

## BIOCHEMISTRY

# Zooming in on protons: Neutron structure of protein kinase A trapped in a product complex

Oksana Gerlits<sup>1</sup>, Kevin L. Weiss<sup>2</sup>, Matthew P. Blakeley<sup>3</sup>, Gianluigi Veglia<sup>4,5</sup>, Susan S. Taylor<sup>6,7\*</sup>, Andrey Kovalevsky<sup>2</sup>

The question vis-à-vis the chemistry of phosphoryl group transfer catalyzed by protein kinases remains a major challenge. The neutron diffraction structure of the catalytic subunit of cAMP-dependent protein kinase (PKA-C) provides a more complete chemical portrait of key proton interactions at the active site. By using a high-affinity protein kinase substrate (PKS) peptide, we captured the reaction products, dephosphorylated nucleotide [adenosine diphosphate (ADP)] and phosphorylated PKS (pPKS), bound at the active site. In the complex, the phosphoryl group of the peptide is protonated, whereas the carboxyl group of the catalytic Asp<sup>166</sup> is not. Our structure, including conserved waters, shows how the peptide links the distal parts of the cleft together, creating a network that engages the entire molecule. By comparing slow-exchanging backbone amides to those determined by the NMR analysis of PKA-C with ADP and inhibitor peptide (PKI), we identified exchangeable amides that likely distinguish catalytic and inhibited states.

## INTRODUCTION

The protein kinase superfamily regulates much of biology in eukaryotic cells, and because dysfunctional protein kinases drive many diseases that include cancer, diabetes, and neuropathies, they have become major therapeutic targets. The first protein kinase structure to be solved was the catalytic subunit of 3',5'-cyclic adenosine monophosphate (cAMP)-dependent protein kinase (PKA-C) in 1991 (1, 2). The structure of the PKA-C bound to a pseudosubstrate inhibitor peptide provided a new conceptual understanding of the eukaryotic protein kinase (EPK) superfamily, and PKA has continued to serve as a template for all protein kinases, although many kinase structures have now been solved. The PKA-C subunit structure not only defined the conserved fold that was predicted to be shared by all EPKs but also showed the precise alignment of the catalytic residues in the active-site cleft and gave functional importance to the conserved motifs that had been defined by Hanks and Hunter (3). The first PKA structure also revealed the conformation of a fully active EPK where the activation loop was phosphorylated; it visualized the important structural consequences of the single phosphate. We now know that the assembly of this active conformation is highly regulated and dynamic, and often facilitated by activation loop phosphorylation (4, 5). The precise cis and trans mechanism for regulation is unique for each kinase, but the conserved driving force is the assembly of a conserved hydrophobic regulatory spine (R-spine) (6), which brings together three essential conserved residues at the active-site cleft that position the  $\gamma$ -phosphate for transfer to a protein substrate. The hydrophobic spine architecture was only appreciated much later after many different

protein kinase structures were solved and compared (7, 8). The second PKA structure published in 1993 showed precisely how adenosine triphosphate (ATP) and two magnesium ions are poised for catalysis at the active-site cleft (9). Subsequent structures of the C subunit have trapped all stages of the catalytic cycle in a crystal lattice (10–14).

Understanding, however, how a kinase functions as a catalyst to transfer the  $\gamma$ -phosphate from ATP to a protein substrate is still a major chemical challenge. Specifically, protonation states of PKA-C residues, including the key catalytic residues, and substrate or product functional groups remain uncertain. An important next step for achieving a full chemical understanding of this process is neutron diffraction. Neutron diffraction allows clear and unambiguous determination of the hydrogen atom positions and visualization of hydrogen bonds at room temperature without causing radiation damage. This information not only provides experimental validation of hydrogen-bonding interactions inferred from x-ray structures but also adds over x-ray data in cases where positions of hydrogen atoms were predicted inaccurately or were not known. Furthermore, it describes the organization of water molecules by accurately determining their orientations. Here, we present previously unidentified atomic details of phosphoryl transfer by PKA-C. We determined a room temperature neutron structure of the PKA-C subunit trapped in a product complex that contains adenosine diphosphate (ADP), a phosphorylated protein kinase substrate (pPKS) peptide, and two metal ions. We observed a protonated phosphate group on the product substrate and deprotonated catalytic Asp<sup>166</sup>, whose protonation states were previously unknown. We also experimentally characterized which short distances correspond to hydrogen-bonding interactions, providing a clear view of the PKA-C internal hydrogen-bonding networks, the enzyme association with the phosphoryl transfer products (pPKS and ADP), and the hydration structure. Combined with the earlier x-ray structures, a more complete chemical portrait of this prototypical kinase emerges, including the critical importance of several conserved and structured water molecules. This information will be essential for the subsequent detailed mechanistic quantum chemical studies. In addition, we identified the slow-exchanging backbone amides through the hydrogen/deuterium (H/D) exchange analysis

Copyright © 2019  
The Authors, some  
rights reserved;  
exclusive licensee  
American Association  
for the Advancement  
of Science. No claim to  
original U.S. Government  
Works. Distributed  
under a Creative  
Commons Attribution  
NonCommercial  
License 4.0 (CC BY-NC).

<sup>1</sup>Bredesen Center, University of Tennessee, Knoxville, TN 37996, USA. <sup>2</sup>Neutron Scattering Division, Oak Ridge National Laboratory, Oak Ridge, TN 37831, USA. <sup>3</sup>Large Scale Structures Group, Institut Laue-Langevin, Grenoble Cedex 9, 38042, France. <sup>4</sup>Department of Chemistry, University of Minnesota, Minneapolis, MN 55455, USA. <sup>5</sup>Department of Biochemistry, Molecular Biology, and Biophysics, University of Minnesota, Minneapolis, MN 55455, USA. <sup>6</sup>Department of Pharmacology, University of California at San Diego, La Jolla, CA 92093, USA. <sup>7</sup>Department of Chemistry and Biochemistry, University of California at San Diego, La Jolla, CA 92093, USA.

\*Corresponding author. Email: staylor@ucsd.edu

afforded by the neutron structure and compared them to the slow-exchanging backbone amides identified previously by nuclear magnetic resonance (NMR).

## RESULTS

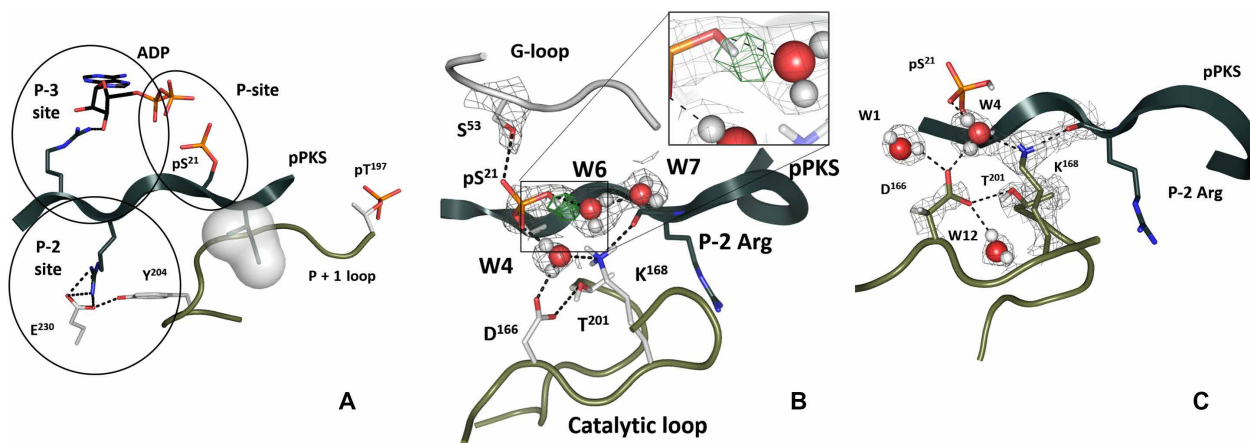
To explore the role that hydrogen atoms play in weaving together the domains of the C subunit and how both products are captured in a complex at the active site, we will begin first with the peptide, specifically with the P-site, where the transfer of the phosphate has occurred. It is important to emphasize that this structure represents a complex with a high-affinity peptide that was generated by creating a substrate version of the protein kinase inhibitor (PKI) peptide (residues 4 to 25). Unlike small peptides, which are immediately released following phosphorylation, the amphipathic helix of PKS allows the peptide to remain tethered to the C subunit following transfer of the phosphate. The high-affinity binding of this unusual substrate version of the inhibitor peptide has allowed us to trap almost every step of the catalytic cycle (10–14). Here, we show the immediate consequences of the phosphoryl transfer between ATP and PKS catalyzed by PKA-C in the solution and captured in the crystalline state in a form of the stable product complex, PKA-C:Sr<sub>2</sub>ADP:pPKS. Figure 1A and fig. S1 show how the peptide unites all parts of the active site, not only nucleating the site of phosphoryl transfer but also weaving together the distal regions of the enzyme that extend across the N- and C-lobes and also include the C-tail helix and the phosphorylated activation loop. In summarizing the active site, we focus first on the P-site, then on the ribose of ADP and the P-3 site where both substrates converge, on the P-2 site in the center of the C-lobe, and finally on the P + 1 site that is created by the activation loop and anchored to the  $\alpha$ C helix. We then show how the N- and C-lobes are synergistically linked by these interactions that are mediated by the polyvalent peptide, which serves as a Velcro strip, locking the protein into a nearly closed conformation even after transfer of the phosphate. We will also comment on some of the water molecules and hydrogen-bonding interactions that were not previously

recognized as important. Our neutron structure reveals and validates previously unknown predicted protonation states of the residues contributing to numerous extended hydrogen-bonding networks seen in the PKA-C complex. Moreover, we determined correct rotamers for hydroxyl groups in Ser, Thr, and Tyr; ammonium groups in Lys; imidazole rings in His; and amide side chains in Asn and Gln.

