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Effects of nicotine and stress exposure across generations in C57BL/6 mice

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Abstract

Chronic administration of nicotine or exposure to stress can produce long-lasting behavioral and physiological changes in humans and animals alike. Further, the impact of nicotine and stress exposure can be inherited by offspring to produce persistent changes in physiology and behavior. To determine if nicotine and stress interact *across* generations to influence offspring behavior we exposed F0 male mice to nicotine and F1 male and female mice to chronic unpredictable stress during adolescence. We then measured locomotor sensitization to repeated nicotine injections in the subsequent F2 and F3 generations. Stress exposure alone (F1) did not influence locomotor sensitization in any lineage. However, in the F1 male lineage, F0 nicotine exposure abrogated locomotor sensitization in F2 male and transiently enhanced locomotor sensitization in F2 female offspring. These effects were not passed down to the F3 generations or observed in the F1 female lineage. F1 stress exposure modulated the effects of prior F0 nicotine exposure in a sex-dependent manner. Specifically, stress blunted the nicotine-induced enhancement in locomotor sensitization observed in F2 female offspring of F1 males. The effect of F0 nicotine and F1 stress exposure in females appears to have skipped a generation and enhanced nicotine sensitization only in the F3 generation, and only in females. This novel multigenerational exposure paradigm examining the inheritance of two different environmental exposures demonstrates that nicotine responses can be modified by nicotine and stress exposure from previous generations, and these effects are strongly influenced by sex.

Lay Summary

An individual's offspring can inherit the adverse effects of exposure to stress or nicotine, the primary addictive substance in tobacco, which may predispose them to nicotine addiction. In this paper, *Yohn* and colleagues investigated whether the “transgenerational” effects of nicotine in one generation are altered by chronic stress exposure that occurs in the next generation. Their results indicate that nicotine and stress appear to interact across generations to alter offspring addiction-related behaviors.

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Conflicts of Interest

The authors declare no conflict of interests

Keywords

transgenerational inheritance; nicotine; chronic unpredictable stress; locomotor sensitization; adolescent; mice

Introduction

In the United States, an estimated 40 million adult (Agaku, King, & Dube, 2014) and 3.5 million youth are smokers (Arrazola et al., 2015). In addition, the national prevalence of smokeless tobacco products continues to rise with the use of electronic cigarettes (Arrazola et al., 2015; Schoenborn & Gindi, 2015). Therefore, with approximately 4 million births occurring each year in the United States (Hamilton, Martin, Osterman, Curtin, & Matthews, 2015), a significant proportion of children will be born to nicotine (NIC) users. NIC produces its reinforcing effects in the brain to promote sustained drug use (Damsma, Day, & Fibiger, 1989; Mereu et al., 1987) concomitant with long-term behavioral and physiological changes. For example, nicotine exposure can increase or decrease anxiety- and depression-like behavior (for review see Picciotto, Brunzell, & Caldarone, 2002), enhance cognition (Heishman, Kleykamp, & Singleton, 2010), mediate changes in metabolism (Grunberg, 1990), and influence additional drug use (Levine et al., 2011).

Nicotine use and abuse is a complex behavior influenced by stress exposure. For example, stress signaling pathways in the CNS can promote NIC addiction (Koob & Volkow, 2010) and withdrawal from NIC is a stressful event that precipitates relapse and increases circulating stress hormones (i.e., cortisol) (Benwell & Balfour, 1979). Further, juvenile stress is associated with increased responses to NIC in late-adolescence and adulthood (Caruso et al., 2018; McCormick, Robarts, Gleason, & Kelsey, 2004). In addition, there is some evidence for the inheritance of stress exposure from mothers and fathers to several generations of offspring (Dietz et al., 2011; Franklin et al., 2010; Rodgers, Morgan, Bronson, Revello, & Bale, 2013; Saavedra-Rodríguez & Feig, 2013; Yehuda, Schmeidler, Wainberg, Binder-Brynes, & Duvdevani, 1998). However, no studies have examined the interaction of NIC and stress across generations and the resulting influence in future generations. Therefore, we sought to determine if two behaviors that closely interact on a physiological and molecular level within a generation, would be additive in their effects on offspring behavior across generations.

We produced four distinct lineages to examine multi- and transgenerational inheritance: F0 vehicle (VEH) and F1 no stress referred to as control (CON) exposure (F0 VEH/F1 CON), F0 VEH and F1 chronic unpredictable stress (CUS) exposure (F0 VEH/F1 CUS), F0 NIC and F1 CON exposure (F0 NIC/F1 CON), and F0 NIC and F1 CUS exposure (F0 NIC/F1 CUS). Two subsequent generations of offspring (F2, F3) from both male and female F1 CON/CUS lineages were assessed. Exposure to NIC and stress occurred during adolescence for several reasons. First, almost all adult smokers initiate NIC use during adolescence (SAMHSA, 2011). Adolescence is also a time when both gametes (Kaati, Bygren, & Edvinsson, 2002; Pembrey, 2010) and the neurobiological circuits that mediate nicotine

reinforcement (Yuan, Cross, Loughlin, & Leslie, 2015) are vulnerable to environmental stimuli.

