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Mechanisms of resistance in estrogen receptor positive breast cancer: overcoming resistance to tamoxifen/aromatase inhibitors

Jamie N Mills¹, Alex C Rutkovsky², and Antonio Giordano¹

¹Medical University of South Carolina, Department of Medicine, Division of Hematology and Oncology, 39 Sabin St. MSC 635, Charleston, SC 29425, USA

²Medical University of South Carolina, Department of Pathology and Laboratory Medicine, 39 Sabin St, Charleston, SC 29425, USA

Abstract

Several mechanisms of resistance have been identified, underscoring the complex nature of estrogen receptor (ER) signaling and the many connections between this pathway and other essential signaling pathways in breast cancer cells. Many therapeutic targets of cell signaling and cell cycle pathways have met success with endocrine therapy and remain an ongoing area of investigation. This review focuses on two major pathways that have recently emerged as important opportunities for therapeutic intervention in endocrine resistant breast tumors: PI3K/AKT/mTOR cell signaling and cyclinD1/cyclin-dependent kinase 4/6 cell cycle pathways. Additionally, we highlight individual and combination strategies in current clinical trials that target these pathways and others under investigation for the treatment of ER positive breast cancer.

Introduction

Endocrine therapy has dramatically improved survival in breast cancer patients over the past several decades, however resistance to these therapies remains one of the major causes of breast cancer mortality today [1]. Late recurrence and death from estrogen receptor positive (ER+) breast cancer can occur for at least 20 years after the original diagnosis even after 5 years of adjuvant endocrine therapy [2●●]. Identifying mechanisms of resistance and strategies by which to combat these mechanisms is paramount to patient survival.

Several mechanisms of resistance to endocrine therapies have been identified, many centered around the structure, activation, and complex functions of ER, as well as cross-talk between the estrogen signaling network and other cellular pathways. The major form of ER in breast cancer is ER α , encoded by *ESR1*, and the major function of ER is as a transcription factor controlling genes associated with cell survival and proliferation [3]. ER function is influenced by circulating estrogens and related molecules, giving ER-targeted therapies their

Corresponding author: Giordano, Antonio (giordana@musc.edu).

Conflict of interest statement

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success. Post-translational modifications also influence function, localization, and interaction with other regulators. In addition to ER α , there also exists transcription factor ER β , encoded by *ESR2*, as well as alternatively spliced and truncated variants of *ESR1/ER α* [4]. The biology of ER is complex and how breast tumors can gain ER function then maintain this despite ER inhibition is not well understood. Thus, the mechanism of action of various endocrine therapies is complicated, varies, and remains an active area of investigation.

Known mechanisms of resistance to hormone therapies are complex and include epigenetic regulation of *ESR1* expression [5,6], *ESR1* mutations [7–12], alternative splicing events [3], *ESR1* truncation and fusion events [13], post-translational modifications [14,15], alterations in the hormone binding domain [7,16], alter-native recruitment sites within the genome [17], differential recruitment of coregulators [18], feedback loops by ER target genes on expression/activity of ER [19[●]], downstream actions of ER target genes on growth factor pathways and other signaling networks [20,21^{●●}], influences of the tumor microenvironment [22], and many others (Figure 1). The details of these mechanisms are beyond the scope of this review but have been thoroughly described by others [23,24]. The complexity of ER function in tumor cells underscores the heterogeneity of breast cancer biology and demonstrates a necessity for continued basic research and clinical demonstration to effectively target the pathways essential to tumor cell survival.

Much literature exists detailing the mechanisms of resistance. This review highlights two major pathways that have recently emerged as important opportunities for therapeutic intervention in endocrine resistant breast tumors: the phosphatidylinositol 3-kinase (PI3K)/AKT/mammalian target of rapamycin (mTOR) cell signaling pathway and the cyclin D1/cyclin-dependent kinase (CDK)4/6 cell cycle pathway. Inhibitors to these pathways have been developed, assessed in preclinical studies, and investigated in multiple clinical trials, each with marked benefit toward improving survival but also with specific challenges and limitations discussed below.

