

Themed Section: Eicosanoids 35 years from the 1982 Nobel: where are we now?

## REVIEW ARTICLE

# Eicosanoids in the gastrointestinal tract

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Eicosanoids play important roles in modulating inflammation throughout the body. The gastrointestinal (GI) tract, in part because of its intimate relationship with the gut microbiota, is in a constant state of low-grade inflammation. Eicosanoids like PGs, lipoxins and leukotrienes play essential roles in maintenance of mucosal integrity. On the other hand, in some circumstances, these mediators can become major drivers of inflammatory processes when the lining of the GI tract is breached. Drugs such as nonsteroidal anti-inflammatories, by altering the production of various eicosanoids, can dramatically impact the ability of the GI tract to respond appropriately to injury. Disorders such as inflammatory bowel disease appear to be driven in part by altered production of eicosanoids. Several classes of drugs have been developed that target eicosanoids.

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### Abbreviations

GI, gastrointestinal; IBD, inflammatory bowel disease; LOX, lipoxygenase; LT, leukotriene; LX, lipoxin; NSAID, nonsteroidal anti-inflammatory drug

The discovery by Sir John Vane that aspirin and other nonsteroidal anti-inflammatory drugs (NSAIDs) inhibited the formation of PGs led to a burst of research into the role of this group of lipid mediators in gastrointestinal (GI) function (Vane, 1971). Prior to that, Andre Robert and his colleagues at The Upjohn Company, in the USA, had discovered that PGs were potent inhibitors of gastric acid secretion, and these lipid mediators could also prevent experimental ulcer formation (Robert *et al.*, 1968). Robert's group went on to characterize the effects of several longer acting synthetic PG analogues in animals and humans (Konturek *et al.*, 1974; Robert *et al.*, 1979), and in a landmark paper in 1979, described the powerful 'cytoprotective' effects of these agents in the stomach and duodenum (Robert *et al.*, 1979). Remarkably, these very low doses of PGs were reported to protect the lining of the rat stomach from damage induced by oral administration of high concentrations of hydrochloric acid or sodium hydroxide, absolute alcohol and even to boiling water (Robert *et al.*, 1979).

The first PG analogue to reach the marketplace was **misoprostol**, a PGE<sub>1</sub> derivative developed by Searle and launched in 1986. Misoprostol was shown to significantly reduce the incidence of NSAID-induced gastric ulcers, but only at doses that suppressed gastric acid secretion (Graham *et al.*, 1988). At such doses, misoprostol significantly increased the incidence of diarrhoea, consistent with known stimulatory effects of PGs on GI epithelial secretion (Matuchansky and Coutrot, 1978).

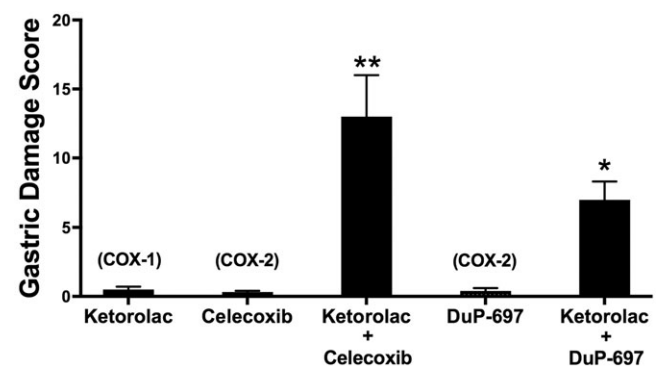
## COX-1 and COX-2

In 1972, another very important discovery was made by Roderick Flower and Sir John Vane that would only be fully appreciated two decades later. They observed that **acetaminophen**, which was known not inhibit peripheral tissue COX activity at therapeutic doses, was able to suppress PG synthesis in the brain (Flower and Vane, 1972). They speculated that there may be more than one form of COX, differentially affected by anti-inflammatory drugs. This notion lingered in the background until 1991, when two groups in the USA reported a previously unrecognized, inducible COX enzyme, which would come to be known as '**COX-2**' (Kujubu *et al.*, 1991; Xie *et al.*, 1991). These publications resulted in a burst of research into the factors regulating expression of the two COX enzymes and their sensitivity to inhibition by the many NSAIDs already on the market, as well as to many new NSAIDs designed to selectively inhibit COX-2 (Masferrer *et al.*, 1994; Mitchell *et al.*, 1994; Warner *et al.*, 1999). The latter was based on the notion that PGs derived from COX-2 accounted for pain and inflammation, while PGs from **COX-1** accounted for the cytoprotective effects that Robert and his colleagues had first observed in the late 1960s (Robert *et al.*, 1968). Several of the world's largest pharmaceutical companies raced to develop selective COX-2 inhibitors. The first to succeed was Searle in 1999 (**celecoxib**; Celebrex<sup>®</sup>), followed less than a year later by Merck (**rofecoxib**; Vioxx<sup>®</sup>). Both were hugely successful from a commercial standpoint, but less than 5 years after its launch, Merck removed rofecoxib from worldwide markets because of evidence that it caused serious cardiovascular events

(Bombardier *et al.*, 2000; Bresalier *et al.*, 2005). Celecoxib, which was less selective for COX-2 than rofecoxib (Warner *et al.*, 1999), has remained on the market and is one of the most commercially successful drugs. Concerns that the selective COX-2 inhibitors increased the risk of serious cardiovascular events led to recommendations from regulators and medical associations for concomitant use of low-dose aspirin in patients deemed to be at increased risk for serious cardiovascular events. However, there is clear evidence that co-administration of aspirin with celecoxib greatly diminishes any reduction of significant GI adverse events that may have been achieved through use of celecoxib versus a conventional NSAID (Silverstein *et al.*, 2000; Wallace *et al.*, 2000).

Animal studies clearly demonstrated that selective inhibition of COX-2 did not result in significant damage in the stomach (Masferrer *et al.*, 1994). However, selective inhibition of COX-1 also did not result in mucosal damage (Wallace *et al.*, 2000). Selective inhibition of one isoform of COX results in rapid up-regulation of the other, at least in the GI tract (Davies *et al.*, 1997; Wallace *et al.*, 2000). Thus, suppression of both COX enzymes was necessary for gastric damage to be induced (Wallace *et al.*, 2000) (Figure 1). These observations from animal models were consistent with the clinical observations on co-administration of celecoxib and low-dose aspirin (Silverstein *et al.*, 2000). The 'COX-2 hypothesis' was also challenged by evidence that COX-1-derived PGs contribute significantly to inflammation (Wallace *et al.*, 1997), and gastric COX-1 is inducible (Ferraz *et al.*, 1997).