Using this novel exposure paradigm, we find that the transgenerational effects of NIC are modulated by a subsequent exposure to stress in the next generation, producing unique phenotypes in subsequent offspring. Specifically, we found NIC and stress exposure across two generations altered locomotor sensitization to NIC in subsequent offspring in a sex- and lineage-dependent manner (i.e., reduced sensitization in F2 males and females or enhanced sensitization in F3 females). This study is the first to our knowledge to explore the interaction of NIC and stress across generations and to track their influence on subsequent generations.

Methods

Animals

Male and Female C57BL/6NTac mice (6–8 weeks of age, 20–30 g) were ordered from Taconic Farms (Hudson, NY), maintained on a 12-h light/dark cycle with food and water ad libitum in accordance with the University of Pennsylvania Animal Care and Use Committee (Philadelphia, PA, USA) and National Institutes of Health care and use of laboratory animals guidelines. Mice were bred at the University of Pennsylvania for two generations to generate the F0 generation. Breeding in house decreased the impact of transportation on mice (Booker, Butt, Wehner, Heinemann, & Collins, 2007) and allowed us to isolate the effects of NIC and CUS exposure in subsequent generations of offspring. Mice remained group-housed with littermates throughout the experiments unless otherwise stated. Experimental groups were comprised of at least 1–2 mice from each litter. Efforts were made to minimize animal suffering and reduce the number of animals used for all experiments.

F0 male nicotine exposure

Adolescent male mice were exposed to chronic NIC (18 mg/kg/day; (–) - Nicotine hydrogen tartrate salt dissolved in 0.9% saline; Sigma-Aldrich, St. Louis, MO) or saline via osmotic mini pump (model 1004; Alzet, Cupertino, CA) for 28 days from PND 28–56 (Figure 1). This dose of nicotine is in the range of doses found to produce comparable levels of NIC in the plasma of smokers following a cigarette (AlSharari et al., 2013; Benowitz & Sharp, 1989; Henningfield & Keenan, 1993; Russell, Wilson, Patel, Feyerabend, & Cole, 1975). In addition, this dose up-regulates nicotinic acetylcholine receptors in the brain of rodents (Yohn, Turner, & Blendy, 2014), a hallmark of chronic NIC use in humans (Staley, 2006). While the minipump does not recapitulate the pulsatile mode of NIC delivery experienced by human smokers, it does provide several experimental advantages. For example, minipumps allow for consistent administration of NIC doses across subjects, precise temporal control over the initiation and termination of NIC treatment, and their ease of use facilitated replication across cohorts.

Mice were anesthetized with an isoflurane/oxygen mixture (1 – 3%), and osmotic minipumps were inserted subcutaneously using aseptic surgery techniques. Minipumps were placed parallel to the spine at shoulder level with the flow moderator directed away from the

surgical incision. The wound was closed with 7-mm stainless steel wound clips (Reflex; Cellpoint Scientific, Gaithersburg, MD). At the end of 28 days a secondary incision site was used to remove the minipump using aseptic surgery techniques. Approximately 1 week following VEH or NIC exposure mice were housed with naive females for 7 days. This time frame allowed for matings to occur well after the elimination NIC withdrawal signs (Damaj, 2003; Turner, Castellano, & Blendy, 2011) and allowed for elimination of any sperm that may have developed prior to NIC exposure. F0 males (PND 70) were then mated with unexposed females. The presence of vaginal plugs was monitored daily, and males were removed when a plug was found. On average females were positive for vaginal plugs within 2–4 days of mating regardless of treatments. Thus, the mating period did not vary between nicotine and vehicle exposed males.

F1 adolescent chronic unpredictable stress (CUS) exposure

NIC exposure in one generation (F0 NIC) was followed with stress exposure in the subsequent generation (F1 CUS) to determine if NIC and stress interact across generations to influence offspring behavior. Both male and female offspring (F1 generation) from F0 VEH or NIC fathers were exposed to CON or CUS conditions for 12 days from PND 28–40 (Figure 1). The CUS paradigm was implemented as previously described (Yohn & Blendy, 2017). The exact stressors, duration of stressor, and sequence of exposures can be found in *Supplementary Information* (Supplementary Table S1) along with detailed descriptions. Briefly, mice were exposed to three stressors a day, in the morning, afternoon, and overnight, for 12 consecutive days in dedicated procedure rooms. Mice were returned to the animal colony between stressors and after the final stressor. Mice were group-housed with littermates during the CUS exposure. Following CUS exposure, male mice (PND 49) were placed with naive animals for 7 days (to allow for elimination of any sperm that may have developed prior to CUS exposure in males). F1 CON and CUS mice (PND 56) were then mated with naive animals. The presence of vaginal plugs was monitored daily, and males were removed when a plug was found. On average naive females mated with CUS males were positive for vaginal plugs within 2–3 days, as were all Control matings. Of interest, CUS females mated with naive males were positive for vaginal plugs within 3–5 days, perhaps reflecting some residual impact of stress exposure. Behaviorally naive F2 male and female mice were mated with naive mice to produce F3 offspring.

Behavioral testing

All behavioral testing sessions were conducted in a room that was separate from the colony room during the lights-on period between 8:00 a.m. and 5:00 p.m. On testing days, mice were transported to the behavior room at least one hour prior to testing.