PI3K/AKT/mTOR pathway

The PI3K/AKT/mTOR pathway is essential for cell growth and survival, protein synthesis, and glucose metabolism. It is dysregulated in many tumor types, prompting investigation of inhibition of this pathway in many cancer models, including the triple negative subtype of breast cancer [25]. Cross-talk between this pathway and estrogen-mediated signaling is known [26,27], prompting investigation of inhibitors targeting this pathway in ER+ breast tumors. Genomic alterations in *PIK3CA*, the gene encoding the catalytic subunit of PI3K, are common in ER+ tumors [28[●],29,30]. It is thought that suppression of the PI3K/AKT/mTOR pathway can lead to increased activity of ER, facilitating resistance, and therefore many groups have used preclinical models to elucidate the mechanisms of PI3K/AKT/mTOR pathway inhibitors and the influence these compounds have on estrogen-mediated signaling [19[●],21^{●●},25,31–33].

Evidence suggests that, while inhibition of the PI3K/AKT/mTOR pathway at all levels results in reduced cell proliferation and survival, the complexity of this signaling pathway

ensures that compensatory mechanisms are activated that confer resistance to single inhibitors [32,33]. For example, inhibition of mTOR results in activation of AKT as well as extracellular signal-regulated kinase (ERK), leading to increased signaling through alternative branches of these pathways [19●,21●●]. Similarly, inhibition of PI3K activates upstream tyrosine kinases, allowing cells to escape the inhibitory effects on cell proliferation [21●●]. AKT, after activation by PI3K, can phosphorylate ER at S167, resulting in ligand-independent activation of ER-mediated transcription [34,35]. Ribas *and colleagues* demonstrated that inhibition of AKT with AZD5363 re-sensitizes cells to tamoxifen, acts synergistically with fulvestrant, and prevents emergence of hormone-independent cells *in vivo*. However, in their model, AKT inhibition activated positive feedback loops driven by MYC, resulting in increased gene expression of human epidermal receptor growth factor 2 and 3 (ERBB2–ERBB3), ERK5, and insulin-like growth factor 1 (IGF1) [19●]. These observations have led to investigation of the use of multiple inhibitors simultaneously and development of novel inhibitors based on alternative iterations of PI3K ligand binding pockets generated by mutations in *PIK3CA*. Cross-talk between the PI3K/AKT/mTOR and ER pathways has immense clinical relevance, but further investigation is required to fully understand the interaction between these pathways. We are not aware of active trials combining a mTOR inhibitor and AKT inhibitor but await the results of combined mTOR inhibitors with PI3K inhibitors.

Several inhibitors of the PI3K/AKT/mTOR pathway have been tested in clinical trials in ER + tumors in conjunction with various endocrine therapies. In 2012, the mTOR inhibitor everolimus was approved by the Food and Drug Administration (FDA) for the treatment of postmenopausal women with advanced ER+/HER2 breast cancer in combination with exemestane. The approval was based on a randomized double-blind multicenter trial (BOLERO-2), where the combination significantly improved disease free survival (DFS) compared to exemestane alone (7.8 months versus 3.2 months) [36]. Recently, the results from a phase 3 trial involving buparlisib, a pan-PI3K inhibitor targeting all four isoforms of class I PI3K (α , β , δ , and γ), were presented and published [37]. Although the efficacy data of buparlisib supports the use of PI3K inhibitors plus endocrine therapy in patients with *PIK3CA* mutations, the safety profile of buparlisib plus fulvestrant did not support its further development in that setting. Pan-PI3K inhibitors such as buparlisib might be limited by adverse events arising from a broad spectrum of off-target effects [38]. PI3K α -selective inhibitors are currently under clinical evaluation in combination with endocrine therapy in patients with ER+, HER2-negative breast cancer. Table 1 summarizes the ongoing clinical trials targeting PI3K/AKT/mTOR.

