



Antibacterial Activity of Lefamulin against Pathogens Most Commonly Causing Community-Acquired Bacterial Pneumonia: SENTRY Antimicrobial Surveillance Program (2015–2016)

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ABSTRACT Lefamulin, the first semisynthetic pleuromutilin antibacterial for intravenous and oral treatment of community-acquired bacterial pneumonia (CABP), and comparators were evaluated for *in vitro* activity against a global collection of pathogens commonly causing CABP ($n = 8595$) from the 2015 and 2016 SENTRY Antimicrobial Surveillance Program. Lefamulin was highly active against the pathogens *Streptococcus pneumoniae*, including multidrug-resistant and extensively drug-resistant strains (MIC_{50/90} for total and resistant subsets, 0.06/0.12 $\mu\text{g/ml}$; 100% inhibited at ≤ 1 $\mu\text{g/ml}$), *Staphylococcus aureus*, including methicillin-resistant *Staphylococcus aureus* (MRSA; both MIC_{50/90}, 0.06/0.12 $\mu\text{g/ml}$; 99.8% and 99.6% inhibited at ≤ 1 $\mu\text{g/ml}$, respectively), *Haemophilus influenzae* (MIC_{50/90}, 0.5/1 $\mu\text{g/ml}$; 93.8% inhibited at ≤ 1 $\mu\text{g/ml}$), and *Moraxella catarrhalis* (MIC_{50/90}, 0.06/0.12 $\mu\text{g/ml}$; 100% inhibited at ≤ 0.25 $\mu\text{g/ml}$), and its activity was unaffected by resistance to other antibacterial classes.

KEYWORDS antimicrobial, community-acquired bacterial pneumonia, lefamulin, pleuromutilin

Community-acquired bacterial pneumonia (CABP) is a potentially serious respiratory infection with incidence rates of approximately 10.6 cases per 1,000 person-years in the United States (1) and ranges from 1.7 to 11.6 cases per 1,000 person-years as reported from Europe (2). It is a leading cause of hospitalization worldwide (3, 4) and, despite antibiotic treatment, is still a relevant cause of death (4, 5). Together with influenza, community-acquired pneumonia is the eighth most common cause of death in the United States (6). *Streptococcus pneumoniae* is the most commonly isolated pathogen associated with CABP; other common etiologic bacterial pathogens include *Haemophilus influenzae*, *Moraxella catarrhalis*, *Staphylococcus aureus*, and the atypical pathogens *Chlamydia pneumoniae*, *Legionella pneumophila*, and *Mycoplasma pneumoniae* (7, 8). Current recommendations for the treatment of CABP include initiation of empirical antibiotic therapies, which vary depending on the treatment setting, and may include monotherapy with a macrolide, doxycycline, respiratory fluoroquinolone, or combination therapy with a β -lactam plus a macrolide or a respiratory fluoroquinolone (9, 10). As a result of the increasing prevalence of resistance to currently available antimicrobials, particularly penicillin and macrolide resistance among *S. pneumoniae* and macrolide resistance among *M. pneumoniae* (11), high treatment failure rates (14% general hospital ward, $\sim 25\%$ outpatient) (12, 13), and adverse effects associated with current treatment options (14, 15), new treatment options for CABP are needed, preferably allowing an intravenous-to-oral switch to reduce the length of hospital stay and hospital-related costs.

Lefamulin is a pleuromutilin antimicrobial in late-stage clinical development for

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intravenous and oral use in patients with CABP. It is an inhibitor of bacterial protein synthesis by binding to the A and P sites of the peptidyl transferase center in the large subunit of the bacterial ribosome (16, 17). The unique interaction in this highly conserved region confers a low propensity for the development of bacterial resistance and is thought to be the reason for the lack of cross-resistance with other antibacterial classes, including macrolides, ketolides, lincosamides, fluoroquinolones, and tetracyclines (17). The antibacterial spectrum of lefamulin covers the typical Gram-positive and fastidious Gram-negative respiratory pathogens known to cause CABP and atypical pathogens, such as *M. pneumoniae* (including macrolide-resistant strains), *C. pneumoniae*, and *L. pneumophila* (18, 19). Pharmacokinetic and pharmacodynamic analyses demonstrated that lefamulin has rapid and predictable penetration into plasma (~1 to 2 $\mu\text{g/ml}$ after a single 150-mg intravenous or 600-mg oral dose) (20) and target tissues, such as the epithelial lining fluid in the lung (21), reaching area under the concentration-time curve (AUC):MIC ratios that support the proposed tentative breakpoints of 1 $\mu\text{g/ml}$ for *S. pneumoniae* and 0.5 $\mu\text{g/ml}$ for *S. aureus* (22). This study evaluated the *in vitro* activity of lefamulin and comparators against a global collection of typical respiratory pathogens that commonly cause CABP, as collected by the SENTRY Antimicrobial Surveillance Program (2015 to 2016).

