

# Maraviroc Intensification Modulates Atherosclerotic Progression in HIV-Suppressed Patients at High Cardiovascular Risk. A Randomized, Crossover Pilot Study

Daniela Francisci,<sup>1,a</sup> Matteo Pirro,<sup>2,a</sup> Elisabetta Schiaroli,<sup>1</sup> Massimo R. Mannarino,<sup>2</sup> Sabrina Cipriani,<sup>1</sup> Vanessa Bianconi,<sup>2</sup> Alessia Alunno,<sup>3</sup> Francesco Bagaglia,<sup>2</sup> Onelia Bistoni,<sup>3</sup> Emanuela Falcinelli,<sup>4</sup> Loredana Bury,<sup>4</sup> Roberto Gerli,<sup>3,⊕</sup> Elmo Mannarino,<sup>2</sup> Raffaele De Caterina,<sup>5</sup> and Franco Baldelli<sup>1</sup>

Units of <sup>1</sup>Infectious Diseases, <sup>2</sup>Internal Medicine, Angiology and Arteriosclerosis, and <sup>3</sup>Rheumatology, and <sup>4</sup>Division of Internal and Cardiovascular Medicine, Department of Medicine, University of Perugia, Perugia, Italy; <sup>5</sup>Institute of Cardiology and Center of Excellence on Aging and Translational Medicine-CeSi-MeT, G.d'Annunzio University- Chieti-Pescara, Chieti, Italy

**Background.** Experimental CCR5 antagonism with maraviroc in atherosclerosis-prone mice and preliminary data in humans suggest an anti-atherosclerotic effect of the drug. We assessed the impact of maraviroc treatment in persons living with HIV on subclinical indicators of atherosclerosis.

**Methods.** Persons living with HIV on effective antiretroviral therapy (ART) including only protease inhibitors were recruited if they had a Framingham risk score >20% and brachial flow-mediated dilation (bFMD) <4%, as indices of high cardiovascular risk. Maraviroc (300 mg per os for 24 weeks) was administered, in addition to ongoing ART, to all patients using a crossover design. Brachial FMD, carotid-femoral pulse wave velocity (cfPWV), and carotid intima-media thickness (cIMT) were measured as markers of atherosclerosis. Vascular competence—as expressed by the ratio of circulating endothelial microparticles (EMPs) to endothelial progenitor cells (EPCs)—and markers of systemic inflammation and monocyte and platelet activation were assessed.

**Results.** Maraviroc treatment significantly improved bFMD, cfPWV, and cIMT by 66%, 11%, and 13%, respectively ( $P = .002$ ,  $P = .022$ ,  $P = .038$ , respectively). We also found a beneficial effect of maraviroc on the EMP/EPC ratio ( $P < .001$ ) and platelet/leucocyte aggregates ( $P = .013$ ). No significant changes in markers of systemic inflammation, monocyte activation, and microbial translocation were observed.

**Conclusions.** Maraviroc led to significant improvements in several markers for cardiovascular risk, endothelial dysfunction, arterial stiffness, and early carotid atherosclerosis, which was accompanied by an increase of vascular competence, without seeming to affect systemic inflammation. Our data support the need for larger studies to test for any effects of maraviroc on preventing atherosclerosis-driven pathologies.

**Keywords.** HIV; atherosclerosis; cardiovascular risk; maraviroc.

Atherosclerosis is an inflammatory disease in which immune mechanisms interact with metabolic risk factors to initiate, propagate, and activate lesion formation and progression [1]. In people living with HIV (PLWH), a residual chronic immune-inflammatory derangement typically persists despite complete viral suppression with ART [2, 3]. In addition, exposure to traditional cardiovascular (CV) risk factors does not completely explain the increased CV risk [4, 5], suggesting that additional conditions, such as chronic inflammation and immune

activation, might play roles in promoting CV diseases (CVD) in such patients [6]. Indeed, markers of inflammation, such as high-sensitivity C-reactive protein (hsCRP) and interleukin-6 (IL-6), can predict coronary events, stroke, and CVD mortality in individuals with or without HIV infection [7, 8].

The chemokine (C-C motif) ligand 5 (CCL5)/C-C chemokine receptor type 5 (CCR5) axis plays a pivotal role in the development and progression of atherosclerotic inflammatory disease [9]. Specifically, CCR5, a G-protein-coupled receptor of the CCL3/MIP-1 $\alpha$ , CCL4/MIP-1 $\beta$ , and CCL5/Regulated on Activation Normal T Cell Expressed and Secreted (RANTES) chemokines regulates the migration of monocytes, natural killer cells, and T helper-1 (Th1) cells into inflammation sites, including atherosclerotic plaques, thus promoting atherosclerosis progression [10]. CCR5 is also the co-receptor for macrophage tropic (R5) HIV-1 [11] and is the target of the CCR5 antagonist maraviroc (MVC), which is used for treating HIV infection [12]. A polymorphism in CCR5, a 32-nucleotide deletion known as CCR5  $\Delta$ 32, has been observed in subjects who remain uninfected despite extensive exposure to HIV-1 [13]

Received 24 September 2018; editorial decision 22 February 2019; accepted 1 March 2019.