CyclinD1/CDK4/6 pathway

Cell cycle progression is regulated by many proteins, including the cyclins and cyclin-dependent kinases (CDK). Cyclin D1 binds to CDK4/6 to regulate progression through the G1 phase of the cell cycle and is a known downstream target of the PI3K/AKT/mTOR pathway [39]. CDK4/6 has also been shown to crosstalk with the ER signaling pathway [40]. Vora *and colleagues* showed that insensitivity to PI3K inhibitors is evident by persistent RB phosphorylation and can be effectively overcome by combining a CDK inhibitor with a PI3K inhibitor [40]. Since tumorigenesis relies heavily on unchecked progression through

the cell cycle, the cyclinD1/CDK4/6 pathway has emerged as a desirable target in the treatment of many cancers, including breast [41–43]. Originally, inhibitors such as palbociclib were hypothesized to be most efficacious in the triple negative subtype of breast cancer, but early studies identified ER expression as highly correlated with response to palbociclib [44]. Over the past 3 years, multiple large-scale clinical trials have demonstrated that inhibiting CDK4/6 leads to significant clinical benefit when combined with standard endocrine therapies in metastatic ER+, HER2-negative breast cancer [45–52]. Currently CDK4/6 inhibitors, including palbociclib, ribociclib and abemaciclib, are being increasingly employed in clinical practice combined with endocrine therapy. Also, abemaciclib was approved as a monotherapy in ER+, HER2-negative metastatic breast cancer whose disease progressed during or after endocrine therapy [50]. However, CDK4/6 inhibitors typically induce tumor stabilization with only modest increased rates of tumor shrinkage and their cytostatic effects are limited by primary and acquired resistance. Primary and acquired resistance to CDK4/6 inhibitors mediated by loss of the RB1 gene has been demonstrated in preclinical models [53,54,55]. In a recent study, the emergence of acquired resistance to palbociclib or ribociclib with the concurrent development of multiple de novo somatic RB1 mutations was shown in metastatic ER+ patients [56]. Table 2 summarizes the ongoing clinical trials with CDK4/6 inhibitors.

Conclusions

Resistance to endocrine therapy remains the most significant challenge in treatment of ER+ breast tumors. Despite this resistance, ER signaling remains biologically significant in these cells, and full understanding of the complex signaling pathways responsible for tumor progression remain to be elucidated. There is a lack of evidence informing optimal sequencing of available therapies in the treatment of advanced ER+ breast cancer [57]. Currently, the working hypothesis within the field is that multiple signaling pathways must converge on essential biological functions such as cell cycle progression, cell survival, and estrogen-independent ER-mediated transcription, such that effective blockade of tumor progression will only be achieved by combination therapies that affect all compensatory mechanisms and alternative pathways. Breast cancer is a highly heterogeneous disease with novel essential factors constantly being discovered at the preclinical level. This is seen here by combination studies that include hormone therapy with addition of inhibitors targeting RAS/rapidly accelerated fibrosarcoma (RAF)/mitogen-activated protein kinase (MAPKinase), MAPK kinase (MEK), Vascular Endothelial Growth Factor (VEGF), other growth factor receptor inhibitors (e.g. IGF, Insulin Growth Factor; FGFR, Fibroblast Growth Factor Receptor) [23], and other immunotherapy inhibitors [58]. Additionally, more potent and bioavailable selective estrogen receptor degraders (SERD) are being tested to overcome resistance induced by ESR1 mutations [10]. Advances in genomic and other technologies that allow deeper understanding of individual tumors and further investigation into the cross-talk between these signaling pathways have provided a plethora of data that require continued mining, both in the preclinical and clinical settings, to personalize therapeutic regimens for each patient.

