

SHORT COMMUNICATION

An EGLN1 mutation may regulate hypoxic response in cyanotic congenital heart disease through the PHD2/HIF-1A pathway

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KEYWORDS

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Abstract Cyanotic congenital heart disease (CCHD), a term describing the most severe congenital heart diseases are characterized by the anatomic malformation of a right to left shunt. Although the incidence of CCHD are far less than the that of congenital heart diseases (CHD), patients with CCHD always present severe clinical features such as hypoxia, dyspnea, and heart failure. Chronic hypoxia induces hypoxemia that significantly contributes to poor prognosis in CCHD. Current studies have demonstrated that the prolyl-4-hydroxylase2 (*PHD2*, encoded by *EGLN1*)/hypoxia-inducible factor-1A (*HIF-1A*) pathway is a key regulator of hypoxic response. Thus, we aim to assess the associations of single polymorphisms (SNPs) of the *EGLN1* gene and hypoxic response in CCHD. A missense variant of *EGLN1* c.380G>C (rs1209790) was found in 46 patients (46/126), with lower hypoxia incidence and higher rate of collateral vessel formation, compared with the wild type ($P < 0.05$). *In vitro* experiments, during hypoxia, *EGLN1* mutation reduced *EGLN1* expression compared with the wild type, with higher *HIF-1A*, *VEGF* and *EPO* expression levels in the mutant. No difference in *HK1* expression was observed between the mutant and wild type. CCHD patients with c.380G>C showed improved response to hypoxia compared with the

Abbreviations: *HIF-1A*, hypoxia inducible factor-1A; *VEGF*, vascular endothelial growth factor; *EPO*, erythropoietin; *HK1*, Hexokinase I.

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wild-type counterparts. The *EGLN1* c.380G>C mutation improves hypoxic response through the *PHD2/HIF-1A* pathway, which may provide a molecular mechanism for hypoxic response in CCHD. The effects of the *EGLN1* c.380G>C mutation on CCHD prognosis deserve further investigation.

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Introduction

Cyanotic congenital heart disease (CCHD), a term describing the most severe congenital heart diseases characterized by the anatomic malformation of a right to left shunt, includes tetralogy of Fallot (TOF), double outlet of right ventricle (DORV), transposition of the great arteries (TGA), and pulmonary artery (PA) defects. Although only about 0.08–0.12% of live births are affected by CCHD, far less than the incidence of congenital heart diseases (CHD), patients with CCHD always present severe clinical features such as hypoxia, dyspnea, and heart failure.^{1–3} Many patients present cyanosis before they could benefit from surgical treatment.⁴

Based on the specific anatomic malformation, patients with CCHD usually develop chronic hypoxia, which induces hypoxemia that significantly contributes to poor prognosis in CCHD, with dysplasia, convulsions, and sudden death, even after an urgent surgical intervention.^{5–7} Consequently, the ability of hypoxic response determines the prognosis of CCHD patients. Current studies have demonstrated that the prolyl-4-hydroxylase2 (*PHD2*, encoded by *EGLN1*)/hypoxia-inducible factor-1A (*HIF-1A*) pathway is a key regulator of hypoxic response. Under normoxia, *PHD2* triggers the oxygen-dependent hydroxylation of ODD within *HIF-1A*. During hypoxia, however, such hydroxylation is blocked, resulting in *HIF-1A* accumulation, angiogenesis, erythropoiesis, and glycolysis. It was reported by many studies that angiogenesis and erythropoiesis enhance hypoxic response through the *PHD2/HIF-1A* pathway, improving cell survival under hypoxic conditions.⁸ These findings indicated that the *PHD2/HIF-1A* pathway plays a crucial role in hypoxic response in CCHD.

Lorenzo FR⁹ assessed *EGLN1* polymorphisms in Tibetan high-altitude adaptation. However, most studies focused on polycythemia; indeed, more than 13 *EGLN1* mutations have been reported in association with polycythemia.^{10–13} Meanwhile, the effects of *EGLN1* polymorphisms on CCHD remain unclear. Thus, we hypothesized that *EGLN1* polymorphisms affect hypoxic response in CCHD through the *PHD2/HIF-1A* pathway.