Nicotine locomotor sensitization—Sensitization of the locomotor response to repeated nicotine administration was assayed in F2 ($n = 5\text{--}8/\text{group}$) and F3 offspring ($n = 3\text{--}8/\text{group}$) on PND 70–84. In order to record baseline activity on the first 2 test days, mice received intraperitoneal (i.p.) injections of 0.9% saline solution and were immediately placed in test cages that had the same dimensions as their home cage ($28.9 \times 17.8 \times 12$ cm) and contained a small layer of clean bedding. On NIC testing days 1–4, mice received 1 mg/kg (–) - Nicotine hydrogen tartrate salt (i.p.) daily and locomotor activity was recorded for 15 min.

Two weeks following the last NIC injection (challenge day), mice received an additional 1mg/kg (i.p.) NIC injection and locomotor activity was recorded. Locomotor activity was detected using a photo beam frame (30 × 24 × 8 cm) with sensors arranged in an eight-beam array strip around the cage and recorded by an activity monitoring system (MED Associates, St. Albans, VT). The primary dependent variable of interest was locomotor activity (beam breaks) over 15 min because this time frame encompasses the near maximal effects of NIC injection on locomotor activity (Marks, Romm, Bealer, & Collins, 1985).

Statistical Analyses

All data are presented as the mean ± standard error of the mean (SEM). Analyses of the results for the nicotine locomotor sensitization testing was analyzed using a two-way mixed factorial analysis of variance with time (day 1, day 4, and challenge) as the within subjects repeated measure and lineage (F0 VEH/F1 CON, F0 VEH/F1 CUS, F0 NIC/F1 CON, and F0 NIC/F1 CUS) as the between subjects independent variable. Outliers were detected using Grubb's outlier test (Grubbs, 1969) and data were excluded (locomotor sensitization: 18/534 data points excluded). Whenever significant main effects of interactions were identified post hoc analyses were performed using a Bonferroni's multiple comparison test. An $\alpha < 0.05$ was considered significant for all statistical analyses including post hoc comparisons. Statistical analyses were performed using Graphpad Prism 7 (Graphpad Software, La Jolla, CA).

Results

F0 nicotine exposure blunted locomotor responses to nicotine in F2 males regardless of F1 male stress exposure

To determine if NIC and CUS interact across generations to influence offspring behavior, half of the F1 male offspring were exposed to CUS during adolescence and half were left undisturbed to serve as the no stress CON group. F2 offspring were then generated from CON- or CUS-exposed F1 (F1 CON and F1 CUS, respectively) males. In addition, to test for transgenerational inheritance of the F0 NIC and F1 CUS exposures, F2 mice were mated with naive partners to produce an F3 generation.

F2 males derived from the male F1 lineages displayed locomotor sensitization as indicated by greater NIC-induced (1.0 mg/kg, i.p.) locomotion on day 4 and the challenge day compared to day 1 (main effect of 'Time': $F_{2,54} = 8.95$, $p < 0.001$) and F2 male offspring of the F0 VEH males derived from the male F1 CON lineage (F0 VEH/F1 CON) displayed greater NIC-induced locomotion than F2 males from the NIC-exposed F0 male lineages through the F1 CON- (F0 NIC/F1 CON) and F1 CUS-exposed (F0 NIC/F1 CUS) males (main effect of 'Lineage': $F_{3,27} = 12.12$; $p < 0.01$). A 'Time x Lineage' interaction (Figure 2A; $F_{6,54} = 2.55$, $p < 0.05$) further revealed that the F2 males from the F0 VEH/F1 CON lineage displayed greater NIC-induced locomotion on day 4 and the challenge day relative to day 1 (p 's < 0.05). F2 males from the F0 VEH- and F1 CUS-exposed (F0 VEH/F1 CUS) lineage exhibited greater NIC-induced locomotion on the challenge day relative to days 1 ($p < 0.001$) and 4 ($p < 0.05$). Alternatively, F2 males from the F0 NIC/F1 CON and F0 NIC/F1 CUS lineages displayed similar levels of NIC-induced locomotion across the test days.

To determine if F2 males derived from the F1 male lineage produced offspring with similar phenotypes we characterized behavior in their F3 male and female offspring. Overall, F3 male (Figure 2B, left; main effect of 'Time': $F_{2,52} = 17.12$, $p < 0.001$) and F3 female (Figure 2B, right; main effect of 'Time': $F_{2,48} = 23.82$, $p < 0.001$) offspring of the F2 males displayed increased NIC-induced locomotion on day 4 and the challenge day, relative to day 1. There were no significant effects of 'Lineage' or 'Time x Lineage' interactions on NIC-induced locomotion for F3 males and females derived from the F2 male lineage.

F0 nicotine exposure resulted in F2 female offspring that display transient locomotor sensitization to nicotine regardless of F1 male stress exposure.

An unusual pattern of locomotor sensitization was observed in F2 females that were derived from the NIC-exposed F0 male and F1 male lineages. F2 females exhibited greater NIC-induced locomotion on day 4 relative to day 1, but this response was significantly reduced by the challenge day, which did not differ from day 1 levels (main effect of 'Time': $F_{2,48} = 6.72$, $p < 0.01$). A 'Time x Lineage' interaction (Figure 2C; $F_{6,48} = 3.99$, $p < 0.01$) also revealed that F2 female offspring of the F0 NIC/F1 CUS male lineage exhibited increased NIC-induced locomotion on day 4 compared to day 1 ($p < 0.001$) and the challenge day ($p < 0.05$). In contrast, F2 female offspring of F0 NIC/F1 CUS male lineage displayed greater NIC-induced locomotion on day 4 compared to the challenge day ($p < 0.05$).

