




Review

# Overview of Human Intervention Studies Evaluating the Impact of the Mediterranean Diet on Markers of DNA Damage

Cristian Del Bo<sup>1,\*</sup> , Mirko Marino<sup>1</sup> , Daniela Martini<sup>2</sup> , Massimiliano Tucci<sup>1</sup>,  
Salvatore Ciappellano<sup>1</sup>, Patrizia Riso<sup>1,†</sup> and Marisa Porrini<sup>1,†</sup>

<sup>1</sup> Department of Food, Environmental and Nutritional Sciences (DeFENS), Università degli Studi di Milano, 20122 Milan, Italy; mirko.marino@unimi.it (M.M.); massimiliano.tucci.mt@gmail.com (M.T.); salvatore.ciappellano@unimi.it (S.C.); patrizia.riso@unimi.it (P.R.); marisa.porrini@unimi.it (M.P.)

<sup>2</sup> Human Nutrition Unit, Department of Veterinary Science, University of Parma, 43125 Parma, Italy; daniela.martini@unipr.it

\* Correspondence: cristian.delbo@unimi.it; Tel.: +39-025-031-6730

† PR and MP equally contributed to this work.

Received: 29 December 2018; Accepted: 11 February 2019; Published: 13 February 2019



**Abstract:** The Mediterranean diet (MD) is characterized by high consumption of fruits, vegetables, cereals, potatoes, poultry, beans, nuts, lean fish, dairy products, small quantities of red meat, moderate alcohol consumption, and olive oil. Most of these foods are rich sources of bioactive compounds which may play a role in the protection of oxidative stress including DNA damage. The present review provides a summary of the evidence deriving from human intervention studies aimed at evaluating the impact of Mediterranean diet on markers of DNA damage, DNA repair, and telomere length. The few results available show a general protective effect of MD alone, or in combination with bioactive-rich foods, on DNA damage. In particular, the studies reported a reduction in the levels of 8-hydroxy-2'-deoxyguanosine and a modulation of DNA repair gene expression and telomere length. In conclusion, despite the limited literature available, the results obtained seem to support the beneficial effects of MD dietary pattern in the protection against DNA damage susceptibility. However, further well-controlled interventions are desirable in order to confirm the results obtained and provide evidence-based conclusions.

**Keywords:** Mediterranean diet; DNA damage; DNA repair; telomere length; dietary intervention study

## 1. Introduction

Oxidative stress is a condition characterized by an imbalance between formation of reactive oxygen species (ROS) and antioxidant defense mechanisms. Overproduction of ROS can cause oxidative damage to lipids, proteins, and DNA [1]. The integrity and stability of DNA is essential to life and for the maintenance of normal cell functions. The most common types of stressors, apart from oxidative species, include chemical agents, ultraviolet/ionizing radiation, and xenobiotics that can contribute to DNA damage and to formation of base deamination, base alkylation, base dimerization, base oxidation, and single/double strand breakage [1]. The resulting DNA damage, if not properly repaired, can increase risk of mutagenesis and bring to the onset or the development of numerous degenerative diseases including cardiovascular diseases (CVDs), diabetes mellitus, Alzheimer's disease, and cancer [2–4]. Chronic oxidative stress has also been reported as a critical mechanism involved in telomere shortening [5]. Telomeres consist of long stretches of TTAGGG-DNA repeats associated with specific proteins and located at the end of chromosomes. They are involved in

the protection of chromosomal stability and have been recognized as potential biomarkers of biological aging [6].

Increasing evidence suggests the crucial role of dietary and lifestyle habits as determinants of DNA oxidative damage [7,8], DNA repair [9], and telomere length [10]. Bioactives and bioactive-rich foods can exert a protective effect against oxidative stress resulting in lower DNA damage. For instance, DNA damage protection has been observed in several human intervention studies following the consumption of tomato [11,12], broccoli [13,14], spinach [15,16], blueberry [17,18], orange juice [19,20], nuts [21,22], green tea [23,24], and coffee [25,26]. Most of these foods are present in the Mediterranean diet (MD) and represent a rich sources of bioactive compounds such as vitamins, carotenoids, glucosinolates, and polyphenols acting as antioxidants or activators of endogenous multiple defense systems [27,28]. In addition, food bioactives have been demonstrated to induce DNA repair activity and to help in maintaining telomere length [10].

The MD has been identified as a sustainable and healthy dietary pattern characterized by a high intake of vegetables, legumes, fruits and nuts, cereals, olive oil, a moderately high intake of fish, a low-to-moderate intake of dairy products, a low intake of meat and poultry, and a regular but moderate intake of alcohol, primarily in the form of wine and generally within meals [29,30]. The adherence to the MD has been recognized to have a favorable effect on blood pressure, insulin sensitivity, lipid profile, inflammation, and oxidative stress, with a consequent decreased risk of numerous non-communicable diseases such as CVDs, cancer, and related deaths [31–33]. The results of PREDIMED (PREvención con DIeta MEDiterránea), a multicenter, randomized, nutritional intervention trial carried out in Spain from 2003 to 2011, demonstrated the protective effect of the traditional MD against CVD in individuals at high cardiovascular risk [34–37].

The exact mechanisms by which MD exerts its protective effects are not yet completely understood since numerous aspects such as lifestyle and environmental factors, characterizing Mediterranean style, can be involved and may interact contributing to the health benefits observed. However, the intake of bioactive compounds with a direct and/or indirect antioxidant action could represent a plausible, even if not the unique, explanation for the apparent benefits. A specific target of this protection could be represented by the defense of DNA from oxidative damage. The present review summarizes the main findings derived from human intervention studies addressing the impact of the MD on DNA protection and telomere length.