Most of the research in the early 'COX-2 era' was focused on ulceration and bleeding in the stomach and proximal duodenum. These are the regions most easy to examine with an endoscope and where the damage is



**Figure 1**

Induction of haemorrhagic, gastric damage by NSAIDs requires inhibition of both COX-1 and COX-2. Ketorolac at a dose (3 mg·kg<sup>-1</sup>) that inhibited COX-1 but not COX-2 did not cause significant gastric damage. Likewise, celecoxib or DuP-697 at doses that selectively inhibit COX-2 (15 mg·kg<sup>-1</sup> and 10 mg·kg<sup>-1</sup>, respectively) did not cause significant gastric damage. However, the combination of a COX-1 and COX-2 inhibitor resulted in extensive haemorrhagic damage in the stomach (\**P* < 0.05, \*\**P* < 0.01 compared with the groups treated only with a single drug). This figure was constructed from previously published data (Wallace *et al.*, 2000).

clearly dependent upon the presence of acid. However, NSAIDs can cause clinically significant ulceration and bleeding throughout the GI tract (Bjarnason *et al.*, 1993; Graham *et al.*, 2005; Lanas *et al.*, 2009; Watanabe *et al.*, 2013; Washio *et al.*, 2016). Several clinical studies demonstrated that there was no reduction in incidence or severity of more distal small intestinal damage with a selective COX-2 inhibitor as compared to that with a conventional NSAID (McCarthy, 2009; Graham *et al.*, 2011). These observations were surprising given that selective COX-2 inhibitors do not block platelet aggregation, which would contribute to GI bleeding. Indeed, endoscopic studies of selective COX-2 inhibitors in healthy volunteers, who typically exhibit greater resistance to NSAID-induced GI damage than arthritis patients, have revealed very high rates of intestinal damage and bleeding (Maiden *et al.*, 2005; Maiden *et al.*, 2007). Also, while selective COX-2 inhibitors were commercially promoted on the basis of vastly improved GI safety, they are commonly co-prescribed with agents that suppress gastric acid secretion. However, administration of selective COX-2 inhibitors with proton pump inhibitors or histamine **H<sub>2</sub> receptor** antagonists has been shown to markedly increase the severity of damage in the distal intestine (Wallace *et al.*, 2011; Satoh *et al.*, 2012; Blackler *et al.*, 2014, Washio *et al.*, 2016). This is in part due to dramatic shifts in the intestinal microbiome (Wallace *et al.*, 2011).

## Leukotrienes and ulcers

When pharmacological tools for studying leukotrienes (LT) first became available, there were numerous studies suggesting a role for these mediators in upper GI injury, but most involved animal models of injury, such as administration of high concentrations of ethanol, with questionable relevance to the injury that is most commonly observed in humans. However, there were some studies identifying increased mucosal levels of **LTB<sub>4</sub>** and **LTC<sub>4</sub>** in biopsies from humans infected with *Helicobacter pylori* (Fukuda *et al.*, 1990; Ahmed *et al.*, 1992). NSAID-induced ulceration was shown to be a neutrophil-dependent process in animal studies (Wallace *et al.*, 1990), leading to the proposal that blockade of COX activity may result in a shunting of **arachidonic acid** to **lipoygenase (LOX)** enzymes and that LTs might be key mediators in activating leukocyte adherence and activation, as well as contributing to mucosal injury. Indeed, NSAID-induced leukocyte adherence was shown to be markedly reduced in rats by a 5-LOX inhibitor and by an **LTB<sub>4</sub> (BLT<sub>1</sub>) receptor** antagonist (Asako *et al.*, 1992), and gastric mucosal synthesis of LTB<sub>4</sub> was shown to be significantly increased in humans taking NSAIDs (Hudson *et al.*, 1993). The availability of selective antagonists of LT receptors facilitated several studies of the role of LTs in animal models of gastric damage. Suppression of LT synthesis by inhibitors of 5-LOX was found to significantly reduce the severity of gastric damage induced by an NSAID (Vaananen *et al.*, 1992). However, treatment with LTB<sub>4</sub> or LTC<sub>4</sub>/**LTD<sub>4</sub>** antagonists had no effect in these studies.

There does not appear to have been any clinical testing of LT antagonists or selective 5-LOX inhibitors with respect to GI safety. Licofelone (ML3000) is a dual COX/5-LOX inhibitor that was proposed to be a 'balanced inhibitor' of 5-LOX and COX with improved GI safety (Jovanovic *et al.*, 2001), but it ultimately failed in phase III clinical development.

## Prostaglandins and intestinal secretion

The 'cytoprotective' effects of PGs can be observed throughout the GI tract in various animal models of injury. Some of these effects may be in part attributable to stimulation of secretion in the gut, which can result in dilution of luminal irritants or toxins, and contribute to the process of expelling such agents. Thus, **PGE<sub>2</sub>** can relax colonic circular muscle *via* actions on **EP<sub>2</sub>** and **EP<sub>4</sub>** receptors (Martinez-Cutillas *et al.*, 2014), while smooth muscle contraction can be increased by PGs acting through the **EP<sub>1</sub> receptor** (Chan and Mashimo, 2013). With respect to the latter, a PGE<sub>1</sub> analogue, **lubiprostone**, has been employed in humans as a treatment for constipation-predominant irritable bowel syndrome (Owen, 2008) and for opioid-induced constipation (Jamal *et al.*, 2015). On the other hand, EP<sub>2</sub> and EP<sub>4</sub> receptor antagonists have been used to suppress diarrhoea, such as that induced by cholera toxin in infected ileal loops (Satitsri *et al.*, 2016).

Duodenal bicarbonate secretion is particularly important for protection of the tissue from high concentrations of gastric acid, and its mechanisms have been well characterized. PGE<sub>2</sub> stimulates duodenal bicarbonate secretion in rats *via* activation of EP<sub>3</sub> and EP<sub>4</sub> receptors (Takeuchi *et al.*, 2010; Said *et al.*, 2015).