To determine if F2 females derived from the F1 male lineage produced offspring with similar phenotypes we characterized behavior in their F3 male and female offspring. Relative to day 1, F3 males derived from the F2 female lineage displayed greater NIC-induced locomotion on day 4 and the challenge day (Figure 2D, left; main effect of 'Time': $F_{2,50} = 13.64$, $p < 0.001$). Similarly, NIC-induced locomotion was greater on the challenge day relative to day 1 in F3 females of the F2 female lineage (Figure 2D, right; main effect of 'Time': $F_{2,44} = 4.59$, $p < 0.05$). There were no significant effects of 'Lineage' or 'Time x Lineage' interactions on NIC-induced locomotion for F3 males and females derived from the F2 female lineage.

F0 nicotine and F1 female stress exposures did not influence the locomotor response to nicotine in subsequent generations of male offspring.

To determine if NIC and stress interact across generations in a sex specific manner, half of the F1 female offspring were exposed to CUS during adolescence and half were left undisturbed to serve as the no stress CON group. F2 offspring were then generated from CON- or CUS-exposed F1 (F1 CON and F1 CUS, respectively) females. In addition, to test for transgenerational inheritance of the F0 NIC and F1 CUS exposures, F2 mice were mated with naive partners to produce an F3 generation. Overall, F2 males derived from the female F1 lineages exhibited locomotor sensitization as indicated by greater NIC-induced (1.0 mg/kg, i.p.) locomotion on day 4 and the challenge day relative to day 1 (Figure 3A; main effect of 'Time': $F_{2,50} = 10.84$, $p < 0.001$). There was no significant effect of 'Lineage' or 'Time x Lineage' interaction for F2 males. Similarly, greater NIC-induced locomotion was exhibited on day 4 and the challenge day, relative to day 1, by F3 males (Figure 3B, left; main effect of 'Time': $F_{2,44} = 20.50$, $p < 0.001$) and females (Figure 3B, right; main effect of

'Time': $F_{2,48} = 18.68$, $p < 0.001$). There were no significant effects of 'Lineage' or 'Time x Lineage' interaction for F3 offspring derived from the F2 male lineages.

F0 nicotine and F1 female stress exposure increased locomotor sensitization to nicotine in F3 female offspring.

As with the males, F2 females displayed greater NIC-induced locomotion on the challenge day, relative to day 1, indicating locomotor sensitization (Figure 3C; main effect of 'Time': $F_{2,40} = 4.87$, $p < 0.05$), but there was no significant effect of 'Lineage' or 'Time x Lineage' interaction. F3 male offspring derived from the F2 females also displayed increased NIC-induced locomotion on day 4 as compared to day 1 and the challenge day relative to days 1 and 4 (Figure 3D, left; main effect of 'Time': $F_{2,46} = 18.60$, $p < 0.001$) with no significant effect of 'Lineage' or 'Time x Lineage' interaction. Finally, F3 female offspring derived from the F2 female lineages displayed greater NIC-induced locomotion on day 4 and the challenge day relative to day 1 (main effect of 'Time': $F_{2,48} = 37.46$, $p < 0.05$). This change in NIC-induced locomotion was dependent on lineage (Figure 3D, right; 'Time x Lineage' interaction: $F_{6,48} = 3.93$, $p < 0.05$). Specifically, NIC-induced locomotion did not differ across the three test days for F3 female offspring of the F0 VEH/F1 CON female lineage. Alternatively, F3 female offspring derived from the F0 VEH/F1 CUS female lineage displayed greater NIC-induced locomotion on the challenge day relative to day 1 ($p < 0.01$). F3 females derived from the F0 NIC-exposed female lineages (i.e., F0 NIC/F1 CON and F0 NIC/F1 CUS) displayed greater NIC-induced locomotion on day 4 and the challenge day compared to day 1 (p 's < 0.001).

Discussion

Transgenerational inheritance of environmental exposures from parent to offspring suggests that the quality of the offspring's life can be affected by the actions and experiences of the parents (Skinner et al., 2014). The use of rodent models of environmental exposures has allowed for substantial progress to be made studying the genetic and epigenetic inheritance of parental exposures to exogenous stimuli. Parental exposure to changes in diet (Ng et al., 2010), environmental toxins (Skinner et al., 2014), and stress (Dietz et al., 2011; Franklin et al., 2010; Rodgers et al., 2013; Saavedra-Rodríguez & Feig, 2013; Yehuda et al., 1998) promote altered behavior, physiology, and disease predisposition in the offspring of future generations. These findings have been extended to drugs of abuse. The effects of parental exposure to cocaine (Vassoler, White, Schmidt, Sadri-Vakili, & Pierce, 2013), morphine (Byrnes, Johnson, Carini, & Byrnes, 2013), cannabinoids (Szutorisz et al., 2014), and alcohol (Finegersh & Homanics, 2014) have been studied and each demonstrates some impact on offspring behavior. NIC exposure may also impact multi- and transgenerational inheritance. For example, mice exposed to NIC *in utero* produce two generations of offspring with hyperactivity (Zhu, Lee, Spencer, Biederman, & Bhide, 2014), altered metabolism (Holloway, Cuu, Morrison, Gerstein, & Tarnopolsky, 2007), and a predisposition to respiratory disease (Rehan et al., 2012).