### Phosphoryl transfer (Fig. 1)

This structure shows the immediate consequences of the phosphoryl transfer from ATP to the substrate analog, PKS, before the products are released. The nanomolar binding affinity of PKS, whose sequence is derived from the physiological protein inhibitor, allows us to trap the products of the reaction: (i) phosphorylated at Ser<sup>21</sup> (pS<sup>21</sup>) substrate, pPKS, and (ii) ADP including two metal ions before dissociation. The neutron structure of the ternary complex unambiguously reveals protonation of one of the phosphoryl group oxygen atoms on the peptide product (Fig. 1B). In this state, the pS<sup>21</sup> phosphate group directly interacts with only one metal ion, M1 (fig. S2A), which is consistent with its negative charge being reduced by protonation, presumably allowing the phosphorylated substrate to dissociate and initiate the release of ADP and the remaining metal. This result agrees well with the previous findings of ADP dissociation being a rate-limiting step at high metal concentrations [reviewed in (13) and references therein]. The pS<sup>21</sup> phosphate group forms five hydrogen-bonding interactions (fig. S2B): three with water molecules (W4, W6, and W9), one with its own main-chain amide, and one with the OH group of Ser<sup>53</sup> from the glycine-rich loop. Two of the water molecules, W4 and W6, are coordinated to M2. W4 also mediates interactions of a phosphate oxygen atom with Asp<sup>166</sup> and Lys<sup>168</sup>, and W6 connects the protonated phosphate's oxygen to the bulk solvent (Fig. 1C and fig. S2). The third water molecule, W9, participates in bridging (through W10) the phosphate groups of pS<sup>21</sup> and ADP.

These observations provide strong evidence that rotation of the pS<sup>21</sup> phosphoryl group away from the active site toward the bulk solvent (fig. S2C) is induced by its protonation. This is consistent with the suggestion that the pS<sup>21</sup> phosphoryl group rotation could



**Fig. 1. Interweaving interactions of pPKS in the vicinity of the PKA-C active site.** (A) Specific sites of the substrate's direct interactions with PKA-C. The space-filled residue signifies the peptide's hydrophobic P + 1 residue, Ile<sup>22</sup>. Here and in all the panels, pPKS is shown as a dark gray ribbon, ADP is colored with black carbon atoms, and the remaining colors are for the enzyme residues. This coloring scheme is kept for all figures throughout the manuscript. (B) P-site showing position of the D atom on pS<sup>21</sup> and interactions with water molecules. The omit  $F_0 - F_C$  difference nuclear density map for the deuterium is shown as green mesh contoured at 3.5 $\sigma$  level. The  $2F_0 - F_C$  nuclear density map contoured at 2.0 $\sigma$  level is shown in gray mesh. (C) The  $2F_0 - F_C$  nuclear density map (gray mesh contoured at 2.0 $\sigma$  level) shows deprotonated Asp<sup>166</sup>, protonated Lys<sup>168</sup>, and the extent of the hydrogen-bonding network.

serve as a trigger for the product peptide's release (14). The protonation and rotation also perturb interactions with the active site and coordination to the metals. Here, to improve neutron diffraction, we used strontium ( $\text{Sr}^{2+}$ ) as the metal instead of physiological  $\text{Mg}^{2+}$ ; however, we can still appreciate how the coordination of the two metal ions changes because of the phosphoryl transfer. Each metal center is surrounded by the same number of ligands; however, M1 has fewer interactions with the active-site residues relative to M2 (fig. S2A). M1 is coordinated only to one active-site residue, Asp<sup>184</sup>, whereas M2 binds with two, Asn<sup>171</sup> and Asp<sup>184</sup>. This observation agrees with the previous suggestion that M1 likely departs before M2 (8, 12), either following the release of the phosphorylated peptide or accompanying it, leaving the active site poised for release of M2 bound to ADP in the last step. This rate-limiting step requires substantial energy to free ADP from its interactions with the enzyme mediated by interactions of the ribose ring that nucleates the hinge and the P-3 Arg (13).

### Nucleotide binding (Fig. 2)

We can consider the ATP binding site as having three distinct regions: (i) the adenine ring, (ii) the phosphates, and (iii) the ribose ring where alteration of the geometry of the M1 metal coordination paves the way for release of the ADP:M2 complex. The P-3 arginine also contributes to this site, as does the C-terminal tail, which is recruited to the ribose ring. The adenine ring is sandwiched between the two parts of the catalytic spine (C-spine): Ala<sup>70</sup> and Val<sup>57</sup> from the N-lobe and Leu<sup>173</sup> from the C-lobe (fig. S1, A and C). Anchoring of the adenine ring synergizes the hydrophobic core of the entire molecule and poises the molecule for transfer of the phosphate (15). The single hydrogen bond to the adenine ring is the Glu<sup>121</sup> backbone interaction with the N6 proton, a contact that is conserved in all kinases (Fig. 2A). The phosphates reach over to the site of phosphoryl transfer, where interactions with the conserved Lys<sup>72</sup> anchor the  $\alpha$  and  $\beta$  phosphates to Glu<sup>91</sup> of the  $\alpha$ C helix. The distance between Lys<sup>72</sup> N $\zeta$  and Glu<sup>91</sup> O $\epsilon$ 1 is 2.6 Å, and two water molecules (W14 and W15) contribute to this salt bridge. This, as well as the more open glycine-rich loop, represents a conformation of the active site, which is intermediate between closed in the PKA-C complex with  $\text{Mg}_2\text{ATP}$  and PKI and fully open in apo-PKA-C (11, 15, 16). Opening of the glycine-rich loop is consistent with this being a product complex, poised for release of the phosphorylated peptide.

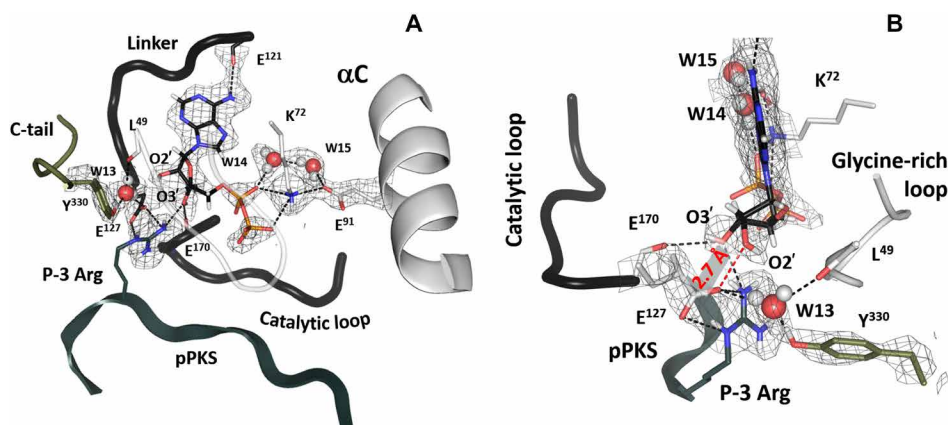
In this structure, we can truly recognize the singular and essential role that the ribose ring plays. It is anchored both to the C-lobe through its interactions with Glu<sup>127</sup>, which immediately follows the linker and is the “gateway” to the C-lobe, and to the N-lobe including Tyr<sup>330</sup> in the C-terminal tail. In PKA, it is also anchored to the substrate via the P-3 Arg. Thus, both substrates, the N-lobe and the C-lobe, converge at this site. This site controls the critical hinge, and water molecules are at the heart of this convergence site. The neutron structure allows us to look more closely at the 2'- and 3'-OH ribose groups. Unexpectedly, neither one participates in strong hydrogen bonding. The oxygen atom of the 2'-OH is 2.7 Å from the carboxylic oxygen of Glu<sup>127</sup>, which implies a strong hydrogen bond. However, as demonstrated in the neutron structure, the D atom of the hydroxyl group is rotated away from the straight line connecting the two oxygens, indicating only a weak interaction instead (Fig. 2B). This might be the result of Glu<sup>127</sup> already making several contacts—a salt bridge with the P-3 Arg residue and a hydrogen bond with a water molecule—shielding its charge and engaging both lone pairs on O $\epsilon$ 2 in hydrogen bonding. Similarly, the 3'-OH weakly hydrogen-bonds to the main-chain carboxyl of Glu<sup>170</sup> (the D...O distance is 2.3 Å). On the basis of chemical logic, dissociation of the products would require weakening of their hydrogen-bonding interactions with the enzyme. Therefore, we hypothesize that the weak interactions observed together with the more open conformation of the glycine-rich loop signify early stages of the cleft opening and reflect products and not substrates being bound at the active site.