<sup>a</sup>Equal contribution

Correspondence: F. Baldelli, MD, Clinica di Malattie Infettive, Università degli Studi di Perugia, Ospedale S. Maria della Misericordia, Piazza Menghini 1, 06129 Perugia ([franco.baldelli@unipg.it](mailto:franco.baldelli@unipg.it)).

## Open Forum Infectious Diseases®

© The Author(s) 2019. Published by Oxford University Press on behalf of Infectious Diseases Society of America. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs licence (<http://creativecommons.org/licenses/by-nc-nd/4.0/>), which permits non-commercial reproduction and distribution of the work, in any medium, provided the original work is not altered or transformed in any way, and that the work is properly cited. For commercial re-use, please contact [journals.permissions@oup.com](mailto:journals.permissions@oup.com) DOI: 10.1093/ofid/ofz112

and has also been associated with reduced risk for severe coronary artery disease (CAD) [14] and myocardial infarction in different clinical settings [15]. The CCR5 ligand CCL5 has been detected in atherosclerotic plaques and is mostly increased in late-stage atherosclerotic lesions [16], where it correlates to an unstable phenotype [17]. Moreover, CCL5 is released by activated degranulating platelets and can trigger shear-resistant monocyte arrest on the inflamed endothelium, therein favoring the early stages of atherosclerosis and its progression [18].

We have previously reported that MVC was able to reduce atherosclerotic progression in 2 different murine models by interfering with the recruitment of inflammatory cells. Moreover, in a ritonavir-treated experimental model, MVC modulated both the inflammatory plaque profile and systemic inflammation [19].

In light of such findings and considering that the safety profile of MVC has been deemed similar to placebo in trials on PLWH, we undertook a pilot study to evaluate the anti-atherosclerotic effectiveness of MVC intensification in suppressed patients at high CVD risk. Early atherosclerosis was investigated by measuring brachial flow-mediated dilation (bFMD), carotid-femoral pulse wave velocity (cfPWV), and carotid intima-media thickness (cIMT); in addition, markers of vascular competence, soluble markers of systemic inflammation, monocyte and platelet activation, and microbial translocation were measured as putative mediators of the anti-atherosclerotic effect of MVC.

## METHODS

### Patients

Patients were consecutively recruited at the Infectious Disease Clinic of Santa Maria della Misericordia Teaching Hospital, Department of Medicine, University of Perugia, from January 15, 2016, to July 15, 2017. Inclusion criteria were age  $\geq 50$  years, on treatment over the past 12 months with an effective protease inhibitor ART regimen (HIV RNA  $< 50$  copies/mL), CD4 T-cell counts  $> 300/\text{mm}^3$  over the past 6 months, a Framingham risk score  $> 20\%$ , and a bFMD  $< 4\%$ . Exclusion criteria included age  $> 70$  years, life expectancy  $< 12$  months, previously recorded platelet functional defects, or any history of chronic alcohol abuse. The study was approved by the Umbria Region Ethics Committee (CEAS registry No. 2166/14; Clinical Trial Registration, NCT 03402815). All patients provided written informed consent.

### Study Design

We performed a 48-week randomized (1:1) crossover study. After giving their informed consent, all patients with the above-mentioned characteristics were randomly allocated with an AB/BA crossover design to either maraviroc 300 mg/d plus ongoing ART for 24 weeks (A) or ongoing ART alone for 24 weeks (B) (Figure 1). At the end of the first 24-week period, all

patients were switched to the alternative arm. A washout period was not performed because of the short half-life of MVC relative to the length of the MVC intervention. At time 0 and every 12 weeks, a clinical check-up was performed for all patients, whereas noninvasive examinations of the studied markers for preclinical atherosclerosis were carried out every 24 weeks. The primary outcome with respect to treatment effect was the change in bFMD after 24 weeks. Secondary outcomes with respect to treatment were cIMT and cfPWV. Additional analyses were performed for endothelial progenitor cells (EPCs), endothelial microparticles (EMPs), soluble markers of systemic inflammation, monocyte and platelet activation, and microbial translocation.

Treating physicians and nurses were not blinded to either study week or treatment assignment, whereas bFMD, cIMT, and cfPWV operators were blinded to study week and treatment assignment.

At the end of the study (48th week), after a preliminary evaluation of any observed efficacy, MVC was again added to only arm A. At the 72nd week, an evaluation of only bFMD was carried out on both arms.

### Clinical and Demographic Characteristics

All data are detailed in the Supplementary Data.

### Blood Sampling and Laboratory Assays

Plasma HIV-RNA, CD4+ and CD8+ T cells, hsCRP, D-Dimer, IL-6, sDC14, sCD163, MCP-1, sVCAM, LBP, EPCs, EMPs, angiogenic T cells (Tangs), platelet microparticles (PMPs), and platelet-leukocyte complexes were evaluated in peripheral blood.

Details of blood samples and laboratory assays are provided in the Supplementary Data.