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

● of special interest

●● of outstanding interest

1. Early Breast Cancer Trialists' Collaborative G: Aromatase inhibitors versus tamoxifen in early breast cancer: patient-level meta-analysis of the randomised trials. *Lancet* 2015, 386:1341–1352. [PubMed: 26211827]
2. ●● Pan H, Gray R, Braybrooke J, Davies C, Taylor C, McGale P, Peto R, Pritchard KI, Bergh J, Dowsett M, Hayes DF et al.: 20-year risks of breast-cancer recurrence after stopping endocrine therapy at 5 years. *N Engl J Med* 2017, 377:1836–1846. [PubMed: 29117498] A meta-analysis combining individual patient data from 88 trials in the Early Breast Cancer Trialists' Collaborative Group (EBCTCG) database of randomized trials.
3. Wang ZY, Yin L: Estrogen receptor alpha-36 (ER-alpha36): a new player in human breast cancer. *Mol Cell Endocrinol* 2015, 418(Pt 3):193–206. [PubMed: 25917453]
4. Ma R, Karthik GM, Lovrot J, Haglund F, Rosin G, Katchy A, Zhang X, Viberg L, Frisell J, Williams C, Linder S et al.: Estrogen receptor beta as a therapeutic target in breast cancer stem cells. *J Natl Cancer Inst* 2017, 109:1–14.
5. De Marchi T, Liu NQ, Stingl C, Timmermans MA, Smid M, Look MP, Tjoa M, Braakman RB, Opdam M, Linn SC, Sweep FC et al.: 4-protein signature predicting tamoxifen treatment outcome in recurrent breast cancer. *Mol Oncol* 2016, 10:24–39. [PubMed: 26285647]
6. Zhang J, Zhou C, Jiang H, Liang L, Shi W, Zhang Q, Sun P, Xiang R, Wang Y, Yang S: Zeb1 induces ER-alpha promoter hypermethylation and confers antiestrogen resistance in breast cancer. *Cell Death Dis* 2017, 8:e2732. [PubMed: 28383555]
7. Toy W, Shen Y, Won H, Green B, Sakr RA, Will M, Li Z, Gala K, Fanning S, King TA, Hudis C et al.: ESR1 ligand-binding domain mutations in hormone-resistant breast cancer. *Nat Genet* 2013, 45:1439–1445. [PubMed: 24185512]
8. Jeselsohn R, Buchwalter G, De Angelis C, Brown M, Schiff R: ESR1 mutations — a mechanism for acquired endocrine resistance in breast cancer. *Nat Rev Clin Oncol* 2015, 12:573–583. [PubMed: 26122181]
9. Fribbens C, O'Leary B, Kilburn L, Hrebien S, Garcia-Murillas I, Beaney M, Cristofanilli M, Andre F, Loi S, Loibl S, Jiang J et al.: Plasma ESR1 mutations and the treatment of estrogen receptor-positive advanced breast cancer. *J Clin Oncol* 2016, 34:2961–2968. [PubMed: 27269946]
10. Bihani T, Patel HK, Arlt H, Tao N, Jiang H, Brown JL, Purandare DM, Hattersley G, Garner F: Elacestrant (RAD1901), a selective estrogen receptor degrader (SERD), has antitumor activity in multiple ER+ breast cancer patient-derived xenograft models. *Clin Cancer Res* 2017, 23:4793–4804. [PubMed: 28473534]
11. Toy W, Weir H, Razavi P, Lawson M, Goepfert AU, Mazzola AM, Smith A, Wilson J, Morrow C, Wong WL, De Stanchina E et al.: Activating ESR1 mutations differentially affect the efficacy of ER antagonists. *Cancer Discov* 2017, 7:277–287. [PubMed: 27986707]
12. Takeshita T, Yamamoto Y, Yamamoto-Ibusuki M, Tomiguchi M, Sueta A, Murakami K, Omoto Y, Iwase H: Analysis of ESR1 and PIK3CA mutations in plasma cell-free DNA from ER-positive breast cancer patients. *Oncotarget* 2017, 8:52142–52155. [PubMed: 28881720]
13. Hartmaier RJ, Trabucco SE, Priedigkeit N, Chung JH, Parachoniak CA, Vanden Borre P, Morley S, Rosenzweig M, Gay LM, Goldberg ME, Suh J et al.: Recurrent hyperactive ESR1 fusion proteins in endocrine therapy resistant breast cancer. *Ann Oncol* 2018.