### Substrate peptide binding

The peptide bridges the active-site cleft from the P-3 Arg that extends to the ribose, the P-2 Arg that nucleates the center of the C-lobe, and the P + 1 hydrophobic residue that docks onto the P + 1 loop and drags with it the activation loop phosphate and the  $\alpha$ C helix (Figs. 1A, 2, and 3).

### P-3 site

This is the site of convergence between the substrates and the N- and C-lobes, where information from the substrate docking to the C-lobe is conveyed to the N-lobe. The side chain of P-3 Arg reaches the ribose where it integrates and helps to coordinate the closing of the N-lobe. In PKA, this engages three structural elements: the C-tail (Phe<sup>327</sup> and Tyr<sup>330</sup>), which, in the absence of a nucleotide, is



**Fig. 2. Nucleotide binding site and its convergence with the P-3 site of the substrate.** (A) Hydrogen-bonding interactions contributing to the nucleotide binding. The  $2F_o - F_c$  nuclear density map is shown as gray mesh contoured at  $2.0\sigma$  level in both panels. (B) Close-up view of the ribose ring anchoring.

flexible; the hydrophobic C-spine; and the catalytic loop (Fig. 2A and fig. S1C). In the product complex, Tyr<sup>330</sup> does not hydrogen-bond to the ribose ring; instead, it binds directly to a water molecule, W13, which hydrogen-bonds with the Glu<sup>127</sup> side chain and the Leu<sup>49</sup> main-chain carbonyl. The C-spine residues in the kinase core and Phe<sup>327</sup> in the C-tail, which are essential parts of the hydrophobic shell, anchor the adenine ring (17). The catalytic loop is connected to the site through the backbone carbonyl of Glu<sup>170</sup>. The ribose is the hub for receiving feedback from substrate versus product binding and eventually will likely trigger opening of the cleft to release first one Mg ion and then the ADP-Mg (18). Once the M1 metal leaves, water can enter the active-site cleft, which mediates the release of the nucleotide.

### P-2 site (Fig. 3)

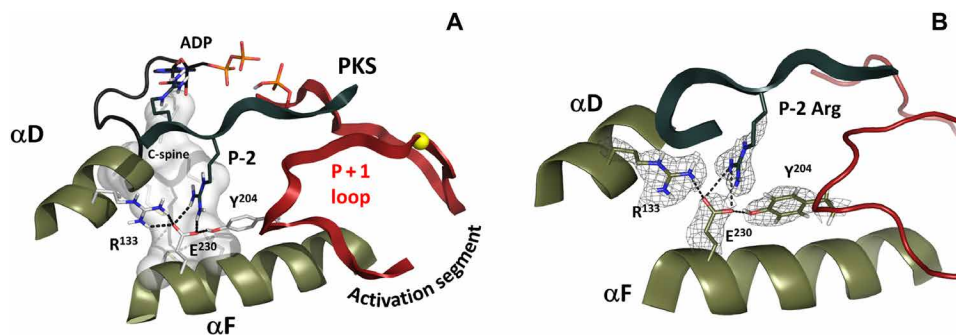
This critical site that bridges from the catalytic loop to the activation segment is nucleated by Glu<sup>230</sup> and recognizes the P-2 Arg in the PKA substrate. Although not all kinases recognize a basic residue at this site, many do, and Glu<sup>230</sup> is highly conserved. It interacts with Tyr<sup>204</sup> in the P + 1 loop that follows the activation loop. This Tyr is highly conserved in Ser/Thr kinases, and it is conserved as a Trp in tyrosine kinases. In PKA, Glu<sup>230</sup> also bridges to Arg<sup>133</sup>, which can serve as a flexible anchor to the C-terminal tail (19). The importance of this site became apparent from several mutations. Replacement of Tyr<sup>204</sup> with Ala, for example, has a major effect on the dynamic properties of the protein. It severely diminishes the ability of the protein to transfer the phosphate from ATP to a peptide substrate; however, its adenosine triphosphatase (ATPase) activity is only marginally reduced. Thus, in the crystal structure of this mutant (20) that bound to ATP and PKI, which largely resembles the wild-type structure, we found both ATP and the products (ADP and PO<sub>4</sub>) trapped in the active-site cleft. The dynamic properties of this mutant, based on NMR data, are also markedly altered (21). Similarly, mutation of Glu<sup>230</sup> to Gln has a major effect not only on catalysis but also on the conformational state of the protein. This mutant crystallizes in a very stable and open apo conformation even when both AMP-PNP (adenylyl-imidodiphosphate) and PKI are present in the crystallization buffer (22). From the current neutron diffraction structure, one can appreciate how the extended hydrogen-bonding network is severed by this simple change, replacing Glu<sup>230</sup> with Gln. It highlights the node as an essential part of the switch mechanism that allows the enzyme to toggle between open and closed conformations as it traverses the catalytic cycle.

### Phosphorylated activation loop (Figs. 4 and 5A and fig. S3)

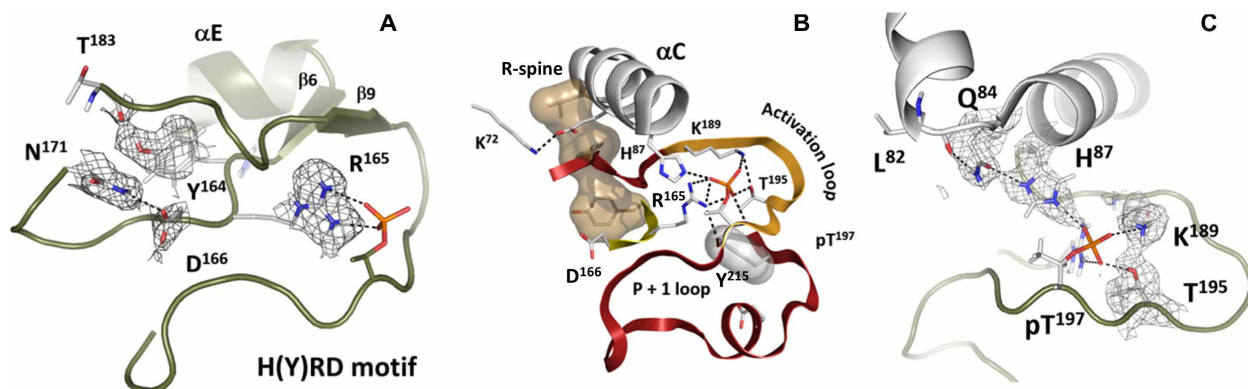
Phosphorylation of the activation loop at Thr<sup>197</sup> (pT<sup>197</sup>) is an important regulatory feature of most EPKs and here, we can observe the consequences of this modification. The phosphate interacts directly with many key residues that link it to other activation loop residues such as Thr<sup>195</sup> and Lys<sup>189</sup>, as well as Arg<sup>165</sup> of the catalytic loop, and His<sup>87</sup> in the  $\alpha$ C helix. However, the hydrogen-bonding and hydrophobic networks that are nucleated by this phosphate extend far beyond the phosphate. The backbone of pT<sup>197</sup> is anchored to the side chain of Tyr<sup>215</sup>, which is also highly conserved in EPKs. A key part of this extended hydrogen-bonding network is the side chain of Arg<sup>165</sup>, which is part of the His-Arg-Asp (HRD) motif in the catalytic loop that is highlighted in Fig. 4. While many of these interactions were appreciated in the original structure of the C subunit, direct visualization of the extended hydrogen-bonding network in the neutron structure allows us to fully appreciate the magnitude of this phosphorylation event and why it is so essential for stabilizing the active conformation of the kinase.