### Noninvasive Markers of Preclinical Atherosclerosis

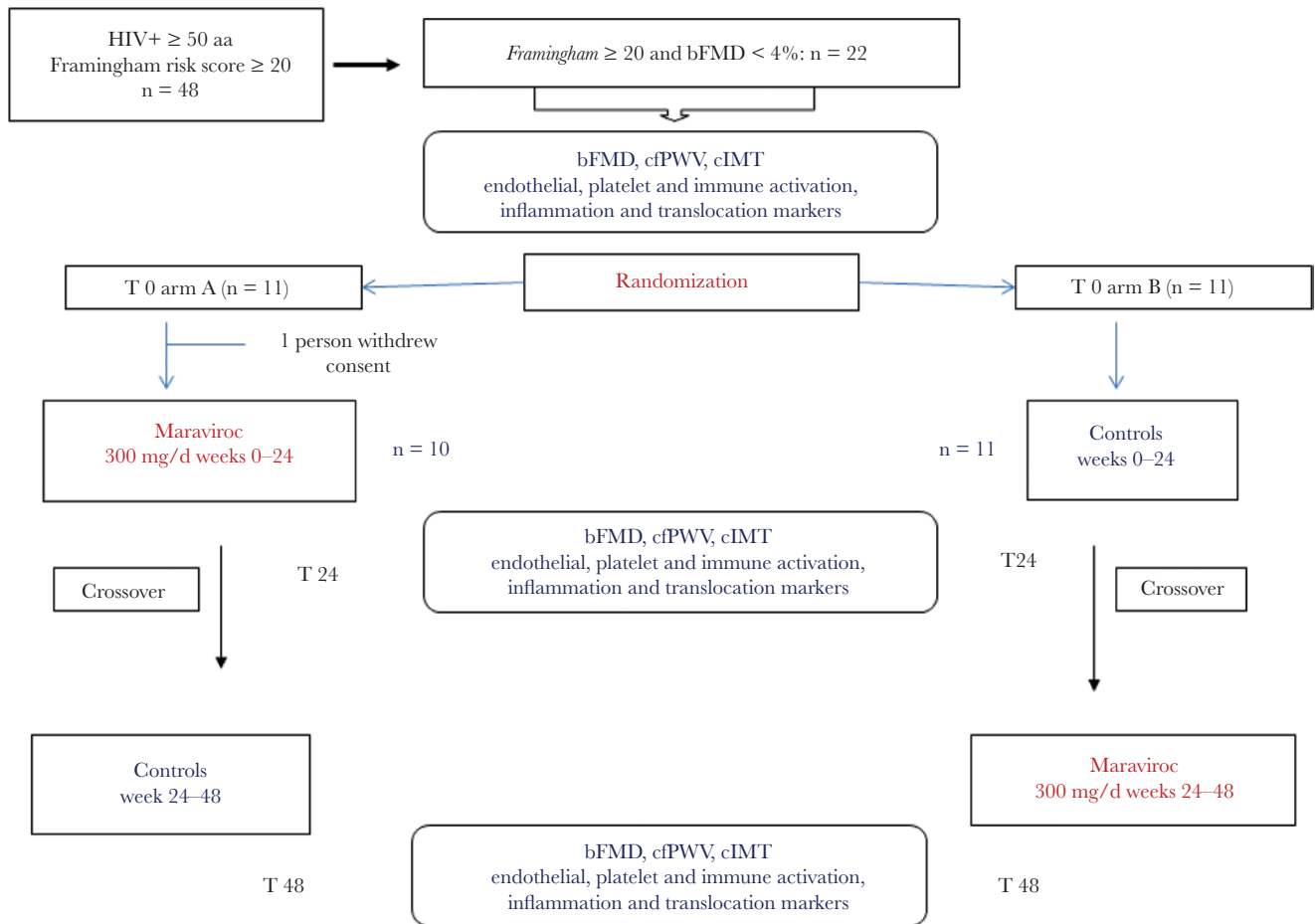
Brachial FMD on the nondominant arm cfPWV and cIMT was evaluated as previously described [20–22].

Details on bFMD, cfPWV, and cIMT measurement are provided in the Supplementary Data.

### Statistical Analysis

We planned a 2-arm crossover study, with restricted randomization (random permuted blocks and phone central randomization), with brachial FMD (primary outcome measure) as a continuous response variable. Random allocation sequence, enrollment of participants, and assignment of treatment were done by different study personnel.

Sample size was estimated assuming a baseline SD for bFMD of 2% and a post-treatment bFMD increase of 2%. A minimum sample size of 18 participants was required to detect statistically significant differences in bFMD with a power of 80% and an  $\alpha$  error of 5%. With a withdrawal/nonevaluable subject rate of 10%, a total of 20 subjects needed to be recruited.



**Figure 1.** Crossover study. The diagram shows screening eligibility, randomization, treatment interventions, and specific laboratory assay evaluations. Abbreviations: bFMD, brachial flow-mediated dilation; cfPWV, carotid-femoral pulse wave velocity; cIMT, carotid intima-media thickness.

We used the SPSS statistical package, release 17.0 (SPSS Inc, Chicago, IL), for all statistical analyses, with data analyzed anonymously and according to the originally assigned groups. Values are expressed as the mean and SD or the median and interquartile range, as appropriate. Base 10 logarithmic (log) transformation was performed for skewed variables, and log-values were used. An independent-sample *t* test and the Mann-Whitney *U* test were used to compare changes in the variables between the treatment orders (AB vs BA). Carryover was assessed by comparing the sum of the variable responses (response/1 + response/2) between the treatment orders (AB vs BA). Correlation analyses were performed using the Pearson's and Spearman's coefficients of correlation.