This study aimed to assess the association of hypoxia with collateral vessel malformation among different *EGLN1* genotypes. In addition, we analyzed the expression levels of target genes involved in the *PHD2/HIF-1A* pathway for various genotypes. The current findings may help further understand the genetic basis of hypoxic response, providing new options for intervention and adjunctive therapy aiming to improve hypoxic response in CCHD.

Materials and methods

Subjects

A total of 126 CCHD patients in Children's Hospital of Chongqing Medical University were screened for *EGLN1* by sequencing (60 males and 66 females; 19.80 ± 1.99 months old). They included 58 TOF, 35 DORV, 10 PA defect, 8 persistent truncus arteriosus (PTA), 7 TGA, 6 anomalous pulmonary venous drainage, and 2 complete atrioventricular septal defect (CAVSD) cases. Mean variabilities of limb SpO₂ were less than 2%. Hypoxia was defined as SpO₂<90%, and normoxia as SpO₂≥90%. Development of collateral vessel was observed by echocardiography during hospitalization. The participants without consanguineous marriages for at least three generations or a family history of congenital heart diseases were Chinese Han individuals. This study was approved by the local ethics committees of Children's Hospital of Chongqing Medical University. All patients provided signed written informed consent. The privacy rights of human subjects were observed.

Sample preparation and genomic analyses

DNA was extracted from peripheral blood with MagPure Blood DNA Mini KF Kit (Magen, NO:MD5111-01). The obtained DNA was submitted to high-throughput sequencing of the *EGLN1* gene, including the 5'UTR and 3'UTR, exons, and the 2 kb region upstream the promoter. This work was completed by Beijing Genomics Institute. Then, the gene variants were identified by Sanger sequencing, with the DNA-Star software used for data analysis.

Cell culture

HEK-293 cells were cultured in Dulbecco's Eagle's Modified Media (DMEM, Gibco) containing 10% fetal bovine serum (FBS) (Gibco) and 100 mg/ml penicillin/streptomycin in a humidified environment with 21% O₂ and 5% CO₂, at 37 °C. HEK-293 cells were co-transfected with 1.8 μg total DNA (variant, wild-type, and vector, respectively) and 4.5 μl Lipofectamine 2000 (Invitrogen). For normoxic experiments, the cells were cultured in a humidified environment containing 21% O₂ and 5% CO₂ at 37 °C. Hypoxic experiments were performed by culture in presence of 3% O₂ and 5% CO₂, also at 37 °C.

Real-time PCR

A total of 48 h after transfection, total RNA was extracted with TRIZOL Reagent (Ambion), and reverse-transcribed into complementary DNA (cDNA) using the Prime Script RT reagent kit containing gDNA Eraser (Takara, NO: RR047A). Then, cDNA was submitted to PCR amplification with primer sets for actin, HIF-1A, VEGF, EPO and HK1. Actin gene primers were a gift from Sangon Biotech. The other primers were: HIF-1A, forward 5'-TGGAGTGCAGTGGAGCAATC-3' and reverse 5'-CAGGCATGGTGGTACATGCT-3'; VEGF, forward 5'-GTCCCTCTTGAATTGGAT-3' and reverse 5'-TGATGTGGGTGGTGTGTC-3'; EPO, forward 5'-CTTCTCCTGTCCCTGCTGTC-3' and reverse 5'-TGATATTCTCGGCCTCCTTG-3'; HK1 forward 5'-TCCTCGTCAAGACAGTGTGC-3' and reverse 5'-CCCACAGTACATTCAGACG-3'.