A key interaction of the activation loop phosphate is with His<sup>87</sup>. In this structure of the semiclosed conformation, His<sup>87</sup> is protonated (Fig. 4C). We hypothesize that it is not protonated in the open conformation, and this is an important part of the switch mechanism that slows down catalysis and makes the release of ADP the rate-limiting step in the catalytic cycle (23). While His<sup>87</sup> is at the N terminus of the  $\alpha$ C helix, the C terminus of this helix is anchored by the assembly of the R-spine. Two coordinated motions contribute to the activation of every protein kinase—the Asp-Phe-Gly (DFG) motif and the  $\alpha$ C helix moving in to the active site. The consequence of these concerted motions is the alignment of the R-spine (Fig. 5A and fig. S4B). There are four spine residues. RS1 is Tyr<sup>164</sup>, which is a part of the HRD motif and is always conserved as either His or Tyr [reviewed in (8) and references therein]. When the spine is aligned, RS1 and RS4 (Leu<sup>106</sup>) are firmly anchored: RS1 to the catalytic loop in the C-lobe and RS4 to the  $\beta$ 4 strand in the N-lobe. In contrast, RS2 (Phe<sup>185</sup>) in the DFG motif and RS3 (Leu<sup>95</sup>) in the  $\alpha$ C helix are dynamically recruited to the active-site cleft as part of the activation mechanism.

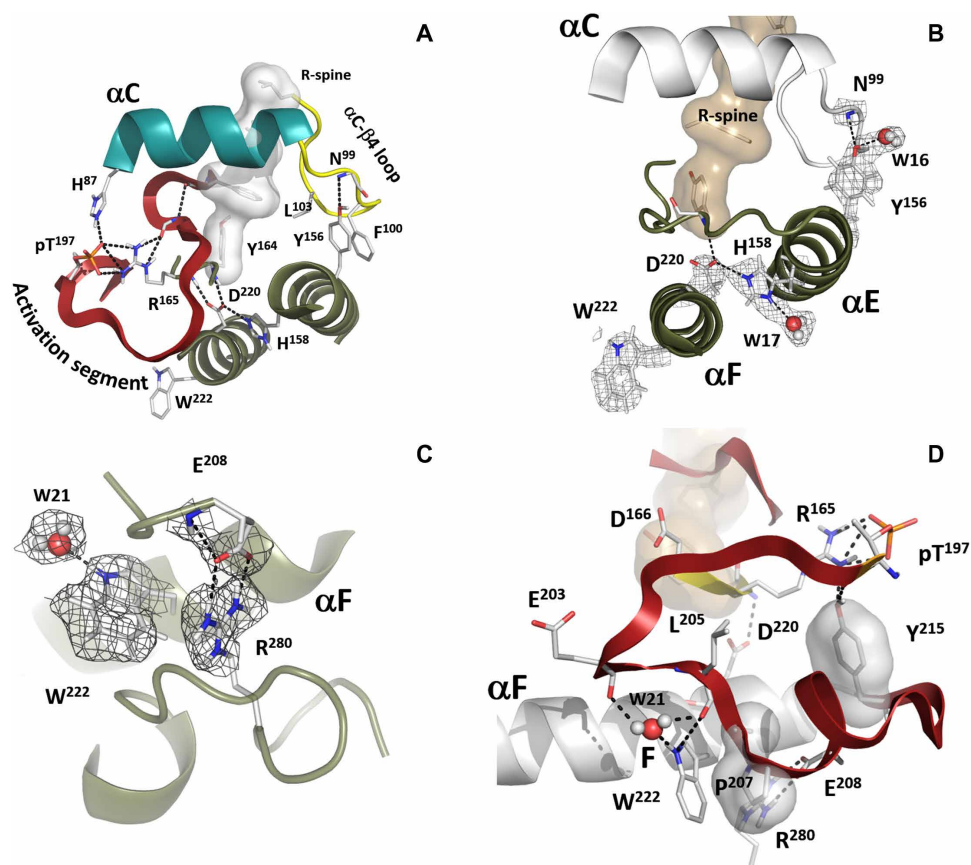
The neutron diffraction structure also shows that Cys<sup>199</sup> in the activation segment is fully protonated, indicating that it has an elevated pK<sub>a</sub> (where K<sub>a</sub> is the acid dissociation constant) value (fig. S3B). In contrast, this Cys is highly reactive, and likely deprotonated, in



**Fig. 3. Integration of P-2 substrate site with the major structural elements at the center of the C-lobe.** (A) The C-spine, C-terminal tail, and P + 1 loop are anchored to the  $\alpha$ F-helix, and the whole architecture of the catalytic subunit is locked in closed conformation through interactions with P-2 Arg of the high-affinity substrate peptide. (B) Recognition of the P-2 Arg is mediated by stabilizing interactions of  $\alpha$ F Glu<sup>230</sup> with Tyr<sup>204</sup> of P + 1 loop and Arg<sup>133</sup>. The  $2F_o - F_c$  nuclear density map is shown as gray mesh contoured at 2.0 $\sigma$  level.



**Fig. 4. Framework of stabilizing interactions at the nucleus of the activation loop—the phosphorylation site pT<sup>197</sup>.** (A) Hydrogen-bonding bridge between phosphorylated Thr<sup>197</sup> and Arg<sup>165</sup> from the HRD motif. (B) Alignment of the C-helix, HRD motif, and activation loop segments critical to activation. (C) Assembly of the charged residues (His<sup>87</sup>, Arg<sup>165</sup>, Lys<sup>189</sup>) around deprotonated phosphate on pT<sup>197</sup>. The  $2F_o - F_c$  nuclear density map is shown as gray mesh contoured at  $2.0\sigma$  level in (A) and (C).



**Fig. 5. Key interactions of the active kinase core.** (A) Relationship of the correctly positioned C-helix to the activation loop and to the HRD motif through  $\alpha$ C- $\beta$ 4 loop tethered to the C-lobe. (B) Close-up look at the interactions anchoring the N- and C-lobes. The  $\alpha$ C- $\beta$ 4 loop is attached to the  $\alpha$ E-helix via conserved Tyr<sup>156</sup>; the  $\alpha$ E links to  $\alpha$ F via interactions between His<sup>158</sup> and Asp<sup>220</sup>, which goes back to the HRD pattern. (C) Allosteric node: Arg<sup>280</sup>-Glu<sup>208</sup> salt bridge. The  $2F_o - F_c$  nuclear density map is shown as gray mesh contoured at  $2.0\sigma$  level. (D) Incorporation of the Arg<sup>280</sup>-Glu<sup>208</sup> node and Trp<sup>222</sup> into the activation segment: activation loop and the HRD motif that precedes the catalytic loop.

the apo state with a  $pK_a$  value of around 6 (24). Cys<sup>199</sup> is protected in the presence of nucleotide (25), in the RI $\alpha$  holoenzymes (26), and in the PKA-C:PKI:Mg<sub>2</sub>ATP complex (27). Modification of Cys<sup>199</sup> by sulfhydryl-specific reagents results in loss of kinase activity, but protonated Cys<sup>199</sup> would be less reactive, ensuring proper kinase functioning.

#### Conserved water molecules (fig. S4)

A conserved water molecule at Asp<sup>184</sup> (W19 in fig. S4B) was recognized early on when Shaltiel *et al.* (28) compared seven different structures of PKA. It was also identified as one of six conserved water molecules when Knight *et al.* (29) carried out a comparison of 13 active-conformation kinases. Most of these water molecules

flank the R-spine, and all are conserved in our neutron diffraction structure of the product complex (fig. S4 and table S1). Potentially, the Asp<sup>184</sup> water molecule can be a fundamental feature of the switch mechanism that allows the synergistic binding of nucleotide and peptide.

While the  $\alpha$ C- $\beta$ 4 loop was largely ignored in the original structure of PKA and of most kinases, it is a classic  $\beta$  turn and is an important element of the secondary structure. Water molecules are an essential feature of  $\beta$  turns, and one of the conserved water molecules in the  $\alpha$ C- $\beta$ 4 loop (W18 in fig. S4) hydrogen-bonds to the backbone carbonyl and amide of Leu<sup>95</sup> (RS3) and Leu<sup>106</sup> (RS4), respectively. This is a conserved feature of the active kinase, whereas the R-spine is broken when the activation loop is not phosphorylated (30). The tip of the  $\alpha$ C- $\beta$ 4 loop is the only element of the N-lobe that is tethered as a rigid body to the C-lobe, as it goes through the opening and closing during the catalytic cycle (31). In AGC kinases, the tip of this loop (Phe<sup>100</sup>-Pro-Phe) is anchored to the beginning of the C-terminal tail at the base of the C-lobe (32). Another important anchor site for the  $\alpha$ C- $\beta$ 4 loop is the Asn<sup>99</sup> backbone amide, which is anchored to the hydroxyl of the Tyr<sup>156</sup> side chain in the  $\alpha$ E helix (Fig. 5, A and B). While the Asn<sup>99</sup> is not conserved, Tyr<sup>156</sup> is conserved as a Tyr or His in most protein kinases. Two residues further on is His<sup>158</sup>, which is the other protonated His in this structure, whose protonation state was previously unappreciated.