14. Zhang X, Tanaka K, Yan J, Li J, Peng D, Jiang Y, Yang Z, Barton MC, Wen H, Shi X: Regulation of estrogen receptor alpha by histone methyltransferase SMYD2-mediated protein methylation. *Proc Natl Acad Sci U S A* 2013, 110:17284–17289. [PubMed: 24101509]
15. Piggott L, da Silva AM, Robinson T, Santiago-Gomez A, Simoes BM, Becker M, Fichtner I, Andera L, Piva M, Vivanco MD, Morris C et al.: Acquired resistance of ER-positive breast cancer to endocrine treatment confers an adaptive sensitivity to trail through post-translational downregulation of c-FLIP. *Clin Cancer Res* 2018.
16. Harrod A, Fulton J, Nguyen VTM, Periyasamy M, Ramos-Garcia L, Lai CF, Metodjeva G, de Giorgio A, Williams RL, Santos DB, Gomez PJ et al.: Genomic modelling of the ESR1 Y537S mutation for evaluating function and new therapeutic approaches for metastatic breast cancer. *Oncogene* 2017, 36:2286–2296. [PubMed: 27748765]
17. Abou-Kandil A, Eisa N, Jabareen A, Huleihel M: Differential effects of HTLV-1 tax oncoprotein on the different estrogen-induced-ER alpha-mediated transcriptional activities. *Cell Cycle* 2016, 15:2626–2635. [PubMed: 27420286]
18. Cottu PH: Systemic neoadjuvant therapy of luminal breast cancer in 2016. *Bull Cancer* 2017, 104:69–78. [PubMed: 27817858]
19. ●● Ribas R, Pancholi S, Guest SK, Marangoni E, Gao Q, Thuleau A, Simigdala N, Polanska UM, Campbell H, Rani A, Lippardi G et al.: Akt antagonist AZD5363 influences estrogen receptor function in endocrine-resistant breast cancer and synergizes with fulvestrant (ICI182780) in vivo. *Mol Cancer Ther* 2015, 14:2035–2048. [PubMed: 26116361] This study assessed the effects of AKT inhibition in endocrine therapy-resistant cells and how inhibition of this pathway affected ER-mediated signaling.
20. Steelman LS, Martelli AM, Cocco L, Libra M, Nicoletti F, Abrams SL, McCubrey JA: The therapeutic potential of mTOR inhibitors in breast cancer. *Br J Clin Pharmacol* 2016, 82:1189–1212. [PubMed: 27059645]
21. ●● Bosch A, Li Z, Bergamaschi A, Ellis H, Toska E, Prat A, Tao JJ, Spratt DE, Viola-Villegas NT, Castel, Minuesa G et al.: PI3K inhibition results in enhanced estrogen receptor function and dependence in hormone receptor-positive breast cancer. *Sci Transl Med* 2015, 7:283ra251. This study demonstrates the cross-talk between PI3K pathway inhibitors and ER-mediated signaling as it relates to resistance and the need to target these pathways simultaneously.