## Eicosanoids and inflammatory bowel disease

The ability of PGs to stimulate colonic secretion led some investigators to propose that PGs may play an important role in inflammatory bowel disease (IBD) (ulcerative colitis and Crohn's disease). As outlined above, PGs may indeed contribute significantly to IBD-associated diarrhoea. However, studies of inhibitors of COX activity in human IBD or in animal models of colitis overwhelmingly suggest important protective (anti-inflammatory) effects of PGs. Several animal studies convincingly demonstrated that selective inhibition of COX-2 led to a marked exacerbation of colitis (Reuter *et al.*, 1996; McCartney *et al.*, 1999; Morteau *et al.*, 2000).

Use of NSAIDs has also been shown to be associated with microscopic colitis, which is characterized by watery diarrhoea (Masclee *et al.*, 2015). It is not clear if this was directly related to suppression of prostanoid synthesis. Proton pump inhibitors are similarly associated with an increased risk of microscopic colitis (Verhaegh *et al.*, 2016). Significant alterations in the gut microbiome may contribute to induction of microscopic colitis by both NSAIDs and proton pump inhibitors (Wallace *et al.*, 2011; Wallace, 2013).

## Leukotrienes in inflammatory bowel disease

The observation by Musch *et al.* (1982) that LOX products of arachidonic acid could potentially stimulate colonic secretion drew the attention of other researchers to the possibility of LTs contributing significantly to the pathogenesis of IBD. Subsequently, a marked increase in colonic synthesis of **LTB<sub>4</sub>** was documented in patients with IBD (Sharon and Stenson, 1984), as was a marked increase in peptido-LTs (i.e. LTC<sub>4</sub>, LTD<sub>4</sub>, LTE<sub>4</sub>) (Peskar *et al.*, 1986). The increase in LTB<sub>4</sub> levels in the colonic mucosa of IBD patients was shown to be due to a significant decrease in ω-hydroxylase activity and a marked increase in 5-LOX activity (Ikehata *et al.*, 1995).

**Mesalamine** (5-aminosalicylic acid) is a commonly used, first-line therapy for IBD, with an unknown mechanism of action. It was shown to markedly reduce peptido-LT synthesis by colonic tissue from patients with IBD (Peskar *et al.*, 1986). As discussed above, NSAIDs are known to exacerbate colitis in many IBD patients. Hudson *et al.* (1993) demonstrated that NSAID administration to IBD patients resulted in a significant enhancement of LTB<sub>4</sub> synthesis and suggested that this could be the mechanism underlying the exacerbating effects of NSAIDs in colitis.

As more selective inhibitors of LT synthesis became available, they were tested in rodent models of IBD and were shown to be very effective in suppressing LT synthesis and accelerating resolution of colitis (Wallace *et al.*, 1989). On the other hand, intracolonic administration of LTB<sub>4</sub> significantly exacerbated experimental colitis (Wallace and Keenan, 1990).

Despite the promising findings in animal models, results from clinical trials of inhibitors of LT synthesis were disappointing. MK-591 was among the first tested. This inhibitor of LTB<sub>4</sub> synthesis was evaluated in a multi-centre clinical trial in ulcerative colitis patients (Roberts *et al.*, 1997). Patients were treated twice daily for 8 weeks and rectal dialysate LTB<sub>4</sub> concentrations and disease activity scores were assessed at the end of that period. The highest dose of MK-591 tested in the study (100 mg twice daily) reduced rectal dialysate LTB<sub>4</sub> levels by ~98%, but lower doses (50 or 12.5 mg twice daily) had no significant effect on LTB<sub>4</sub> levels as compared with placebo-treated patients. None of the doses of MK-591 produced a significant improvement in clinical activity as compared with the placebo-treated group.

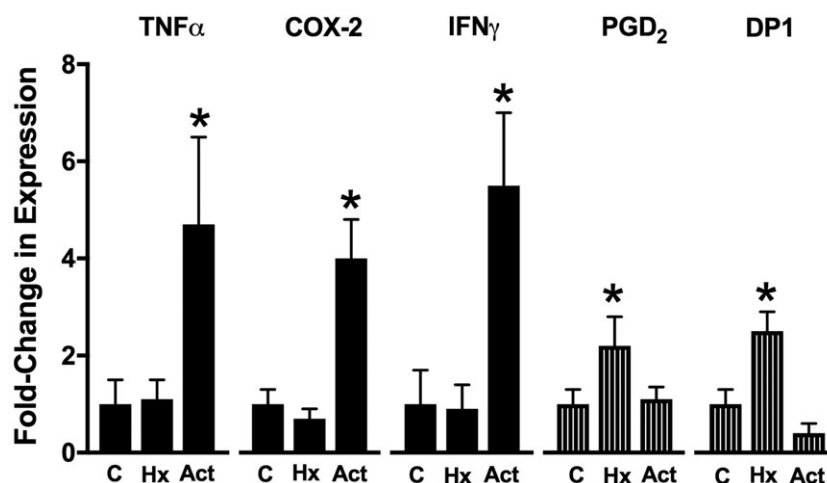
The role of LTB<sub>4</sub> in ulcerative colitis was further examined in a 6-month clinical trial of **zileuton**, another selective 5-LOX inhibitor (Hawkey *et al.*, 1997). The patients were in remission at the start of the study. Treatment with zileuton (600 mg qid) was compared with treatment with placebo or mesalamine (400 mg qid). The primary endpoint was maintenance of remission over the 6-month period. As in the trial of MK-591, the results of the zileuton trial did not support a significant role of LTB<sub>4</sub> in ulcerative colitis. The remission rate in patients treated with zileuton (54%) was not significantly different from that in patients treated with placebo (43%) or mesalamine (63%). The relapse rate for patients treated with mesalamine was significantly lower than the relapse rate for patients on placebo.