The activity of lefamulin and comparators were determined against 8,595 unique bacterial pathogens collected from 65 medical centers from North America (United States [34 states]; $n = 3,240$), 39 from Europe and the Mediterranean region (19 nations, including Turkey and Israel; $n = 3,362$), 15 from the Asia-Pacific region (7 nations; $n = 1,271$), and 10 from Latin America (4 nations; $n = 722$) and submitted to a central monitoring laboratory (JMI Laboratories, North Liberty, IA, USA) for confirmation of bacterial identification (matrix-assisted laser desorption ionization–time of flight mass spectrometry [MALDI-TOF]) and susceptibility testing. Only 1 isolate per patient infection episode was included in the surveillance. All organisms were isolated from documented infections, including 4,667 (54.3%) community-acquired respiratory tract infections (3,124 *S. pneumoniae*, 930 *H. influenzae*, and 613 *M. catarrhalis* isolates), 2,036 (23.7%) pneumonia in hospitalized patients (276 *S. pneumoniae*, 1,585 *S. aureus*, 126 *H. influenzae*, and 49 *M. catarrhalis* isolates), 1,133 (13.2%) bloodstream infections (bacteremia; 370 *S. pneumoniae*, 737 *S. aureus*, 23 *H. influenzae*, and 3 *M. catarrhalis* isolates), 589 (6.9%) skin and skin structure infections (15 *S. pneumoniae*, 571 *S. aureus*, and 3 *H. influenzae* isolates), and 170 (2.0%) infections from other sites.

MICs for lefamulin and comparators were determined against *S. pneumoniae* ($n = 3,923$), *S. aureus* ($n = 2,919$), *H. influenzae* ($n = 1,086$), and *M. catarrhalis* ($n = 667$) strains from frozen-form MIC panels prepared by JMI Laboratories per Clinical and Laboratory Standards Institute (CLSI) broth microdilution methods (23). Susceptibility and comparator categorical interpretations used breakpoint criteria from CLSI M100-S28 and European Committee on Antimicrobial Susceptibility Testing (2018), where available (24, 25). Cation-adjusted Mueller-Hinton broth was used for testing *S. aureus* and *M. catarrhalis* and was supplemented with 2.5% to 5% lysed horse blood for *S. pneumoniae*. *Haemophilus* test medium was used for *H. influenzae*.

Lefamulin demonstrated potent *in vitro* activity against this large contemporary collection of bacterial pathogens that commonly cause CABP. Lefamulin at $\leq 1 \mu\text{g/ml}$, the proposed tentative susceptible breakpoint for *S. pneumoniae* based on clinical and nonclinical studies (22), inhibited 99.2% of all isolates tested, including 100% of *S. pneumoniae* isolates, 99.8% of *S. aureus* isolates, 93.8% of *H. influenzae* isolates, and 100% of *M. catarrhalis* isolates (Table 1). The activity of lefamulin against *S. pneumoniae*, the most frequently isolated bacterial CABP pathogen, was unaffected by resistance to other antibacterial classes, including β -lactams, fluoroquinolones, and macrolides as well as multidrug-resistant (MDR) or extensive drug-resistant strains; all MIC_{50/90} values were 0.06/0.12 $\mu\text{g/ml}$, except among levofloxacin nonsusceptible isolates (0.06/0.25 $\mu\text{g/ml}$) (Table 1). Lefamulin was among the most potent agents tested and had the lowest MIC_{50/90} values against MDR *S. pneumoniae* (0.06/0.12 $\mu\text{g/ml}$) (Table 1). Among comparators, *S. pneumoniae* isolates showed high rates of susceptibility ($\geq 98\%$) to

