



Alternaria yunnanensis sp. nov., a New *Alternaria* Species Causing Foliage Spot of Rubber Tree in China

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ABSTRACT

A new species of *Alternaria* causing leaf spots on the rubber tree (*Hevea brasiliensis*) in Yunnan, China, was isolated, examined, and illustrated. Morphologically, it belongs to the section *Porri* of *Alternaria*, which produces relatively large conidia and a simple or branched, filamentous long beak. It is, however, characterized by conidiophores gradually enlarging near the apex into a clavate conidiogenous cell and long ellipsoid to obclavate, smooth-walled conidia with a long filamentous beak. Molecular phylogenetic analyses based on *ITS* rDNA, *GAPDH*, and *TEF1-alpha* sequences demonstrate that the phytopathogen falls in the clade of the section *Porri*, being most closely related to *A. sidae*, *A. sennae*, *A. deseritcola*, *A. cyamopsisidis*, *A. rostellata*, *A. nitrimali*, *A. crassa*, and *A. thunbergiae*.

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1. Introduction

The *Alternaria* is saprobic and ubiquitous, and some species are well-known as major plant pathogens. They can also cause many human health disorders, while many can play an important role in decay and decomposition [1]. *Alternaria* was divided into 24 sections on the basis of morphological and multi-loci molecular phylogeny (18S, *ITS* and 28S rDNA, *GAPDH*, *RPB2*, and *TEF1-alpha*) [2–5]. The section *Porri* is the largest *Alternaria* section with species characterized by medium to large conidia with a simple or branched, filamentous long beak [2,3]. This section includes some important phytopathogens, such as *A. porri*, *A. bataticola*, *A. solani*, and *A. tomatophila*. *A. porri* causes purple blotch of onion [6], *A. bataticola* causes leaf petiole and stem blight of sweet potato [7], *A. solani* is the causative agent of early blight of potato [8], and *A. tomatophila* is known for causing early blight of tomato [9].

Earlier Martin [10] reported *Alternaria* sp. infection on *Hevea brasiliensis* in Mexico. In 2006, another *Alternaria* leaf blight with similar symptom on rubber trees caused by *A. alternata* was identified by Roy et al. [11] in India. Our research group reported that *A. heveae* and *A. alternata* were able to incite black leaf spot of rubber tree in China [12,13]. The disease symptoms initially appear as minute spots on leaves. The spots are circular with

black center surrounded by a yellow halo, and lesions slightly sunken.

During a survey on rubber tree diseases, a fungus isolated from rubber tree leaf spot lesions was morphologically identified to be an *Alternaria* species in the section *Porri* but which did not fit any known species of this genus and further confirmed molecularly by grouping within the section of *Porri*. Herein, we propose this new species with morphological description, illustration, and molecular phylogeny.

2. Materials and methods

2.1. Isolation and cultures

Fungus-infected leaves of the rubber tree were collected in May 2014 from Hula village, Ruili city, Yunnan Province, China. Segmented leaf lesions between symptomatic and healthy tissue were surface sterilized with 70% ethanol for 2 min and then air-dried, plated on potato dextrose agar (PDA; 20 g white potato boiled and filtered, 20 g dextrose, 15 g agar, and 1 L distilled water), and incubated at 28 °C with natural day/night cycles of light for 5–7 days. Single spores were isolated by following the method of Ho and Ko [14] and cultured and transferred onto a new PDA plate to establish pure cultures.

Isolates were maintained at the Herbarium of Department of Plant Pathology (YITC 5109 and YITC

5114), Yunnan institute of Tropic Crops. Ex-type was stored in the Agricultural Culture Collection of China, Institute of Plant Protection, Chinese Academy of Agricultural Sciences (ACCC 39327) and also in the China General Microbial Culture Collection, Institute of Microbiology, Chinese Academy of Sciences, Beijing, China (CGMCC3.1890). The type was deposited in the Herbarium Mycologicum Academiae Sinicae, Institute of Microbiology, Chinese Academy of Sciences, Beijing, China (HMAS247784).

2.2. Pathogenicity assay

Pathogenicity assays using a field rubber tree inoculation method were conducted as previously described by Cai et al. [13]. Fungal conidia were harvested by flooding 7-day-old cultures with sterile water from single conidial cultures, centrifuging, and adjusting the concentration to 1×10^4 conidia/ml, and spraying the spore suspension to the copper-color leaves (approximately 5–7 cm in length) of rubber tree using manual pressure sprayer. Rubber plants sprayed with sterile distilled water were used as controls. After inoculation, the plants were covered with plastic bags for 48 h. The plastic bags were removed after 2 days postinoculation and monitored daily for symptom development.

2.3. Morphological observations

The isolate YITC 5109 was used for morphological descriptions. Colonial characteristics (color, size, and texture) were assessed after 7 days of growth on the PDA plates and V-8 juice agar (V-8; 100 mL V-8 juice, 0.2 g CaCO₃, 20 g agar, and 900 mL distilled water) at 25 °C under a fluorescent light/dark cycle of 8/16 h. Conidial morphology was examined according to the method of Simmons [15,16]. Isolates were inoculated on potato carrot agar (PCA; 20 g white potato boiled and filtered, 20 g carrot, and 20 g agar in 1 L distilled water) and V-8 juice agar. The plates were stored at 22 °C for 7 days in a chamber without humidity control (a gradually drying atmosphere in unsealed plates). A total of 100 conidia were randomly chosen and measured by using an OLYMPUS CH30RF200 light microscope (Olympus, Tokyo, Japan) with a Moticam 5000 digital camera (Motic Deutschland GmbH, Wetzlar, Germany).