## RESULTS

### Patient Population

A total of 48 patients with Framingham risk scores  $\geq 20\%$  were screened for bFMD, and 22 underwent randomization. One patient withdrew his consent before MVC intensification and was excluded from the study. Patient demographic and clinical characteristics at

baseline are reported in [Table 1](#). At enrollment, patients had a long history of HIV infection (mean, 18 years), a mean CD4 T-cell nadir of  $149/\text{mm}^3$ , a median length of undetectable viral load of 5 years, a good control of HIV replication (with 90% of patients having  $<20$  copies/mL), and a good immunological status (76% of patients had  $>500$  CD4 T cell/ $\text{mm}^3$ , a mean CD4/CD8 ratio of 0.847). Twelve patients were taking boosted darunavir, 8 boosted atazanavir, and 1 boosted lopinavir. Three patients were taking abacavir too. Sixty-two percent were current smokers, 33% had a history of diabetes, and 43% were hypertensive (33% on antihypertensive therapy). Patients had, on average, borderline-high fasting glucose and triglyceride levels. Total and low-density lipoprotein cholesterol and creatinine clearance levels were in range.

Overall, baseline functional and structural vascular parameters, vascular competence markers, inflammation, platelet and monocyte activation indices, and microbial translocation parameters are reported in [Table 2](#).

### Effect of Maraviroc on Outcome Measures

Clinical and biochemical parameters before and after 24 weeks in both the treatment and control groups are reported

**Table 1. Demographic and Clinical Baseline Characteristics**

Baseline Characteristics	Total Patients (n = 21)
Male, No. (%)	20 (95)
Mean age (SD), y	61 (9)
Median BMI (IQR), kg/m <sup>2</sup>	26.87 (24.28–28.17)
Current smoking, No. (%)	13 (62)
Diabetes, No. (%)	7 (33)
Hypertension, No. (%)	9 (43)
History of CVD, No. (%)	5 (24)
Mean systolic blood pressure (SD), mmHg	127 (14)
Mean diastolic blood pressure (SD), mmHg	78 (9)
Mean blood glucose (SD), mg/dL	104 (34)
Mean total cholesterol (SD), mg/dL	197 (40)
Mean HDL cholesterol (SD), mg/dL	43 (10)
Mean LDL cholesterol (SD), mg/dL	108 (41)
Median triglycerides (IQR), mg/dL	172 (144–232)
Mean clearance creatinine (SD), mL/min	90 (17)
Mean known HIV (SD), y	17.81 (7.903)
Median nadir CD4 cell count, cells/mm <sup>3</sup> (IQR)	120 (81–260)
Median CD4 T-cell count, cells/mm <sup>3</sup> (IQR)	642 (497–852)
HIV-RNA <20 copies/mL, No. (%)	19 (90)
Boosted darunavir, No.	12
Boosted atazanavir, No.	8
Boosted lopinavir, No.	1
Abacavir, No.	3
Statin use, No. (%)	6 (28.5)
Acetylsalicylic acid treatment, No. (%)	5 (24)
Antihypertensive treatment, No. (%)	7 (33.3)

Abbreviations: BMI, body mass index; CVD, cardiovascular disease; HDL, high-density lipoprotein cholesterol; IQR, interquartile range; LDL, low-density lipoprotein cholesterol.

in Supplementary Table 1. No changes were observed in body mass index, current smoking habits, glucose, cholesterol, or triglyceride levels or in renal function within and between groups.

#### Noninvasive Markers of Preclinical Atherosclerosis

The treatment effect of MVC treatment weighted for control treatment is reported in Table 3. Noninvasive markers of atherosclerosis significantly improved with MVC treatment. Specifically, a 2.6% increase in bFMD was observed with MVC treatment ( $P = .002$ ). Of note, the median brachial artery diameter at 24 and 48 weeks did not change over 5% compared with baseline values, showing the reproducibility assessment of bFMD (at 24 weeks, 4.6; interquartile range [IQR], 4.20–4.93; at 48 weeks, 4.56; IQR, 4.11–4.9). A 1.0 m/s reduction in cFPWV was observed ( $P = .022$ ). A significant reduction in cIMT max of about 13% was observed in the between-group evaluation ( $-90 \mu\text{m}$ ;  $P = .038$ ). The carryover effects were not significant ( $P > .1$ ) for the above investigated markers.

Extending the evaluation of FMD, we observed a consistent bFMD improvement for arm A, which had resumed the drug therapy from weeks 48 to 72, whereas a worsening was observed for arm B, which had discontinued it at the 48th week (Supplementary Figure 1).