Immunoblotting

A total of 52 h after transfection, total cellular protein was extracted on ice using $1 \times$ lysis buffer (KeyGEN BioTECH, NO: KGP250) supplemented with a protease inhibitor and a phosphatase inhibitor. Equal amounts of protein were resolved by 10% SDS-polyacrylamide gel electrophoresis and transferred onto PVDF membranes. Non-specific bands were blocked with TBS-T containing 5% nonfat-dried milk for 1 h. Then, the membranes were successively incubated with specific primary antibodies at 4 °C for 15 h, and secondary antibodies at room temperature for 1.5 h. The membranes were washed three times with TBS-T, and target protein bands were visualized by ECL (KeyGEN BioTECH, NO: KGP1127). The primary antibodies used were: rabbit anti-EGLN1 mAb (Cell Signaling Technology, NO:48355, 1:1000), rabbit anti-HIF-1 alpha mAb (Cell Signaling Technology, NO:14179S, 1:500), anti-VEGF antibody (Arigo, NO:10513, 1:500), anti-EPO antibody (ab126876, 1:500), rabbit anti-hexokinase I mAb (Cell Signaling Technology, NO:2024S), and anti-actin antibody (4A Biotech Co, NO: ICM001-050, 1:1000). Secondary antibodies were goat anti-mouse IgG (ATGene) and goat anti-Rabbit IgG (Sino biological Inc, NO:HO10AP0801).

Statistics

SPSS 20.0 was used for statistical analyses. Data are mean \pm standard error (SE). Enumeration data were assessed the Chi-square test. Differences among groups were analyzed by one way analysis of variance (ANOVA). $P < 0.05$ was considered statistically significant.

Results

Genomic testing for hypoxic adaptation related mutations in patients with CCHD

The missense mutation c.380G>C was found in the EGLN1 gene in 46 patients with CCHD (21 males and 25 females; variant and wild-type bearing patients were 22.82 ± 3.89 months and 18.06 ± 2.20 months old, respectively; Fig. 1A and B). A total of 36.96% (17/46) of these carriers had

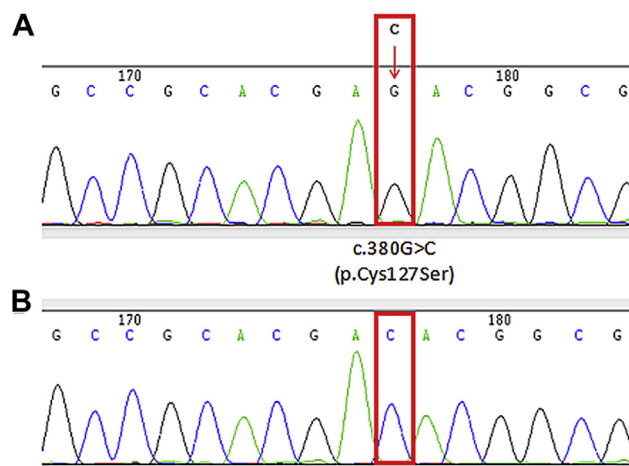


Figure 1 Identification of *EGLN1* mutation in patients with cyanotic congenital heart disease. Results are presented with template strand which is opposite to the coding strand, so the remarked sign is changing C into G. (A). Sequencing results of *EGLN1* c.380G>C mutation in 46 patients. (B) Sequencing results of wild-type DNA mutation in 80 patients.

hypoxemia, indicating a lower rate compared with the wild-type group (52/80) ($p = 0.003$, Table 1). In addition, angiogenesis is known to benefit hypoxic response. Thus, to further assess the association of c.380G>C mutation with hypoxic response, collateral vessel formation was observed. Interestingly, 50% (23/46) of mutation carriers showed collateral vessel formation, a higher rate compared with that of the wild-type group (22/80) ($p = 0.01$, Table 1). Collateral vessel formation is a compensatory mechanism for response to the hypoxic environment. In this study, high incidence of collateral vessel formation was observed in patients with the c.380G>C mutation, suggesting this gene alteration may enhance hypoxic response.

Plasmid validation and transfection

Recombinant plasmids (c.380G>C mutation, wild-type, and vector) were confirmed by sequencing. A total of 48 h after transfection, GFP expression by the plasmid vector was observed on a fluorescence microscope, suggesting successful transfection.

The c.380G>C mutation reduces PHD2 protein expression under hypoxic conditions

To assess the effect of the c.380G>C mutation, PHD2 protein expression was analyzed by immunoblot. Unlike the empty vector and control (untreated cells) groups, the wild-type and c.380G>C mutation groups expressed the exogenous PHD2 protein, providing further evidence for successful transfection and encoding. As shown in Fig. 2A, PHD2 levels in the c.380G>C mutation group decreased by 53.34% compared with the wild-type group under hypoxic conditions ($p < 0.05$, Fig. 2A). However, there was no significant difference between the c.380G>C mutation and wild type groups in normoxia ($P > 0.05$, Fig. 2B).