2.4. DNA extraction, amplification and sequencing

Isolates were grown on PDA for 5–7 days and then collected for DNA extraction. Genomic DNA was extracted following a previously described method [17] with some modifications. The nuclear

ribosomal internal transcribed spacer of ribosomal DNA (*ITS rDNA*) region was amplified with the universal primers ITS1 and ITS4 [18], the glyceraldehyde-3-phosphate dehydrogenase gene (*GAPDH*) with the primers gpd1 and gpd2 [19], the *TEF1-alpha* gene (*TEF1*) with the primers EF1-728F and EF1-986R [20]. The PCR mixtures included 9.5 μL of ddH₂O, 2 μL of each primer (10 μM), 1 μL of genomic DNA, 12.5 μL of 2 × EsTaq Master Mix containing 0.5 units of EsTaq DNA Polymerase, 3 mM of MgCl₂ and 200 μM of each dNTP and 1× PCR buffer (Cwbio, Beijing, China). The PCR reactions were carried out in a Veriti 96-well thermo cycler (Applied Biosystems, Carlsbad, CA). The cycling program was initiated at 94 °C for 5 min, and then followed by 35 cycles of denaturation at 94 °C for 30 sec, annealing at 58 °C for 30 sec, and elongation at 72 °C for 30 sec, and finalized with an extension at 72 °C for 10 min. PCR products were sequenced by BGI (Beijing Genomics Institute, Beijing, China). The obtained sequences (*ITS*, *GAPDH*, and *TEF1*) were deposited in GenBank with the accession numbers shown in Table 1.

2.5. Phylogenetic analyses

The obtained sequences and relevant ones available in the GenBank database (Table 1) were aligned using CLUSTAL_X ver. 2.0.11 (EMBL-EBI, Cambridgeshire, UK), and adjusted manually [21]. The combined dataset of *ITS*, *GAPDH*, and *TEF1* gene sequences was analyzed using MEGA 6 software [22]. Maximum likelihood analysis was performed with 1000 bootstrap replicates and the General Time Reversible model of nucleotide substitution. The node reliability was assessed by no less than 70% of parsimony bootstrap support values *A. gypsophilae* (CBS107.41) were used as outgroups for the phylogeny analysis.

3. Results and discussion

3.1. Phylogenetic analysis

The combined dataset resulted in an alignment containing a total of 1434 characters, which comprised 583, 575, and 276 characters of *ITS rDNA*, *GAPDH*, and *TEF1*, respectively. Maximum likelihood analysis was used to generate the phylogenetic tree shown in Figure 1. The two isolates YITC 5109 and YITC 5114 of *A. yunnanensis* sp. nov. were identical in all these three genes, and formed a strongly supported clade in the phylogram, being most closely related to *A. sidae*, *A. sennae*, *A. deserticola*. These two species further clustered together with *A. cyamopsisidis*, *A. rostellata*, *A. nitrimali*, *A. thunbergiae*, and *A. crassa*.

Table 1. Isolates used in this study and their GenBank accession numbers.