### Role of His<sup>158</sup>

His<sup>158</sup> is highly conserved, although its functional importance has only recently been appreciated (33). Here, we see that it is directly hydrogen-bonded to the backbone carbonyl of Asp<sup>220</sup> (Fig. 5, A and B), which is an essential anchor to the active site. The side chain of Asp<sup>220</sup> docks onto the backbone carbonyl and nitrogen of Tyr<sup>164</sup>, which, in turn, is the anchoring residue for the assembly of the R-spine. In addition to its role as a spine residue, Tyr<sup>164</sup> side chain bridges across the catalytic loop to anchor against Asn<sup>171</sup>. The side chain of Asn<sup>171</sup> not only coordinates to the metal ion bound to ADP but also bridges the catalytic loop to anchor with the backbone carbonyl of Asp<sup>166</sup>, which is also part of the HRD motif. The importance of His<sup>158</sup> takes on added significance based on the recent NMR work (34), where it is shown how important the  $\alpha$ C- $\beta$ 4 residues are in terms of mediating and/or nucleating the allosteric network. Val<sup>104</sup>, for example, is an essential residue that docks to the adenine ring of ATP, while Leu<sup>103</sup> bridges to the network that links directly to His<sup>158</sup> at the active site. The  $\alpha$ C- $\beta$ 4 loop provides an extended network that allows the catalytic site to sense ATP binding.

### Signal integration motifs

We previously identified “signal integration motifs” (SIMs) as small peptide segments that integrate at least three distal conserved motifs (16). With the neutron structure, we now experimentally validate the protonation states of the residues that contribute to these extended networks. The turn of the  $\alpha$ C helix (Glu<sup>91</sup>-Lys<sup>92</sup>-Arg<sup>93</sup>) is one such motif (fig. S5), where Glu<sup>91</sup> bridges to Lys<sup>72</sup> in  $\beta$ 3, Lys<sup>92</sup> anchors the C-terminal hydrophobic motif to the  $\alpha$ C helix, which is a key step in assembling the active conformation of the R-spine, and Arg<sup>93</sup> bridges to Trp<sup>30</sup> in the  $\alpha$ A helix. The  $\alpha$ A helix, in turn, is anchored to Arg<sup>190</sup> in the activation loop, a critical interaction that is missing in most protein kinases.

The single turn that initiates the  $\alpha$ F helix (Asp<sup>220</sup> through Trp<sup>222</sup>) is another highly conserved SIM (Fig. 5, C and D). This segment

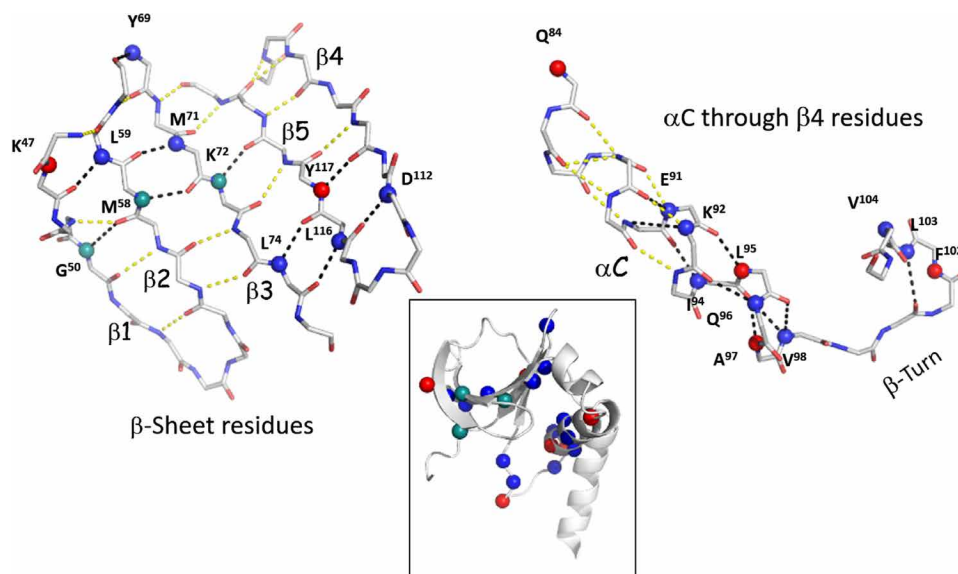
interacts with the HRD motif in the catalytic loop and with His<sup>158</sup> in the  $\alpha$ E helix (Fig. 5, A and B) and nucleates the Arg<sup>280</sup>-Glu<sup>208</sup> electrostatic node, which is a defining feature that distinguishes the EPKs from their evolutionary precursors, the eukaryotic-like kinases (35, 36). The neutron structure not only highlights this motif as a SIM but also reveals an ordered water molecule (W21 in Fig. 5, C and D, and fig. S4B) as a part of the motif.

### EPK-specific allosteric node, Arg<sup>280</sup>-Glu<sup>208</sup> (Fig. 5, C and D)

While the evolutionary-related, eukaryotic-like kinases share many of the same catalytic residues as the EPKs, they lack the highly regulated activation loop and the  $\alpha$ H- $\alpha$ I loop motifs that are firmly anchored both to the hydrophobic  $\alpha$ F helix and to the activation loop. The key node that links the two EPK-specific motifs is the Arg<sup>280</sup>-Glu<sup>208</sup> electrostatic pair. Both residues are firmly anchored by hydrophobic interactions to the N terminus of the F-helix, which begins with Asp<sup>220</sup>. We have already shown that the buried His<sup>158</sup> is protonated and anchored to Asp<sup>220</sup>. Here, we see how the highly conserved Trp<sup>222</sup> contributes to this hydrophobic node. In addition, one of the stable and buried water molecules, W21, anchors the side chain of Trp<sup>222</sup> to the backbone of Glu<sup>203</sup> and Leu<sup>205</sup> in the P + 1 loop (Fig. 5D). The other stable water molecules in this structure are at the active site (fig. S4B), but, unlike the Arg<sup>280</sup>-Glu<sup>208</sup> node, the active site will be exposed to solvent when the cleft opens. The side chain of Tyr<sup>215</sup> is also highly conserved in EPKs, and it hydrogen-bonds to the backbone carbonyl of Thr<sup>197</sup> (Fig. 5D). Figure 5 summarizes the remarkable interactions that link this Arg<sup>280</sup>-Glu<sup>208</sup> node to the phosphorylated activation loop, including how Asp<sup>220</sup> feeds back directly to the HRD motif that proceeds the catalytic loop.

### Comparison of the slow-exchanging backbone amides with NMR H/D fractionation data (Fig. 6)

As a final analysis, it is instructive to compare H/D exchange patterns of the backbone amides in the neutron structure with our earlier NMR analysis of the amides of the similar protein kinase complex, PKA-C:Mg<sub>2</sub>ADP:PKI (37). In a neutron structure, the H/D exchange of each backbone amide is classified on the basis of the occupancy of the corresponding D atom. Amides with deuterium occupancies less than 0.5 can be categorized as slow-exchanging and those with 0.51 to 1 as fully exchanged. Using NMR, it is possible to probe the strength of hydrogen bonds using H/D fractionation factors,  $\phi$ . Fractionation factors are equilibrium constants that can be directly correlated with the free energy of the H-bond. Low  $\phi$  values ( $\phi < 0.5$ ) are indicative of stronger H-bonds, and high  $\phi$  values are associated with weaker hydrogen bonds (38). Weaker bonds are more accessible and accumulate more deuterium relative to stronger and less accessible ones. For this comparison, we focused on the group of slow-exchanging amides ( $\phi < 0.5$ ). Fifty-five residues belonging to this category were identified in the NMR set of the PKA-C ternary complex and checked against the neutron data set for the D atom occupancies of the corresponding backbone amides (table S2). Figure 6 summarizes the backbone amides that could be compared between the neutron diffraction and NMR data. It is striking to detect similarities in the distribution of the slow-exchanging amides in the N- and C-lobes and to correlate them with the secondary and tertiary elements of the core (fig. S6). The stable hydrogen bonds in the N-lobe are distributed between the  $\beta$  sheet, which is the dominant structural element of the N-lobe, and the  $\alpha$ C helix, which communicates with the activation loop in the C-lobe. In contrast to the



**Fig. 6. Distribution of the backbone amides in the neutron structure, identified as slow-exchanging by NMR.** The amide groups are represented as spheres and color-coded according to the occupancy of the corresponding deuterium, D, atom: Blue and green are amides with a D occupancy of less than 0.5, classified as slow-exchanging and are the same in the NMR dataset; red amides have an occupancy of 0.51 to 1, classified as fully exchanged and are different compared to the NMR dataset.