## 2. Materials and Methods

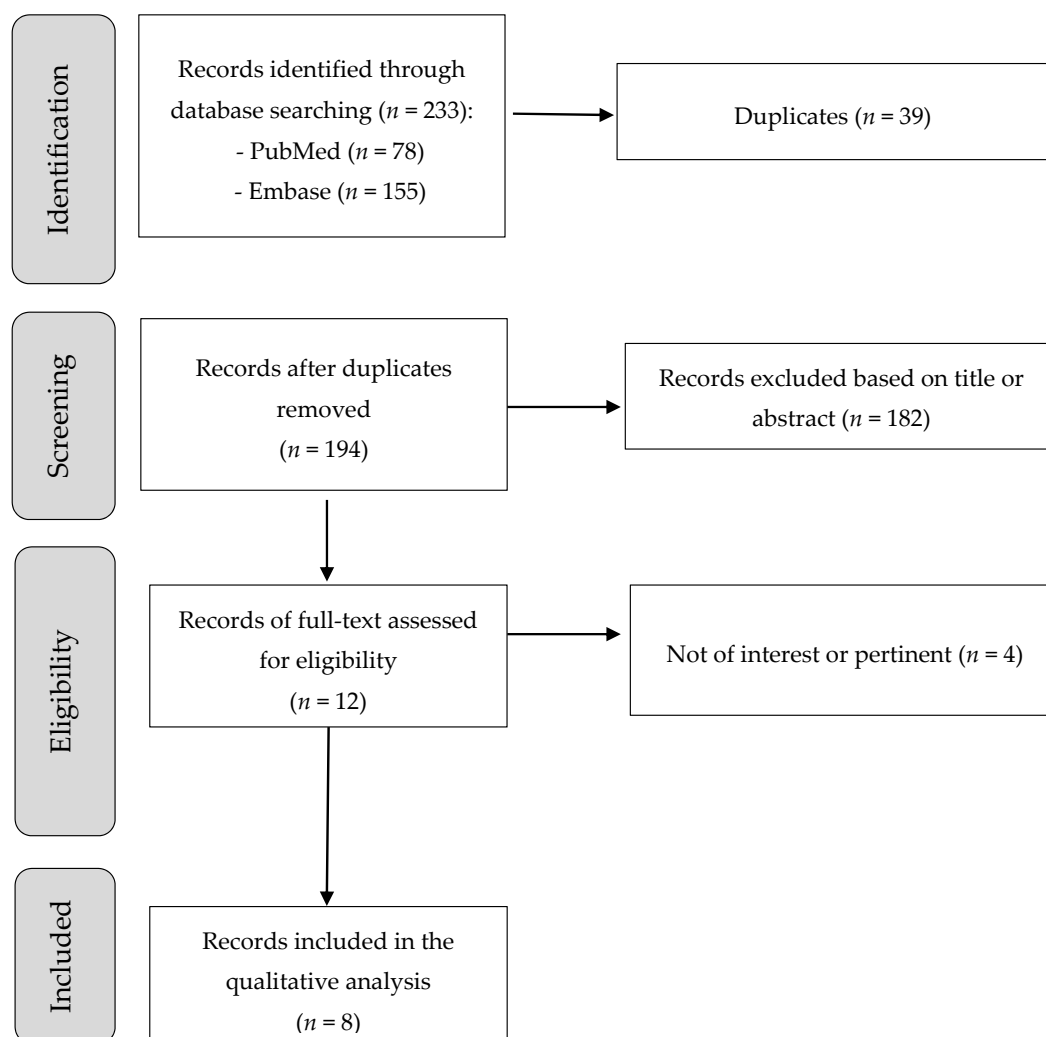
### 2.1. Search Strategy and Study Selection

PUBMED (<http://www.ncbi.nlm.nih.gov/pubmed>) and EMBASE (<http://www.embase.com/>) databases (updated December 2018) were searched to identify pertinent articles. The searches used the combination of the following terms: (Diet, Mediterranean OR Mediterranean diet OR diets, Mediterranean OR Mediterranean diets) AND (DNA damage OR DNA damages OR damage, DNA OR damages, DNA OR DNA injury OR DNA injuries OR injuries, DNA OR injury, DNA OR genotoxic stress OR genotoxic stresses OR stresses, genotoxic OR stress, genotoxic OR telomeric DNA damage OR telomere length). The retrieved papers were also screened for additional papers. The search strategy is summarized in Figure 1.

The search was limited to human intervention studies. No other specific restrictions for the selection of the studies have been used with the exception of the language. Only papers written in English have been considered. Two independent reviewers (MM and DM) conducted the literature search in the scientific databases and assessed and verified the eligibility of the studies based on the title and abstract. Disagreement between reviewers was resolved through consultation with a third independent reviewer (CDB) to reach a consensus.

Data extraction of the papers meeting the inclusion criteria was performed by two reviewers (MM and MT). The following information was reported: (1) first author name and year of publication; (2) the

country where the study was performed; (3) study design; (4) subjects' characteristics; (5) characteristics of the control group; (6) characteristics of the intervention group; (7) list of markers of DNA damage; and (8) main findings.



**Figure 1.** Flowchart of the study selection process.

## 2.2. Risk of Bias in Individual Studies

Risk of bias in individual studies was assessed independently by two review authors (MM and DM) by considering the following components to produce the resulting scores: (1) Selection Bias—Sequence generation and allocation concealment; (2) Performance Bias—Blinding of participants and personnel; (3) Detection Bias—Blinding of outcome assessment; (4) Attrition Bias—Incomplete outcome data; (5) Reporting Bias—Selective reporting; and (6) Other Bias. Scores were assessed considering three potential bias risks: “Low risk of bias”, when the study presented the considered characteristics; “High risk of bias”, when the study did not fully highlight the considered characteristics; and “Unclear risk of bias”, when it was not possible to attribute one of the two other scores (“Low risk” or “High risk”) due to missing information. All disagreements were resolved by consensus with a third reviewer (CDB).

### 3. Results and Discussion

#### 3.1. Study Selection

A total of 233 records were identified from PubMed and EMBASE database search. After removing 39 duplicate articles, 194 studies were screened and 182 excluded based on title or abstract. The full text of eligible studies ( $n = 12$ ) was read; a total of four records were excluded because not of interest or pertinent. At the end of the selection process a total of 8 human intervention studies were included in the review [38–45].

#### 3.2. Study Characteristics

The main characteristics of the studies included in the review are reported in Table 1. Six out of eight studies were conducted in Spain [39–42,44,45], one in New Zealand [38], and one in Chile [43]. The study population included males and females of different age (young, adults and older subjects), and with different characteristics: healthy subjects [39,41], individuals with high cardiovascular risk [44,45], metabolic syndrome [40], and prostate cancer [38]. All the studies showed a randomized controlled design with the exception of one non-randomized and non-controlled study [38]. The duration of the intervention varied from a minimum of 4 weeks [39,41,43] up to 5 years [44,45]. Most of the investigations did not analyze the effects of MD alone but in combination with olive oil [40,42], nuts [40], wine [43], or MD supplemented with coenzyme Q10 [39,41]. Among the various markers of DNA oxidative damage, 8-hydroxy-2'-deoxyguanosine (8-oxo-dG) was the main evaluated, followed by the expression of DNA repair genes and telomere length. Only one study quantified the levels of strand breaks and oxidatively-induced DNA damage in peripheral blood mononuclear cells (PBMCs) [38].

#### 3.3. Risk of Bias of the Studies

Risks of bias within individual studies are reported in Figures S1 and S2. On the whole, the results show an unclear risk of bias for most of the types of bias. The performance (blinding of participants and personnel) and the selection (i.e., random sequence generation) represent the highest risks of bias.

#### 3.4. Main Findings

The characteristics of the intervention, the type of markers analyzed and the main findings of the studies are reported in Table 2. On the whole, the results obtained show that MD alone or in combination with specific bioactive-rich foods (i.e. olive oil, nuts, red wine) or antioxidant compounds (i.e., coenzyme Q10), may exert a protection against DNA damage and play an important role in the modulation of DNA repair genes (e.g. down-regulation of growth arrest and DNA-damage-inducible alpha, polymerase (DNA directed)). In particular, three studies reported a reduction of the levels of 8-oxo-dG in urine [40], plasma [41], or peripheral blood leukocytes [43] following the intervention with the MD alone or in combination with others foods/dietary components, while one study did not report a significant effect [42]. Conversely, one study showed a significant decrease in the levels of oxidatively-induced, but not endogenous, DNA damage [38]. Regarding DNA repair capacity, two studies showed a significant modulation of the genes involved in this repair pathway activity [39,41], while one study failed to report a significant effect [42]. The effect of MD on telomere length has been investigated within the PREDIMED-NAVARRA study. One paper did not show any effect on telomere length after MD intervention [44], while the other reported a significant increase in telomeres depending on different individual gene variants [45].