TABLE 1 Frequency of occurrence of lefamulin MICs for all pathogens tested

Organism (no. of isolates)	Cumulative % of isolates inhibited at lefamulin MIC ($\mu\text{g/ml}$) of:												MIC ₅₀ ($\mu\text{g/ml}$)	MIC ₉₀ ($\mu\text{g/ml}$)	
	≤ 0.008	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	>8			
<i>S. pneumoniae</i> (3,923)	0.1	1.8	11.4	55.1	93.7	99.6	99.9	100.0						0.06	0.12
Penicillin nonsusceptible, nonmeningitis ($\geq 4 \mu\text{g/ml}$) (189)	0.0	1.1	7.9	64.0	98.4	100.0								0.06	0.12
Ceftriaxone nonsusceptible ($\geq 2 \mu\text{g/ml}$) (155)	0.0	0.6	10.3	63.9	99.4	100.0								0.06	0.12
Erythromycin nonsusceptible ($\geq 0.5 \mu\text{g/ml}$) (1,348)	0.2	2.2	12.4	52.9	93.1	99.0	99.7	100.0						0.06	0.12
Levofloxacin nonsusceptible ($\geq 4 \mu\text{g/ml}$) (47)	0.0	8.5	23.4	68.1	89.4	97.9	97.9	100.0						0.06	0.25
MDR ^a (821)	0.4	2.9	15.8	61.1	96.3	99.8	99.8	100.0						0.06	0.12
XDR ^a (181)	0.0	0.6	7.2	64.6	98.9	100.0								0.06	0.12
<i>S. aureus</i> (2,919)			26.0	88.9	99.2	99.6	99.7	99.8	99.8	99.8	99.8	100.0	0.06	0.12	
Methicillin susceptible (1,981)			25.6	95.2	99.7	99.7	99.8	99.9	>99.9	>99.9	>99.9	100.0	0.06	0.06	
Methicillin resistant (938)			26.9	75.7	98.2	99.4	99.5	99.6	99.7	99.8	99.8	100.0	0.06	0.12	
<i>H. influenzae</i> (1,086)					1.7	20.4	69.4	93.8	99.1	99.9	100.0		0.5	1	
β -lactamase negative (835)					1.9	20.0	67.5	93.2	98.9	100.0			0.5	1	
β -lactamase positive (251)					1.2	21.9	75.7	96.0	99.6	99.6	100.0		0.5	1	
<i>M. catarrhalis</i> (667)	1.0	2.4	11.1	88.3	99.9	100.0							0.06	0.12	

^aMDR and XDR status was based on nonsusceptibility to ≥ 3 and ≥ 5 classes, respectively, of the following antimicrobial agents, as described by Golden et al. (30) and applying the following breakpoints: penicillin (MIC, $\geq 4 \mu\text{g/ml}$), ceftriaxone (MIC, $\geq 2 \mu\text{g/ml}$), erythromycin (MIC, $\geq 0.5 \mu\text{g/ml}$), clindamycin (MIC, $\geq 0.5 \mu\text{g/ml}$), levofloxacin (MIC, $\geq 4 \mu\text{g/ml}$), tetracycline (MIC, $\geq 2 \mu\text{g/ml}$), and trimethoprim-sulfamethoxazole (MIC, $\geq 1 \mu\text{g/ml}$). MDR, multidrug resistant; XDR, extensive drug resistant.

ceftaroline, levofloxacin, linezolid, moxifloxacin, and vancomycin (Table 2). Moderate resistance (range, 16.8% to 34.3%) was seen to erythromycin, azithromycin, tetracycline, trimethoprim-sulfamethoxazole, and clindamycin; among the 20.9% of MDR *S. pneumoniae* isolates, rates of resistance to these same agents were higher (range, 46.9% to 98.8%) (Table 2).

Lefamulin was also among the most active compounds against *S. aureus* (MIC_{50/90}, 0.06/0.12 $\mu\text{g/ml}$), including methicillin-resistant *S. aureus* (MRSA; MIC_{50/90}, 0.06/0.12 $\mu\text{g/ml}$) (Table 1 and 2). Among comparators, *S. aureus* isolates showed high rates of susceptibility ($\geq 95\%$) to ceftaroline, doxycycline, linezolid, trimethoprim-sulfamethoxazole, and vancomycin (Table 2). High resistance rates were observed among MRSA isolates, particularly to macrolides (azithromycin, 75.5% to 76.2%) and fluoroquinolones (levofloxacin, 72.3% to 74.0%) (Table 2).