22. Andre F, Bachelot T, Campone M, Dalenc F, Perez-Garcia JM, Hurvitz SA, Turner N, Rugo H, Smith JW, Deudon S, Shi M et al.: Targeting FGFR with dovitinib (TKI258): preclinical and clinical data in breast cancer. *Clin Cancer Res* 2013, 19:3693–3702. [PubMed: 23658459]
23. Osborne CK, Schiff R: Mechanisms of endocrine resistance in breast cancer. *Annu Rev Med* 2011, 62:233–247. [PubMed: 20887199]
24. Gu G, Dustin D, Fuqua SA: Targeted therapy for breast cancer and molecular mechanisms of resistance to treatment. *Curr Opin Pharmacol* 2016, 31:97–103. [PubMed: 27883943]
25. Politz O, Siegel F, Barfacker L, Bommer U, Hagebarth A, Scott WJ, Michels M, Ince S, Neuhaus R, Meyer K, Fernandez-Montalvan AE et al.: Bay 1125976, a selective allosteric AKT1/2 inhibitor, exhibits high efficacy on akt signaling-dependent tumor growth in mouse models. *Int J Cancer* 2017, 140:449–459. [PubMed: 27699769]
26. Boulay A, Rudloff J, Ye J, Zumstein-Mecker S, O'Reilly T, Evans DB, Chen S, Lane HA: Dual inhibition of mTOR and estrogen receptor signaling in vitro induces cell death in models of breast cancer. *Clin Cancer Res* 2005, 11:5319–5328. [PubMed: 16033851]
27. Miller TW, Hennessy BT, Gonzalez-Angulo AM, Fox EM, Mills GB, Chen H, Higham C, Garcia-Echeverria C, Shyr Y, Arteaga CL: Hyperactivation of phosphatidylinositol-3 kinase promotes escape from hormone dependence in estrogen receptor-positive human breast cancer. *J Clin Invest* 2010, 120:2406–2413. [PubMed: 20530877]
28. ● Zardavas D, Te Marvelde L, Milne RL, Fumagalli D, Fountzilias G, Kotoula V, Razis E, Papaxoinis G, Joensuu H, Moynahan ME, Hennessy BT et al.: Tumor PIK3CA genotype and prognosis in early-stage breast cancer: a pooled analysis of individual patient data. *J Clin Oncol* 2018 JCO2017748301. Clinical study from over 10 000 patients assessing the prognostic value of PIK3CA mutations in different subtypes of breast cancer.
29. Kang S, Bader AG, Vogt PK: Phosphatidylinositol 3-kinase mutations identified in human cancer are oncogenic. *Proc Natl Acad Sci U S A* 2005, 102:802–807. [PubMed: 15647370]