**Table 1.** Characteristics of the studies

Reference	Country	Subject Characteristics	Study Design	Control
Erdrich et al. [38]	New Zealand	Subjects = 20 (F = 0; M = 20) with prostate cancer, Age = 52–74 years; BMI = 23–33 kg/m <sup>2</sup> Non-smokers = 7; former-smokers = 13	One arm intervention	N.A.
Gutierrez-Mariscal et al. [39]	Spain	Subjects = 20 (F = 10; M = 10); Age = > 65 years; BMI = 20–40 kg/m <sup>2</sup> Non-smokers = 20	Randomized, controlled crossover trial	Western diet rich in SFA (6 subjects)
Mitjavila et al. [40]	Spain	Subjects = 110 (F = 110; M = 0) with MetS; Age = 55–80 years; BMI = < 35 kg/m <sup>2</sup> Non-smokers	Multicentric, randomized, controlled, parallel clinical trial PREDIMED	Low-fat diet (37 subjects)
Gutierrez-Mariscal et al. [41]	Spain	Subjects = 20 (F = 10; M = 10); Age = > 65 years; BMI = 20–40 kg/m <sup>2</sup> Non-smokers=20	Randomized, controlled crossover trial	Western diet rich in SFA (6 subjects)
Konstantinidou et al. [42]	Spain	Subjects = 90 (F = 64; M = 26); Age = 20–50 years; BMI = < 30 kg/m <sup>2</sup> Smokers = N.A.	Randomized, parallel, controlled clinical trial	Habitual diet (30 subjects)
Urquiaga et al. [43]	Chile	Subjects = 42 (F = 0; M = 42); Age = 20–27 years; BMI = N.A. Smokers = N.A.	Partially randomized, controlled, trial	Occidental diet (21 subjects)
García-Calzon et al. [44]	Spain	Subjects = 520 (F = 286; M = 234) at high CV risk; Age = 55–80 years; BMI = 25–35 kg/m <sup>2</sup> Non-smokers = 326; former-smokers = 117; smokers = 77	Multicentric, randomized, controlled, parallel clinical trial PREDIMED-NAVARRA	Low-fat diet (140 subjects)
García-Calzon et al. [45]	Spain	Subjects = 521 at high CV risk Pro/pro group = 451 (F = 244; M = 207); Ala carrier group = 70 (F = 64; M = 25) Age = 60–80 years F, 55–80 years M; BMI = 25–35 kg/m <sup>2</sup>	Multicentric, randomized, controlled, parallel clinical trial PREDIMED-NAVARRA	Low-fat diet (140 subjects)

Legend: BMI: body mass index; CV: cardiovascular; EVOO: extra virgin olive oil; F: female; M: male; MD: Mediterranean diet; MetS: metabolic syndrome; N.A: not available; PREDIMED: PREvención con DIeta MEDiterránea project; SFA: saturated fatty acid.

**Table 2.** Main findings about the role of Mediterranean diet in the modulation of markers of DNA damage, DNA repair, and telomere length.

Reference	Intervention	Markers of DNA Damage	Main Findings
Erdrich et al. [38]	Adherence to Mediterranean-style diet consisting of: extra virgin olive oil, fresh frozen salmon (200 g/week), unsweetened pure pomegranate juice (1 L/week) and samples of a variety of canned legumes 3 months	Percentage DNA in the tail	↓cell DNA damage ( $p = 0.013$ )
Gutierrez-Mariscal et al. [39]	MD supplemented with Q10 (7 subjects) Only MD (7 subjects) 4 weeks Each diet	Gadd45a Ogg1 APE-1/Ref-1 DNA pol $\beta$ XPC	↓ Gadd45a mRNA levels MD plus Q10 vs. SFA ( $p = 0.044$ ) ↓ nuclear Gadd45a in fasting and at 4 h MD vs. SFA ( $p = 0.023$ and $p = 0.038$ , respectively) ↑ Ogg1 mRNA levels during postprandial period SFA vs. MD plus Q10 ( $p = 0.048$ ) ↑ nuclear APE-1/ Ref-1 protein level during the postprandial period and long-term consumption SFA vs. MD plus Q10 ( $p = 0.038$ and $p = 0.028$ , respectively) ↓ DNApol $\beta$ mRNA levels MD plus Q10 vs. SFA ( $p = 0.041$ ) ↑ nuclear DNApol $\beta$ protein levels SFA vs. MD plus Q10 ( $p = 0.044$ ) ↑ XPC mRNA levels during postprandial period SFA vs. MD plus Q10 ( $p = 0.019$ )
Mitjavila et al. [40]	MD plus EVOO (38 subjects) MD plus nuts (35 subjects) 1 year	8-OH-dG	↓ Urinary 8-OH-dG concentrations MD groups vs. Control ( $p < 0.001$ )
Gutierrez-Mariscal et al. [41]	MD supplemented Q10 (7 subjects) Only MD (7 subjects) 4 weeks	8-OH-dG P53 p-p53 (Ser20) p53R2	↓ 8-OH-dG plasma concentrations MD and MD plus Q10 vs. SFA ( $p < 0.0001$ ) ↓ 8-OH-dG plasma concentrations MD vs. MD plus Q10 ( $p < 0.001$ ) ↓ postprandial levels of cytoplasmic p53 MD plus Q10 vs. SFA and MD ( $p < 0.05$ ) ↓ nuclear p-p53 (Ser20) postprandial levels MD plus Q10 vs. SFA and MD ( $p = 0.0013$ ). ↑ p53 mRNA levels postprandial and after 2 h SFA vs. MD ( $p = 0.047$ ). ↔ mRNA p53R2 MD plus Q10 vs. SFA vs. MD ( $p > 0.05$ )
Konstantinidou et al. [42]	MD plus VOO (30 subjects) MD plus WOO (30 subjects) 3 months	8-OH-dG CCNG1 POLK TP53 DCLRE1C DNA ERCC5 XRCC5	↓ polymerase (DNA directed)- (POLK) MD vs. control group ( $p < 0.05$ ) ↔ 8-OH-dG, CCNG1, TP53, DCLRE1C, ERCC5, XRCC5 MD vs. control group ( $p > 0.05$ )
Urquiaga et al. [43]	MD (21 subjects) 3 months	8-OH-dG	↓ 8-OH-dG in DNA from peripheral blood leukocytes MD group vs. OD group ( $p < 0.008$ )
García-Calzón et al. [44]	MD plus EVOO (210 subjects) MD plus nuts (170 subjects) 5 years	Telomere length	↔ telomere length MD plus EVOO vs. Control ↓ telomere length MD plus nuts vs. Control ( $p < 0.001$ )
García-Calzón et al. [45]	MD plus EVOO (212 subjects) MD plus nuts (169 subjects) 5 years	Telomere length	↑ telomere length Ala carriers group plus MD vs. Pro/pro group plus MD ( $p < 0.01$ )

Legend: APE-1/Ref-1: Reduction-oxidation factor 1-apurinic/aprimidinic endonuclease; CCNG1: Cyclin G1; DCLRE1C: DNA cross-link repair 1C; DNA pol  $\beta$ : DNA polymerase beta; EVOO: extra virgin olive oil; ERCC5: excision repair cross-complementing rodent repair deficiency, complementation group 5; Gadd45a: growth arrest and DNA-damage-inducible alpha; MD: Mediterranean diet; OD: Occidental diet; Ogg1: 8-oxoguanine DNA glycosylase; P53: protein 53; p-p53 (Ser20): pospho-p53 (serine20); p53R2: p53 inducible ribonucleotide reductase gene; POLK: polymerase (DNA directed); Q10: Coenzyme Q10; SFA: saturated fatty acids; TP53: tumor protein p53; VOO: virgin olive oil; WOO: washed virgin olive oil; XRCC5: X-ray repair complementing defective repair in Chinese hamster cells 5 (double-strand-break rejoining; Ku autoantigen, 80 kDa); XPC: xeroderma pigmentosum, complementation group C; 8-OH-dG: 8-hydroxy-2'-deoxyguanosine.