## PGD<sub>2</sub>: a stop signal in colitis?

Experimental and clinical data suggest that PGs, while contributing to diarrhoea, can also play an important role in dampening inflammation during bouts of colitis. On the other hand, NSAIDs can exacerbate colitis, presumably due to suppression of 'cytoprotective' PGE<sub>2</sub> synthesis. However, while increased PGE<sub>2</sub> production is associated with the promotion of oedema formation and pain (Murakami and Kudo, 2006), **PGD<sub>2</sub>** acts in the opposite direction, exerting significant anti-inflammatory effects (Gilroy *et al.*, 1999; Ajuebor *et al.*, 2000; Rajakariar *et al.*, 2007). For example, we demonstrated that in experimental colitis, markedly elevated PGD<sub>2</sub> synthesis was a rapid response to tissue injury, acting as a 'braking mechanism' that countered the pro-inflammatory effects of PGE<sub>2</sub> and other chemotaxins (Ajuebor *et al.*, 2000). Remarkably, during the resolution phase of colitis, when tissue structure and function returned to normal, we observed a marked elevation of tissue PGD<sub>2</sub> synthesis that persisted for months after the acute colitis had resolved (Ajuebor *et al.*, 2000; Zamuner *et al.*, 2003). The PGD<sub>2</sub> produced in this setting was derived primarily *via* COX-2, and selective inhibition of that enzyme resulted in marked increase in granulocyte infiltration into the mucosa. This infiltration was significantly reduced by administration of either PGD<sub>2</sub> or a **DP receptor** agonist (Ajuebor *et al.*, 2000). The prolonged up-regulation of PGD<sub>2</sub> synthesis after resolution of a bout of colitis was also accompanied by significant changes in colonic epithelial function. Thus, colonic chloride secretory responses (*in vitro*) were markedly diminished relative to those in controls, but could be recovered to normal by treatment with a selective COX-2 inhibitor (celecoxib), but not selective COX-1 inhibitor (SC-560) (Zamuner *et al.*, 2003). This hyporesponsiveness could be mimicked in normal colonic tissue by exposure to PGD<sub>2</sub>, but not to its metabolite, 15-deoxy-Δ<sup>12-14</sup>PGJ<sub>2</sub>. There was also further evidence for substantially altered colonic epithelial function in rats that appeared to have fully recovered from colitis: although appearing healthy and normal, these rats exhibited a 10-fold increase in bacterial colonization of the colon and more than a threefold increase in bacterial translocation. The latter could be reversed by treatment for 1 week with a selective COX-2 inhibitor (rofecoxib). These studies demonstrate an important role for COX-2, apparently *via* generation of PGD<sub>2</sub>, in mediating the prolonged barrier and epithelial secretory dysfunction that is evident after apparent resolution of colitis in the rat (Zamuner *et al.*, 2003).

There are also human data suggestive of an important role for PGD<sub>2</sub> in the resolution of colitis and particularly in maintenance of remission. Vong *et al.* (2010) analysed colonic biopsies from patients with active ulcerative colitis, looking at pro- and anti-inflammatory PG production, expression of the major enzymes of synthesis and expression of the key receptors. They used two groups as controls: healthy subjects with no prior history of ulcerative colitis and healthy individuals who had previously had ulcerative colitis, but had been in clinical remission for at least 4 years. As expected, the patients with active colitis exhibited significant elevations of colonic synthesis of PGE<sub>2</sub> and increased expression (fourfold to sixfold) of COX-2, **IFN-γ** and **TNF-α** (Figure 2). However, there were some remarkable findings related to PGD<sub>2</sub> when





**Figure 2**

Expression of pro- and anti-inflammatory factors in biopsies from healthy volunteers and ulcerative colitis patients. The 'healthy' groups include subjects with no history (Hx) of ulcerative colitis ('C') and subjects who had previously had ulcerative colitis, but were in long-term remission (more than 4 years). Pro-inflammatory markers such as TNF- $\alpha$ , COX-2 and IFN- $\gamma$  were significantly elevated in the patients with active (Act) colitis. Anti-inflammatory markers such as PGD $_2$  and the DP $_1$  receptor (DP1) were significantly elevated in patients in long-term remission. \* $P < 0.05$  versus controls. This figure was constructed from previously published data (Vong *et al.*, 2010).

the group of patients who had been in long-term remission were examined. In that group alone, there were significant increases in PGD $_2$  synthesis and in expression of mRNA for the **DP $_1$  receptor**, but not for the **DP $_2$  receptor**. PGD $_2$  preferentially binds to DP $_1$  receptors, and its activation is largely thought to be responsible for the anti-inflammatory effects of PGD $_2$ . These data suggest that the same long-term increase in colonic PGD $_2$  synthesis that had been observed in rats after resolution of colitis (Ajuebor *et al.*, 2000; Zamuner *et al.*, 2003) are also evident in humans who are in long-term remission after resolution of colitis. Sturm *et al.* (2014) suggested that there may be a role for modulation of colitis through blockade of the other DP receptor, the DP $_2$  receptor also known as chemoattractant receptor-homologous molecule expressed on T-helper type 2 cells (CRTH2). They demonstrated that blockade of the DP $_2$  receptor (CRTH2) significantly reduced the severity of colitis in a murine model.

## Lipoxins in the GI tract: anti-inflammatory and protective

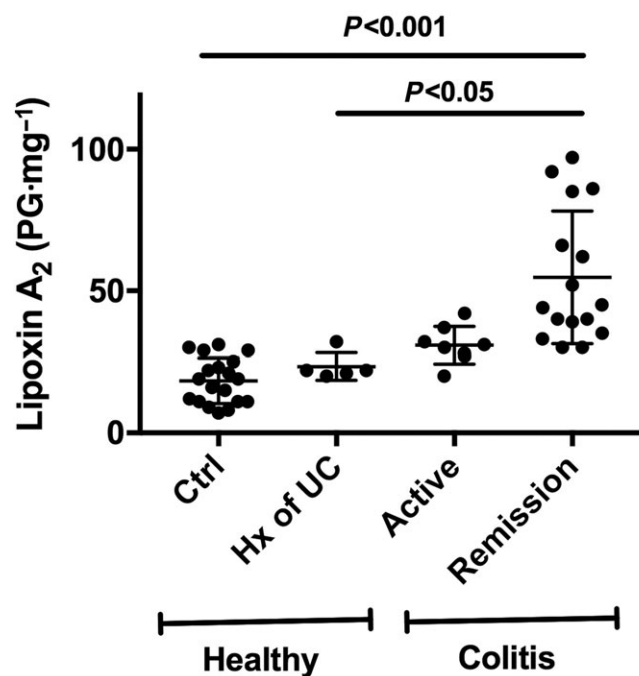
Lipoxins (LX) are endogenous, anti-inflammatory molecules that contribute significantly to reducing tissue injury and progression from acute to chronic inflammation (Serhan *et al.*, 2007; 2015). The 'classical' LX-generating pathways involve (1) the sequential actions of 5-LOX (in leukocytes) and 12-LOX (in platelets) to produce **LXA $_4$**  and (2) the sequential actions of 15-LOX (in epithelial cells) and 5-LOX (in leukocytes) to produce **LXB $_4$**  (Serhan *et al.*, 2015). A third pathway for LX synthesis, involving aspirin, is described below.