Table 1 Associations of *EGLN1* gene c.380G>C mutation with CCHD features.

parameter	Total (n = 126)	c.380G>C mutation (n = 46)	Wild-type (n = 80)	P-value
Hypoxia, n (%)	69 (54.76%)	17 (36.96%)	52 (65.00%)	0.003
CV, n (%)	45 (35.71%)	23 (50.00%)	22 (7.50%)	0.01

CV: collateral vessel.

The c.380G>C mutation activates the PHD2/HIF-1A pathway and induces HIF-1A accumulation during hypoxia

PHD2 is a hydroxylase that interacts with HIF-1A and hydroxylates its ODD region, resulting in acute degradation of the HIF-1A protein. Thus, we hypothesized that the c.380G>C mutation may affect the degradation of the HIF-1A protein by altering PHD2 mediated hydroxylation. To test this hypothesis, HIF-1A expression was analyzed in cells transfected with different recombinant plasmids. Under hypoxic conditions, HIF-1A protein expression levels in the c.380G>C mutation group were overtly increased (by 1.81-fold) compared with the wild-type group ($P < 0.01$, Fig. 2C). These results suggested that the c.380G>C mutation may activate the PHD2/HIF-1A pathway and induce HIF-1A protein expression. Under normoxic conditions, HIF-1A protein was degraded immediately after production. Thus, the HIF-1A protein under normoxic conditions was barely detectable. In addition, there were no significant differences among groups under hypoxic and normoxic conditions (data not shown).

The c.380G>C mutation stimulates *VEGF* expression in association with the PHD2/HIF-1A pathway under hypoxic conditions

VEGF, a crucial component of the PHD2/HIF-1A pathway, is induced by the HIF-1A protein through PHD2/HIF-1A

signaling under hypoxic conditions. Meanwhile, it was shown that the c.380G>C mutation induces HIF-1A protein expression. Hence, it is conceivable that the c.380G>C mutation may upregulate *VEGF* in association with the PHD2/HIF-1A pathway under hypoxic conditions. Indeed, our results showed that *VEGF* mRNA levels in the c.380G>C mutation group were higher (1.41-fold) compared with the wild-type group in hypoxia ($p < 0.05$, Fig. 3A); the *VEGF* protein was also up-regulated by 3.19-fold ($p < 0.05$, Fig. 3C). However, these beneficial effects were not observed in normoxia ($p > 0.05$, Fig. 4B; $p < 0.05$, Fig. 3B and D). These findings indicated that the c.380G>C mutation upregulated *VEGF* in association with the PHD2/HIF-1A pathway in hypoxia.

The c.380G>C mutation increases *EPO* expression in association with PHD2/HIF-1A signaling under hypoxic conditions

EPO, another critical component of the PHD2/HIF-1A pathway, is upregulated by HIF-1A in hypoxia. Next, *EPO* expression levels were detected by immunoblot and RT-PCR, respectively, to further assess the effects of the c.380G>C mutation on hypoxic response associated with PHD2/HIF-1A signaling. As expected, the gene expression levels of *EPO* were increased by 0.42-fold in c.380G>C mutation harboring cells versus wild-type counterparts ($p < 0.05$, Fig. 4A). Similarly, *EPO* protein levels in the c.380G>C mutation group were significantly increased by