To the best of our knowledge, this is the first review aimed at providing evidence on the effects of MD in the modulation of DNA damage, DNA repair genes, and telomere length. The availability of data on protection from DNA damage is significant, since it has been reported to play a crucial role in the development of degenerative diseases, while the adherence to MD can represent a protective dietary pattern.

It is noteworthy that most of the studies focused their attention on the dietary fat quality as possible determinant of DNA damage and numerous diseases. In particular, it has been suggested that the amount and quality of dietary fats could be important for the maintenance of DNA stability. Some researchers found that a polyunsaturated fatty acid (PUFA)-rich diet was associated with reduced DNA damage [46], while saturated fatty acid (SFA) intake was demonstrated to increase DNA damage [47]. In this regard, a recent study evaluated the association between fat intake, as part of a modified Mediterranean style dietary intervention study, and several markers of inflammation and oxidative stress including DNA damage [48]. The authors found an inverse correlation between DNA damage and monounsaturated fatty acids (MUFAs), particularly oleic acid, while a positive correlation was observed between DNA damage and the intake of dairy products and red meat (possibly due to SFAs) [48].

The role of MUFAs and PUFAs in the modulation of the levels of DNA damage and DNA repair has been poorly investigated *in vivo* and results, in particular for PUFA, are quite controversial. For example, Bishop et al. [48] found a positive correlation between DNA damage and circulating levels of omega-6 PUFA. Similar findings were found in other *in vivo* studies hypothesizing a process of lipid peroxidation as determinant of DNA damage [49–51]. Conversely, numerous other studies have reported a protection of MUFAs and PUFAs against oxidative stress and DNA damage [46, 52, 53]. In the present review, the role of olive oil and nuts as food sources of MUFA and PUFAs respectively, but not limited to them, has been evaluated within a context of Mediterranean diet. Mitjavila and colleagues [40] performed a randomized, controlled, parallel clinical trial in which 110 female subjects with metabolic syndrome, recruited within the PREDIMED study, were randomly assigned to three intervention groups: (1) MD *plus* nuts; (2) MD *plus* extra virgin olive oil; and (3) control diet (advice on low-fat diet). The effects of these three interventions on the levels of DNA damage and other parameters related to cardiovascular health were evaluated after 1-year follow-up. The results obtained showed an overall significant improvement of cardiovascular health outcomes and a reduction in the urinary levels of 8-OH-dG in the MD group as compared to the control group. In another study, Konstantinidou and coworkers [42] investigated the effects of the traditional MD, associated with the consumption of two different virgin olive oils, on the expression of atherosclerosis markers and related genes including DNA damage and DNA repair genes. To this aim, the authors performed a randomized, parallel, controlled clinical trial with three different interventions: (1) MD *plus* virgin olive oil; (2) MD *plus* washed virgin olive oil (low content in polyphenols compared to normal virgin olive oil); (3) control diet consisting in the habitual diet of the participants. The impact of the dietary interventions was evaluated in 90 subjects (30 subjects for each arm of intervention) after 3 months. The results have shown an overall general improvement of the MD intervention on the genes and markers related to atherosclerosis, while the effect on the urinary levels of 8-OH-dG was significant only after the intervention with virgin olive oil but not washed virgin olive oil. Conversely, the effect on the modulation of DNA repair genes was significant only when considering the global effect of both the interventions with the MD and olive oil. The results showed a down-regulation of polymerase (DNA directed)  $\kappa$  (POLK) gene expression, suggesting a protective role of the MD on DNA oxidation and damage. In fact, POLK is a specialized DNA polymerase that catalyzes the translesion DNA synthesis, which allows DNA replication in the presence of lesions. The authors attributed the beneficial effects of the intervention to presence of MUFAs and other components (i.e. polyphenols) provided by olive oil and the MD.

SFAs may promote cell transformation by negatively regulating the DNA damage response pathway [49]. Gutierrez-Mariscal et al. [41] evaluated the impact of a 4-week intervention with MD

and MD *plus* coenzyme Q10 (a fat-soluble bioactive with antioxidant activity), compared to a SFA-rich diet, on different markers of DNA damage and DNA repair in a group of elderly subjects. The authors found a significant decrease in p53 protein levels (a transcription factor which mediates the cellular response to DNA damage), as well as plasma circulating 8-OH-dG levels (as marker of DNA damage), following both the Mediterranean dietary approaches but not with the diet rich in SFAs. The authors also demonstrated, in the same population, that the adherence to MD was able to control the expression of DNA repair genes, while the intervention with the SFA diet triggered the p53-dependent DNA repair machinery, as defense mechanism in response to a stress condition [26]. Similarly, Urquiaga et al. [43] investigated the effect of a 3-month MD intervention versus an Occidental diet (OD, resembling a Western or U.S. diet) richer in fat, in particular SFAs, in a group of young healthy subjects. During the first and the third month, subjects received the prepared diets alone, while during the second month they were asked also to add 240 mL of red wine (corresponding to 2 glasses per day). Subjects on MD, compared to those on OD (higher in SFAs), had reduced levels of 8-OH-dG in peripheral blood leucocytes. However, since both the diets included a moderate consumption of red wine for a period of 30 days, a positive or negative contribution of this product in the results obtained cannot be excluded.

DNA damage has been positively associated with cancer risk. In fact, it has been clearly documented that DNA lesions can alter the primary structure of the double helix thereby affecting transcription and replication [53]. Moreover, erroneous DNA repair of lesions can lead to mutations or chromosomal aberrations affecting oncogenes and tumor suppressor genes; the effect is that cells undergo malignant transformation resulting in cancerous growth with deleterious consequences for individual's health [53]. Several epidemiological studies reported an inverse association between MD adherence and neoplastic diseases [54–56] at the level of different sites such as breast [57], colorectal [58], bladder [59], and prostate cancer [60]. In the present review, we included a dietary intervention study performed on 20 subjects with prostate cancer in which the effect of a modified Mediterranean-style diet, consisting in extra-virgin olive oil, fresh frozen salmon, unsweetened pure pomegranate juice and canned legumes, was evaluated [38]. The intervention was a 3-month non-randomized controlled trial in which the levels of DNA damage in individual cells were evaluated by the comet assay [61,62]. The authors documented that dietary changes towards a modified Mediterranean-style pattern could have beneficial effects in terms of reduction of oxidatively-induced DNA damage in this specific target group [38]. This effect was attributed to an undeterminable synergistic effect of dietary components provided through the MD intervention.