We utilized a novel multigenerational exposure paradigm to identify the effects of two environmental exposures during a vulnerable developmental window on behavior in future

generations of offspring. F0 NIC and F1 CUS exposure produced striking changes in the F2 and F3 offspring's NIC responses. By including both males and females in our analysis, we considered sex as a relevant biological variable that could affect exposure inheritance across generations (Clayton, 2016). As a result, our work is the first to assess transgenerational the effects of NIC and stress exposure across three generations in both male and female mice.

Despite epidemiological evidence supporting the inheritance of NIC exposure in offspring (Hillemacher et al., 2008; Mill & Petronis, 2008), only a few studies have utilized rodent models to examine the multi- and transgenerational effects of NIC exposure (Holloway et al., 2007; Rehan et al., 2012; Zhu et al., 2014). For the most part these previous studies used *in utero* NIC administration paradigms and produced mice that were directly exposed to drug. NIC is known to cross the placental barrier and thus can directly affect the developing fetus (Jordanov, 1990). In addition, *in utero* exposure paradigms include maternal responses and distress (Lambers & Clark, 1996) that could influence fetal development and have a major impact on behavior. To eliminate these factors and evaluate the inheritance of postnatal NIC exposure, we exposed F0 male mice to NIC via osmotic minipumps during adolescence, removed the pumps following 1-month of exposure, and mated them to produce a generation of F1 offspring.

We focused exclusively on paternal NIC exposure in the F0 generation. When drug exposure occurs in F0 males, the germ cells that produce the F1 generation are also “exposed”. Therefore, phenotypes found in F0 and F1 animals are multigenerational. In contrast, phenotypes in F2 animals and all subsequent generations are transgenerational; F2 animals are the first generation whose cells have not been exposed to drug (for review see Yohn, Bartolomei, & Blendy, 2015). Further, males were no longer administered NIC at mating and were removed from the mating cage prior to the birth of pups. By ensuring no interaction with NIC-exposed fathers and a short mating window, we attempted to minimize the impact of paternal NIC exposure on maternal care (Curley, Mashoodh, & Champagne, 2011).

The experience of stress is a particularly salient environmental exposure in both humans and animals. Chronic stress promotes maladaptive responses and disease states in the individual exposed to stress (McEwen & Stellar, 1993) as well as altered physiology and behavior in several generations of offspring in both humans and animals (Dietz et al., 2011; Franklin et al., 2010; Rodgers et al., 2013; Saavedra-Rodríguez & Feig, 2013; Yehuda et al., 1998). However, we wanted to know if stress exposure across generations would interact with NIC to influence subsequent behavior. Previous work by Zhu and colleagues (2014) identified a hyperactive phenotype in the male and female offspring of mice that were exposed to NIC *in utero*. Thus, we decided to focus on NIC-induced locomotor activity.

The influence of F0 NIC exposure on subsequent generations was dependent on both the sex of the F1 parents and their offspring. When derived from the F1 male CON/CUS lineage, the F2 females displayed increased sensitivity the locomotor effects of NIC, as indicated by a robust increase in NIC-induced locomotion on day 4. These results reflect an alteration in the *induction of sensitization*. However, the *expression of sensitization* was not affected by F0 NIC exposure, as the F2 females' NIC-induced locomotor responses on the challenge day

were similar to that of day 1. Notably, there are unique NIC-induced neurobiological adaptations that contribute to the induction, but not the expression, of sensitization. Furthermore, these adaptations are required, but they are not sufficient for the expression of sensitization (DiFranza & Wellman, 2007). As such, it is possible that the decreased response to nicotine on the challenge day results from a lack of maintenance of the sensitized response to repeated NIC exposures and may reflect the development of tolerance (Tapper et al., 2004). The influence on the induction of sensitization was transient and no changes were observed in the F3 offspring.

Surprisingly, when offspring were derived from the F1 female CON/CUS lineage the effects of F0 NIC exposure manifested as an enhancement in the induction and expression of sensitization one generation removed in the F3 offspring, and then only in females. Sex differences in the regulation of central dopaminergic neurotransmission may be a contributing factor in modulating nicotine locomotor responses. For example, the increase in the extracellular dopamine concentration in the nucleus accumbens has been reported to be higher in female rats than in male rats following systemic nicotine administration (Pogun, 2001). The long-lasting facilitation of locomotor sensitization in these F3 females suggests that NIC exposure might target unique neurobiological processes when transmitted through the female lineage. The two main phases of locomotor sensitization are the induction and expression (Robinson, Browman, Crombag, & Badiani, 1998; Todtenkopf, Mihalakopoulos, & Stellar, 2002). The induction phase relies on behavioral and physiological events elicited by the repeated psychostimulant administration, resulting in enduring neurochemical, molecular and, depending on drug class, morphological alterations in mesolimbic-cortical pathways. During expression, it is thought that the long-term changes developed during induction are now consolidated giving rise to the subsequent sensitization responses observed weeks to months after the last drug exposure (Robinson & Berridge, 1993). Thus, prior exposure to nicotine and stress, may differentially impact these mechanisms in males and females.