Name	Old name	Strain number ^a	Host/Substrate	Locality	GenBank accession numbers	
					ITS	GAPDH
<i>Alternaria acalypicola</i>		CBS 541.94; E.G.S. 38.100	<i>Acalypha indica</i>	Seychelles	KJ718097	KJ718446
<i>Alternaria agerati</i>		CBS 117221; E.G.S. 30.001	<i>Ageratum houstonianum</i>	USA, Illinois	KJ718098	KJ718447
<i>Alternaria aripestis</i>		CBS 577.94; E.G.S. 41.034	<i>Euphorbia esula</i> , stem lesion	Canada, Saskatchewan	KJ718099	KJ718448
<i>Alternaria allii</i>	<i>Alternaria porri</i>	CBS 107.28; E.G.S. 48.084	<i>Allium cepa</i> , leaf spot	Puerto Rico	KJ718100	KJ717954
	<i>Alternaria vanuatuensis</i>	CBS 121345; E.G.S. 45.018	<i>Allium cepa</i> , leaf	Vanuatu	KJ718104	KJ717958
	<i>Alternaria solani</i>	CBS 105.51; ATCC 11078;	<i>Solanum lycopersicum</i> , fruit	UK, England	KJ718105	KJ717959
<i>Alternaria alternariaciada</i> sp. nov.		CBS 101004	<i>Anagallis arvensis</i> , leaf spot	New Zealand, Auckland	KJ718107	KJ718454
<i>Alternaria anagallidis</i>		CBS 117128; E.G.S. 42.074	<i>Anagallis arvensis</i> , leaf spot	New Zealand, Auckland	KJ718108	KJ718455
<i>Alternaria anodae</i>		PPRI 12376	<i>Anoda cristata</i> , leaf	South Africa, Gauteng	KJ718110	KJ717963
<i>Alternaria aragakii</i>		CBS 594.93; E.G.S. 29.016	<i>Passiflora edulis</i>	USA, Hawaii	KJ718111	KJ717964
<i>Alternaria angustifolia</i>		CBS 117222; E.G.S. 35.122	<i>Argyroxiphium</i> sp.	USA, Hawaii	KJ718112	KJ718460
<i>Alternaria argyroxiphii</i>		PPRI 11848	<i>Ipomoea batatas</i> , stem lesion	South Africa, Gauteng	KJ718113	KJ718461
<i>Alternaria azadirachtae</i>		PPRI 11971	<i>Ipomoea batatas</i> , leaf	China, Japan	KJ718114	KJ717966
<i>Alternaria azadirachtae</i>		CBS 116444; E.G.S. 46.195;	<i>Azadirachta indica</i> , leaf spot	Australia, Queensland	KJ718115	KJ718463
		CBS 116445; E.G.S. 46.196;	<i>Azadirachta indica</i> , leaf spot	Australia, Queensland	KJ718116	KJ718464
<i>A. yunnanensis</i> sp. nov.		YITC 5109	<i>Hevea brasiliensis</i> , leaf spot	China, Thailand	MH044725	MH044727
		YITC 5114	<i>Hevea brasiliensis</i> , leaf spot	China, Thailand	MH044728	MH044730
<i>Alternaria bataticola</i>		CBS 531.63; IFO 6187	<i>Ipomoea batatas</i>	Japan	KJ718117	KJ718465
<i>Alternaria blumeae</i>	<i>Alternaria brasiliensis</i>	CBS 117096; E.G.S. 42.158;	<i>Ipomoea batatas</i> , leaf spot	Australia, Queensland	KJ718120	KJ718468
		CBS 117215; E.G.S. 39.116	<i>Phaseolus vulgaris</i> , leaf spot	Brazil, Espirito Santo	KJ718125	KJ718473
		CBS 117364; E.G.S. 40.149	<i>Blumea aurita</i>	Thailand, Yala Province	KJ718126	KJ718474
<i>Alternaria calendulae</i>		CBS 224.76; ATCC 38903;	<i>Calendula officinalis</i> , leaf spot	Germany	KJ718127	KJ718475
		CBS 116439; E.G.S. 42.197	<i>Rosa</i> sp., leaf spot	New Zealand, Auckland	KJ718129	KJ717977
<i>Alternaria carthami</i>		CBS 635.80	<i>Carthamus tinctorius</i> , leaf	Italy, Perugia	KJ718131	KJ718479
		CBS 116440; E.G.S. 3.143;	<i>Helianthus annuus</i> , leaf	Canada, Saskatchewan	KJ718132	KJ718480
		CBS 117092; E.G.S. 37.057	<i>Carthamus tinctorius</i>	Iraq	KJ718134	KJ718482
<i>Alternaria carthamicola</i>		CBS 478.81; E.G.S. 33.147	<i>Senna obtifolia</i> , seedling	USA, Mississippi	KJ718135	KJ718483
<i>Alternaria cassiae</i>		CBS 116119; E.G.S. 47.112	<i>Sauvagesia androgynus</i>	Malaysia, Sarawak	KJ718136	KJ718484
		CBS 117369; E.G.S. 50.166	<i>Hibiscus sabdariffa</i> , leaf	Fiji	KJ718138	KJ717988
		CBS 137456; PD 013/05703936	<i>Catananche caerulea</i>	Netherlands	KJ718139	KJ717989
		CBS 116446; E.G.S. 47.119	<i>Centaurea sstitialis</i> , leaf spot	USA, California	KJ718140	KJ717990
		CBS 102.33; E.G.S. 07.017	<i>Cichorium intybus</i> , leaf spot	Cyprus	KJ718141	KJ717991
		CBS 113261; E.G.S. 41.136	<i>Cirsium arvense</i> , stem lesion	Canada, Saskatchewan	KJ718143	KJ717993
		CBS 103.32; VKM F-1881	<i>Citrullus vulgaris</i> , fruit	Cyprus	KJ718144	KJ717994
		CBS 137457	–	Netherlands	KJ718145	KJ717995
<i>Alternaria catananches</i> sp. nov.		CBS 110.38	<i>Datura stramonium</i> , leaf spot	Cyprus	KJ718147	KJ717997
<i>Alternaria centaureae</i>		CBS 116114; E.G.S. 35.123	<i>Luffa acutangula</i>	USA, Hawaii	KJ718153	KJ718501
<i>Alternaria cichorii</i>		CBS 364.67; E.G.S. 17.065;	<i>Cyamopsis tetragonoloba</i> , leaf spot	USA, Maryland	KJ718156	KJ718503
<i>Alternaria cirrhoxia</i>		CBS 117219; E.G.S. 13.120;	<i>Cyamopsis tetragonoloba</i> , leaf spot	USA, Georgia	KJ718157	KJ718504
<i>Alternaria citrullicola</i> sp. nov.		CBS 111.38	<i>Dichondra repens</i> , leaf spot	Italy	KJ718158	KJ718505
<i>Alternaria conidiophora</i> sp. nov.		CBS 101592	<i>Daucus carota</i> , seed	Netherlands	KJ718162	KJ718506
<i>Alternaria crassa</i>	<i>Alternaria loofahae</i>	CBS 117099; E.G.S. 47.131	<i>Daucus carota</i> , seed	USA, California	KJ718164	KJ718512
<i>Alternaria deserticola</i> sp. nov.		CBS 110799	desert soil	Namibia	KJ718249	KJ718595
<i>Alternaria dichondrae</i>		CBS 199.74; E.G.S. 38.007	<i>Dichondra repens</i> , leaf spot	Italy	KJ718166	KJ718514
<i>Alternaria dauci</i>		CBS 200.74; E.G.S. 38.008	<i>Echinacea</i> sp., leaf lesion	New Zealand, isborne	KJ718167	KJ718515
<i>Alternaria cucumerina</i>		CBS 116117; E.G.S. 46.081	<i>Echinacea</i> sp., leaf lesion	New Zealand, isborne	KJ718170	KJ718518
<i>Alternaria centaureae</i>		CBS 116118; E.G.S. 46.082	<i>Echinacea</i> sp., leaf lesion	New Zealand, isborne	KJ718171	KJ718519

(continued)

Table 1. Continued.