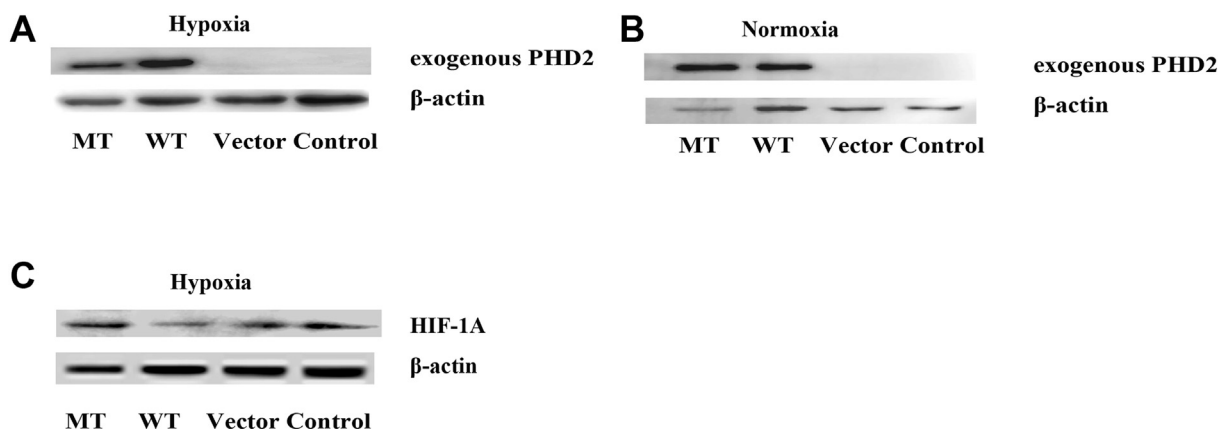


Figure 2 Functional study of *EGLN1* mutation using western blot. Expression of exogenous PHD2 protein (encoded by *EGLN1*) was observed only in c.380G>C mutation and wild-type, providing a further evidence for the successful transfection. (A) Cells were transfected in a 6-well format with 1.8 μ g recombinant plasmids under hypoxic conditions (3% O₂). c.380G>C mutation reduced the expression of PHD2 compared with wild-type under hypoxic conditions. β -actin was used as a loading control. (B) There is no significant difference of PHD2 protein expression between c.380G>C mutation and wild-type during normoxia. (C) Expression of *HIF-1A* in cells transfected with recombinant plasmids under hypoxic condition. Cells were infected with c.380G>C mutation, wild-type and empty vector. Under hypoxic conditions, increased expression levels of *HIF-1A* in c.380G>C mutation were observed in four independent experiments. MT: c.380G>C mutation; WT: wild-type; vector: empty vector with no PHD2 expression; control: untreated group.