LXA $_4$  analogues have been synthesized and have shown promising anti-inflammatory effects in models of colitis. For example, Gewirtz *et al.* (2002) demonstrated that LXA $_4$  analogues reduced pro-inflammatory gene expression in intestinal epithelial cells and significantly attenuated the

severity of experimental colitis in mice. LXA $_4$  and LX analogues have been shown to down-regulate leukocyte degranulation (Gewirtz *et al.*, 1999) and chemokine release (Gewirtz *et al.*, 1998). Moreover, LXA $_4$  has been shown to dampen intestinal inflammation by inhibiting NF- $\kappa$ B activity (Kure *et al.*, 2010). LXA $_4$  has also been reported to be important for driving **IL-10** release, thereby producing additional anti-inflammatory effects (Souza *et al.*, 2007), as well as mediating the protective effects of an n-6 fatty acid-enriched diet in ischaemia-reperfusion injury (Gobbetti *et al.*, 2015).

The oral efficacy of ZK-192, a  $\beta$ -oxidation-resistant analogue of aspirin-triggered lipoxin, was examined in experimental colitis. Daily oral administration of ZK-192 at 300 or 1,000  $\mu$ g.kg $^{-1}$  markedly reduced the severity of colitis in rodents, whether given in preventive or therapeutic treatment regimens. This included significant reductions of macroscopic and histological colon injury, weight loss and mucosal neutrophil infiltration. The observed beneficial effects of ZK-192 were comparable with those observed with 3 to 10 mg.kg $^{-1}$  of prednisolone. Expression of a mRNA for several pro-inflammatory mediators (e.g. **inducible NOS**, **macrophage inflammatory protein 2**, COX-2) was significantly decreased by treatment with ZK-192, along with significantly reduced mucosal mRNA and protein levels of TNF- $\alpha$ , **IL-2** and IFN- $\gamma$ .

A significant role for LXA $_2$  in maintenance of remission in patients with ulcerative colitis was suggested by Vong *et al.* (2012). They observed that there were markedly elevated levels of colonic LXA $_2$  in the patients who were in remission, but not in healthy controls or in patients with active ulcerative colitis (Figure 3). They also observed significant elevations of 5-LOX expression in the patients with active colitis, but no change in 12- or 15-LOX expression (Vong *et al.*, 2012). These results demonstrated a specific up-regulation of a pro-resolution circuit in patients who were in remission



**Figure 3**

Mucosal lipoxin A<sub>2</sub> levels in biopsies from healthy volunteers and ulcerative colitis patients. The 'healthy' group included subjects with no history (Hx) of ulcerative colitis (Ctrl) and subjects who previously had ulcerative colitis (UC), but had been in long-term remission (more than 4 years). The 'colitis' patients were subdivided into those with active disease and those in medically induced remission. The patients in remission exhibited markedly elevated lipoxin A<sub>2</sub> levels as compared with the two groups of healthy subjects. This figure was constructed from previously published data (Vong *et al.*, 2012).

from ulcerative colitis, with significant contributions of LXA<sub>2</sub> in promoting mucosal homeostasis.

The ability of aspirin to cause damage in the GI tract, particularly in the stomach, is well known. Acetylation of COX-1 by aspirin results in an irreversible inhibition of that enzyme, preventing PG synthesis. However, acetylation of COX-2, while also preventing PG synthesis, does not completely block the function of the enzyme. Arachidonic acid can still be modified by acetylated COX-2, leading to the formation of 15R-HETE (15R-hydroxyeicosatetraenoic acid), which can be further transformed to 'aspirin-triggered lipoxin' (such as 15-epi-LXA<sub>4</sub>) *via* the action of 5-LOX (Serhan *et al.*, 2007). In the GI tract, aspirin-triggered lipoxins have been shown to contribute significantly to GI mucosal defence, as well as exhibiting significant anti-inflammatory effects (Fiorucci *et al.*, 2002, 2004).

## Closing Comment

As in other tissues, eicosanoids are involved in a wide range of physiological and pathological processes in laboratory animals and humans. There are a wide range of drugs that have been developed to enhance or inhibit the actions of eicosanoids in the GI tract. Despite extensive research for the past five decades, there remains a need for better understanding of the role of eicosanoids in resolution of inflammation in the GI tract. Such research will be an important part of shifting treatment regimens from a target of

reducing the magnitude of inflammatory processes (largely symptom control) to an orderly promotion of resolution of inflammation.

## Nomenclature of targets and ligands

Key protein targets and ligands in this article are hyperlinked to corresponding entries in <http://www.guidetopharmacology.org>, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY (Harding *et al.*, 2018), and are permanently archived in the Concise Guide to PHARMACOLOGY 2015/16 (Alexander *et al.*, 2017a,b).

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## Conflict of interest

The author declares no conflicts of interest.

## References

Ahmed A, Holton J, Vaira D, Smith SK, Houlst JR (1992). Eicosanoid synthesis and *Helicobacter pylori* associated gastritis: increase in

leukotriene C<sub>4</sub> generation associated with *H. pylori* colonization. *Prostaglandins* 44: 75–86.

Ajuebor MN, Singh A, Wallace JL (2000). Cyclooxygenase-2-derived prostaglandin D<sub>2</sub> and resolution of acute inflammation through PGD<sub>2</sub> and 15-deoxy-Δ<sup>12,14</sup> PGJ<sub>2</sub>. *Am J Physiol Liver Physiol* 279: G238–G244.

Alexander SPH, Christopoulos A, Davenport AP, Kelly E, Marrion NV, Peters JA *et al.* (2017a). The Concise Guide to PHARMACOLOGY 2017/18: G protein-coupled receptors. *Br J Pharmacol* 174: S17–S129.

Alexander SPH, Fabbro D, Kelly E, Marrion NV, Peters JA, Faccenda E *et al.* (2017b). The Concise Guide to PHARMACOLOGY 2017/18: Enzymes. *Br J Pharmacol* 174: S272–S359.

Asako H, Kubes P, Wallace J, Gaginella T, Wolf RE, Granger DN (1992). Indomethacin-induced leukocyte adhesion in postcapillary venules: role of lipoxygenase products. *Am J Physiol* 262: G903–G908.

Bjarnason I, Hayllar J, MacPherson AJ, Russell AS (1993). Side effects of nonsteroidal anti-inflammatory drugs on the small and large intestine in humans. *Gastroenterology* 104: 1832–1847.