$\alpha$ C helix, which is solvent exposed, amides of the buried  $\alpha$ F and  $\alpha$ E helices are much more exchangeable. Clustering of the slow-exchanging amides in the C terminus of the  $\alpha$ C helix and the  $\alpha$ C- $\beta$ 4 loop can be an indication of the quenched dynamics, signifying remarkable stability of these structural elements, critical for the proper assembly of the active site. This is the node where the N- and C-lobes come together in the community map analysis of the ternary complex that is fully committed to catalysis. It is worth noting that one-third of the 55 backbone amides identified as slow-exchanging in the NMR set are fully exchanged in the neutron structure. An increase in the  $\phi$  factors, for the same group of amides, was observed in the NMR sets of apo and/or binary (with  $\text{Mg}_2\text{ADP}$ ) PKA-C complexes relative to the ternary, PKA-C: $\text{Mg}_2\text{ADP}$ :PKI, cluster. Globally, this correlates with the enzyme transitioning from closed (ternary) to open (apo) or semiopen (binary) conformations, weakening of hydrogen bonds, and different dynamics (15, 21, 37). In the neutron structure of PKA-C: $\text{Sr}_2\text{ADP}$ :pPKS, the enzyme is in its intermediate or semiopen conformation. We suggest that the difference in the H/D pattern observed in the neutron dataset is due to the neutron structure representing products rather than substrates and might be an indication of altered dynamics between the two complexes. This comparison provides a hypothesis that can be tested in future studies.

## DISCUSSION

In the neutron diffraction structure, we can see how the peptide from the P-3 to the P + 1 residues traverses the entire active-site region like a Velcro strip from the nucleotide binding site to the P + 1 loop (Fig. 1). The mechanism for opening and closing of the catalytic cleft, which brings the N- and C-lobes together to facilitate transfer of the phosphate and then to mediate product release, is embedded in this network. The expanded chemical portrait that is provided by the neutron diffraction structure also shows how conserved water molecules contribute strategically to

this interface and potentially to opening and closing of the active-site cleft. This electrostatic and hydrogen-bonding network also synergizes with the conserved hydrophobic architecture of the kinase core, which was defined by the hydrophobic R- and C-spines and seen experimentally with NMR when the methyl side chains were labeled (34).

Since its structure was first solved in 1991, the PKA-C subunit has served as the prototypical protein kinase, and this neutron diffraction structure adds a valuable new dimension to our chemical understanding of protein kinase structure and function. It paves the way for improved quantum chemistry calculations that include products as well as inhibitor and transition state complexes. In addition, we can couple the diffraction data with experimental techniques that can now track residue-specific dynamics on microsecond to millisecond time scales (NMR) and on second to minute time scales with H/D exchange mass spectrometry. This allows us to compare in real time the dynamic features of specific residues as the enzyme toggles between its catalytic states. By labeling methyl side chains, we can also track the correlated motions of the hydrophobic side chains, which reveals how entropic changes in the conserved hydrophobic core drive catalysis. Using powerful computational simulations, we can also track correlated motions in new ways that allow us to identify “communities” that are dedicated to specific functions. This goes beyond traditional assignments of function to secondary and tertiary structural elements and instead defines an entropy-driven allosteric framework for driving catalysis. Comparison of the exchangeable backbone amides in this neutron diffraction structure of the PKA-C: $\text{Sr}_2\text{ADP}$ :pPKS complex with the NMR structure of the PKA-C: $\text{Mg}_2\text{ADP}$ :PKI complex allows us to build and test hypotheses about the processes that drive opening and closing of the catalytic cleft. The combined integration of information from computational and experimental systems is beginning to define a more comprehensive way of appreciating how entropy and enthalpy contribute to the finely tuned allosteric regulation of this molecular switch.

## MATERIALS AND METHODS

### Protein expression, purification, and crystallization

For deuteration of His<sub>6</sub>-tagged recombinant mouse PKA-C subunit, BL21 (DE3) cells were grown in LB medium. Once the culture reached OD<sub>600</sub> (optical density at 600 nm) of ~2, it was spun down, and the pellet was resuspended into 80% (v/v) D<sub>2</sub>O minimal medium with carbenicillin (100 µg/ml). The resulting preculture (~100 ml) was used to inoculate 3 liters of fresh 80% (v/v) D<sub>2</sub>O medium in a benchtop bioreactor (BioFlo 3000, New Brunswick Scientific) set at 35°C. The initial OD<sub>600</sub> read 0.02, and the cells were grown to OD<sub>600</sub> of ~10, induced with 1 mM isopropyl-β-D-thiogalactopyranoside, and cooled down to 18°C. Upon consumption of the initial glycerol in the batch, the culture was fed with a solution of 10% (w/v) hydrogenous glycerol and 0.2% (w/v) MgSO<sub>4</sub>; 10% (w/v) NaOH was added on demand to control pD (>6.9 uncorrected, pD = pH+0.4). After the 20-hour induction, the cell suspension was collected and pelleted at 6000g via centrifugation at 4°C for 30 min to yield ~30 g/liter wet weight of deuterated cell paste. All solutions were prepared with 80% (v/v) D<sub>2</sub>O and filter-sterilized into dry, sterile containers before use. The enzyme was first purified by metal affinity chromatography using a 5-ml HisTrap FF column, then concentrated to 8 to 10 mg/ml, and loaded onto a gel filtration column (HiLoad 16/60 Superdex 75) equilibrated with 50 mM tris-HCl (pH 6.5), 20 mM SrCl<sub>2</sub>, 250 mM NaCl, and 2 mM dithiothreitol (DTT). Eluted protein fractions were concentrated to ~10 mg/ml and used for crystallization experiments. Crystals were grown by vapor diffusion in nine-well glass plates at 14°C. The product complex was prepared before crystallization. The PKA-C:Na<sub>2</sub>ATP:PKS molar ratio was kept at 1:10:10. The nucleotide and the peptide substrate were added as solids to the protein solution. The reaction mixture was incubated at 4°C for 30 to 40 min, filtered through 0.2-µm VWR spin filters, and combined at a 1:1 ratio with the crystallization solution containing 19.5% (w/v) polyethylene glycol (PEG) 6000, 0.1 M MES (pH 5), 5 mM DTT, and 50 mM SrCl<sub>2</sub>. Several crystals (~0.1 mm<sup>3</sup>) from the same crystallization drop were mounted in quartz capillaries containing the reservoir solution made with 100% D<sub>2</sub>O. Several weeks were allowed for the H/D vapor exchange process before starting data collection.

### Data collection and structure refinement

The preliminary neutron diffraction data at room temperature were collected on the IMAGINE (39) instrument located at the High Flux Isotope Reactor (Oak Ridge National Laboratory) from a 0.1-mm<sup>3</sup> crystal of deuterated PKA-C:Sr<sub>2</sub>ADP:pPKS. The complete room temperature quasi-Laue dataset to 2.5 Å resolution was collected on the LADI-III beamline at the Institut Laue-Langevin (40). A summary of the experimental and refinement statistics is given in table S3. Table S4 provides the references for the software used in data collection and structure refinement.