Name	Old name	Strain number ^a	Host/Substrate	Locality	GenBank accession numbers	
					TTS	GAPDH
<i>Alternaria grandis</i>		CBS 109158; E.G.S. 44.106	<i>Solanum tuberosum</i> , leaf spot	USA, Pennsylvania	KJ718239	JQ646341
		CBS 116695; E.G.S. 44.108	<i>Solanum tuberosum</i> , leaf spot	USA, Pennsylvania	KJ718241	EU130547
		CBS 198.86; E.G.S. 38.382	<i>Euphorbia pulcherrima</i>	USA, Florida	KJ718172	KJ718547
<i>Alternaria euphorbiicola</i>		CBS 119410; E.G.S. 41.029	<i>Euphorbia pulcherrima</i>	USA, Hawaii	KJ718173	KJ718520
<i>Alternaria gypsophilae</i>		CBS 107.41; E.G.S. 07.025;	<i>Gypsophila elegans</i> , seed	Netherlands	KC584199	KJ718521
<i>Alternaria ipomoeae</i> sp. nov.	<i>Alternaria cucumerina</i>	CBS 219.79	<i>Ipomoea batatas</i> , stem	Ethiopia	KJ718175	KC584660
		PPRI 8988	<i>Ipomoea batatas</i> , stem	South Africa, Gauteng	KJ718020	KJ718523
<i>Alternaria jesenskae</i>		CBS 133855; CCM 8361	<i>Fumana procumbens</i> , seed	Slovakia	KJ718176	KJ718524
<i>Alternaria limicola</i>		CBS 483.90; E.G.S. 39.070	<i>Citrus aurantiifolia</i> , leaf spot	Mexico, Colima	KJ718177	KJ718525
<i>Alternaria lineariae</i>		CBS 117360; E.G.S. 43.009	<i>Citrus</i> sp.	Mexico, Jalisco	KJ718178	KJ718526
		CBS 109156; E.G.S. 42.156	<i>Solanum lycopersicum</i> , leaf spot	USA, Indiana	KJ718183	KJ718527
<i>Alternaria macrospora</i>	<i>Alternaria tomatophila</i>	CBS 109161; E.G.S. 45.113	<i>Solanum lycopersicum</i> var. <i>cerasiforme</i> , leaf spot	USA, Louisiana	JQ646347	KJ718531
	<i>Alternaria subcylindrica</i>	CBS 109164; E.G.S. 46.188	<i>Solanum lycopersicum</i> , leaf spot	Greece, Crete	KJ718184	KJ718532
	<i>Alternaria cretica</i>	CBS 116438; E.G.S. 41.057	<i>Cucumis sativus</i> , leaf spot	New Zealand	KJ718185	JQ646342
	<i>Alternaria acuminicola</i>	CBS 117228; E.G.S. 50.190	<i>Gossypium barbadense</i>	USA, Arizona	KJ718186	KJ718533
<i>Alternaria montanica</i>		CBS 121343; E.G.S. 44.112	<i>Cirsium arvense</i>	USA, Montana	KC584204	KC584668
		CBS 712.68; ATCC 18515	<i>Richardia scabra</i> , floral bract	USA, Georgia	KJ718194	KJ718541
		CBS 713.68; ATCC 18517;	<i>Ipomoea batatas</i> , stem	South Africa, Gauteng	KJ718195	KJ718542
		PPRI 11845	<i>Richardia scabra</i> , floral bract	USA, Georgia	KJ718196	KJ718542
		PPRI 11847	<i>Ipomoea batatas</i> , stem	South Africa, Mpumal	KJ718198	KJ718544
<i>Alternaria neopomoeae</i> sp. nov.		CBS 116120; E.G.S. 46.151	<i>Solanum viarum</i> , leaf spot	Puerto Rico	KJ718199	KJ718545
		CBS 109163; E.G.S. 46.151	<i>Citrus</i> sp., dry leaf	Papua New Guinea	KJ718201	JU646362
		CBS 12171	<i>Galinsoga parviflora</i> , leaf	South Africa, Gauteng	KJ718202	KJ718034
		CBS 117367; E.G.S. 42.063	<i>Euphorbia pulcherrima</i> , leaf	USA, California	KJ718203	KJ718549
		CBS 134278; E.G.S. 42.064	<i>Euphorbia pulcherrima</i>	USA, California	KJ718204	KJ718550
		CBS 116652; E.G.S. 47.157;	<i>Linum usitatissimum</i> , seed	Canada, Manitoba	KJ718205	KJ718551
		CBS 630.93; E.G.S. 29.020;	<i>Passiflora edulis</i>	USA, Hawaii	KJ718206	KJ718552
		CBS 116333; E.G.S. 50.121	<i>Gaura lindheimeri</i> , leaf	New Zealand,uckland	KJ718210	JQ646352
		CBS 116115; E.G.S. 40.096	<i>Cajanus cajan</i> , seed	India	KJ718211	KJ718556
		CBS 116649; E.G.S. 17.082	<i>Allium cepa</i> , leaf	USA, Nebraska	KJ718214	KJ718557
		CBS 116699; E.G.S. 48.152	<i>Allium cepa</i> , leaf spot	USA, New York	KJ718215	KJ718560
		CBS 116437; E.G.S. 32.076	<i>Hordeum vulgare</i> , seed	New Zealand	KJ718218	KJ718564
		CBS 116651; E.G.S. 45.020	<i>Solanum tuberosum</i> , tuber	USA, California	KJ718220	KJ718566
		CBS 119411; E.G.S. 42.060	<i>Euphorbia pulcherrima</i>	USA, California	KC584217	KC584688
		CBS 116330; E.G.S. 38.039	<i>Ranunculus asiaticus</i> , seed	Israel	JN383483	KC584680
		CBS 215.31	<i>Ricinus communis</i>	Japan	KJ718225	KJ718571
		CBS 353.86	<i>Ricinus communis</i>	Italy, Sardinia	KJ718226	KJ718572
		CBS 117366; E.G.S. 42.061	<i>Euphorbia pulcherrima</i> , leaf	USA, California	KJ718227	KJ718573
		CBS 103.46; Elliot No. 45-190C	<i>Linum usitatissimum</i>	UK, Scotland	KJ718229	JQ646332
		CBS 478.83; E.G.S. 38.011	<i>Scorzonera hispanica</i> , leaf spot	Netherlands, Reuse	KJ718190	KJ718563
		CBS 116703; E.G.S. 36.110	<i>Linum usitatissimum</i> , seed	UK, Derbyshire	KJ718191	KJ718567
		CBS 477.81; E.G.S. 34.030	<i>Senna corymbosa</i> , leaf	India, Uttar Pradesh	KJ718230	JQ646344
		CBS 240.73	<i>Sesamum indicum</i> , seedling	Egypt	KJ718231	JQ646343
		CBS 115264;	<i>Sida fallax</i> , leaf spot	India	JF780939	KJ718576
		CBS 117730; E.G.S. 12.129	<i>Silybum marianum</i> , leaf	Kiribati, Phoenix islands	KJ718232	KJ718577
		CBS 134092; VKM F-4109	<i>Silybum marianum</i> , leaf	Russia, Vladivostok	KJ718233	KJ718578
		CBS 134093; VKM F-4117	<i>Silybum marianum</i> , leaf	Russia, Vladivostok	KJ718234	KJ718580

(continued)

Table 1. Continued.