Blackler RW, Gemici B, Manko A, Wallace JL (2014). NSAID-gastroenteropathy: new aspects of pathogenesis and prevention. *Curr Opin Pharmacol* 19: 11–16.

Bombardier C, Laine L, Reicin A, Shapiro D, Burgos-Vargas R, Davis B *et al.* (2000). Comparison of upper gastrointestinal toxicity of rofecoxib and naproxen in patients with rheumatoid arthritis. VIGOR Study Group. *N Engl J Med* 343: 1520–1528.

Bresalier RS, Sandler RS, Quan H, Bolognese JA, Oxenius B, Horgan K *et al.* (2005). Cardiovascular events associated with rofecoxib in a colorectal adenoma chemoprevention trial. *N Engl J Med* 352: 1092–1102.

Chan WW, Mashimo H (2013). Lubiprostone increases small intestinal smooth muscle contractions through a prostaglandin E receptor 1 (EP1)-mediated pathway. *J Neurogastroenterol Motil* 19: 312–318.

Davies NM, Sharkey KA, Asfaha S, MacNaughton WK, Wallace JL (1997). Aspirin induces a rapid up-regulation of cyclooxygenase-2 expression in the rat stomach. *Aliment Pharmacol Ther* 11: 1101–1108.

Ferraz JG, Sharkey KA, Reuter BK, Asfaha S, Tigley AW, Brown ML *et al.* (1997). Induction of cyclooxygenase 1 and 2 in the rat stomach during endotoxemia: role in resistance to damage. *Gastroenterology* 113: 195–204.

Fiorucci S, Menezes de Lima O, Mencarelli A, Palazzetti B, Distrutti E, McKnight *Wet al.* (2002). COX-2-derived lipoxin A<sub>4</sub> increases gastric resistance to aspirin-induced damage. *Gastroenterology* 123: 1598–1606.

Fiorucci S, Wallace JL, Mencarelli A, Distrutti E, Rizzo G, Farneti S *et al.* (2004). A beta-oxidation-resistant lipoxin A<sub>4</sub> analog treats hapten-induced colitis by attenuating inflammation and immune dysfunction. *Proc Natl Acad Sci U S A* 104: 20979–20984.

Flower RJ, Vane JR (1972). Inhibition of prostaglandin synthetase in brain explains the anti-pyretic activity of paracetamol (4-acetamidophenol). *Nature* 240: 410–411.

Fukuda T, Kimura S, Arakawa T, Kobayashi K (1990). Possible role of leukotrienes in gastritis associated with *Campylobacter pylori*. *J Clin Gastroenterol* 12 (Suppl 1): 131–134.

Gewirtz AT, Collier-Hyams LS, Young AN, Kucharzik T, Guilford WJ, Parkinson JF *et al.* (2002). Lipoxin A<sub>4</sub> analogs attenuate induction of intestinal epithelial proinflammatory gene expression and reduce the

severity of dextran sodium sulfate-induced colitis. *J Immunol* 168: 5260–5267.

Gewirtz AT, McCormick B, Neish AS, Petasis NA, Gronert K, Serhan CN *et al.* (1998). Pathogen-induced chemokine secretion from model intestinal epithelium is inhibited by lipoxin A<sub>4</sub> analogs. *J Clin Invest* 101: 1860–1869.

Gewirtz AT, Fokin VV, Petasis NA, Serhan CN, Madara JL (1999). LXA<sub>4</sub>, aspirin-triggered 15-epi-LXA<sub>4</sub>, and their analogs selectively downregulate PMN azurophilic degranulation. *Am J Physiol* 276: C988–C994.

Gilroy DW, Colville-Nash PR, Willis D, Chivers J, Paul-Clark MJ, Willoughby DA (1999). Inducible cyclooxygenase may have anti-inflammatory properties. *Nat Med* 5: 698–701.

Gobbetti T, Ducheix S, le Faouder P, Perez T, Riols F, Boue J *et al.* (2015). Protective effects of n-6 fatty acids-enriched diet on intestinal ischaemia/reperfusion injury involve lipoxin A<sub>4</sub> and its receptor. *Br J Pharmacol* 172: 910–923.

Graham DY, Agrawal NM, Roth SH (1988). Prevention of NSAID-induced gastric ulcer with misoprostol: multicentre, double-blind, placebo-controlled trial. *Lancet* 2: 1277–1280.

Graham DY, Opekun AR, Willingham FF, Qureshi WA (2005). Visible small-intestinal mucosal injury in chronic NSAID users. *Clin Gastroenterol Hepatol* 3: 55–59.

Graham DY, Jewell NP, Chan FKL (2011). Rofecoxib and clinically significant upper and lower GI events revisited based on documents from recent litigation. *Am J Med Sci* 342: 356–364.

Harding SD, Sharman JL, Faccenda E, Southan C, Pawson AJ, Ireland S *et al.* (2018). The IUPHAR/BPS Guide to PHARMACOLOGY in 2018: updates and expansion to encompass the new guide to IMMUNOPHARMACOLOGY. *Nucl Acids Res* 46: D1091–D1106.

Hawkey CJ, Dube LM, Rountree LV, Linnen PJ, Lancaster JF (1997). A trial of zileuton versus mesalazine or placebo in the maintenance of remission of ulcerative colitis. The European Zileuton Study Group For Ulcerative Colitis. *Gastroenterology* 112: 718–724.

Hudson N, Balsitis M, Everitt S, Hawkey CJ (1993). Enhanced gastric mucosal leukotriene B<sub>4</sub> synthesis in patients taking non-steroidal anti-inflammatory drugs. *Gut* 34: 742–747.

Ikehata A, Hiwatashi N, Kinouchi Y, Yamazaki H, Ito K, Toyota T (1995). Altered leukotriene B<sub>4</sub> metabolism in colonic mucosa with inflammatory bowel disease. *Scand J Gastroenterol* 30: 44–49.

Jamal MM, Adams AB, Jansen JP, Webster LR (2015). A randomized, placebo-controlled trial of lubiprostone for opioid-induced constipation in chronic noncancer pain. *Am J Gastroenterol* 110: 725–732.

Jovanovic DV, Fernandes JC, Martel-Pelletier J, Jolicoeur FC, Reboul P, Laufer S *et al.* (2001). The *in vivo* dual inhibition of cyclooxygenase and lipoxygenase by ML-3000 reduces the progression of experimental osteoarthritis. Suppression of collagenase-1 and interleukin-1β synthesis. *Arthritis Rheum* 44: 2320–2330.