Future studies are required to address mechanisms underlying lineage-dependent effects of F0 NIC exposure. Epigenetic modifications could underlie the longevity of psychiatric conditions such as drug abuse both within, as well as across generations (Nielsen, Utrankar, Reyes, Simons, & Kosten, 2012). Studies show that non-imprinted genes and repetitive genomic elements can escape loss of methylation patterning that typically occurs following reprogramming events (Lane et al., 2003; Orozco et al., 2014). Thus, the retention of genomic methylation patterns in sperm of exposed parents and brains of offspring may occur following generational stress (Franklin et al., 2010) or drug-exposure (Govorko, Bekdash, Zhang, & Sarkar, 2012). Interestingly, Dai et al. (2017) found that paternal nicotine exposure downregulates miR-15b expression, due to DNA hypermethylation, in both F0 sperm and the thalamus of F1 offspring. Further, virally-mediated overexpression of thalamic miR-15b also prevented the reduction in anxiety- and depression-like behavior that was displayed by F1 males following F0 NIC exposure. As such, our findings may reflect the contribution of similar epigenetic modifications that arise from the female germline. To date, few studies have examined the transgenerational effects of F0 exposure in females, possibly due to the confound of maternal care, even if offspring are cross-fostered, and the effect on F1 generations.

Overall, our findings raise important questions about the impact of parental exposure of drugs on their offspring's susceptibility to responses to similar agents. However, caution must be exercised when interpreting these results. Given that sensitization is associated with neurobiological adaptations in the same brain reward circuits that are implicated in addiction (DiFranza & Wellman, 2007; Robinson & Berridge, 1993), it is tempting to speculate that enhanced NIC sensitization translates to increased risk for NIC addiction in F2/F3 offspring. However, it is unknown whether sensitization occurs in human smokers. Theories postulate NIC sensitization enhances the salience of reward-related cues associated with NIC. These sensitized cues may drive compulsive craving and NIC-seeking behavior (Robinson & Berridge, 1993). Alternatively, sensitization may enhance NIC's ability to suppress the generation of cravings. Repeat NIC exposure would therefore result in a homeostatic imbalance that, in the absence of NIC, enhances craving and drives NIC-seeking behavior (DiFranza & Wellman, 2007). Few, if any, studies have tested these theories experimentally. In lieu of empirical evidence supporting a role for sensitization in NIC addiction, any such interpretation remains speculative.

Conclusion

In this study we exposed F0 males to nicotine and F1 males to stress and determined the transgenerational interaction of nicotine and stress on offspring behavior. Remarkably, we found that environmental exposures are subject to cross-generational inheritance and produce unique phenotypes in offspring. In addition, we identified novel phenotypes in several generations of offspring derived from paternal nicotine exposure. When considering these results, we might infer that an epigenetic mechanism is in play as our behaviors occur over multiple generations. Importantly, the occurrence of a behavioral phenotype may occur proximal to the exposure (e.g., from F1 male CON/CUS to the next F2 generation) or it may skip generations (e.g., from F1 female CON/CUS to the F3 generation). Future work to mechanistically identify and probe cellular changes that mediate the phenotypes characterized for functional significance will greatly add to our knowledge of transgenerational interactions and reprogramming of the offspring brain.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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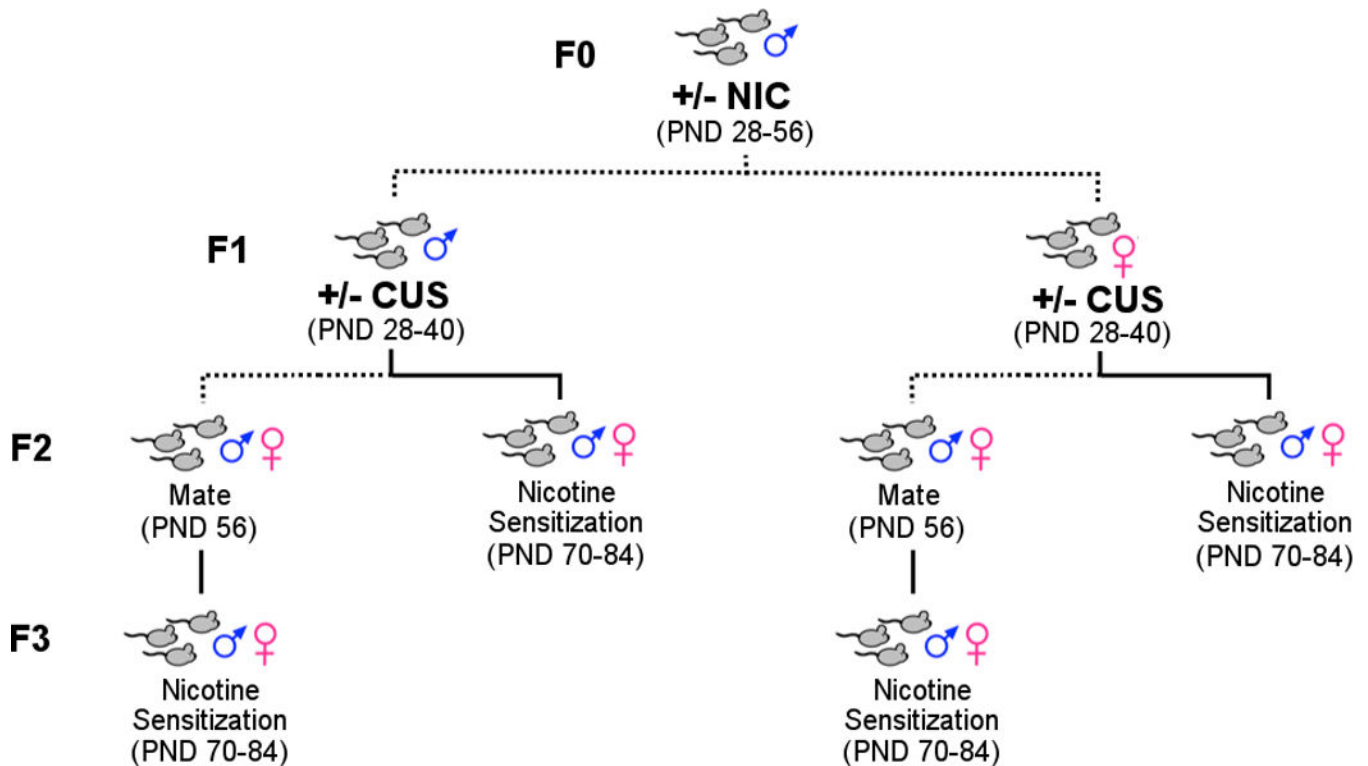