Name	Old name	Strain number ^a	Host/Substrate	GenBank accession numbers		
				ITS	GAPDH	TEF1
<i>Alternaria solani</i>	<i>Alternaria danida</i>	CBS 111.44; E.G.S. 07.029	<i>Ageratum houstonianum</i> , seed	Y17070	KJ718068	KJ718584
	<i>Alternaria viciae-tabiae</i>	CBS 116442; E.G.S. 46.162	<i>Vicia faba</i>	KJ718240	KJ718069	KJ718586
	<i>Alternaria cyphomandrae</i>	CBS 109155; E.G.S. 40.058	<i>Cyphomandra betacea</i> , fruit	KJ718242	JQ646360	KJ718588
	<i>Alternaria herbiculinae</i>	CBS 116332; E.G.S. 49.180	<i>Petroselinum crispum</i> , plant	KJ718244	KJ718072	KJ718590
	<i>Alternaria glyceriae</i>	CBS 116334; E.G.S. 51.107	<i>Glyceria maxima</i> , leaf spot	KJ718245	KJ718073	KJ718591
<i>Alternaria steviae</i>		CBS 631.88; IFO 31/212	<i>Stevia rebaudiana</i> , leaf spot	KJ718250	KJ718078	KJ718596
		CBS 117362; E.G.S. 37.019	<i>Stevia rebaudiana</i> , leaf spot	KJ718252	KJ718079	KJ718598
<i>Alternaria tagetica</i>		CBS 297.79; GST AM2	<i>Taraxacum officinale</i> , seed	KJ718253	KJ718080	KJ718599
		CBS 479.81; E.G.S. 33.081	<i>Tagetes erecta</i> , seed	KC584221	KC584143	KC584692
		CBS 117217; E.G.S. 44.045	<i>Tagetes</i> sp., leaf spot	KJ718256	KJ718083	KJ718602
<i>Alternaria thunbergiae</i>		CBS 116331; E.G.S. 41.073	<i>Thunbergia alata</i> , leaf spot	KJ718257	KJ718084	KJ718603
		CBS 120986; E.G.S. 51.075	<i>Allium cepa</i> , leaf	KJ718258	KJ718085	KJ718604
		CBS 122597	<i>Thunbergia alata</i>	KJ718259	KJ718086	KJ718605
<i>Alternaria tillandsiae</i>		CBS 116116; E.G.S. 43.074	<i>Tillandsia usneoides</i>	KJ718260	KJ718087	KJ718606
		CBS 631.93; E.G.S. 39.126	<i>Passiflora edulis</i> , fruit	KJ718261	KJ718088	KJ718607
<i>Alternaria tropica</i>		CBS 117216; E.G.S. 39.125	<i>Passiflora edulis</i> , fruit	KJ718262	KJ718089	KJ718608
		CBS 116121; E.G.S. 48.065	<i>Phaseolus vulgaris</i> , leaf spot	KJ718263	KJ718090	KJ718609
		CBS 117223; E.G.S. 44.035	<i>Zinnia elegans</i> , leaf spot	KJ718270	KJ718096	KJ718616
<i>Alternaria venezuelensis</i>			New Zealand, Auckland			
<i>Alternaria zimiae</i>						

^aATCC: American Type Culture Collection, Manassas, VA, USA; BRIP: Queensland Plant Pathology Herbarium, Queensland, Australia; CBS: Culture collection of the Centralbureau voor Schimmelcultures, Fungal Biodiversity Centre, Utrecht, Netherlands; CCM: Czech Collection of Microorganisms, Brno, Czech Republic; CECT: Spanish Type Culture Collection, Valencia, Spain; CPC: Personal collection of P.W. Crous, Utrecht, Netherlands; DAOM: Canadian Collection of Fungal Cultures, Ottawa, Canada; DSM: German Collection of Microorganisms and Cell Cultures, Leibniz Institute, Braunschweig, Germany; E.G.S.: Personal collection of Dr. E.G. Simmons; Elliott: Personal collection of M.E. Elliott; GST: Personal collection of G.S. Taylor; ICMP: International Collection of Micro-organisms from Plants, Aalst, Belgium; IFO: Institute for Fermentation Culture Collection, Osaka, Japan; IMI: Culture collection of CABI Europe UK Centre, Egham UK; LEV: Plant Health and Diagnostic Station, Levin, New Zealand; MUCL: (Agro)Industrial Fungi and Yeast Collection of the Belgian Co-ordinated Collections of Micro-organisms (BCCM), Louvain-la Neuve, Belgium; Nattrass: Personal collection of R.M. Nattrass; PD: Plant Protection Service, Wageningen, Netherlands; PPRI: ARC-Plant Protection Research Institute, Roodeplaat, South Africa; QM: Quarter Master Culture Collection, Amherst, MA, USA; VKM: All-Russian Collection of Microorganisms, Moscow, Russia.