Konturek SJ, Radecki T, Demitrescu T, Kwiecień N, Pucher A, Robert A (1974). Effect of synthetic 15-methyl analog of prostaglandin E<sub>2</sub> on gastric secretion and peptic ulcer formation. *J Lab Clin Med* 84: 716–725.

Kujubu DA, Fletcher BS, Varnum BC, Lim RW, Herschman HR (1991). TIS10, a phorbol ester tumor promoter-inducible mRNA from Swiss 3T3 cells, encodes a novel prostaglandin synthase/cyclooxygenase homologue. *J Biol Chem* 266: 12866–12872.

Kure I, Nishiumi S, Nishitani Y, Tanoue T, Ishida T, Mizuno M *et al.* (2010). Lipoxin A<sub>4</sub> reduces lipopolysaccharide-induced

- inflammation in macrophages and intestinal epithelial cells through inhibition of nuclear factor- $\kappa$ B activation. *J Pharmacol Exp Ther* 332: 541–548.
- Lanas A, García-Rodríguez LA, Polo-Tomás M, Ponce M, Alonso-Abreu I, Perez-Aisa MA *et al.* (2009). Time trends and impact of upper and lower gastrointestinal bleeding and perforation in clinical practice. *Am J Gastroenterol* 104: 1633–1641.
- Maiden L, Thjodleifsson B, Theodors A, Gonzalez J, Bjarnason I (2005). A quantitative analysis of NSAID-induced small bowel pathology by capsule endoscopy. *Gastroenterology* 128: 1172–1178.
- Maiden L, Thjodleifsson B, Seigal A, Bjarnason II, Scott D, Birgisson S *et al.* (2007). Long-term effects of nonsteroidal anti-inflammatory drugs and cyclooxygenase-2 selective agents on the small bowel: a cross-sectional capsule enteroscopy study. *Clin Gastroenterol Hepatol* 5: 1040–1045.
- Martinez-Cutillas M, Mañe N, Gallego D, Jimenez M, Martin MT (2014). EP2 and EP4 receptors mediate PGE2 induced relaxation in murine colonic circular muscle: pharmacological characterization. *Pharmacol Res* 90: 76–86.
- Masclee GM, Coloma PM, Kuipers EJ, Sturkenboom MC (2015). Increased risk of microscopic colitis with use of proton pump inhibitors and non-steroidal anti-inflammatory drugs. *Am J Gastroenterol* 110: 749–759.
- Masferrer JL, Zweifel BS, Manning PT, Hauser SD, Leahy KM, Smith WG *et al.* (1994). Selective inhibition of inducible cyclooxygenase 2 *in vivo* is antiinflammatory and nonulcerogenic. *Proc Natl Acad Sci U S A* 91: 3228–3232.
- Matuchansky C, Coutrot S (1978). The role of prostaglandins in the study of intestinal water and electrolyte transport in man. *Biomedicine* 1978: 143–148.
- McCarthy DM (2009). GI bleeding: problems that persist. *Gastrointest Endosc* 70: 225–228.
- McCartney SA, Mitchell JA, Fairclough PD, Farthing MJ, Warner TD (1999). Selective COX-2 inhibitors and human inflammatory bowel disease. *Aliment Pharmacol Ther* 13: 1115–1117.
- Mitchell JA, Belvisi MG, Akaraserenont P, Robbins RA, Kwon OJ, Croxtall J *et al.* (1994). Induction of cyclo-oxygenase-2 by cytokines in human pulmonary epithelial cells: regulation by dexamethasone. *Br J Pharmacol* 113: 1008–1014.
- Morteau O, Morham SG, Sellon R, Dieleman LA, Langenbach R, Smithies O *et al.* (2000). Impaired mucosal defense to acute colonic injury in mice lacking cyclooxygenase-1 or cyclooxygenase-2. *J Clin Invest* 105: 469–478.
- Murakami M, Kudo I (2006). Prostaglandin E synthase: a novel drug target for inflammation and cancer. *Curr Pharm Des* 12: 943–954.
- Musch MW, Miller RJ, Field M, Siegel MI (1982). Stimulation of colonic secretion by lipoxigenase metabolites of arachidonic acid. *Science* 217: 1255–1256.
- Owen RT (2008). Lubiprostone—a novel treatment for irritable bowel syndrome with constipation. *Drugs Today (Barc)* 2008 44: 645–652.
- Peskar BM, Dreyling KW, Peskar BA, May B, Goebell H (1986). Enhanced formation of sulfidopeptide-leukotrienes in ulcerative colitis and Crohn's disease: inhibition by sulfasalazine and 5-aminosalicylic acid. *Agents Actions* 18: 381–383.
- Rajakariar R *et al.* (2007). Hematopoietic prostaglandin D<sub>2</sub> synthase controls the onset and resolution of acute inflammation through PGD<sub>2</sub> and 15-deoxyDelta<sup>12</sup> 14 PGJ<sub>2</sub>. *Proc Natl Acad Sci U S A* 101: 15736–15741.
- Reuter BK, Asfaha S, Buret A, Sharkey KA, Wallace JL (1996). Exacerbation of inflammation-associated colonic injury in rat through inhibition of cyclooxygenase-2. *J Clin Invest* 98: 2076–2085.
- Robert A, Nezamis JE, Lancaster C, Hanchar AJ (1979). Cytoprotection by prostaglandins in rats. Prevention of gastric necrosis produced by alcohol, HCl, NaOH, hypertonic NaCl, and thermal injury. *Gastroenterology* 77: 433–443.
- Robert A, Nezamis JE, Phillips JP (1968). Effect of prostaglandin E1 on gastric secretion and ulcer formation in the rat. *Gastroenterology* 55: 481–487.
- Roberts WG, Simon TJ, Berlin RG, Haggitt RC, Snyder ES, Stenson WF *et al.* (1997). Leukotrienes in ulcerative colitis: results of a multicenter trial of a leukotriene biosynthesis inhibitor, MK-591. *Gastroenterology* 112: 725–732.
- Said H, Kaji I, Kaunitz JD (2015). Gastroduodenal mucosal defense mechanisms. *Curr Opin Gastroenterol* 31: 486–491.
- Satitsri S, Pongkorpsakol P, Srimanote P, Chatsudthipong V, Muanprasat C (2016). Pathophysiological mechanisms of diarrhea caused by the *Vibrio cholerae* O1 El Tor variant: an *in vivo* study in mice. *Virulence* 7: 789–805.
- Satoh H, Amagase K, Takeuchi K (2012). Exacerbation of nonsteroidal anti-inflammatory drug-induced small intestinal lesions by antisecretory drugs in rats: the role of intestinal motility. *J Pharmacol Exp Ther* 343: 270–277.
- Serhan CN, Brain SD, Buckley C, Gilroy DW, Haslett C, O'Neill LA *et al.* (2007). Resolution of inflammation: state of the art definitions and terms. *FASEB J* 21: 325–332.
- Serhan CN, Chiang N, Dalli J (2015). The resolution code of acute inflammation: novel pro-resolving lipid mediators in resolution. *Semin Immunol* 27: 200–215.
- Sharon P, Stenson WF (1984). Enhanced synthesis of leukotriene B<sub>4</sub> by colonic mucosa in inflammatory bowel disease. *Gastroenterology* 86: 453–460.
- Silverstein FE, Faich G, Goldstein JL, Simon LS, Pincus T, Whelton A *et al.* (2000). Gastrointestinal toxicity with celecoxib vs nonsteroidal anti-inflammatory drugs for osteoarthritis and rheumatoid arthritis: the CLASS study: a randomized controlled trial. Celecoxib Long-term Arthritis Safety Study. *JAMA* 284: 1247–1255.
- Souza DG, Fagundes CT, Amaral FA, Cisalpino D, Sousa LP, Vieira AT *et al.* (2007). The required role of endogenously produced lipoxin A<sub>4</sub> and annexin-1 for the production of IL-10 and inflammatory hyporesponsiveness in mice. *J Immunol* 179: 8533–8543.
- Sturm EM, Radnai B, Jandl K, Stancic A, Parzmair GP, Hogenauer C *et al.* (2014). Opposing roles of prostaglandin D<sub>2</sub> receptors in ulcerative colitis. *J Immunol* 193: 827–839.
- Takeuchi K, Koyama M, Hayashi S, Aihara E (2010). Prostaglandin EP receptor subtypes involved in regulating HCO<sub>3</sub><sup>-</sup> secretion from gastroduodenal mucosa. *Curr Pharm Des* 16: 1241–1251.
- Vaananen PM, Keenan CM, Grisham MB, Wallace JL (1992). Pharmacological investigation of the role of leukotrienes in the pathogenesis of experimental NSAID gastropathy. *Inflammation* 16: 227–240.
- Vane JR (1971). Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nature new biology* 231: 232–235.
- Verhaegh BP, de Vries F, Masclee AA, Keshavarzian A, de Boer A, Souverein PC *et al.* (2016). High risk of drug-induced microscopic colitis with concomitant use of NSAIDs and proton pump inhibitors. *Aliment Pharmacol Ther* 43: 1004–1013.