Figure 1.

Schematic of experimental design. F0 male mice were exposed to vehicle (VEH, n=5) or nicotine (NIC, n=7) and mated (dashed line) to produce F1 offspring. Resulting F1 males and females were randomly distributed to chronic unpredictable stress (CUS) or no stress control (CON) condition and mated to produce F2 offspring. Of note, mice from a given litter were distributed evenly into CUS or CON groups, such that the resulting CUS or CON groups were comprised of mice from at least 5–7 different litters. One half of the F2 offspring were tested for locomotor sensitization to NIC (solid line) while the other half of the behaviorally-naïve F2 mice were mated to produce the F3 offspring. As before, F3 mice from a single litter were evenly distributed to nicotine or saline treatments and tested for locomotor sensitization.

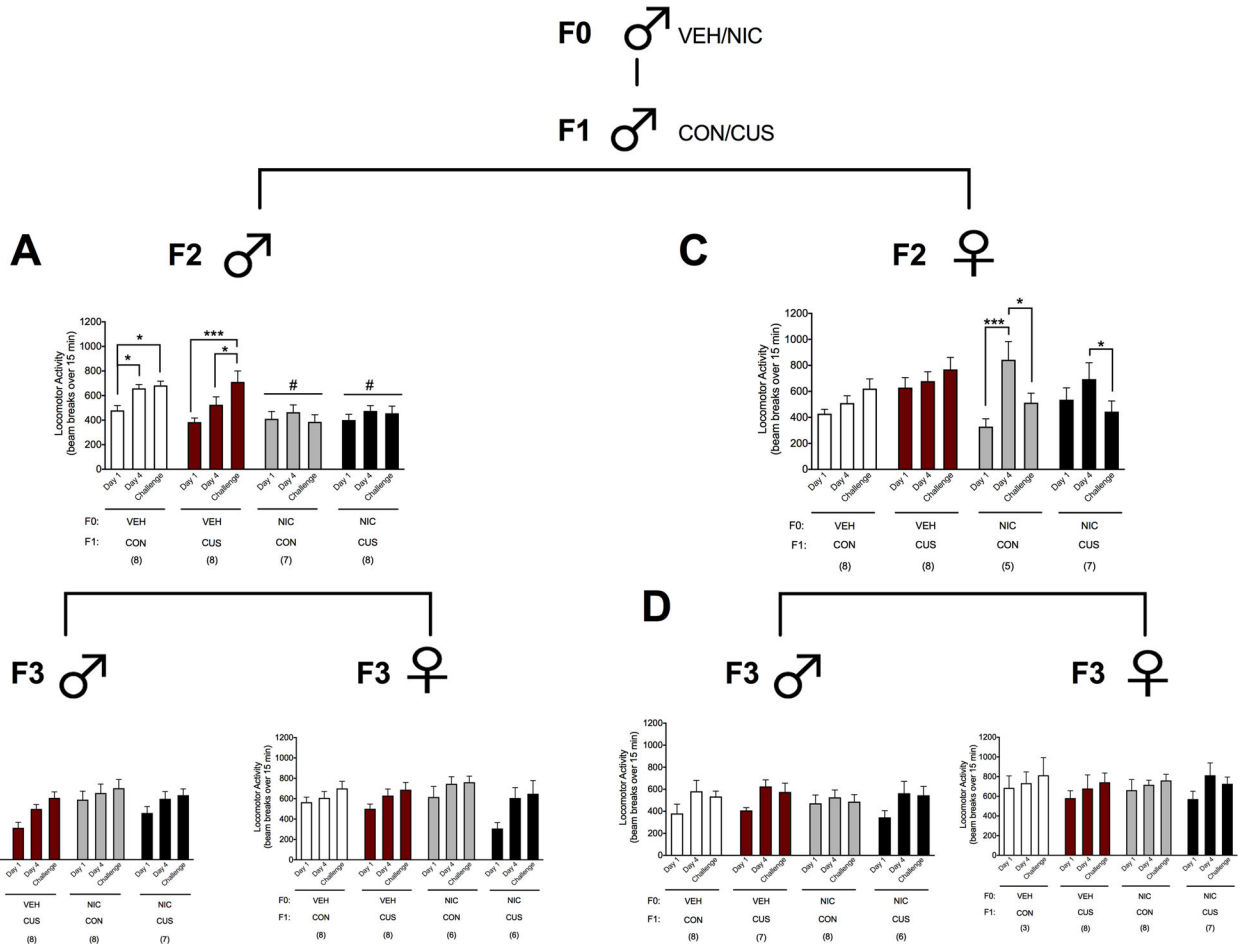


Figure 2. F0 NIC exposure prevents locomotor sensitization to NIC in male and female F2, but not F3, offspring regardless of male F1 CUS exposure. Data (mean ± SEM) represent NIC-induced (1 