**Table 2. Baseline Markers of Inflammation, Platelet and Monocyte Activation, Microbial Translocation, Vascular Homeostasis, and Preclinical Atherosclerosis**

Variables	Baseline Median Value (IQR)
hsCRP, mg/L	1.81 (1.02–3.56)
IL6, pg/mL	3.8 (2.6–6.0)
D-Dimer, ng/mL	116.0 (74.0–156.0)
sCD14, ng/mL	5815 (5024–6723)
sCD163, ng/mL	2009 (1166–2287)
MCP-1, pg/mL	173.4 (147.3–286.2)
LBP, ng/mL	18 544 (13 458–22 353)
sVCAM, ng/mL	936.6 (653.1–1474.0)
Platelets/leukocyte aggregates, %	15.2 (11.0–20.8)
Platelet-derived microparticles, n/ $\mu\text{L}$	60 481 (28 686–88 547)
EMP, n/ $\mu\text{L}$	362.0 (310.0–606.5)
EPC, n/mL	130.6 (75.4–215.3)
Tang, n/mL	597.7 (402.6–934.4)
Brachial artery diameter, mm	4.73 (4.21–4.95)
bFMD, %	3.1 (1.7–4.9)
cfPWV, m/s	9.0 (7.1–9.5)
cIMT max, mm	0.690 (0.605–0.755)

Abbreviations: bFMD, brachial flow-mediated dilation; cFPWV, carotid-femoral pulse wave velocity; cIMT max, maximum carotid intima-media thickness; EMP, endothelial microparticle; EPC, endothelial microparticle; hsCRP, high-sensitivity C-reactive protein; IL6, interleukin-6; IQR, interquartile range; LBP, lipopolysaccharide binding protein; MCP, monocyte chemoattractant protein 1; sVCAM, soluble vascular cell adhesion molecule-1; Tang, angiogenic T cell.

#### Markers of Inflammation, Microbial Translocation, Endothelial Homeostasis, and Platelet and Monocyte Activation

A significant decrease in the EMP/EPC ratio was observed ( $-0.22$ ,  $P < .001$ ). No modifications in Tang counts were seen. Also, significant reductions in platelet/leukocytes aggregates ( $-3.2\%$ ,  $P < .001$ ) were observed. No significant changes, either for the maraviroc group or the cumulative analysis, were observed for the other investigated inflammatory markers and immune activation parameters.

#### DISCUSSION

We had previously reported that MVC reduced the atherosclerosis burden by modulating the inflammatory plaque recruitment in 2 ApoE knockout mice models with either early ritonavir-induced atherogenesis or late spontaneous atherosclerotic progression. Moreover, MVC reversed ritonavir-induced systemic inflammation in the former but not in the latter model [19]. In light of these results, a comprehensive evaluation of any potential anti-atherosclerotic effect associated with MVC in patients at high CV risk was warranted.

In this study, we observed significant improvements for all the surrogate noninvasive markers of early atherosclerosis in the maraviroc-treated patients, without any reductions of plasma cholesterol levels or any modifications for markers of systemic inflammation and immune activation.

Virally suppressed PLWH at high CV risk (Framingham risk score  $\geq 20\%$ ) with evidence of impaired bFMD at baseline and

**Table 3. Treatment Effect of Maraviroc, Weighted for Control Treatment, on Markers of Inflammation, Platelet and Monocyte Activation, Microbial Translocation, Vascular Homeostasis, and Preclinical Atherosclerosis**

Variables	Mean Change, SD	P
hsCRP, mg/L	-6.39 ± 29	.888
IL6, pg/mL	-5.0 ± 24	.324
D-Dimer, ng/mL	1.3 ± 32	.525
sCD14, ng/mL	786 ± 3924	.944
sCD163, ng/mL	63.0 ± 624	.231
MCP-1, pg/mL	-20.6 ± 102	.101
LBP, ng/mL	-1577 ± 7445	.573
sVCAM, ng/mL	90.3 ± 389	.491
Platelets/leukocyte aggregates, %	-3.2 ± 4.6	.013
Platelet-derived microparticles, n/μL	-14765 ± 35573	.132
EMP, n/μL	-311 ± 472	<.001
EPC, n/mL	40 ± 192	.014
EMP/EPC ratio	-0.22	<.001
Tang, n/mL	13 ± 474	.833
Brachial artery diameter, mm	0.02 ± 0.22	.600
bFMD, %	2.6 ± 3.0	.002
cfPWV, m/s	-1.0 ± 1.6	.022
cIMT, mm	-0.09 ± 0.18	.038

Change was measured as the difference of the variable responses in the treatment orders as follows: (response/2 – response/1) in the AB order and (response/1 – response/2) in the BA order.

Abbreviations: bFMD, brachial flow-mediated dilation; cfPWV, carotid-femoral pulse wave velocity; cIMT, carotid intima-media thickness; EMP, endothelial microparticle; EPC, endothelial microparticle; hsCRP, high-sensitivity C-reactive protein; IL6, interleukin-6; LBP, lipopolysaccharide binding protein; MCP, monocyte chemoattractant protein 1; sVCAM, soluble vascular cell adhesion molecule-1; Tang, angiogenic T cell.

taking only protease inhibitor–boosted regimens, thus allowing once-daily maraviroc administration, were enrolled. Indeed, maraviroc was not administered to control viral replication but to evaluate anti-atherosclerotic effectiveness. Moreover, in real life, once-daily MVC with boosted protease inhibitors (PIs) has been often administered, although the approved dosage was 150 mg twice daily in people taking PIs.

Sixty-three percent of these were current smokers, and 43% and 33% had a history of hypertension and diabetes, respectively. Moreover, 47.6% had plasma hsCRP levels  $\geq 2$  mg/L, and 61.9% had an IL-6  $>3.01$  pg/mL; the latter was associated with a CV risk over 12% at 48 months according to the SMART study results [8]. In our study population, we investigated for any impact from MVC use on several markers of early atherosclerosis (ie, endothelial dysfunction, aortic stiffness, cIMT). Additionally, markers of systemic inflammation (hsCRP, IL-6, MCP-1, LBP, sCD163, and sCD14), endothelial homeostasis (circulating EMPs, EPCs, and VCAM levels), platelet reactivity (PMPs and platelet-leukocyte aggregates), and microbial translocation were also investigated.