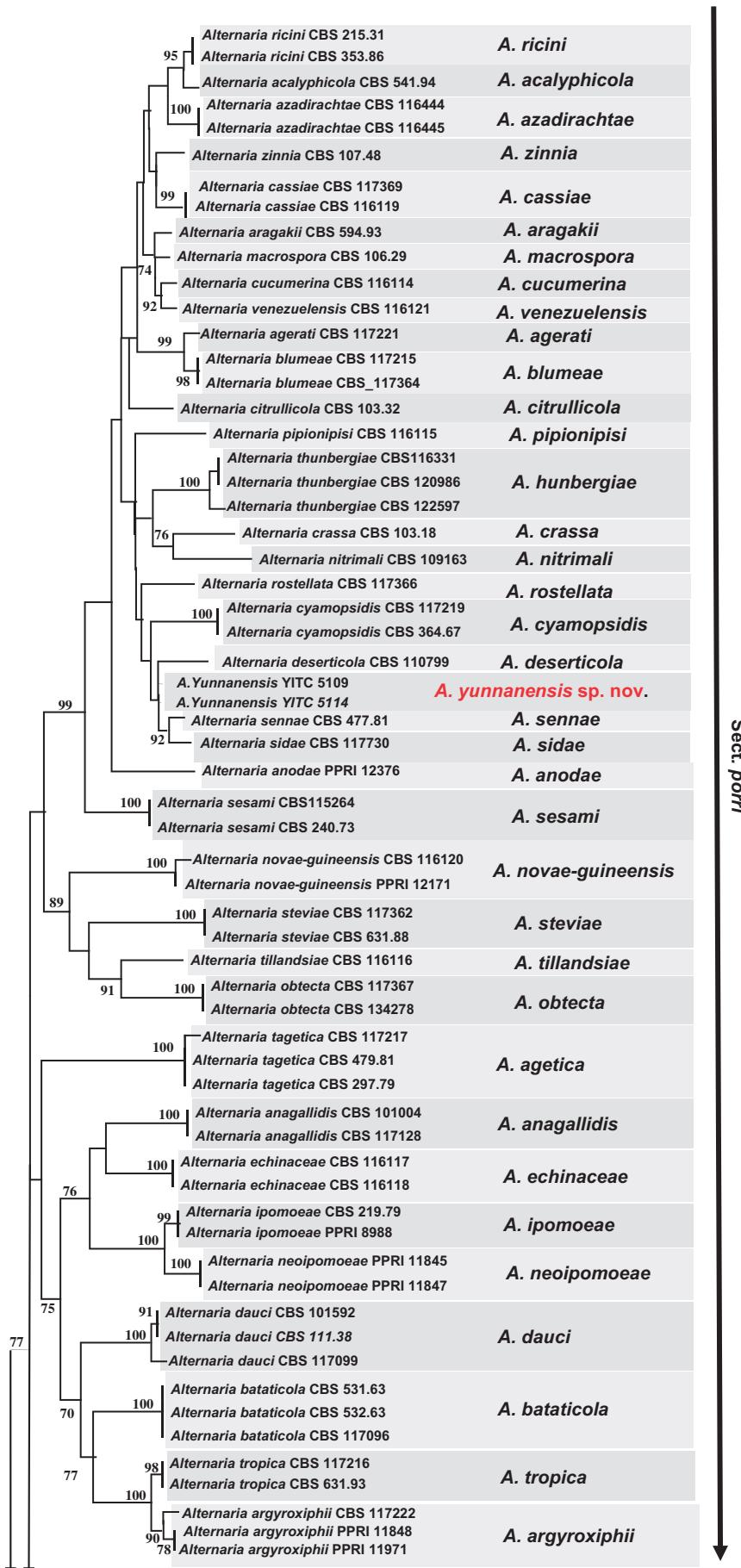


Figure 1. The Maximum-likelihood tree derived from combined *ITS rDNA*, *GAPDH*, and *TEF1* gene sequences of *Alternaria* species. Bootstrap support values above 50% are shown at the nodes. *Alternaria gypsophila*e serve as outgroup. The strains YITC 5109 and YITC 5114 of a new species *A. yunnanensis* proposed in this study are emphasized in red.

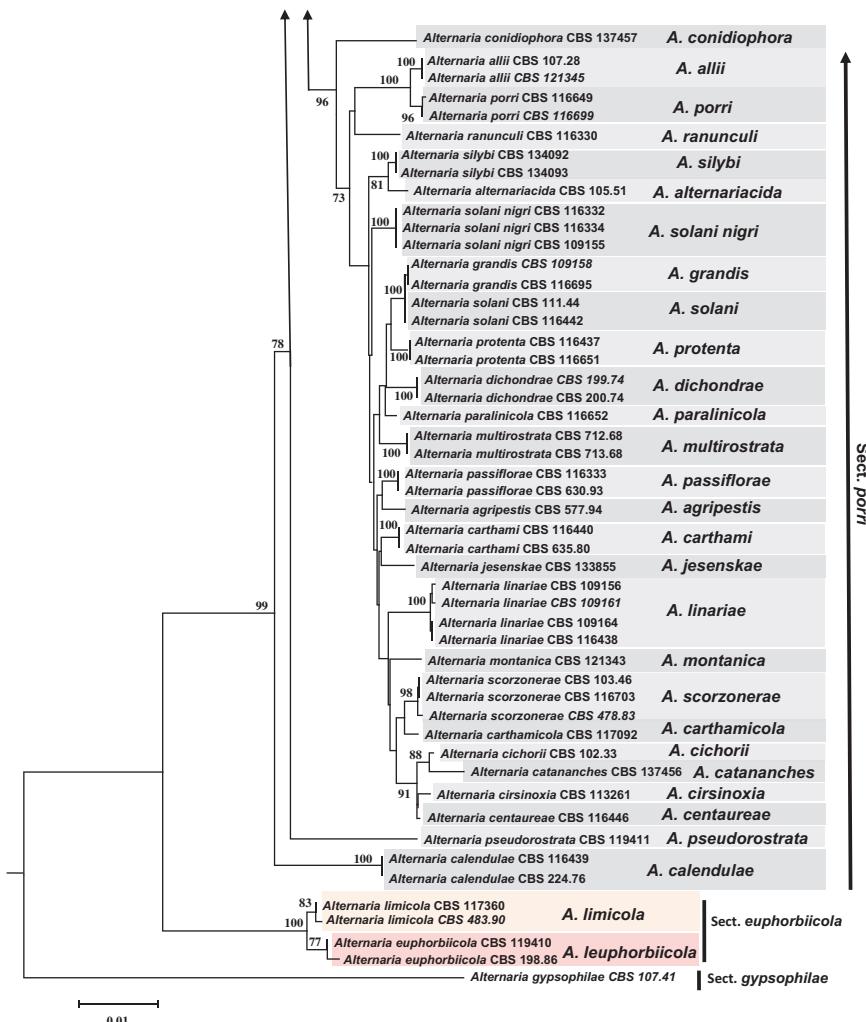


Figure 1. Continued.

3.2. Taxonomy

Alternaria yunnanensis Z.Y. Cai, X.Y. Liu, Y.X. Liu & Y.P. Shi, sp. nov. (Figure 2).

Etymology: In reference to the location of Yunnan Province from where the fungus was isolated.

Fungal Name: FN570554.