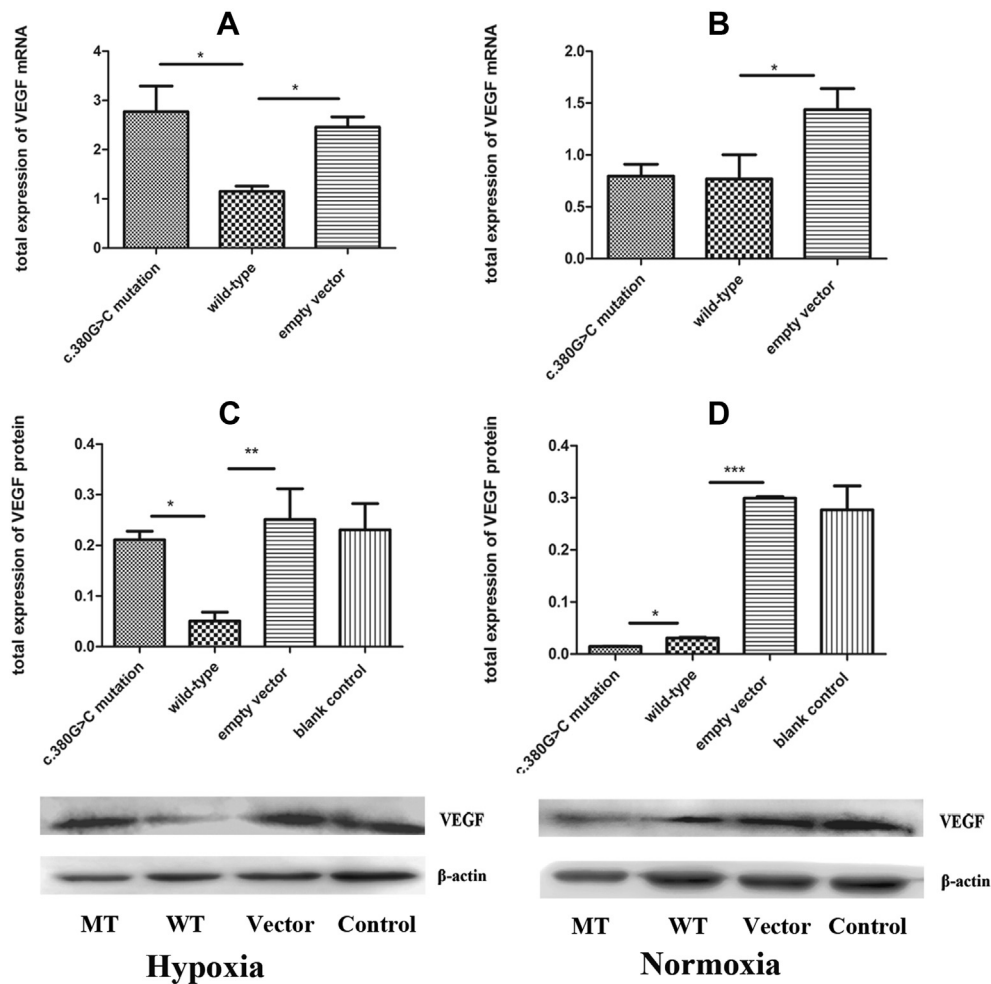


Figure 3 Expression of *VEGF* in cells transfected with c.380G>C mutation, wild-type and empty vector. (A) Under hypoxic conditions, increased expression levels of *VEGF* mRNA in c.380G>C mutation were observed compared with wild-type. (B) No significant difference of *VEGF* mRNA expression was observed between c.380G>C mutation and wild-type during normoxia. (C) Under hypoxic conditions, increased expression levels of *VEGF* protein in c.380G>C mutation were observed compared with wild-type. (D) Under normoxic conditions, expression of *VEGF* protein in c.380G>C mutation is lower than that in wild-type. MT: c.380G>C mutation; WT: wild-type; vector: empty vector with no PHD2 expression; control: untreated group.

2.05-fold compared with wild-type counterparts ($p = 0.011$, Fig. 4C). Next, *EPO* expression in normoxic conditions was analyzed, and upregulation was not observed ($p > 0.05$, Fig. 4B; $p > 0.05$, Fig. 4D). These findings suggested that the c.380G>C mutation increased *EPO* expression in association with the PHD2/HIF-1A pathway during hypoxia.

HK1 expression is associated with the PHD2/HIF-1A pathway

It is well-known that hypoxia stimulates glycolysis. Next, to comprehensively assess the effects of the c.380G>C mutation on glycolysis associated with the PHD2/HIF-1A pathway, the expression levels of *HK1*, a critical enzyme involved in glycolysis, were determined by Western blot and RT-PCR, respectively. No significant differences were observed among all groups under hypoxic or normoxic conditions (data not shown).

Discussion

This study revealed a missense variant of the *EGLN1* gene (c.380G>C) in patients with CCHD, with lower hypoxia incidence in CCHD patients with the c.380G>C mutation compared with wild-type subjects. In addition, the incidence of collateral vessel formation, considered a critical mechanism of hypoxic response, was higher in the c.380G>C mutation group compared with wild-type patients. Next, *in vitro* experiments were performed to assess the effects of the c.380G>C mutation on hypoxic response. Our results demonstrated that the c.380G>C mutation reduced PHD2 protein expression, which resulted in alleviated hydroxylation mediated by PHD2 and increased accumulation of the HIF-1A protein. *VEGF* and *EPO* are crucial components of the PHD2/HIF-1A pathway. Many studies have reported that PHD2 alteration indeed affects *VEGF* and *EPO* expression. In this study, the c.380G>C mutation upregulated *HIF-1A*, *VEGF*, and *EPO* in association with the PHD2/HIF-1A pathway under hypoxic conditions.