We found that adding MVC to the ART regimens of patients with viral suppression improved bFMD, cfPWV, and cIMT by 66%, 11%, and 13%, respectively. Moreover, the EMP/EPC ratio, which is considered a marker of vascular competence [22, 23], and platelet/leukocyte aggregates were significantly reduced.

Throughout the study, neither changes in lifestyle, in particular, current smoking, nor reductions in lipid levels were observed. After MVC intensification, hsCRP, IL-6, d-Dimer, sCD14, sCD163, and MCP-1 did not change, whereas microbial translocation was blunted without reaching statistical significance.

To date, evidence regarding the impact of MVC intensification on surrogate markers of atherosclerosis has been limited [24, 25]. Krikke et al. [24] have reported a mild improvement in endothelial function following MVC intensification in patients with HIV on abacavir. This improvement persisted after MVC discontinuation. However, the duration of this crossover study (16 weeks) was shorter than ours. Piconi et al. [25] have reported that MVC improved cfPWV, cIMT, IL-6, microbial translocation, and VCAM levels in 6 PLWH with a Framingham risk score of 10%–20%. However, the small sample size and the retrospective enrollment of control subjects might have biased the results. Conversely, a preliminary report by Hsue et al. did not provide evidence of a clinically relevant effect of MVC intensification on bFMD in persons living with HIV on treatment [26]; however, the characteristics of the recruited population were not fully described [26].

From a pathophysiological perspective, the beneficial effects of 24-week MVC intensification that we observed on bFMD, cfPWV, and cIMT are supported by several lines of evidence.

First, brachial FMD is a reliable indicator of endothelial function and a significant predictor of CV risk [27]. Endothelial fragmentation into microparticles and a reduced number of circulating EPCs have both been associated with impaired bFMD and increased CV risk [28–30]. Hence, the observed improvement of bFMD following MVC intensification was related to its positive impact on the balance between endothelial injury and repair. Given that nitric oxide (NO) is involved in the regulation of endothelial function and has been associated with both reduced endothelial cell injury and higher circulating EPC counts [23, 30], it is plausible that increased NO bioavailability could have mediated our observed MVC-induced improvements in endothelial function and overall vascular structural integrity.

Second, cfPWV has been reported to be influenced by NO bioavailability [31] and by endothelial function as well. Despite elastic fiber degeneration and a reduced elastin/collagen ratio being key factors in inducing arterial stiffening [32], endothelial dysfunction and reduced NO bioavailability may promote arterial stiffening by modulation of vascular smooth muscle cell relaxation and arterial tone [31, 33]. As elastic fiber degeneration and the elastin/collagen ratio are rather difficult to revert [32], the rapid improvement observed in our study in cfPWV is more likely attributable to an endothelium-dependent NO-mediated effect of MVC on vascular tone.

Third, several studies have reported a time-dependent cIMT progression in persons living with HIV [34] similar to what we observed in our control subjects; a 20- $\mu$ m progression of cIMT

was seen over a 24-week period. Notably, in our study, cIMT was reduced by 60  $\mu\text{m}$  with MVC intensification. This result is in agreement with a study exploring the effect of CCR5 inhibition by MVC in persons with HIV/HCV coinfection [35]. In that study, a 48-week intensification with maraviroc was associated with a reduction in atherosclerotic plaque progression. Improvements in endothelial function and vascular competence might mediate the beneficial effects of maraviroc on cIMT, as both reduced bFMD and increased EMP/EPC ratio have been associated with early atherosclerosis in the carotid arteries [22, 36].

Several hypotheses may further explain the observed maraviroc-induced efficacy on major markers of early atherosclerosis. First, an endothelium-protective impact from CCR5 inhibition might be involved. This is suggested by a recent *in vitro* study reporting that MVC incubation with coronary artery endothelium resulted in inhibiting vasoconstriction and stimulation of intimal hyperplasia [37]. Moreover, CCR5 antagonism can downregulate inflammatory cell recruitment into vascular walls, leading to a further improvement in major markers of atherosclerosis. This local anti-inflammatory effect with MVC was also observed in our previous study, where we reported a 50% reduction in plaque monocyte/macrophage CCR5+ infiltration following MVC treatment in animal models [19]. Hence, we hypothesize that there is a bidirectional causative association between endothelial injury and arterial wall inflammation [38]; if so, MVC treatment might play a key role in interfering with this pro-atherogenic loop.

Strengths of our study include (1) its homogeneous population of persons living with HIV and near-complete ART-induced viral suppression; (2) its design, which measured several recognized major early indicators for atherosclerotic risk and putative biochemical and cellular mediators within the atherosclerotic process; (3) its observation period of up to 72 weeks; and (4) the widely documented safety of MVC in HIV-infected patients.