mg/kg, i.p.) locomotion in (A) F2 male offspring derived from the F1 male lineage, (B) F3 male and female offspring of the F2 male lineage, (C) F2 female offspring derived from the F1 male lineage, and (D) F3 male and female offspring of the F2 female lineage. Significant main effect of ‘Lineage’: # $p < 0.01$. Significant ‘Time x Lineage’ interaction: * $p < 0.05$, *** $p < 0.001$. Sample sizes are reported in parentheses in the figure.

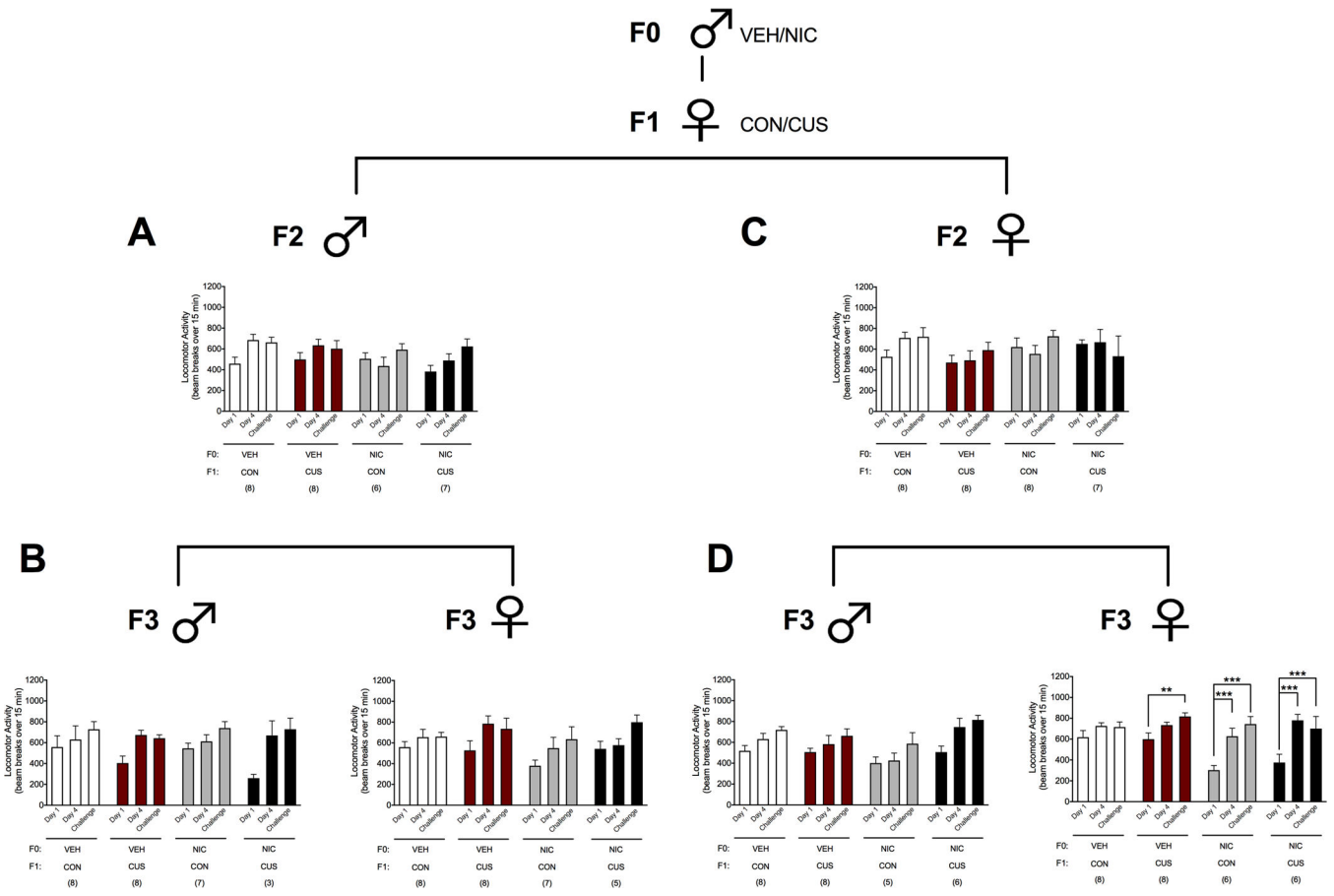


Figure 3. F0 NIC and female F1 CUS exposure enhances locomotor sensitization to NIC in female F3, but not F2, offspring. Data (mean ± SEM) represent NIC-induced (1 mg/kg, i.p.) locomotion in (A) F2 male offspring derived from the F1 female lineage, (B) F3 male and female offspring of the F2 male lineage, (C) F2 female offspring derived from the F1 male lineage, and (D) F3 male and female offspring of the F2 female lineage. Significant ‘Time x Lineage’ interaction: ***p* < 0.01, ****p* < 0.001. Sample sizes are reported in parentheses in the figure.

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