Lefamulin displayed potent activity against *H. influenzae*, including β -lactamase-positive isolates (all MIC_{50/90}, 0.5/1 $\mu\text{g/ml}$). Compared with the Gram-positive pathogens, the overall MIC distribution of lefamulin shifted to higher MIC values as observed for macrolide antibiotics. However, most *H. influenzae* isolates (93.8%), including β -lactamase negative (93.2%) and β -lactamase positive (96.0%), were inhibited at concentrations of $\leq 1 \mu\text{g/ml}$ (Table 1). *H. influenzae* isolates showed high rates of susceptibility ($\geq 90\%$) to all comparator agents tested, with the exception of trimethoprim-sulfamethoxazole and ampicillin (Table 2). Resistance rates were high to trimethoprim-sulfamethoxazole (39.8% to 41.8%) and to ampicillin (99.6% to 100%) among β -lactamase-positive isolates, whereas resistance rates were lower among β -lactamase-negative isolates (trimethoprim-sulfamethoxazole, 28.3% to 30.3%; ampicillin, 2.9% to 9.3%).

All *M. catarrhalis* isolates were inhibited at lefamulin concentrations of $\leq 0.25 \mu\text{g/ml}$ (MIC_{50/90}, 0.06/0.12 $\mu\text{g/ml}$) (Table 1). *M. catarrhalis* isolates showed high rates of susceptibility (97.3% to 100%) to all comparators (Table 2). β -Lactamase activity was detected in 97% of tested isolates ($n = 335$).

Analysis of lefamulin results by infection type and by geographic region showed no

TABLE 2 Activity of lefamulin and comparators against pathogens commonly causing community-acquired bacterial pneumonia

