00019
<i>C1orf173</i>	1	75,037,039	75,102,074	11	10	0.00102	4.41E-05	M2	15	12	1.38E-01	1.50E-05
<i>AARD</i>	8	117,954,812	117,954,935	16	2	0.00148	4.84E-05	M2	3	0	6.16E-03	1.01E-06
High/Moderate Impact Variants (SWS threshold - $p < 3.12 \times 10^{-6}$)												
<i>IGH16</i>	14	106,329,417	106,329,468	10	9	0.00093	3.46E-07	M0	0	0	NA	NA
<i>DTYMK</i>	2	242,615,564	242,625,278	15	13	0.00138	2.95E-11	M2	0	3	6.60E-01	3.40E-08

Chr = chromosome; cMAC = cumulative minor allele account; cMAF = cumulative minor allele frequency; EA = European Ancestry; CH = Caribbean Hispanic

M0 = Model 0; adjustment for PCs and sequencing center; M1 = Model 1; same adjustments as Model 0 + age and sex; M2 = Model 2; same adjustments as Model 1 + APOE ε4 status and APOE ε2 status
 Study-wide significant (SWS) results highlighted in **bold**