Some observational and ex-vivo studies have shown that a greater adherence to MD was associated not only with an improvement of health status, but also with longer telomeres and higher telomerase activity in the elderly [63–66]. Garcia-Calzón and colleagues [44] found that a higher baseline adherence to the MD was associated with longer telomerases in women, but not in men, from the PREDIMED-NAVARRA study. After the 5-year intervention, the results obtained showed that the MD failed to prevent telomere shortening in this target group of older subjects. However, further data elaboration, by considering some gene variants (i.e., Pro/Ala polymorphisms of the peroxisome proliferator-activated receptor  $\gamma$ 2), showed that subjects with Ala carrier variant had increased telomeres length following MD intervention [45]. Moreover, the research team documented that the adherence to MD intervention improved obesity parameters [67] and dietary inflammatory index [68], likely slowing down telomere shortening.

#### 4. Conclusions

Undoubtedly, the MD represents a balanced and healthy dietary pattern associated with a reduced risk of major chronic diseases. However, the role of the MD in the modulation of DNA damage, as potential contributor in disease onset, is poorly investigated. Apart from PREDIMED, most of the studies did not focus on the role of the MD as such, but in combination with specific bioactives and/or bioactive-rich foods; thus, specific contributions are difficult to discern. Although preliminary, the papers reviewed seem to strengthen the hypothesis of a potential role of MD in the protection of



DNA damage and in the modulation of DNA repair genes and telomere length. However, the studies show several limitations due to the numerous bias and confounding factors apparently present in the experimental designs. For these reasons, the results in the present review can represent a starting point for further well-controlled intervention studies targeted on the specific effects of traditional and/or revised MD pattern in the modulation of DNA damage. These studies will be useful to provide more evidence-based proof of MD protective activity and to reveal the molecular mechanisms at the base of the protection observed.

**Supplementary Materials:** The following are available online at <http://www.mdpi.com/2072-6643/11/2/391/s1>, Figure S1: Risk of bias graph: review authors' judgements about each risk of bias item presented as percentages across all included studies, Figure S2: Risk of bias summary: review authors' judgements about each risk of bias item for each included study.

**Author Contributions:** M.M., D.M. and M.T. made the literature search, reviewed the abstracts of the studies selected, performed the analysis of risk of bias and prepared the tables; C.D.B. acted as a third independent reviewer and wrote the first draft of the manuscript. S.C. improved the quality of the manuscript. P.R. and M.P. critically revised the scientific contents of the manuscript and improved the quality of the manuscript.

**Funding:** This research received no external funding.

**Acknowledgments:** This work was supported by the European Cooperation for Science and Technology (CA COST Action CA15132, The comet assay as a human biomonitoring tool (hCOMET)). The authors are grateful for support granted by Ministero delle Politiche Agricole, Alimentari, Forestali e del Turismo (MIPAAFT) and the work was performed within the framework of the European Joint Programming Initiative "A Healthy Diet for a Healthy Life" (JPI HDHL) MaPLE. We thank Donato Angelino for its help in the management of risk of bias.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Thanan, R.; Oikawa, S.; Hiraku, Y.; Ohnishi, S.; Ma, N.; Pinlaor, S.; Yongvanit, P.; Kawanishi, S.; Murata, M. Oxidative stress and its significant roles in neurodegenerative diseases and cancer. *Int. J. Mol. Sci.* **2014**, *16*, 193–217. [[CrossRef](#)] [[PubMed](#)]
2. Mahat, R.K.; Singh, N.; Gupta, A.; Rathore, V. Oxidative DNA damage and carotid intima media thickness as predictors of cardiovascular disease in pre-diabetic subjects. *J. Cardiovasc. Dev. Dis.* **2018**, *5*, 15. [[CrossRef](#)] [[PubMed](#)]
3. Basu, A.K. DNA damage, mutagenesis and cancer. *Int. J. Mol. Sci.* **2018**, *19*, 970. [[CrossRef](#)] [[PubMed](#)]
4. Blasiak, J.; Arabski, M.; Krupa, R.; Wozniak, K.; Zadrozny, M.; Kasznicki, J.; Zurawska, M.; Drzewoski, J. DNA damage and repair in type 2 diabetes mellitus. *Mutat. Res.* **2004**, *554*, 297–304. [[CrossRef](#)] [[PubMed](#)]
5. Coluzzi, E.; Colamartino, M.; Cozzi, R.; Leone, S.; Meneghini, C.; O'Callaghan, N.; Sgura, A. Oxidative stress induces persistent telomeric DNA damage responsible for nuclear morphology change in mammalian cells. *PLoS ONE* **2014**, *9*, e110963. [[CrossRef](#)] [[PubMed](#)]
6. Prasad, K.N.; Wu, M.; Bondy, S.C. Telomere shortening during aging: Attenuation by antioxidants and anti-inflammatory agents. *Mech. Ageing Dev.* **2017**, *164*, 61–66. [[CrossRef](#)] [[PubMed](#)]
7. Giovannelli, L.; Saieva, C.; Masala, G.; Testa, G.; Salvini, S.; Pitozzi, V.; Riboli, E.; Dolaro, P.; Palli, D. Nutritional and lifestyle determinants of DNA oxidative damage: A study in a Mediterranean population. *Carcinogenesis* **2002**, *23*, 1483–1489. [[CrossRef](#)] [[PubMed](#)]
8. Fenech, M.; Bonassi, S. The effect of age, gender, diet and lifestyle on DNA damage measured using micronucleus frequency in human peripheral blood lymphocytes. *Mutagenesis* **2011**, *26*, 43–49. [[CrossRef](#)] [[PubMed](#)]
9. Prado, R.P.; dos Santos, B.F.; Pinto, C.L.; de Assis, K.R.; Salvadori, D.M.; Ladeira, M.S. Influence of diet on oxidative DNA damage, uracil misincorporation and DNA repair capability. *Mutagenesis* **2010**, *25*, 483–487. [[CrossRef](#)]
10. Freitas-Simoes, T.M.; Ros, E.; Sala-Vila, A. Nutrients, foods, dietary patterns and telomere length: Update of epidemiological studies and randomized trials. *Metabolism* **2016**, *65*, 406–415. [[CrossRef](#)]
11. Riso, P.; Pinder, A.; Santangelo, A.; Porrini, M. Does tomato consumption effectively increase the resistance of lymphocyte DNA to oxidative damage? *Am. J. Clin. Nutr.* **1999**, *69*, 712–718. [[CrossRef](#)] [[PubMed](#)]