Antibacterial agent by pathogen ^a (no. organisms tested)	MIC ₅₀ (μg/ml)	MIC ₉₀ (μg/ml)	MIC range (μg/ml)	Susceptibility rates (%) according to:					
				CLSI ^b			EUCAST ^b		
				S	I	R	S	I	R
<i>S. pneumoniae</i>									
Lefamulin (3,923)	0.06	0.12	≤0.008 to 1						
Amoxicillin-clavulanic acid (3,902)	≤0.03	2	≤0.03 to >4	93.6	2.9	3.5			
Azithromycin (3,921)	0.06	>4	≤0.03 to >4	65.7	0.7	33.6	65.3	0.4	34.3
Ceftaroline (3,905)	≤0.008	0.12	≤0.008 to >1	99.8			99.5		0.5
Ceftriaxone (3,905)	0.03	1	≤0.015 to >2	86.5	9.5	4.0 ^c	86.5	12.5	0.9
				96.0	3.0	0.9 ^d			
Clindamycin (3,906)	≤0.25	>1	≤0.25 to >1	82.7	0.5	16.8	83.2		16.8
Erythromycin (3,907)	0.03	>2	≤0.015 to >2	65.5	0.4	34.1	65.5	0.4	34.1
Levofloxacin (3,923)	1	1	≤0.12 to >4	98.8	0.1	1.1	98.8		1.2
Moxifloxacin (2,088 ^e)	0.12	0.25	≤0.03 to >4	98.9	0.5	0.6	98.8		1.2
Penicillin (3,923)	≤0.06	2	≤0.06 to >8	65.6	22.4	12.0 ^f	65.6		34.4 ^c
				65.6		34.4 ^g	65.6	29.5	4.8 ^d
				95.2	4.4	0.4 ^h			
Tetracycline (3,922)	≤0.25	>4	≤0.25 to >4	76.7	0.5	22.8	76.7	0.5	22.8
Trimethoprim-sulfamethoxazole (3,921)	≤0.5	>4	≤0.5 to >4	71.2	10.8	18.1	77.8	4.2	18.1
MDR <i>S. pneumoniae</i>									
Lefamulin (821)	0.06	0.12	≤0.008 to 1						
Amoxicillin-clavulanic acid (820)	1	>4	≤0.03 to >4	74.1	11.7	14.1			
Azithromycin (821)	>4	>4	0.015 to >4	1.2	1.1	97.7	1.0	0.2	98.8
Ceftaroline (821)	0.06	0.25	≤0.008 to >1	99.3			97.8		2.2
Ceftriaxone (821)	0.5	2	≤0.015 to >2	55.8	25.8	18.4 ^c	55.8	39.8	4.4
				81.6	14.0	4.4 ^d			
Clindamycin (821)	>1	>1	≤0.25 to >1	22.4	1.8	75.8	24.2		75.8
Erythromycin (821)	>2	>2	≤0.015 to >2	0.6	0.6	98.8	0.6	0.6	98.8
Levofloxacin (821)	1	1	0.25 to >4	96.5	0.2	3.3	96.5		3.5
Moxifloxacin (427 ^e)	0.12	0.25	≤0.03 to >4	97.2	1.6	1.2	97.0		3.0
Penicillin (821)	1	4	≤0.06 to >8	16.3	41.4	42.3 ^f	16.3		83.7 ^c
				16.3		83.7 ^g	16.3	62.1	21.6 ^d
				78.4	19.5	2.1 ^h			
Tetracycline (821)	>4	>4	≤0.25 to >4	4.5	1.1	94.4	4.5	1.1	94.4
Trimethoprim-sulfamethoxazole (821)	2	>4	≤0.5 to >4	32.9	20.2	46.9	45.3	7.8	46.9
<i>S. aureus</i>									
Lefamulin (2,919)	0.06	0.12	≤0.03 to >32						
Azithromycin (2,919)	0.5	>4	≤0.03 to >4	59.3	1.4	39.4	58.7	0.6	40.7
Ceftaroline (2,919)	0.25	1	≤0.06 to >8	95.0	4.8	0.1	95.0	4.8	0.1 ⁱ
							95.0		5.0 ^j
Clindamycin (2,919)	≤0.25	>2	≤0.25 to >2	85.8	0.2	14.0	85.5	0.3	14.2
Doxycycline (2,919)	≤0.06	0.25	≤0.06 to >8	97.9	2.0	0.1	95.1	0.9	4.0
Erythromycin (2,919)	0.25	>8	≤0.06 to >8	58.9	5.3	35.8	59.5	1.8	38.7
Levofloxacin (2,919)	0.25	>4	≤0.03 to >4	72.2	0.9	26.9	72.2		27.8
Linezolid (2,919)	1	1	≤0.12 to 8	>99.9		<0.1	>99.9		<0.1
Moxifloxacin (1,646 ^e)	≤0.06	4	≤0.06 to >4	73.4	7.5	19.1	73.0		27.0
Oxacillin (2,919)	0.5	>2	≤0.25 to >2	67.9		32.1	67.9		32.1
Trimethoprim-sulfamethoxazole (2,919)	≤0.5	≤0.5	≤0.5 to >4	98.2		1.8	98.2	0.2	1.6
Vancomycin (2,919)	0.5	1	≤0.12 to 2	100.0	0.0	0.0	100.0		0.0
MRSA									
Lefamulin (938)	0.06	0.12	≤0.03 to >32						
Azithromycin (938)	>4	>4	0.06 to >4	23.8	0.7	75.5	23.3	0.4	76.2
Ceftaroline (938)	1	2	0.25 to >8	84.5	15.0	0.4	84.5	15.0	0.4 ⁱ
							84.5		15.5 ^j
Clindamycin (938)	≤0.25	>2	≤0.25 to >2	62.0	0.0	38.0	61.9	0.1	38.0
Doxycycline (938)	≤0.06	2	≤0.06 to >8	94.5	5.3	0.2	89.4	1.4	9.2
Erythromycin (938)	>8	>8	≤0.06 to >8	23.3	4.7	72.0	23.8	1.2	75.1
Levofloxacin (938)	>4	>4	0.06 to >4	26.0	1.7	72.3	26.0		74.0
Linezolid (938)	1	1	≤0.12 to 2	100.0		0.0	100.0		0.0
Moxifloxacin (536 ^e)	2	>4	≤0.06 to >4	29.7	18.5	51.9	28.7		71.3
Oxacillin (938)	>2	>2	>2 to >2	0.0		100.0	0.0		100.0
Trimethoprim-sulfamethoxazole (938)	≤0.5	≤0.5	≤0.5 to >4	94.9		5.1	94.9	0.6	4.5
Vancomycin (938)	0.5	1	≤0.12 to 2	100.0	0.0	0.0	100.0		0.0

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TABLE 2 (Continued)