Colonies of *A. yunnanensis* on PDA agar develop well, attaining a diameter of 7.7–8.5 cm under a daily fluorescent light/dark cycle of 8/16 h at 25 °C in 7 days, circular, with evident concentric rings and a whitish aerial-mycelium margin, cottony, yellowish to light gray, secreting an orangish pigment into the medium (Figure 2(A)). Mycelia produced on V-8 agar are light to dark gray (Figure 2(B)). Conidiophores are simple or sparingly branched, septate, pale brown to mid brown, smooth, arising beneath the agar surface or directly from hyphae, ca. 40–104 × 5–7 µm, and gradually enlarge near the apex into a clavate conidiogenous cell that produces conidia (Figure 2(F)). Conidia are abundant on V-8 agar, and few on PDA and PCA, smooth, dilute dull brown, 41–97 × 13–29 µm, with 5–10 transverse septa and 1–6 longitudinal or oblique septa, often constricted at the septa, usually solitary or rarely in

chains of two (Figure 2(C,E)), straight or slightly curved, long ellipsoid to obclavate, tapering to the beak which is equal in length to or up to fourfold as long as the body. The apical beaks are simple, pale, septate, up to 252 µm long and 2.5 µm wide (Figure 2(E)).

3.3. Holotype

China, isolated from rubber tree leaf, Hula village, Ruili city, Yunnan Province, China, May 2014, Z.Y. Cai (dried culture HMAS 247784), ex-holotype cultures (ACCC 39327 and CGMCC3.18901).

3.4. Pathogenicity assay

Four days after inoculation, typical black leaf spots observed, which resembles the field symptom with round, black, foliar spots, surrounded by yellow halos, and lesions sometimes slightly sink (Figure 2(D)). A fungus reisolated from the leaf lesions on inoculated plants has the same colonial and conidial morphology as the inocula *A. yunnanensis*, but not from asymptomatic leaves of control rubber plants, fulfilling Koch's postulates. The result indicates that

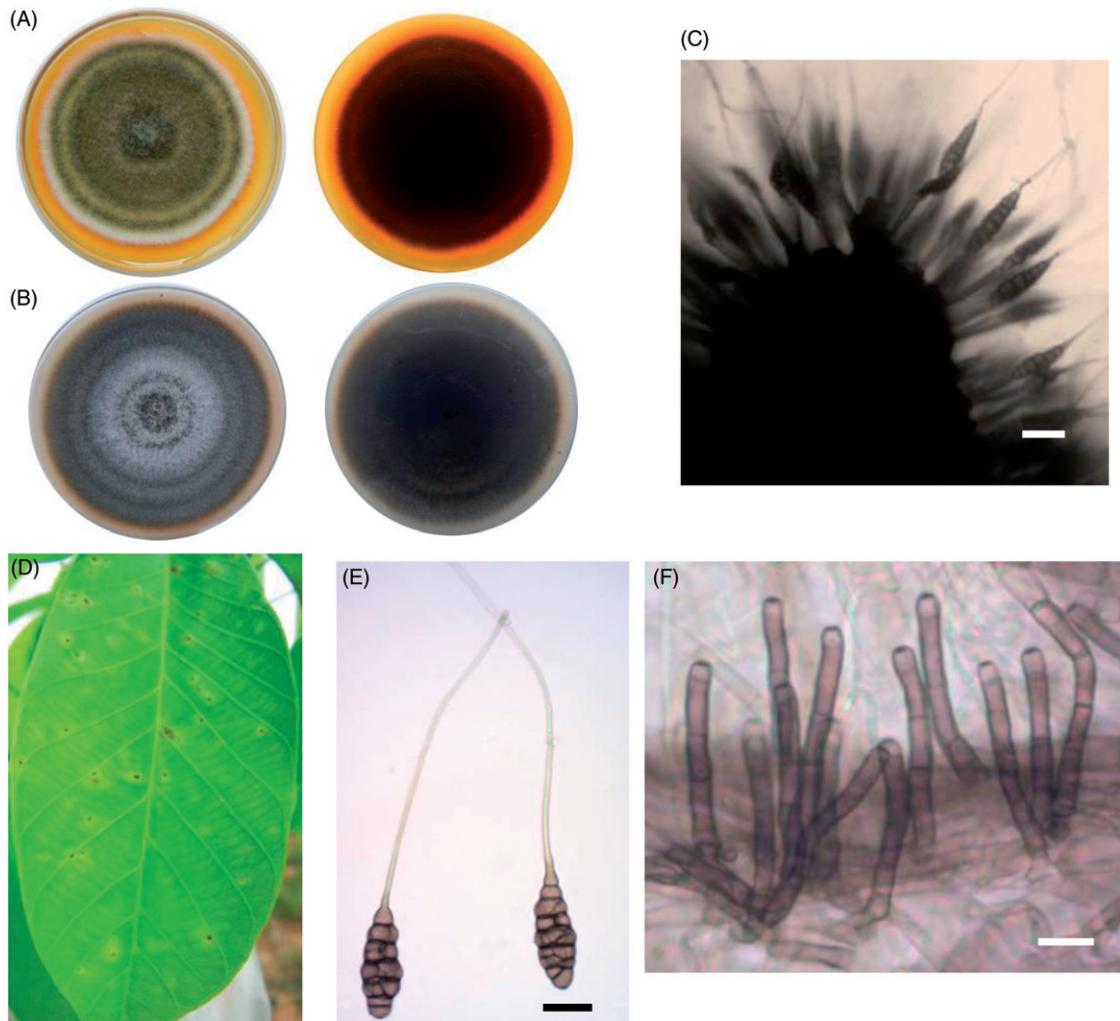


Figure 2. *Alternaria yunnanensis* (ACCC 39327: ex-holotype). (A) Obverse (left) and reverse (right) of colony on PDA at 25 °C for 7 days; (B) Obverse (left) and reverse (right) of colony on V-8 juice agar at 25 °C for 7 days; (C) features of the sporulation apparatus; (D) Symptoms on *Hevea brasiliensis* leaves; (E) Conidia produced on V-8 juice agar at 22 °C for 7 days; (F) Conidiophores produced on V-8 juice agar at 22 °C for 7 days. Scale bars: 50 µm for (C), 40 µm for (E), 10 µm for (F).

the species is the causal agent of leaf spot diseases in rubber tree.