12. Chen, L.; Stacewicz-Sapuntzakis, M.; Duncan, C.; Sharifi, R.; Ghosh, L.; van Breemen, R.; Ashton, D.; Bowen, P.E. Oxidative DNA damage in prostate cancer patients consuming tomato sauce-based entrees as a whole-food intervention. *J. Natl. Cancer Inst.* **2001**, *93*, 1872–1879. [[CrossRef](#)]
13. Riso, P.; Martini, D.; Møller, P.; Loft, S.; Bonacina, G.; Moro, M.; Porrini, M. DNA damage and repair activity after broccoli intake in young healthy smokers. *Mutagenesis* **2010**, *25*, 595–602. [[CrossRef](#)] [[PubMed](#)]
14. Riso, P.; Martini, D.; Visioli, F.; Martinetti, A.; Porrini, M. Effect of broccoli intake on markers related to oxidative stress and cancer risk in healthy smokers and nonsmokers. *Nutr. Cancer* **2009**, *61*, 232–237. [[CrossRef](#)] [[PubMed](#)]
15. Porrini, M.; Riso, P.; Oriani, G. Spinach and tomato consumption increases lymphocyte DNA resistance to oxidative stress but this is not related to cell carotenoid concentrations. *Eur. J. Nutr.* **2002**, *41*, 95–100. [[CrossRef](#)] [[PubMed](#)]
16. Moser, B.; Szekeres, T.; Bieglmayer, C.; Wagner, K.H.; Mišák, M.; Kundi, M.; Zakerska, O.; Nersesyan, A.; Kager, N.; Zahrl, J.; et al. Impact of spinach consumption on DNA stability in peripheral lymphocytes and on biochemical blood parameters: Results of a human intervention trial. *Eur. J. Nutr.* **2011**, *50*, 587–589. [[CrossRef](#)] [[PubMed](#)]
17. Riso, P.; Klimis-Zacas, D.; Del Bo', C.; Martini, D.; Campolo, J.; Vendrame, S.; Møller, P.; Loft, S.; De Maria, R.; Porrini, M. Effect of a wild blueberry (*Vaccinium angustifolium*) drink intervention on markers of oxidative stress, inflammation and endothelial function in humans with cardiovascular risk factors. *Eur. J. Nutr.* **2013**, *52*, 949–961. [[CrossRef](#)] [[PubMed](#)]
18. Del Bo', C.; Riso, P.; Campolo, J.; Møller, P.; Loft, S.; Klimis-Zacas, D.; Brambilla, A.; Rizzolo, A.; Porrini, M. A single portion of blueberry (*Vaccinium corymbosum* L) improves protection against DNA damage but not vascular function in healthy male volunteers. *Nutr. Res.* **2013**, *33*, 220–227. [[CrossRef](#)] [[PubMed](#)]
19. Guarnieri, S.; Riso, P.; Porrini, M. Orange juice vs vitamin C: Effect on hydrogen peroxide-induced DNA damage in mononuclear blood cells. *Br. J. Nutr.* **2007**, *97*, 639–643. [[CrossRef](#)]
20. Rangel-Huerta, O.D.; Aguilera, C.M.; Martin, M.V.; Soto, M.J.; Rico, M.C.; Vallejo, F.; Tomas-Barberan, F.; Perez-de-la-Cruz, A.J.; Gil, A.; Mesa, M.D. Normal or high polyphenol concentration in orange juice affects antioxidant activity, blood pressure, and body weight in obese or overweight adults. *J. Nutr.* **2015**, *145*, 1808–1816. [[CrossRef](#)]
21. Guaraldi, F.; Deon, V.; Del Bo', C.; Vendrame, S.; Porrini, M.; Riso, P.; Guardamagna, O. Effect of short-term hazelnut consumption on DNA damage and oxidized LDL in children and adolescents with primary hyperlipidemia: A randomized controlled trial. *J. Nutr. Biochem.* **2018**, *57*, 206–211. [[CrossRef](#)] [[PubMed](#)]
22. López-Uriarte, P.; Nogués, R.; Saez, G.; Bulló, M.; Romeu, M.; Masana, L.; Tormos, C.; Casas-Agustench, P.; Salas-Salvadó, J. Effect of nut consumption on oxidative stress and the endothelial function in metabolic syndrome. *Clin. Nutr.* **2010**, *29*, 373–380. [[CrossRef](#)]
23. Erba, D.; Riso, P.; Bordoni, A.; Foti, P.; Biagi, P.L.; Testolin, G. Effectiveness of moderate green tea consumption on antioxidative status and plasma lipid profile in humans. *J. Nutr. Biochem.* **2005**, *16*, 144–149. [[CrossRef](#)] [[PubMed](#)]
24. Ho, C.K.; Choi, S.W.; Siu, P.M.; Benzie, I.F. Effects of single dose and regular intake of green tea (*Camellia sinensis*) on DNA damage, DNA repair, and heme oxygenase-1 expression in a randomized controlled human supplementation study. *Mol. Nutr. Food Res.* **2014**, *58*, 1379–1383. [[CrossRef](#)] [[PubMed](#)]
25. Bakuradze, T.; Lang, R.; Hofmann, T.; Schipp, D.; Galan, J.; Eisenbrand, G.; Richling, E. Coffee consumption rapidly reduces background DNA strand breaks in healthy humans: Results of a short-term repeated uptake intervention study. *Mol. Nutr. Food Res.* **2016**, *60*, 682–686. [[CrossRef](#)] [[PubMed](#)]
26. Bakuradze, T.; Lang, R.; Hofmann, T.; Eisenbrand, G.; Schipp, D.; Galan, J.; Richling, E. Consumption of a dark roast coffee decreases the level of spontaneous DNA strand breaks: A randomized controlled trial. *Eur. J. Nutr.* **2015**, *54*, 149–156. [[CrossRef](#)] [[PubMed](#)]
27. Kiokias, S.; Proestos, C.; Oreopoulou, V. Effect of natural food antioxidants against LDL and DNA oxidative changes. *Antioxidants* **2018**, *7*, 133. [[CrossRef](#)] [[PubMed](#)]
28. Azqueta, A.; Collins, A. Polyphenols and DNA damage: A mixed blessing. *Nutrients* **2016**, *8*, 785. [[CrossRef](#)] [[PubMed](#)]
29. Willett, W.C.; Sacks, F.; Trichopoulou, A.; Drescher, G.; Ferro-Luzzi, A.; Helsing, E.; Trichopoulos, D. Mediterranean diet pyramid: A cultural model for healthy eating. *Am. J. Clin. Nutr.* **1995**, *61* (Suppl. 6), S1402–S1406. [[CrossRef](#)] [[PubMed](#)]