Antibacterial agent by pathogen ^a (no. organisms tested)	MIC ₅₀ (μg/ml)	MIC ₉₀ (μg/ml)	MIC range (μg/ml)	Susceptibility rates (%) according to:					
				CLSI ^b			EUCAST ^b		
				S	I	R	S	I	R
<i>H. influenzae</i>									
Lefamulin (1,086)	0.5	1	≤0.12 to 8						
Amoxicillin-clavulanic acid (1,086)	0.5	2	≤0.12 to >8	98.0		2.0	94.3		5.7
Ampicillin (1,086)	0.5	>8	0.12 to >8	69.7	5.1	25.2	69.7		30.3
Azithromycin (1,086)	1	1	0.12 to >4	98.8			98.8 ^c		
Cefepime (1,086)	0.06	0.25	≤0.015 to >2	99.8			96.6		3.4
Ceftriaxone (1,086)	≤0.015	≤0.015	≤0.015 to 0.5	100.0			98.5		1.5
Clarithromycin (1,086)	8	8	0.25 to >16	91.9	6.4	1.7	100.0 ^k		
Levofloxacin (1,086)	≤0.015	0.03	≤0.015 to >2	99.6			98.2		1.8
Moxifloxacin (550 ^e)	0.03	0.03	0.008 to >1	99.6			98.9		1.1
Tetracycline (1,086)	0.5	0.5	≤0.12 to >8	98.3	0.1	1.6	98.2	0.2	1.7
Trimethoprim-sulfamethoxazole (1,086)	0.12	>4	≤0.06 to >4	65.7	3.4	30.9	65.7	1.4	33.0
<i>M. catarrhalis</i>									
Lefamulin (667)	0.06	0.12	≤0.008 to 0.25						
Amoxicillin-clavulanic acid (667)	0.12	0.25	≤0.06 to 0.5	100.0		0.0	100.0		0.0
Azithromycin (662)	0.015	0.03	0.002 to 0.06	100.0			100.0	0.0	0.0
Ceftriaxone (667)	0.25	0.5	≤0.015 to 2	100.0			99.7	0.3	0.0
Clarithromycin (662)	≤0.12	≤0.12	≤0.12 to 0.25	100.0			100.0	0.0	0.0
Erythromycin (662)	0.12	0.12	≤0.015 to 1	100.0			98.9	0.8	0.3
Levofloxacin (667)	0.03	0.06	≤0.015 to 1	100.0			99.4		0.6
Moxifloxacin (221 ^e)	0.06	0.06	0.03 to 0.5				99.5		0.5
Tetracycline (667)	0.25	0.25	≤0.03 to 0.5	100.0	0.0	0.0	100.0	0.0	0.0
Trimethoprim-sulfamethoxazole (667)	0.12	0.25	≤0.06 to 2	97.3	2.7	0.0	97.3	2.1	0.6

^aMDR, multidrug resistant; MRSA, methicillin-resistant *S. aureus*.

^bCriteria as published by CLSI 2018 and EUCAST 2018. CLSI, Clinical and Laboratory Standards Institute; EUCAST, European Committee on Antimicrobial Susceptibility Testing; I, intermediate; R, resistant; S susceptible.

^cUsing meningitis breakpoints.

^dUsing nonmeningitis breakpoints.

^eMoxifloxacin was tested in 2016 but not in 2015.

^fUsing oral breakpoints.

^gUsing parenteral, meningitis breakpoints.

^hUsing parenteral, nonmeningitis breakpoints.

ⁱUsing other than pneumonia breakpoints.

^jUsing pneumonia breakpoints.

^kPercentage of wild type based on epidemiological cutoff value. EUCAST version 8.0 (2018).

apparent differences across isolate sources and across Asia-Pacific, Europe, Latin America, or North America.

Our results are consistent with previous studies reporting on the *in vitro* activity of lefamulin against typical and atypical respiratory pathogens (e.g., *M. pneumoniae*, *C. pneumoniae*, and *L. pneumophila*) (18, 19, 26, 27). Furthermore, the *in vitro* activity of lefamulin has translated to clinical efficacy in two phase 3 clinical trials in adults with CABP, demonstrating the noninferiority of 5 to 10 days of lefamulin versus 7 to 10 days of moxifloxacin given in intravenous-to-oral or oral administration (28, 29).

In conclusion, lefamulin was highly active against pathogens commonly causing CABP collected globally between 2015 and 2016, with activity consistent across geographic regions and unaffected by resistance to other antibacterial classes. These data support the ongoing clinical development of lefamulin for the treatment of CABP and other respiratory tract infections.