- Vong L, Ferraz JGP, Panaccione R, Beck PL, Wallace JL (2010). A pro-resolution mediator, prostaglandin D<sub>2</sub>, is specifically up-regulated in individuals in long-term remission from ulcerative colitis. *Proc Natl Acad Sci U S A* 107: 12023–12027.
- Vong L, Ferraz JG, Dufton N, Panaccione R, Beck PL, Perretti M *et al.* (2012). Spatio-temporal up-regulation of annexin-A1 and lipoxin A<sub>4</sub> in individuals with ulcerative colitis may promote mucosal homeostasis. *PLoS One* 7: e39244.
- Wallace JL (2013). Polypharmacy of osteoarthritis: the perfect intestinal storm. *Dig Dis Sci* 58: 3088–3093.
- Wallace JL, Keenan CM (1990). An orally active inhibitor of leukotriene synthesis accelerates healing in a rat model of chronic colitis. *Am J Physiol* 258: G527–G534.
- Wallace JL, Bak A, McKnight W, Asfaha S, Sharkey KA, MacNaughton WK (1997). Cyclooxygenase-1 contributes to inflammatory responses in rats and mice: implications for GI toxicity. *Gastroenterology* 115: 101–109.
- Wallace JL, Keenan CM, Granger DN (1990). Gastric ulceration induced by non-steroidal anti-inflammatory drugs is a neutrophil-dependent process. *Am J Physiol* 259: G462–G467.
- Wallace JL, MacNaughton WK, Morris GP, Beck PL (1989). Inhibition of leukotriene synthesis markedly accelerates healing in a rat model of inflammatory bowel disease. *Gastroenterology* 96: 29–36.
- Wallace JL, McKnight W, Reuter BK, Vergnolle N (2000). NSAID-induced gastric damage in the rat: requirement for inhibition of both cyclooxygenase-1 and -2. *Gastroenterology* 119: 706–714.
- Wallace JL, Denou E, Syer S, Vong L, McKnight W, Jury J *et al.* (2011). Proton pump inhibitors exacerbate NSAID-induced small intestinal injury via induction of dysbiosis. *Gastroenterology* 141: 1314–1322.
- Warner TD, Giuliano F, Vojnovic I, Bukasa A, Mitchell JA, Vane JR (1999). Nonsteroid drug selectivities for cyclo-oxygenase-1 rather than cyclo-oxygenase-2 are associated with human gastrointestinal toxicity: a full *in vitro* analysis. *Proc Natl Acad Sci U S A* 96: 7563–7568.
- Washio E, Esaki M, Maehata Y, Miyazaki M, Kobayashi H, Ishikawa H *et al.* (2016). Proton pump inhibitors increase incidence of nonsteroidal anti-inflammatory drug-induced small bowel injury: a randomized, placebo-controlled trial. *Clin Gastroenterol Hepatol* 14: 809–815.
- Watanabe T, Tanigawa T, Nadatani Y, Nagami Y, Sugimori S, Okazaki H *et al.* (2013). Risk factors for severe nonsteroidal anti-inflammatory drug-induced small intestinal damage. *Dig Liver Dis* 45: 390–395.
- Xie WL, Chipman JG, Robertson DL, Erikson RL, Simmons DL (1991). Expression of a mitogen-responsive gene encoding prostaglandin synthase is regulated by mRNA splicing. *Proc Natl Acad Sci U S A* 88: 2692–2696.
- Zamuner SR, Warriar N, Buret AG, MacNaughton WK, Wallace JL (2003). Cyclooxygenase 2 mediates post-inflammatory colonic secretory and barrier dysfunction. *Gut* 52: 1714–1720.