We recognize several limitations. First, the sample size was small, although sufficiently powered to reach statistical significance, qualifying this as a pilot, hypothesis-generating study. Second, our crossover design did not include a washout period. This decision was made based on the following: MVC's half-life is about 16 hours, and the study treatment period lasted 24 weeks. Of note, a carryover effect was evaluated and found not to be significant. Third, we did not perform a direct evaluation of vascular wall inflammation or a direct measure of NO bioavailability, thus limiting our ability to fully interpret the results. Fourth, we did not evaluate any markers of oxidative stress, which may have contributed to the observed anti-atherosclerotic effects of MVC. Fifth, we did not investigate endothelial progenitor cell homing. In fact, the observed trend toward an increased EPC count following MVC intensification does not necessarily imply active participation of these cells in endothelial repair. It

has been reported that CCR5 expressed by EPCs facilitates EPC recruitment and exerts anti-atherosclerotic effects in ApoE<sup>-/-</sup> mice [39]. Hence, maraviroc-induced CCR5 inhibition might contribute to an increased circulating availability of EPCs due to reduced vascular homing. Finally, only PI-treated individuals were included in our study; thus these results cannot be generalized to PLWH treated with non-PI-containing regimens.

In conclusion, in this pilot study of persons living with HIV, treated with PIs, and having complete viral suppression, but at a high risk of CVD, MVC intensification was associated with significant and consistent improvements in several major indicators of increased CV risk, namely EMP/EPC ratio, endothelial dysfunction, arterial stiffness, and cIMT.

The nondocumented interference of MVC in systemic inflammation and its widely documented tolerability may support its use as an off-label anti-atherosclerotic therapy, as drugs targeting systemic inflammation can be burdened by higher incidence of serious fatal infections [40]. Further investigations with MVC are warranted in larger samples of patients treated also with other antiretroviral drugs or even individuals without HIV infection but at high risk of cardiovascular diseases.

#### Supplementary Data

Supplementary materials are available at *Open Forum Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

#### Acknowledgments

We would like to thank Thomas Charles Kilcline for his important editorial assistance, Verena De Angelis for her expert advice on the statistical analysis, and both Patrizia Sforza and Sabrina Bastianelli for technical support.

**Financial support.** This work was supported by Fondazione Cassa di Risparmio di Perugia, Italy (Award No. 2014.0339011 to F.B.).

**Potential conflicts of interest.** All authors: no reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

#### References

1. Ross R. Atherosclerosis—an inflammatory disease. *N Engl J Med* **1999**; 340:115–26.
2. Deeks SG. HIV infection, inflammation, immunosenescence, and aging. *Annu Rev Med* **2011**; 62:141–55.
3. Wada NI, Jacobson LP, Margolick JB, et al. The effect of HAART-induced HIV suppression on circulating markers of inflammation and immune activation. *AIDS* **2015**; 29:463–71.
4. Freiberg MS, Chang CC, Kuller LH, et al. HIV infection and the risk of acute myocardial infarction. *JAMA Intern Med* **2013**; 173:614–22.
5. Post WS, Budoff M, Kingsley L, et al. Associations between HIV infection and subclinical coronary atherosclerosis. *Ann Intern Med* **2014**; 160:458–67.
6. Hsue PY, Deeks SG, Hunt PW. Immunologic basis of cardiovascular disease in HIV-infected adults. *J Infect Dis* **2012**; 205(Suppl 3):S375–82.
7. Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med* **2005**; 352:1685–95.
8. Duprez DA, Neuhaus J, Kuller LH, et al; INSIGHT SMART Study Group. Inflammation, coagulation and cardiovascular disease in HIV-infected individuals. *PLoS One* **2012**; 7:e44454.
9. Jones KL, Maguire JJ, Davenport AP. Chemokine receptor CCR5: from AIDS to atherosclerosis. *Br J Pharmacol* **2011**; 162:1453–69.
10. Charo IF, Ransohoff RM. The many roles of chemokines and chemokine receptors in inflammation. *N Engl J Med* **2006**; 354:610–21.