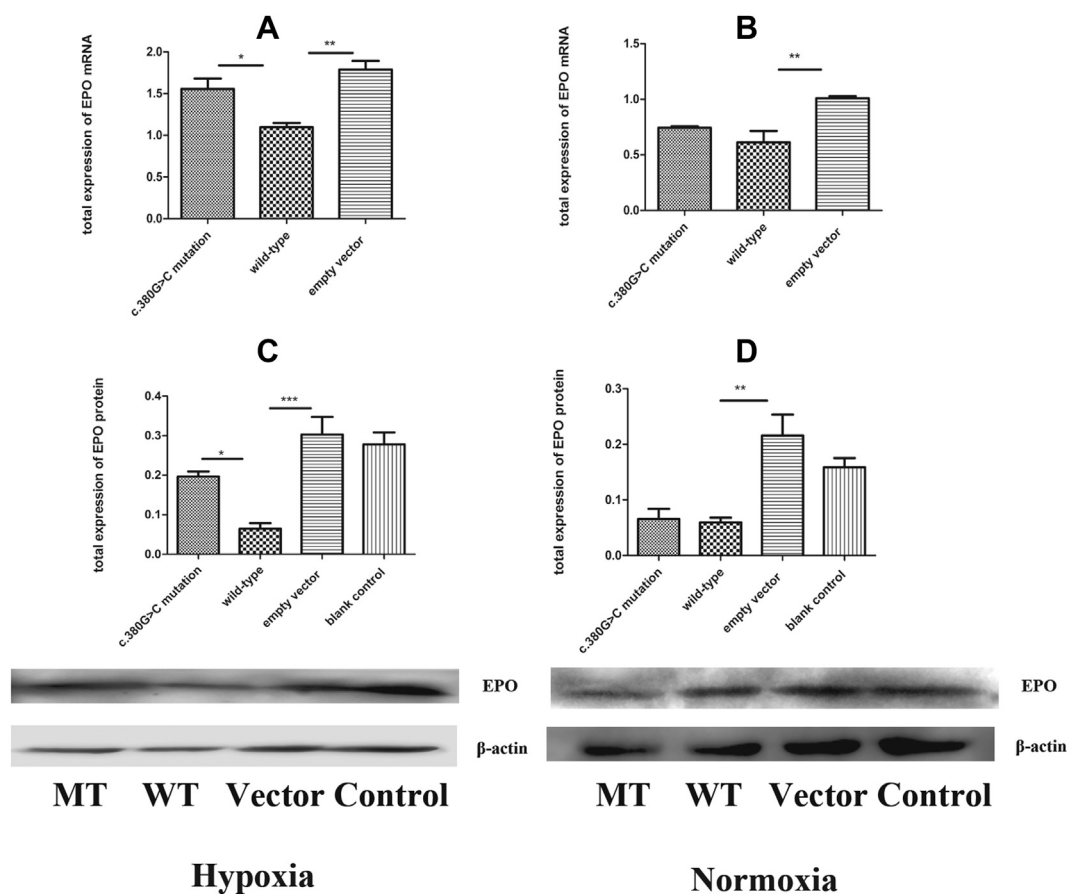


Figure 4 Expression of *EPO* in cells transfected with c.380G>C mutation, wild-type and empty vector. (A) Under hypoxic conditions, increased expression levels of *EPO* mRNA in c.380G>C mutation were observed compared with wild-type. (B) No significant difference of *EPO* mRNA expression between c.380G>C mutation and wild-type was observed during normoxia. (C) Under hypoxic conditions, increased expression level of *EPO* protein in c.380G>C mutation were observed compared with wild-type. (D) Under normoxic conditions, expression of *EPO* protein in c.380G>C mutation is lower than that in wild-type. MT: c.380G>C mutation; WT: wild-type; vector: empty vector with no PHD2 expression; control: untreated group.

The PHD2/HIF-1A pathway in the pathogenesis of hypoxemia and CCHD have been described by many experimental and clinical studies.^{14,15} Under hypoxic conditions, upregulating the adaptive gene (e.g. *VEGF*, *EPO*, and several glycolytic enzymes) by the accumulated *HIF-1A* protein plays an important role in response to hypoxia, and improves the outcome of CCHD patients.^{16–20} Overexpression of *VEGF* increases neovascularization and peripheral oxygen delivery, improving hypoxic response in individuals with hypoxemia.^{17,21} Likewise, *EPO* maintains oxygen homeostasis by stimulating the proliferation of erythroid progenitors for erythropoiesis under hypoxic conditions. Increased erythrocyte amounts facilitate O₂ transport in response to hypoxia.²² These evidences demonstrate that overexpression of *VEGF* and *EPO* indeed could improve hypoxemia and CCHD prognosis in association with the PHD2/HIF-1A pathway. In this study, compared with wild-type individuals, the c.380G>C group showed a lower hypoxia incidence and a higher collateral vessel formation rate. Angiogenesis is one of the known mechanisms of hypoxia response. The current findings suggested that the c.380G>C mutation might benefit hypoxic response. In *in vitro* experiments, we found that the