In total, 30 diffraction images were collected (with an average exposure time of 24 hours per image) from two different crystal orientations. The neutron data were processed using the Daresbury Laboratory LAUE suite program LAUEGEN modified to account for the cylindrical geometry of the detector. The program LSCALE was used to determine the wavelength-normalization curve using the intensities of symmetry-equivalent reflections measured at different wavelengths. No explicit absorption corrections were applied. These data were then merged in SCALA. Monochromatic

x-ray diffraction data were collected on a different crystal from the same crystallization drop using an in-house Rigaku HomeFlux system, equipped with a MicroMax-007 HF generator, Osmic VariMax optics, and an R-AXIS IV++ image-plate detector. Diffraction data were collected, integrated, and scaled with the HKL-3000 software suite. The structure was solved by molecular replacement with CCP4 suite using the hydrogenous PKA-C:Sr<sub>2</sub>ADP:pPKS complex [Protein Data Bank (PDB) ID 4IAY] as a starting model. The room temperature x-ray structure of deuterated PKA-C:Sr<sub>2</sub>ADP:pPKS at a resolution of 2.0 Å was refined using SHELX-97 and served as a starting model in the joint x-ray/neutron (XN) refinement. The joint XN structure of deuterated PKA-C:Sr<sub>2</sub>ADP:pPKS was determined using nCNS and manipulated in Coot. The initial rigid-body refinement was followed by several cycles of positional, atomic displacement parameter, and occupancy refinement. Between each cycle, the structure was checked, side-chain conformations were modified, and water molecule orientations were built based on the  $F_O - F_C$  difference neutron scattering length density map. The  $2F_O - F_C$  and  $F_O - F_C$  neutron scattering length density maps were then examined to determine the correct orientation of hydroxyl (Ser, Thr, Tyr), ammonium (Lys), and sulfhydryl (Cys) groups and protonation states of His, Lys, Asp, Cys, and phosphorylated residues. The level of H/D exchange at OH, NH, and SH sites was refined. All H positions in deuterated PKA-C and labile H positions in the ligands (ADP and pPKS) were modeled as D, and then the occupancies of D were refined within the range of -0.56 to 1.00 (the scattering length of H is -0.56 times the scattering length of D; the amount of D at each position is then calculated as [(D occupancy + 0.56)/1.56]). Before depositing the final structure to the PDB, a script was run that converts a record for the coordinate of D atom into two records corresponding to H and D atoms partially occupying the same site, both with positive partial occupancies that add up to unity. The joint XN structure of deuterated PKA-C:Sr<sub>2</sub>ADP:pPKS has been deposited to the PDB (ID 6E21).

### SUPPLEMENTARY MATERIALS

Supplementary material for this article is available at <http://advances.sciencemag.org/cgi/content/full/5/3/eaav0482/DC1>

Fig. S1. Neutron structure describes the PKA-C product complex, PKA-C:Sr<sub>2</sub>ADP:pPKS.

Fig. S2. The phosphorylation site.

Fig. S3. Positive  $2F_O - F_C$  electron (red) and nuclear (gray) density maps at 2.0σ demonstrate complementary features of X-ray and neutron diffraction techniques.

Fig. S4. Stable water molecules.

Fig. S5. The turn of the αC helix, Glu<sup>91</sup>-Lys<sup>92</sup>-Arg<sup>93</sup>, is a signal integration motif.

Fig. S6. Distribution of the identified backbone amides deconvoluted according to the occupancy of corresponding D atoms and structure.

Table S1. Stable water molecules.

Table S2. Backbone H/D exchange comparison.

Table S3. Room temperature crystallographic data collection and joint XN refinement statistics.

Table S4. The software used in data collection and refinement.

References (41–50)

### REFERENCES AND NOTES

1. D. R. Knighton, J. H. Zheng, L. F. Ten Eyck, V. A. Ashford, N. H. Xuong, S. S. Taylor, J. M. Sowadski, Crystal structure of the catalytic subunit of cyclic adenosine monophosphate-dependent protein kinase. *Science* **253**, 407–414 (1991).
2. D. R. Knighton, J. H. Zheng, L. F. Ten Eyck, N. H. Xuong, S. S. Taylor, J. M. Sowadski, Structure of a peptide inhibitor bound to the catalytic subunit of cyclic adenosine monophosphate-dependent protein kinase. *Science* **253**, 414–420 (1991).
3. S. K. Hanks, T. Hunter, Protein kinases 6. The eukaryotic protein kinase superfamily: Kinase (catalytic) domain structure and classification. *FASEB J.* **9**, 576–596 (1995).