Based on phylogenetic analyses and morphological characteristics, the fungus isolated from the leaf spots of the rubber tree was identified. Both data indicate that the fungus is a novel species belonging to the genus *Alternaria*. With its long history of rubber cultivation, rubber trees have had ample opportunity to attract a wide variety of diseases. A number of significant new diseases have been discovered. At present, the *Alternaria* leaf disease is not very serious. However, identifying *Alternaria* species on rubber tree is important to control rubber tree disease in case the disease outbreaks. In China among the three known *Alternaria* disease of rubber tree, *A. alternata* is the most widely distributed, which can be found at every rubber planting region. *Alternaria heveae* and *A. yunnanensis* were only found at very few rubber plantations.

Along with the *A. yunnanensis*, 112 *Alternaria* strains including 63 species of the section *Porri* of

Alternaria were analyzed. The phylogram (Figure 1) generated from the three-gene combined dataset (ITS, GAPDH, and TEF1) exhibits similar topology to previously reported one [3]. The phylogenetic analyses indicate that the new species, *A. yunnanensis*, belongs to the *Porri* species-group, all of which produce relatively large conidia with a simple or branched, filamentous long beak [2,3].

The *Porri* species-group includes 63 species [3]. Phylogenetic analysis showed that *A. yunnanensis* was most closely related to *A. sidae*, *A. sennae*, *A. deserticola*, and relevant to *A. cyamopsisidis*, *A. rostellata*, *A. thunbergiae*, *A. nitrimali*, and *A. crassa* in the section *Porri*.

Based on conidial morphology, *A. yunnanensis* is most similar to *A. sennae*. Conidia of *A. yunnanensis* have 5–10 transverse septa and often evidently constricted at the septa, whereas those of *A. sennae* have 7–8 transverse septa, slightly constricted near some transverse septa. Besides the conidia of *A. sennae* (55–62 × 10–12 µm) are narrower than that of *A. yunnanensis* (41–97 × 13–29 µm) [3]. The culture

Table 2. Morphological characteristics of the present isolates and relevant species.

Species/MB or FN No.	Shape	Size (μm)	Conidia		
			Septation	Ornamentation	Beak
<i>A. yunnanensis</i> FN 570554	Straight or curved, long ellipsoid to obclavate	41–97 × 13–29	5–10 transverse, 1–6 longitudinal or oblique septa	Smooth	A filamentous beak, up to 252 μm long and 2.5 μm wide
<i>A. sidae</i> MB 489129	Variant of a long-elliptical or sometimes cylindrically narrow ovoid	70–110 × 18–27	7–10 transverse septa and 1–2 longisepta	Conidia with a pustulate surface	A filamentous beak reach 95–110 × 1.5 μm
<i>A. sennae</i> MB 809000	Broadly ovoid, ellipsoid, or sometimes obovoid	55–62 × 10–12	7–8 transverse distosepta and 2–3 longitudinal septa	Smooth	A filamentous beak, occasionally branches once, ca. 99–163 × 2 μm
<i>A. cyamopsisidis</i> MB 292412	Ellipsoid to broadly ovoid	60–90 × 17–25	7–10 transverse distosepta with 1–3 longisepta	Conidia bear a scattering of granules	A filamentous beak up to 100–170 × 1.5–2.0 μm
<i>A. rostellata</i> MB 414600	Broadly ovoid, ellipsoid, or obclavate	50–80 × 20–30	7–9 transverse septa and 1–3 longitudinal septa	Smooth to moderately punctate-rough	A narrow beak, ca. 30–55 × 2 μm or usually shorter
<i>A. thunbergiae</i> MB 504986	Ovoid or ellipsoid,	96–112 × 27–32	8–12 transverse septa and 1–2 longitudinal septa	Either smooth or, closely punctulate	Tapered beak up to 225–320 × 2 μm
<i>A. nitrimali</i> MB 467495	Ovoid or ellipsoid,	70–87 × 14–17.5	7–10 transverse distosepta and only 1 longiseptum	Smooth or inconspicuously punctulate	A long filamentous beak 115–186 × 2 μm

of *A. deserticola* (CBS 110799) is sterile. However, Conidia of *A. yunnanensis* are abundant on V-8 agar. Furthermore *A. yunnanensis* differs from *A. deserticola* of its closest phylogenetic neighbour based on alleles of all three loci (positions derived from respective alignments of the separate loci: *ITS* positions 389(C), 390(C), and 504(T); *GAPDH* positions 111(G), 172(C), 332(C), 482(T), and 500(C); *TEF1* positions 264(C) and 310(A) [3]. Conidium outer walls of *A. yunnanensis* are smooth, while those of relevant species (*A. sidae*, *A. cyamopsisidis*, *A. rostellata*, *A. thunbergiae* and *A. nitrimali*) are punctulate to strongly punctuate ornamentation. The conidia of *A. yunnanensis* are obviously smaller than that of *A. thunbergiae* (96–112 × 27–32 μm). Additionally, the apical beaks of *A. yunnanensis* are simple and long filamentous up to fourfold as long as the body, which differs from *A. rostellata* with a narrow beak, ca. 30–55 × 2 μm or usually shorter, *A. sennae* with a single beak which occasionally branches once. A detailed comparison between *A. yunnanensis* and members of the *Porri* species-group is provided in Table 2.

Compared with other *Alternaria* pathogens on rubber tree, including *A. alternata* and *A. heveae*, the conidia of *A. yunnanensis* are quite different, which are obviously larger than that of *A. alternata* (22.5–67.5 × 10–15 μm) and *A. heveae* (22.75–49.5 × 8–20 μm) [12,13]. In GenBank, there are only *ITS* sequences of *A. heveae* and *A. alternata* (KF953884 and KM111289) of these known *Alternaria* pathogens of rubber tree. We compared our sequences with them, and they were different (data not shown). Thus, we confirm the novelty of *A. yunnanensis*.

Disclosure statement

No potential conflict of interest was reported by the authors.

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