30. Banerji S, Cibulskis K, Rangel-Escareno C, Brown KK, Carter SL, Frederick AM, Lawrence MS, Sivachenko AY, Sougnez C, Zou L, Cortes ML et al.: Sequence analysis of mutations and translocations across breast cancer subtypes. *Nature* 2012, 486:405–409. [PubMed: 22722202]
31. Estevez LG, Garcia E, Hidalgo M: Inhibiting the PI3K signaling pathway: buparlisib as a new targeted option in breast carcinoma. *Clin Transl Oncol* 2016, 18:541–549. [PubMed: 26510854]
32. Choi AR, Kim JH, Woo YH, Cheon JH, Kim HS, Yoon S: Co-treatment of LY294002 or MK-2206 with AZD5363 attenuates AZD5363-induced increase in the level of phosphorylated AKT. *Anticancer Res* 2016, 36:5849–5858. [PubMed: 27793908]
33. Lui A, New J, Ogony J, Thomas S, Lewis-Wambi J: Everolimus downregulates estrogen receptor and induces autophagy in aromatase inhibitor-resistant breast cancer cells. *BMC Cancer* 2016, 16:487. [PubMed: 27421652]
34. Campbell RA, Bhat-Nakshatri P, Patel NM, Constantinidou D, Ali S, Nakshatri H: Phosphatidylinositol 3-kinase/AKT-mediated activation of estrogen receptor alpha: a new model for anti-estrogen resistance. *J Biol Chem* 2001, 276:9817–9824. [PubMed: 11139588]
35. Yamnik RL, Digilova A, Davis DC, Brodt ZN, Murphy CJ, Holz MK: S6 kinase 1 regulates estrogen receptor alpha in control of breast cancer cell proliferation. *J Biol Chem* 2009, 284:6361–6369. [PubMed: 19112174]
36. Yardley DA, Noguchi S, Pritchard KI, Burris HA III, Baselga J, Gnant M, Hortobagyi GN, Campone M, Pistilli B, Piccart M, Melichar B et al.: Everolimus plus exemestane in postmenopausal patients with HR(+) breast cancer: BOLERO-2 final progression-free survival analysis. *Adv Ther* 2013, 30:870–884. [PubMed: 24158787]
37. Di Leo A, Johnston S, Lee KS, Ciruelos E, Lonning PE, Janni W, O'Regan R, Mouret-Reynier MA, Kalev D, Egle D, Csozsi T et al.: Buparlisib plus fulvestrant in postmenopausal women with hormone-receptor-positive, HER2-negative, advanced breast cancer progressing on or after mTOR inhibition (BELLE-3): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol* 2018, 19:87–100. [PubMed: 29223745]
38. Baselga J, Im SA, Iwata H, Cortes J, De Laurentiis M, Jiang Z, Arteaga CL, Jonat W, Clemons M, Ito Y, Awada A et al.: Buparlisib plus fulvestrant versus placebo plus fulvestrant in postmenopausal, hormone receptor-positive, HER2-negative, advanced breast cancer (BELLE-2): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol* 2017, 18:904–916. [PubMed: 28576675]
39. Muise-Helmericks RC, Grimes HL, Bellacosa A, Malstrom SE, Tschlis PN, Rosen N: Cyclin D expression is controlled post-transcriptionally via a phosphatidylinositol 3-kinase/Akt-dependent pathway. *J Biol Chem* 1998, 273:29864–29872. [PubMed: 9792703]
40. Vora SR, Juric D, Kim N, Mino-Kenudson M, Huynh T, Costa C, Lockerman EL, Pollack SF, Liu M, Li X, Lehar J et al.: CDK 4/6 inhibitors sensitize PIK3CA mutant breast cancer to PI3K inhibitors. *Cancer Cell* 2014, 26:136–149. [PubMed: 25002028]
41. Kenny FS, Hui R, Musgrove EA, Gee JM, Blamey RW, Nicholson RI, Sutherland RL, Robertson JF: Overexpression of cyclin D1 messenger RNA predicts for poor prognosis in estrogen receptor-positive breast cancer. *Clin Cancer Res* 1999, 5:2069–2076. [PubMed: 10473088]
42. Kilker RL, Planas-Silva MD: Cyclin D1 is necessary for tamoxifen-induced cell cycle progression in human breast cancer cells. *Cancer Res* 2006, 66:11478–11484. [PubMed: 17145896]
43. Wilcken NR, Prall OW, Musgrove EA, Sutherland RL: Inducible overexpression of cyclin D1 in breast cancer cells reverses the growth-inhibitory effects of antiestrogens. *Clin Cancer Res* 1997, 3:849–854. [PubMed: 9815758]
44. Finn RS, Aleshin A, Slamon DJ: Targeting the cyclin-dependent kinases (CDK) 4/6 in estrogen receptor-positive breast cancers. *Breast Cancer Res* 2016, 18:17. [PubMed: 26857361]
45. Finn RS, Martin M, Rugo HS, Jones S, Im SA, Gelmon K, Harbeck N, Lipatov ON, Walshe JM, Moulder S, Gauthier E et al.: Palbociclib and letrozole in advanced breast cancer. *N Engl J Med* 2016, 375:1925–1936. [PubMed: 27959613]
46. Finn RS, Crown JP, Lang I, Boer K, Bondarenko IM, Kulyk SO, Ettl J, Patel R, Pinter T, Schmidt M, Shparyk Y et al.: The cyclin-dependent kinase 4/6 inhibitor palbociclib in combination with letrozole versus letrozole alone as first-line treatment of oestrogen receptor-positive, HER2-