30. Bach-Faig, A.; Berry, E.M.; Lairon, D.; Reguant, J.; Trichopoulou, A.; Dernini, S.; Medina, F.X.; Battino, M.; Belahsen, R.; Miranda, G.; et al. Mediterranean Diet Foundation Expert Group. Mediterranean diet pyramid today. Science and cultural updates. *Public Health Nutr.* **2011**, *14*, 2274–2284. [[CrossRef](#)] [[PubMed](#)]
31. Koloverou, E.; Panagiotakos, D.B.; Pitsavos, C.; Chrysohoou, C.; Georgousopoulou, E.N.; Grekas, A.; Christou, A.; Chatzigeorgiou, M.; Skoumas, I.; Tousoulis, D.; et al. ATTICA Study Group. Adherence to Mediterranean diet and 10-year incidence (2002–2012) of diabetes: Correlations with inflammatory and oxidative stress biomarkers in the ATTICA cohort study. *Diabetes Metab. Res. Rev.* **2016**, *32*, 73–81. [[CrossRef](#)] [[PubMed](#)]
32. Dinu, M.; Pagliai, G.; Casini, A.; Sofi, F. Mediterranean diet and multiple health outcomes: An umbrella review of meta-analyses of observational studies and randomised trials. *Eur. J. Clin. Nutr.* **2018**, *72*, 30–43. [[CrossRef](#)] [[PubMed](#)]
33. Bonaccio, M.; Di Castelnuovo, A.; Costanzo, S.; Gialluisi, A.; Persichillo, M.; Cerletti, C.; Donati, M.B.; de Gaetano, G.; Iacoviello, L. Mediterranean diet and mortality in the elderly: A prospective cohort study and a meta-analysis. *Br. J. Nutr.* **2018**, *120*, 841–854. [[CrossRef](#)] [[PubMed](#)]
34. Buil-Cosiales, P.; Toledo, E.; Salas-Salvadó, J.; Zazpe, I.; Farràs, M.; Basterra-Gortari, F.J.; Diez-Espino, J.; Estruch, R.; Corella, D.; Ros, E.; et al. Association between dietary fibre intake and fruit, vegetable or whole-grain consumption and the risk of CVD: Results from the PREvención con Dieta MEDiterránea (PREDIMED) trial. *Br. J. Nutr.* **2016**, *116*, 534–546. [[CrossRef](#)] [[PubMed](#)]
35. Garcia-Arellano, A.; Ramallal, R.; Ruiz-Canela, M.; Salas-Salvadó, J.; Corella, D.; Shivappa, N.; Schröder, H.; Hébert, J.R.; Ros, E.; Gómez-García, E.; et al. Dietary Inflammatory Index and incidence of cardiovascular disease in the PREDIMED Study. *Nutrients* **2015**, *7*, 4124–4138. [[CrossRef](#)]
36. Martínez-González, M.A.; Salas-Salvadó, J.; Estruch, R.; Corella, D.; Fitó, M.; Ros, E.; PREDIMED Investigators. Benefits of the Mediterranean Diet: Insights from the PREDIMED Study. *Prog. Cardiovasc. Dis.* **2015**, *58*, 50–60.
37. Guasch-Ferré, M.; Salas-Salvadó, J.; Ros, E.; Estruch, R.; Corella, D.; Fitó, M.; Martínez-González, M.A.; PREDIMED Investigators. The PREDIMED trial, Mediterranean diet and health outcomes: How strong is the evidence? *Nutr. Metab. Cardiovasc. Dis.* **2017**, *27*, 624–632.
38. Erdrich, S.; Bishop, K.S.; Karunasinghe, N.; Han, D.Y.; Ferguson, L.R. A pilot study to investigate if New Zealand men with prostate cancer benefit from a Mediterranean-style diet. *Peer J.* **2015**, *3*, e1080. [[CrossRef](#)]
39. Gutierrez-Mariscal, F.M.; Yubero-Serrano, E.M.; Rangel-Zúñiga, O.A.; Marín, C.; García-Rios, A.; Perez-Martinez, P.; Delgado-Lista, J.; Malagón, M.M.; Tinahones, F.J.; Pérez-Jimenez, F.; et al. Postprandial activation of p53-dependent DNA repair is modified by Mediterranean diet supplemented with coenzyme Q10 in elderly subjects. *J. Gerontol. A Biol. Sci. Med. Sci.* **2014**, *69*, 886–893. [[CrossRef](#)] [[PubMed](#)]
40. Mitjavila, M.T.; Fandos, M.; Salas-Salvadó, J.; Covas, M.I.; Borrego, S.; Estruch, R.; Lamuela-Raventós, R.; Corella, D.; Martínez-González, M.Á.; Sánchez, J.M.; et al. The Mediterranean diet improves the systemic lipid and DNA oxidative damage in metabolic syndrome individuals. A randomized, controlled, trial. *Clin. Nutr.* **2013**, *32*, 172–178. [[CrossRef](#)]
41. Gutierrez-Mariscal, F.M.; Perez-Martinez, P.; Delgado-Lista, J.; Yubero-Serrano, E.M.; Camargo, A.; Delgado-Casado, N.; Cruz-Teno, C.; Santos-Gonzalez, M.; Rodriguez-Cantalejo, F.; Castaño, J.P.; et al. Mediterranean diet supplemented with coenzyme Q10 induces postprandial changes in p53 in response to oxidative DNA damage in elderly subjects. *Age* **2012**, *34*, 389–403. [[CrossRef](#)] [[PubMed](#)]
42. Konstantinidou, V.; Covas, M.I.; Muñoz-Aguayo, D.; Khymenets, O.; de la Torre, R.; Saez, G.; del Carmen Tormos, M.; Toledo, E.; Marti, A.; Ruiz-Gutiérrez, V.; et al. In vivo nutrigenomic effects of virgin olive oil polyphenols within the frame of the Mediterranean diet: A randomized controlled trial. *FASEB J.* **2010**, *24*, 2546–2557. [[CrossRef](#)] [[PubMed](#)]
43. Urquiaga, I.; Strobel, P.; Perez, D.; Martinez, C.; Cuevas, A.; Castillo, O.; Marshall, G.; Rozowski, J.; Leighton, F. Mediterranean diet and red wine protect against oxidative damage in young volunteers. *Atherosclerosis* **2010**, *211*, 694–699. [[CrossRef](#)] [[PubMed](#)]
44. García-Calzón, S.; Martínez-González, M.A.; Razquin, C.; Arós, F.; Lapetra, J.; Martínez, J.A.; Zalba, G.; Marti, A. Mediterranean diet and telomere length in high cardiovascular risk subjects from the PREDIMED-NAVARRA study. *Clin. Nutr.* **2016**, *35*, 1399–1405. [[CrossRef](#)] [[PubMed](#)]
45. García-Calzón, S.; Martínez-González, M.A.; Razquin, C.; Corella, D.; Salas-Salvadó, J.; Martínez, J.A.; Zalba, G.; Marti, A. Pro12Ala polymorphism of the PPAR $\gamma$ 2 gene interacts with a mediterranean diet to