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REFERENCES

- Broulette J, Yu H, Pyenson B, Iwasaki K, Sato R. 2013. The incidence rate and economic burden of community-acquired pneumonia in a working-age population. *Am Health Drug Benefits* 6:494–503.
- Gibson J. 2013. The European lung white book: respiratory health and disease in Europe, 2nd ed. European Respiratory Society, Sheffield, United Kingdom.
- Centers for Disease Control and Prevention. 2015. Pneumococcal disease. In Hamborsky J, Kroger A, Wolfe S (ed), *Epidemiology and prevention of vaccine-preventable diseases*, 13th ed. Public Health Foundation, Washington, DC.
- File TM, Marrie TJ. 2010. Burden of community-acquired pneumonia in North American adults. *Postgrad Med* 122:130–141. <https://doi.org/10.3810/pgm.2010.03.2130>.
- Joya-Montosa C, Delgado-Amaya MD, Trujillo-Garcia E, Curiel-Balsera E. 2015. Assessment of specific risk scores for patients admitted to the ICU for severe community-acquired pneumonia. *Critical Care* 19:S3.
- Xu J, Murphy SL, Kochanek KD, Bastian B, Arias E. 2018. Deaths: final data for 2016, vol. 67, no. 5. National Center for Health Statistics, Hyattsville, MD.
- Jain S, Self WH, Wunderink RG, Fakhran S, Balk R, Bramley AM, Reed C, Grijalva CG, Anderson EJ, Courtney DM, Chappell JD, Qi C, Hart EM, Carroll F, Trabue C, Donnelly HK, Williams DJ, Zhu Y, Arnold SR, Ampofo K, Waterer GW, Levine M, Lindstrom S, Winchell JM, Katz JM, Erdman D, Schneider E, Hicks LA, McCullers JA, Pavia AT, Edwards KM, Finelli L, CDC EPIC Study Team. 2015. Community-acquired pneumonia requiring hospitalization among U.S. adults. *N Engl J Med* 373:415–427. <https://doi.org/10.1056/NEJMoa1500245>.
- Welte T, Torres A, Nathwani D. 2012. Clinical and economic burden of community-acquired pneumonia among adults in Europe. *Thorax* 67:71–79. <https://doi.org/10.1136/thx.2009.129502>.
- Mandell LA, Wunderink RG, Anzueto A, Bartlett JG, Campbell GD, Dean NC, Dowell SF, File TM, Jr, Musher DM, Niederman MS, Torres A, Whitney CG, Infectious Diseases Society of America, American Thoracic Society. 2007. Infectious Diseases Society of America/American Thoracic Society consensus guidelines on the management of community-acquired pneumonia in adults. *Clin Infect Dis* 44:S27–S72.
- File TM. 2009. The science of selecting antimicrobials for community-acquired pneumonia (CAP). *J Manag Care Pharm* 15:S5–S11.
- Peyrani P, Mandell L, Torres A, Tillotson GS. 2018. The burden of community-acquired bacterial pneumonia in the era of antibiotic resistance. *Expert Rev Respir Med* <https://doi.org/10.1080/17476348.2019.1562339>.
- Hess G, Hill JW, Raut MK, Fisher AC, Mody S, Schein JR, Chen CC. 2010. Comparative antibiotic failure rates in the treatment of community-acquired pneumonia: results from a claims analysis. *Adv Ther* 27:743–755. <https://doi.org/10.1007/s12325-010-0062-1>.
- Oster G, Berger A, Edelsberg J, Weber DJ. 2013. Initial treatment failure in non-ICU community-acquired pneumonia: risk factors and association with length of stay, total hospital charges, and mortality. *J Med Econ* 16:809–819. <https://doi.org/10.3111/13696998.2013.794805>.
- Llop CJ, Tuttle E, Tillotson GS, LaPlante K, File TM, Jr. 2017. Antibiotic treatment patterns, costs, and resource utilization among patients with community acquired pneumonia: a US cohort study. *Hosp Pract (1995)* 45:1–8. <https://doi.org/10.1080/21548331.2017.1279012>.
- U.S. Food and Drug Administration. 2016. FDA Drug Safety Communication: FDA updates warnings for oral and injectable fluoroquinolone antibiotics due to disabling side effects. U.S. Food and Drug Administration, Silver Spring, MD. <https://www.fda.gov/Drugs/DrugSafety/ucm511530.htm>.
- Eyal Z, Matzov D, Krupkin M, Paukner S, Riedl R, Rozenberg H, Zimmerman E, Bashan A, Yonath A. 2016. A novel pleuromutilin antibacterial compound, its binding mode and selectivity mechanism. *Sci Rep* 6:39004. <https://doi.org/10.1038/srep39004>.