4. B. Nolen, S. Taylor, G. Ghosh, Regulation of protein kinases; controlling activity through activation segment conformation. *Mol. Cell* **15**, 661–675 (2004).
5. M. Huse, J. Kuriyan, The conformational plasticity of protein kinases. *Cell* **109**, 275–282 (2002).
6. A. P. Kornev, N. M. Haste, S. S. Taylor, L. F. Ten Eyck, Surface comparison of active and inactive protein kinases identifies a conserved activation mechanism. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 17783–17788 (2006).
7. A. P. Kornev, S. S. Taylor, L. F. Ten Eyck, A helix scaffold for the assembly of active protein kinases. *Proc. Natl. Acad. Sci. U.S.A.* **105**, 14377–14382 (2008).
8. S. S. Taylor, A. P. Kornev, Protein kinases: Evolution of dynamic regulatory proteins. *Trends Biochem. Sci.* **36**, 65–77 (2011).
9. J. Zheng, E. A. Trafny, D. R. Knighton, N. H. Xuong, S. S. Taylor, L. F. Ten Eyck, J. M. Sowadski, 2.2 Å refined crystal structure of the catalytic subunit of cAMP-dependent protein kinase complexed with MnATP and a peptide inhibitor. *Acta Crystallogr. D Biol. Crystallogr.* **49**, 362–365 (1993).
10. Madhusudan, P. Akamine, N. H. Xuong, S. S. Taylor, Crystal structure of a transition state mimic of the catalytic subunit of cAMP-dependent protein kinase. *Nat. Struct. Biol.* **9**, 273–277 (2002).
11. P. Akamine, Madhusudan, J. Wu, N. H. Xuong, L. F. Ten Eyck, S. S. Taylor, Dynamic features of cAMP-dependent protein kinase revealed by apoenzyme crystal structure. *J. Mol. Biol.* **327**, 159–171 (2003).
12. A. C. Bastidas, M. S. Deal, J. M. Steichen, Y. Guo, J. Wu, S. S. Taylor, Phosphoryl transfer by protein kinase A is captured in a crystal lattice. *J. Am. Chem. Soc.* **135**, 4788–4798 (2013).
13. A. C. Bastidas, J. Wu, S. S. Taylor, Molecular features of product release for the PKA catalytic cycle. *Biochemistry* **54**, 2–10 (2015).
14. O. Gerlits, J. Tian, A. Das, P. Langan, W. T. Heller, A. Kovalevsky, Phosphoryl transfer reaction snapshots in crystals: Insights into the mechanism of protein kinase A catalytic subunit. *J. Biol. Chem.* **290**, 15538–15548 (2015).
15. L. R. Masterson, C. Cheng, T. Yu, M. Tonelli, A. Kornev, S. S. Taylor, G. Veglia, Dynamics connect substrate recognition to catalysis in protein kinase A. *Nat. Chem. Biol.* **6**, 821–828 (2010).
16. D. A. Johnson, P. Akamine, E. Radzio-Andzelm, M. Madhusudan, S. S. Taylor, Dynamics of cAMP-dependent protein kinase. *Chem. Rev.* **101**, 2243–2270 (2001).
17. J. Yang, E. J. Kennedy, J. Wu, M. S. Deal, J. Pennypacker, G. Ghosh, S. S. Taylor, Contribution of non-catalytic core residues to activity and regulation in protein kinase A. *J. Biol. Chem.* **284**, 6241–6248 (2009).
18. I. V. Khavrutskii, B. Grant, S. S. Taylor, J. A. McCammon, A transition path ensemble study reveals a linchpin role for Mg<sup>2+</sup> during rate-limiting ADP release from protein kinase A. *Biochemistry* **48**, 11532–11545 (2009).
19. X. Cheng, C. Phelps, S. S. Taylor, Differential binding of cAMP-dependent protein kinase regulatory subunit isoforms I $\alpha$  and I $\beta$  to the catalytic subunit. *J. Biol. Chem.* **276**, 4102–4108 (2001).
20. J. Yang, L. F. Ten Eyck, N. H. Xuong, S. S. Taylor, Crystal structure of a cAMP-dependent protein kinase mutant at 1.26 Å: New insights into the catalytic mechanism. *J. Biol. Chem.* **336**, 473–487 (2004).
21. L. R. Masterson, A. Mascioni, N. J. Traaseth, S. S. Taylor, G. Veglia, Allosteric cooperativity in protein kinase A. *Proc. Natl. Acad. Sci. U.S.A.* **105**, 506–511 (2008).
22. J. Wu, J. Yang, N. Kannan, Madhusudan, N.-H. Xuong, L. F. Ten Eyck, S. S. Taylor, Crystal structure of the E230Q mutant of cAMP dependent protein kinase reveals an unexpected apoenzyme conformation and an extended N-terminal  $\alpha$ helix. *Protein Sci.* **14**, 2871–2879 (2005).
23. H. S. Meharena, X. Fan, L. G. Ahuja, M. M. Keshwani, C. L. McClendon, A. M. Chen, J. A. Adams, S. S. Taylor, Decoding the interactions regulating the active state mechanics of eukaryotic protein kinases. *PLOS Biol.* **14**, e2000127 (2016).
24. J. S. Jiménez, A. Kupfer, V. Gani, S. Shaltiel, Salt-induced conformational changes in the catalytic subunit of adenosine cyclic 3',5'-phosphate dependent protein kinase. Use for establishing a connection between one sulfhydryl group and the  $\gamma$ -P subsite in the ATP site of this subunit. *Biochemistry* **21**, 1623–1630 (1982).
25. E. A., First, D. A. Johnson, S. S. Taylor, Fluorescence energy transfer between cysteine 199 and cysteine 343: Evidence for MgATP-dependent conformational change in the catalytic subunit of cAMP-dependent protein kinase. *Biochemistry* **28**, 3606–3613 (1989).
26. N. C. Nelson, S. S. Taylor, Selective protection of sulfhydryl groups in cAMP-dependent protein kinase II. *J. Biol. Chem.* **258**, 10981–10987 (1983).
27. K. M. Humphries, C. Juliano, S. S. Taylor, Regulation of cAMP-dependent protein kinase activity by glutathionylation. *J. Biol. Chem.* **277**, 43505–43511 (2002).
28. S. Shaltiel, S. Cox, S. S. Taylor, Conserved water molecules contribute to the extensive network of interactions at the active site of protein kinase A. *Proc. Natl. Acad. Sci. U.S.A.* **95**, 484–491 (1998).
29. J. D. Knight, D. Hamelberg, J. A. McCammon, R. Kothary, The role of conserved water molecules in the catalytic domain of protein kinases. *Proteins* **76**, 527–535 (2009).
30. J. M. Steichen, G. H. Iyer, S. Li, S. A. Saldanha, M. S. Deal, V. L. Woods Jr., S. S. Taylor, Global consequences of activation loop phosphorylation on protein kinase A. *J. Biol. Chem.* **285**, 3825–3832 (2010).
31. I. Tsigelny, J. P. Greenberg, S. Cox, W. L. Nichols, S. S. Taylor, L. F. Ten Eyck, 600 ps molecular dynamics reveals stable substructures and flexible hinge points in cAMP dependent protein kinase. *Biopolymers* **50**, 513–524 (1999).
32. N. Kannan, N. Haste, S. S. Taylor, A. F. Neuwald, The hallmark of AGC kinase functional divergence is its C-terminal tail, a cis-acting regulatory module. *Proc. Natl. Acad. Sci. U.S.A.* **104**, 1272–1277 (2007).
33. N. Kannan, A. F. Neuwald, Did protein kinase regulatory mechanisms evolve through elaboration of a simple structural component? *J. Mol. Biol.* **351**, 956–972 (2005).
34. J. Kim, L. G. Ahuja, F. A. Chao, Y. Xia, C. L. McClendon, A. P. Kornev, S. S. Taylor, G. Veglia, A dynamic hydrophobic core orchestrates allostery in protein kinases. *Sci. Adv.* **3**, e1600663 (2017).
35. S. S. Taylor, M. M. Keshwani, J. M. Steichen, A. P. Kornev, Evolution of the eukaryotic protein kinases as dynamic molecular switches. *Philos. Trans. R. Soc. B Biol. Sci.* **367**, 2517–2528 (2012).
36. J. Yang, J. Wu, J. M. Steichen, A. P. Kornev, M. S. Deal, S. Li, B. Sankaran, V. L. Woods Jr., S. S. Taylor, A conserved Glu-Arg salt bridge connects coevolved motifs that define the eukaryotic protein kinase fold. *J. Mol. Biol.* **415**, 666–679 (2012).
37. G. C. Li, A. K. Srivastava, J. Kim, S. S. Taylor, G. Veglia, Mapping the hydrogen bond networks in the catalytic subunit of protein kinase A using H/D fractionation factors. *Biochemistry* **54**, 4042–4049 (2015).
38. M. M. Kreevoy, T.-M. Liang, K.-C. Chiang, Structures and isotopic fractionation factors of complexes AHA-1. *J. Am. Chem. Soc.* **99**, 5207–5209 (1977).
39. F. Meilleur, P. Munshi, L. Robertson, A. D. Stoica, L. Crow, A. Kovalevsky, T. Koritsanszky, B. C. Chakoumakos, R. Blessing, D. A. Myles, The IMAGINE instrument: First neutron protein structure and new capabilities for neutron macromolecular crystallography. *Acta Crystallogr. D Biol. Crystallogr.* **69**, 2157–2160 (2013).
40. M. P. Blakeley, S. C. Teixeira, I. Petit-Haertlein, I. Hazemann, A. Mitschler, M. Haertlein, E. Howard, A. D. Podjarny, Neutron macromolecular crystallography with LADI-III. *Acta Crystallogr. D Biol. Crystallogr.* **66**, 1198–1205 (2010).
41. J. W. Campbell, LAUEGEN, an X-windows-based program for the processing of Laue diffraction data. *J. Appl. Cryst.* **28**, 228–236 (1995).
42. J. W. Campbell, Q. Hao, M. M. Harding, N. D. Nguti, C. Wilkinson, LAUEGEN version 6.0 and INTLDM. *J. Appl. Cryst.* **31**, 496–502 (1998).
43. S. Arzt, J. W. Campbell, M. M. Harding, Q. Hao, J. R. Helliwell, LSCALE - the new normalization, scaling and absorption correction program in the Daresbury Laue software suite. *J. Appl. Cryst.* **32**, 554–562 (1999).
44. M. S. Weiss, Global indicators of X-ray data quality. *J. Appl. Cryst.* **34**, 130–135 (2001).
45. W. Minor, M. Cymborowski, Z. Otwinowski, M. Chruszcz, HKL3000: The integration of data reduction and structure solution—From diffraction images to an initial model in minutes. *Acta Crystallogr.* **62**, 859–866 (2006).
46. Collaborative Computational Project, Number 4, The CCP4 suite: Programs for protein crystallography. *Acta Crystallogr. D Biol. Crystallogr.* **50** (Pt 5), 760–763 (1994).
47. O. Gerlits, M. J. Waltman, S. Taylor, P. Langan, A. Kovalevsky, Insights into the phosphoryl transfer catalyzed by cAMP-dependent protein kinase: An X-ray crystallographic study of complexes with various metals and peptide substrate SP20. *Biochemistry* **52**, 3721–3727 (2013).
48. G. M. Sheldrick, A short history of SHELX. *Acta Crystallogr.* **A64**, 112–122 (2008).
49. P. D. Adams, M. Mustyhalimov, P. V. Afonine, P. Langan, Generalized X-ray and neutron crystallographic analysis: More accurate and complete structures for biological macromolecules. *Acta Crystallogr.* **D65**, 567–573 (2009).
50. P. Emsley, B. Lohkamp, W. G. Scott, K. Cowtan, Features and development of Coot. *Acta Crystallogr.* **66**, 486–501 (2010).

**Acknowledgments:** This research at ORNL's High Flux Isotope Reactor (IMAGINE beamline) was sponsored by the Scientific User Facilities Division, Office of Basic Energy Sciences, U.S. Department of Energy. We thank Institut Laue-Langevin (beamline LADI-III) for awarded neutron beamtime. The Office of Biological and Environmental Research supported research at Oak Ridge National Laboratory's Center for Structural Molecular Biology (CSMB) involving protein deuteration, using facilities supported by the Scientific User Facilities Division, Office of Basic Energy Sciences, U.S. Department of Energy. This manuscript has been authored by UT-Battelle LLC under DOE contract no. DE-AC05-00OR22725. We thank Dr. A. Kornev (University of California at San Diego) for technical assistance with the manuscript preparation and critical analysis of the paper, and Y. Wang (University of Minnesota at Minneapolis) for help in retrieving the NMR data for the H/D exchange comparison. **Funding:** S.S.T. and G.V. were partly supported by NIH grants GM19301 and GM100310, respectively. O.G., S.S.T., and A.K. were partly supported by a UCOP grant. O.G. and A.K. were partly supported by the U.S. Department of Energy's (DOE) Office of Basic Energy Sciences. **Author contributions:**

O.G., S.S.T., and A.K. designed research; O.G. and A.K. performed research; O.G. and K.L.W. prepared deuterated protein; M.P.B. collected and processed neutron diffraction data; O.G., G.V., S.S.T., and A.K. analyzed data; O.G., S.S.T., and A.K. wrote the paper. **Competing interests:** The authors declare that they have no competing interests. **Data and materials availability:** All data needed to evaluate the conclusions in the paper are present in the paper and/or the Supplementary Materials. The joint XN structure of deuterated PKA-C:Sr<sub>2</sub>ADP:pPKS has been deposited to the PDB (ID 6E21). Additional data related to this paper may be requested from the authors.

Submitted 9 August 2018

Accepted 31 January 2019

Published 20 March 2019

10.1126/sciadv.aav0482

**Citation:** O. Gerlits, K. L. Weiss, M. P. Blakeley, G. Veglia, S. S. Taylor, A. Kovalevsky, Zooming in on protons: Neutron structure of protein kinase A trapped in a product complex. *Sci. Adv.* **5**, eaav0482 (2019).