c.380G>C mutation increased HIF-1A protein accumulation. In addition *VEGF* and *EPO* were upregulated in the c.380G>C mutation group, in association with PHD2/HIF-1A signaling. Overexpression of *VEGF* and *EPO* enhances tolerance to hypoxia and improves the outcome of CCHD patients, as elucidated previously. Therefore, our findings suggested that the c.380G>C mutation protects patients with CCHD from hypoxia in association with the PHD2/HIF-1A pathway. This may provide a molecular mechanism for hypoxic response in patients with CCHD.

Although the c.380G>C mutation has been assessed for Tibetan high-altitude adaptation, its function has not been analyzed alone. On the one hand, Lorenzo FR⁹ analyzed the two missense variants c. [12C > G; 380G > C] together. On the other hand, the molecular mechanisms and pathophysiology of hypoxic response between CCHD and Tibetan may be different. We first assessed the association of the c.380G>C mutation with CCHD, also analyzing the function of the c.380G>C mutation *in vitro*. Recently, associations of gene polymorphisms with diseases have been explored in many studies, with significant findings reported. Indeed, gene polymorphisms play important roles in many diseases, e.g. effects of R530X *MIB1* on left ventricular non-

compaction (LVNC),²³ c.300delA *DNAJC19* on dilated cardiomyopathy syndrome (DCMA),²⁴ c.609C > G *EGLN1* on polycythemia,¹² et al. However, the associations of gene polymorphisms with CCHD remain unclear. Polymorphisms in *EGLN1* were reported in polycythemia and high-altitude adaptation. The associations of *EGLN1* polymorphisms with CCHD remain undefined. Thus, this may be the first study assessing the effects of *EGLN1* gene polymorphisms on hypoxic response in CCHD. Our findings may help further understand the genetic basis of hypoxic response, and provide insights for developing new approaches in reducing CCHD mortality, improving patient prognosis, earning more time before surgery, and improving the patients' quality of life.

Following hypoxia, the HIF-1A protein accumulates at an alarming rate, but is rapidly degraded by a mechanism involving PHD2. PHD2 protein levels directly affect *HIF-1A* expression by controlling the hydroxylation of ODD within *HIF-1A*, but not transcription. Thus, PHD2 expression does not alter the gene expression of *HIF-1A*, which fully accounts for the findings of no significant differences in *HIF-1A* mRNA expression among groups. Another characteristic of response to chronic hypoxia is a shift from lipid oxidation to glycolysis. As a critical glycolytic enzyme, *HK1* is upregulated under hypoxic conditions.²⁵ Surprisingly, discordant changes were found in the expression levels of *HK1* and *HIF-1A*. *HK1* expression showed no significant difference between the c.380G>C mutation and wild-type groups. This may be explained by the fact that *HK1* is regulated by the AMP-dependent protein kinase (AMPK) signaling pathway, not PHD2/HIF-1A signaling.^{25,26} Inevitably, this study had several limitations, including small sample size, which also limited sub-group analysis by severity of congenital heart defects.

Conclusion

Hypoxemia incidence in CCHD patients with the c.380G>C mutation in *EGLN1* is lower than in wild type counterparts. Meanwhile, the c.380G>C mutation contributes to collateral vessel formation. The c.380G>C upregulates *VEGF* and *EPO* in association with the PHD2/HIF-1A pathway to improve hypoxic response in patients with CCHD. This study may provide a new approach for intervention and adjunctive therapy to improve hypoxic response in CCHD.

Conflict of interest

There are no financial or other interests of conflict exist.

Acknowledgements

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