11. Dragic T, Litwin V, Allaway GP, et al. HIV-1 entry into CD4+ cells is mediated by the chemokine receptor CC-CKR-5. *Nature* **1996**; 381:667–73.
12. Jones KL, Maguire JJ, Davenport AP. Chemokine receptor CCR5: from AIDS to atherosclerosis. *Br J Pharmacol* **2011**; 162:1453–69.
13. Huang Y, Paxton WA, Wolinsky SM, et al. The role of a mutant CCR5 allele in HIV-1 transmission and disease progression. *Nat Med* **1996**; 2:1240–3.
14. Szalai C, Duba J, Prohászka Z, et al. Involvement of polymorphisms in the chemokine system in the susceptibility for coronary artery disease (CAD). Coincidence of elevated Lp(a) and MCP-1 -2518 G/G genotype in CAD patients. *Atherosclerosis* **2001**; 158:233–9.
15. Rodríguez-Rodríguez L, González-Juanatey C, García-Bermúdez M, et al. CCR5Δ32 variant and cardiovascular disease in patients with rheumatoid arthritis: a cohort study. *Arthritis Res Ther* **2011**; 13:R133.
16. Wilcox JN, Nelken NA, Coughlin SR, et al. Local expression of inflammatory cytokines in human atherosclerotic plaques. *J Atheroscler Thromb* **1994**; 1(Suppl 1):S10–3.
17. Herder C, Peeters W, Illig T, et al; CARDIoGRAM Consortium. RANTES/CCL5 and risk for coronary events: results from the MONICA/KORA Augsburg case-cohort, Athero-Express and CARDIoGRAM studies. *PLoS One* **2011**; 6:e25734.
18. von Hundelshausen P, Weber KS, Huo Y, et al. RANTES deposition by platelets triggers monocyte arrest on inflamed and atherosclerotic endothelium. *Circulation* **2001**; 103:1772–7.
19. Cipriani S, Francisci D, Mencarelli A, et al. Efficacy of the CCR5 antagonist maraviroc in reducing early, ritonavir-induced atherogenesis and advanced plaque progression in mice. *Circulation* **2013**; 127:2114–24.
20. Pirro M, Schillaci G, Romagno PF, et al. Influence of short-term rosuvastatin therapy on endothelial progenitor cells and endothelial function. *J Cardiovasc Pharmacol Ther* **2009**; 14:14–21.
21. Pirro M, Schillaci G, Mannarino MR, et al. Circulating immature osteoprogenitor cells and arterial stiffening in postmenopausal osteoporosis. *Nutr Metab Cardiovasc Dis* **2011**; 21:636–42.
22. Pirro M, Stingeni L, Vaudo G, et al. Systemic inflammation and imbalance between endothelial injury and repair in patients with psoriasis are associated with preclinical atherosclerosis. *Eur J Prev Cardiol* **2015**; 22:1027–35.
23. Sabatier F, Camoin-Jau L, Anfosso F, et al. Circulating endothelial cells, microparticles and progenitors: key players towards the definition of vascular competence. *J Cell Mol Med* **2009**; 13:454–71.
24. Krikke M, Tesselaar K, Arends JE, et al. Maraviroc intensification improves endothelial function in abacavir-treated patients, an open-label randomized cross-over pilot study. *Infect Dis Ther* **2016**; 5:389–404.
25. Piconi S, Pocaterra D, Rainone V, et al. Maraviroc reduces arterial stiffness in PI-treated HIV-infected patients. *Sci Rep* **2016**; 6:28853.
26. Hsue P, Scherzer R, Gilman L, et al. Effect of maraviroc intensification on endothelial function in treated HIV infection. Abstract 123. In: 19th Conference on Retroviruses and Opportunistic Infections; March 5–8, **2012**; Seattle, WA.
27. Ghiadoni L, Salvetti M, Muiesan ML, Taddei S. Evaluation of endothelial function by flow mediated dilation: methodological issues and clinical importance. *High Blood Press Cardiovasc Prev* **2015**; 22:17–22.
28. Sinning JM, Losch J, Walenta K, et al. Circulating CD31+/Annexin V+ microparticles correlate with cardiovascular outcomes. *Eur Heart J* **2011**; 32:2034–41.
29. Werner N, Kosiol S, Schiegl T, et al. Circulating endothelial progenitor cells and cardiovascular outcomes. *N Engl J Med* **2005**; 353:999–1007.
30. Mannarino E, Pirro M. Endothelial injury and repair: a novel theory for atherosclerosis. *Angiology* **2008**; 59:69S–72S.
31. Bellien J, Favre J, Jacob M, et al. Arterial stiffness is regulated by nitric oxide and endothelium-derived hyperpolarizing factor during changes in blood flow in humans. *Hypertension* **2010**; 55:674–80.
32. Wagenseil JE, Mecham RP. Elastin in large artery stiffness and hypertension. *J Cardiovasc Transl Res* **2012**; 5:264–73.
33. Kinlay S, Creager MA, Fukumoto M, et al. Endothelium-derived nitric oxide regulates arterial elasticity in human arteries in vivo. *Hypertension* **2001**; 38:1049–53.
34. Longenecker CT, Sattar A, Gilkeson R, McComsey GA. Rosuvastatin slows progression of subclinical atherosclerosis in patients with treated HIV infection. *AIDS* **2016**; 30:2195–203.
35. Maggi P, Bruno G, Perilli F, et al. Effects of therapy with maraviroc on the carotid intima media thickness in HIV-1/HCV co-infected patients. *In Vivo* **2017**; 31:125–31.
36. Juonala M, Viikari JS, Laitinen T, et al. Interrelations between brachial endothelial function and carotid intima-media thickness in young adults: the cardiovascular risk in Young Finns Study. *Circulation* **2004**; 110:2918–23.
37. Maguire JJ, Jones KL, Kuc RE, et al. The CCR5 chemokine receptor mediates vasoconstriction and stimulates intimal hyperplasia in human vessels in vitro. *Cardiovasc Res* **2014**; 101:513–21.
38. De Caterina R, Massaro M, Scoditti E, Annunziata Carluccio M. Pharmacological modulation of vascular inflammation in atherothrombosis. *Ann N Y Acad Sci* **2010**; 1207:23–31.
39. Zhang Z, Dong J, Lobe CG, et al. CCR5 facilitates endothelial progenitor cell recruitment and promotes the stabilization of atherosclerotic plaques in ApoE<sup>-/-</sup> mice. *Stem Cell Res Ther* **2015**; 6:36.
40. Ridker PM, Everett BM, Thuren T, et al; CANTOS Trial Group. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med* **2017**; 377:1119–31.