- prevent telomere shortening in the PREDIMED-NAVARRA randomized trial. *Circ. Cardiovasc. Genet.* **2015**, *8*, 91–99. [[CrossRef](#)]
46. Müllner, E.; Brath, H.; Pleifer, S.; Schiermayr, C.; Baierl, A.; Wallner, M.; Fastian, T.; Millner, Y.; Paller, K.; Henriksen, T.; et al. Vegetables and PUFA-rich plant oil reduce DNA strand breaks in individuals with type 2 diabetes. *Mol. Nutr Food Res.* **2013**, *57*, 328–338. [[CrossRef](#)] [[PubMed](#)]
47. Zeng, L.; Wu, G.Z.; Goh, K.J.; Lee, Y.M.; Ng, C.C.; You, A.B.; Wang, J.; Jia, D.; Hao, A.; Yu, Q.; et al. Saturated fatty acids modulate cell response to DNA damage: Implication for their role in tumorigenesis. *PLoS ONE* **2008**, *3*, e2329. [[CrossRef](#)] [[PubMed](#)]
48. Bishop, K.S.; Erdrich, S.; Karunasinghe, N.; Han, D.Y.; Zhu, S.; Jesuthasan, A.; Ferguson, L.R. An investigation into the association between DNA damage and dietary fatty acid in men with prostate cancer. *Nutrients* **2015**, *7*, 405–422. [[CrossRef](#)]
49. Jenkinson, A.; Franklin, M.F.; Wahle, K.; Duthie, G.G. Dietary intakes of polyunsaturated fatty acids and indices of oxidative stress in human volunteers. *Eur. J. Clin. Nutr.* **1999**, *53*, 523–528. [[CrossRef](#)]
50. Kimura, Y.; Sato, M.; Kurotani, K.; Nanri, A.; Kawai, K.; Kasai, H.; Imaizumi, K.; Mizoue, T. PUFAs in serum cholesterol ester and oxidative DNA damage in Japanese men and women. *Am. J. Clin. Nutr.* **2012**, *95*, 1209–1214. [[CrossRef](#)]
51. Sakai, C.; Ishida, M.; Ohba, H.; Yamashita, H.; Uchida, H.; Yoshizumi, M.; Ishida, T. Fish oil omega-3 polyunsaturated fatty acids attenuate oxidative stress-induced DNA damage in vascular endothelial cells. *PLoS ONE* **2017**, *12*, e0187934. [[CrossRef](#)] [[PubMed](#)]
52. Kikugawa, K.; Yasuhara, Y.; Ando, K.; Koyama, K.; Hiramoto, K.; Suzuki, M. Protective effect of supplementation of fish oil with high n-3 polyunsaturated fatty acids against oxidative stress-induced DNA damage of rat liver in vivo. *J. Agric. Food Chem.* **2003**, *51*, 6073–6079. [[CrossRef](#)] [[PubMed](#)]
53. Dizdaroglu, M. Oxidatively induced DNA damage and its repair in cancer. *Mutat. Res. Rev. Mutat. Res.* **2015**, *763*, 212–245. [[CrossRef](#)] [[PubMed](#)]
54. Grosso, G.; Buscemi, S.; Galvano, F.; Mistretta, A.; Marventano, S.; La Vela, V.; Drago, F.; Gangi, S.; Basile, F.; Biondi, A. Mediterranean diet and cancer: Epidemiological evidence and mechanism of selected aspects. *BMC Surg.* **2013**, *13* (Suppl. 2), S14. [[CrossRef](#)] [[PubMed](#)]
55. Schwingshackl, L.; Schwedhelm, C.; Galbete, C.; Hoffmann, G. Adherence to Mediterranean diet and risk of cancer: An updated systematic review and meta-analysis. *Nutrients* **2017**, *9*, 1063. [[CrossRef](#)] [[PubMed](#)]
56. Giacosa, A.; Barale, R.; Bavaresco, L.; Gatenby, P.; Gerbi, V.; Janssens, J.; Johnston, B.; Kas, K.; La Vecchia, C.; Mainguet, P.; et al. Cancer prevention in Europe: The Mediterranean diet as a protective choice. *Eur. J. Cancer Prev.* **2013**, *22*, 90–95. [[CrossRef](#)] [[PubMed](#)]
57. Turati, F.; Carioli, G.; Bravi, F.; Ferraroni, M.; Serraino, D.; Montella, M.; Giacosa, A.; Toffolutti, F.; Negri, E.; Levi, F.; et al. Mediterranean diet and breast cancer risk. *Nutrients* **2018**, *10*, 326. [[CrossRef](#)]
58. Grosso, G.; Biondi, A.; Galvano, F.; Mistretta, A.; Marventano, S.; Buscemi, S.; Drago, F.; Basile, F. Factors associated with colorectal cancer in the context of the Mediterranean diet: A case-control study. *Nutr. Cancer* **2014**, *66*, 558–565. [[CrossRef](#)]
59. Bravi, F.; Spei, M.E.; Polesel, J.; Di Maso, M.; Montella, M.; Ferraroni, M.; Serraino, D.; Libra, M.; Negri, E.; La Vecchia, C.; et al. Mediterranean diet and bladder cancer risk in Italy. *Nutrients* **2018**, *10*, 1061. [[CrossRef](#)]
60. Russo, G.I.; Solinas, T.; Urzi, D.; Privitera, S.; Campisi, D.; Cocci, A.; Carini, M.; Madonia, M.; Cimino, S.; Morgia, G. Adherence to Mediterranean diet and prostate cancer risk in Sicily: Population-based case-control study. *Int. J. Impot. Res.* **2018**. [[CrossRef](#)]
61. Collins, A.R. Measuring oxidative damage to DNA and its repair with the comet assay. *Biochim. Biophys. Acta* **2014**, *1840*, 794–800. [[CrossRef](#)] [[PubMed](#)]
62. Møller, P. The comet assay: Ready for 30 more years. *Mutagenesis* **2018**, *33*, 1–7. [[CrossRef](#)] [[PubMed](#)]
63. Boccardi, V.; Esposito, A.; Rizzo, M.R.; Marfella, R.; Barbieri, M.; Paolisso, G. Mediterranean diet, telomere maintenance and health status among elderly. *PLoS ONE* **2013**, *8*, e62781. [[CrossRef](#)] [[PubMed](#)]
64. Crous-Bou, M.; Fung, T.T.; Prescott, J.; Julin, B.; Du, M.; Sun, Q.; Rexrode, K.M.; Hu, F.B.; De Vivo, I. Mediterranean diet and telomere length in nurses' health study: Population based cohort study. *BMJ* **2014**, *349*, g6674. [[CrossRef](#)] [[PubMed](#)]
65. Gu, Y.; Honig, L.S.; Schupf, N.; Lee, J.H.; Luchsinger, J.A.; Stern, Y.; Scarmeas, N. Mediterranean diet and leukocyte telomere length in a multi-ethnic elderly population. *Age* **2015**, *37*, 24. [[CrossRef](#)] [[PubMed](#)]

66. Marin, C.; Delgado-Lista, J.; Ramirez, R.; Carracedo, J.; Caballero, J.; Perez-Martinez, P.; Gutierrez-Mariscal, F.M.; Garcia-Rios, A.; Delgado-Casado, N.; Cruz-Teno, C.; et al. Mediterranean diet reduces senescence-associated stress in endothelial cells. *Age* **2012**, *34*, 1309–1316. [[CrossRef](#)] [[PubMed](#)]
67. García-Calzón, S.; Gea, A.; Razquin, C.; Corella, D.; Lamuela-Raventós, R.M.; Martínez, J.A.; Martínez-González, M.A.; Zalba, G.; Marti, A. Longitudinal association of telomere length and obesity indices in an intervention study with a Mediterranean diet: The PREDIMED-NAVARRA trial. *Int. J. Obes.* **2014**, *38*, 177–182. [[CrossRef](#)] [[PubMed](#)]
68. García-Calzón, S.; Zalba, G.; Ruiz-Canela, M.; Shivappa, N.; Hébert, J.R.; Martínez, J.A.; Fitó, M.; Gómez-Gracia, E.; Martínez-González, M.A.; Marti, A. Dietary inflammatory index and telomere length in subjects with a high cardiovascular disease risk from the PREDIMED-NAVARRA study: Cross-sectional and longitudinal analyses over 5 y. *Am. J. Clin. Nutr.* **2015**, *102*, 897–904. [[CrossRef](#)] [[PubMed](#)]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).