- Paukner S, Riedl R. 2016. Pleuromutilins: potent drugs for resistant bugs—mode of action and resistance. In Silver L, Bush K (ed), *Antibiotics and antibiotic resistance*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- Waites KB, Crabb DM, Duffy LB, Jensen JS, Liu Y, Paukner S. 2017. *In vitro* activities of lefamulin and other antimicrobial agents against macrolide-susceptible and macrolide-resistant *Mycoplasma pneumoniae* from the United States, Europe, and China. *Antimicrob Agents Chemother* 61:e02008-16. <https://doi.org/10.1128/AAC.02008-16>.
- Sader HS, Paukner S, Ivezic-Schoenfeld Z, Biedenbach DJ, Schmitz FJ, Jones RN. 2012. Antimicrobial activity of the novel pleuromutilin antibiotic BC-3781 against organisms responsible for community-acquired respiratory tract infections (CARTIs). *J Antimicrob Chemother* 67:1170–1175. <https://doi.org/10.1093/jac/dks001>.
- Wicha WW, Prince WT, Lell C, Heilmayer W, Gelone SP. Pharmacokinetics and tolerability of lefamulin following intravenous and oral dosing. *J Antimicrob Chemother*, in press.
- Zeitlinger M, Schwameis R, Burian A, Burian B, Matzneller P, Muller M, Wicha WW, Strickmann DB, Prince W. 2016. Simultaneous assessment of the pharmacokinetics of a pleuromutilin, lefamulin, in plasma, soft tissues and pulmonary epithelial lining fluid. *J Antimicrob Chemother* 71:1022–1026. <https://doi.org/10.1093/jac/dkv442>.
- Bhavnani SM, Zhang L, Hammel JP, Rubino CM, Bader JC, Sader HS, Gelone SP, Wicha WW, Ambrose PG. Pharmacokinetic-pharmacodynamic target attainment analyses to support intravenous and oral lefamulin dose selection for the treatment of patients with community-acquired bacterial pneumonia. *J Antimicrob Chemother*, in press.
- Clinical and Laboratory Standards Institute. 2015. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard M07-A10, 10th ed. Clinical and Laboratory Standards Institute, Wayne, PA.
- Clinical and Laboratory Standards Institute. 2018. Performance standards for antimicrobial susceptibility testing; 28th informational supplement, M100Ed28E. Clinical and Laboratory Standards Institute, Wayne, PA.
- European Committee on Antimicrobial Susceptibility Testing. 2018. Breakpoint tables for interpretation of MICs and zone diameters, version 8.0. European Committee on Antimicrobial Susceptibility Testing, Växjö, Sweden. http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Breakpoint_tables/v_8.0_Breakpoint_Tables.pdf.
- Paukner S, Sader HS, Ivezic-Schoenfeld Z, Jones RN. 2013. Antimicrobial activity of the pleuromutilin antibiotic BC-3781 against bacterial pathogens isolated in the SENTRY Antimicrobial Surveillance Program in 2010. *Antimicrob Agents Chemother* 57:4489–4495. <https://doi.org/10.1128/AAC.00358-13>.
- Mendes RE, Farrell DJ, Flamm RK, Talbot GH, Ivezic-Schoenfeld Z, Paukner S, Sader HS. 2016. *In vitro* activity of lefamulin tested against *Streptococcus pneumoniae* with defined serotypes, including multidrug-resistant isolates causing lower respiratory tract infections in the United States. *Antimicrob Agents Chemother* 60:4407–4411. <https://doi.org/10.1128/AAC.00627-16>.
- File T, Goldberg L, Das A, Sweeney C, Saviski J, Gelone S, Seltzer E, Talbot G, Gasink L. 2018. Lefamulin is non-inferior to moxifloxacin in adults with community-acquired bacterial pneumonia (CABP): the phase 3 Lefamulin Evaluation Against Pneumonia (LEAP 1) study. *ASM Microbe*, Atlanta, GA.
- Alexander E, Goldberg L, Das A, Moran GJ, Sandrock C, Gasink LB, Spera P, Sweeney C, Paukner S, Wicha WW, Schranz J. 2018. Oral lefamulin is safe and effective in the treatment of adults with community-acquired bacterial pneumonia (CABP): results of Lefamulin Evaluation Against Pneumonia (LEAP 2) study, abstr 74297. *IDWeek*, San Francisco, CA.
- Golden AR, Rosenthal M, Fultz B, Nichol KA, Adam HJ, Gilmour MW, Baxter MR, Hoban DJ, Karlowitsky JA, Zhanel GG. 2015. Characterization of MDR and XDR *Streptococcus pneumoniae* in Canada, 2007–13. *J Antimicrob Chemother* 70:2199–2202. <https://doi.org/10.1093/jac/dkv107>.