- negative, advanced breast cancer (PALOMA-1/TRIO-18): a randomised phase 2 study. *Lancet Oncol* 2015, 16:25–35. [PubMed: 25524798]
47. Turner NC, Ro J, Andre F, Loi S, Verma S, Iwata H, Harbeck N, Loibl S, Huang Bartlett C, Zhang K, Giorgetti C et al.: Palbociclib in hormone-receptor-positive advanced breast cancer. *N Engl J Med* 2015, 373:209–219. [PubMed: 26030518]
 48. Cristofanilli M, Turner NC, Bondarenko I, Ro J, Im SA, Masuda N, Colleoni M, DeMichele A, Loi S, Verma S, Iwata H et al.: Fulvestrant plus palbociclib versus fulvestrant plus placebo for treatment of hormone-receptor-positive, HER2-negative metastatic breast cancer that progressed on previous endocrine therapy (PALOMA-3): final analysis of the multicentre, double-blind, phase 3 randomised controlled trial. *Lancet Oncol* 2016, 17:425–439. [PubMed: 26947331]
 49. Hortobagyi GN, Stemmer SM, Burris HA, Yap YS, Sonke GS, Paluch-Shimon S, Campone M, Blackwell KL, Andre F, Winer EP, Janni W et al.: Ribociclib as first-line therapy for HR-positive, advanced breast cancer. *N Engl J Med* 2016, 375:1738–1748. [PubMed: 27717303]
 50. Dickler MN, Tolane SM, Rugo HS, Cortes J, Dieras V, Patt D, Wildiers H, Hudis CA, O'Shaughnessy J, Zamora E, Yardley DA et al.: MONARCH 1, a phase II study of abemaciclib, a CDK4 and CDK6 inhibitor, as a single agent, in patients with refractory HR(+)/HER2() metastatic breast cancer. *Clin Cancer Res* 2017, 23:5218–5224. [PubMed: 28533223]
 51. Sledge GW Jr, Toi M, Neven P, Sohn J, Inoue K, Pivot X, Burdaeva O, Okera M, Masuda N, Kaufman PA, Koh H et al.: MONARCH 2: Abemaciclib in combination with fulvestrant in women with HR+/HER2 advanced breast cancer who had progressed while receiving endocrine therapy. *J Clin Oncol* 2017, 35:2875–2884. [PubMed: 28580882]
 52. Goetz MP, Toi M, Campone M, Sohn J, Paluch-Shimon S, Huober J, Park IH, Tredan O, Chen SC, Manso L, Freedman OC et al.: Monarch 3: Abemaciclib as initial therapy for advanced breast cancer. *J Clin Oncol* 2017, 35:3638–3646. [PubMed: 28968163]
 53. Dean JL, Thangavel C, McClendon AK, Reed CA, Knudsen ES: Therapeutic CDK4/6 inhibition in breast cancer: key mechanisms of response and failure. *Oncogene* 2010, 29:4018–4032. [PubMed: 20473330]
 - 54 ●●. Herrera-Abreu MT, Palafox M, Asghar U, Rivas MA, Cutts RJ, Garcia-Murillas I, Pearson A, Guzman M, Rodriguez O, Grueso J, Bellet M et al.: Early adaptation and acquired resistance to CDK4/6 inhibition in estrogen receptor-positive breast cancer. *Cancer Res* 2016, 76:2301–2313. [PubMed: 27020857] This original work illustrates convergent mechanisms of early adaptation and acquired resistance to CDK4/6 inhibitors and supports the clinical development of combinations of CDK4/6 and PI3K/mTOR inhibitors in ER + cancers.
 55. Johnson J, Thijssen B, McDermott U, Garnett M, Wessels LF, Bernards R: Targeting the RB-E2F pathway in breast cancer. *Oncogene* 2016, 35:4829–4835. [PubMed: 26923330]
 56. Condorelli R, Spring L, O'Shaughnessy J, Lacroix L, Bailleux C, Scott V, Dubois J, Nagy RJ, Lanman RB, Iafrate AJ, Andre F et al.: Polyclonal RB1 mutations and acquired resistance to CDK4/6 inhibitors in patients with metastatic breast cancer. *Ann Oncol* 2017.
 - 57 ●. Pritchard KI, Chia SK, Simmons C, McLeod D, Paterson A, Provencher L, Rayson D: Enhancing endocrine therapy combination strategies for the treatment of postmenopausal HR+/HER2 advanced breast cancer. *Oncologist* 2017, 22:12–24. [PubMed: 27864574] This article reviewed phase III clinical trials evaluating endocrine therapy combination therapy in postmenopausal patients with ER+ HER2-negative breast cancer, with the purpose of providing practical clinical guidance.
 58. Polk A, Svane IM, Andersson M, Nielsen D: Checkpoint inhibitors in breast cancer — current status. *Cancer Treat Rev* 2018, 63:122–134. [PubMed: 29287242]

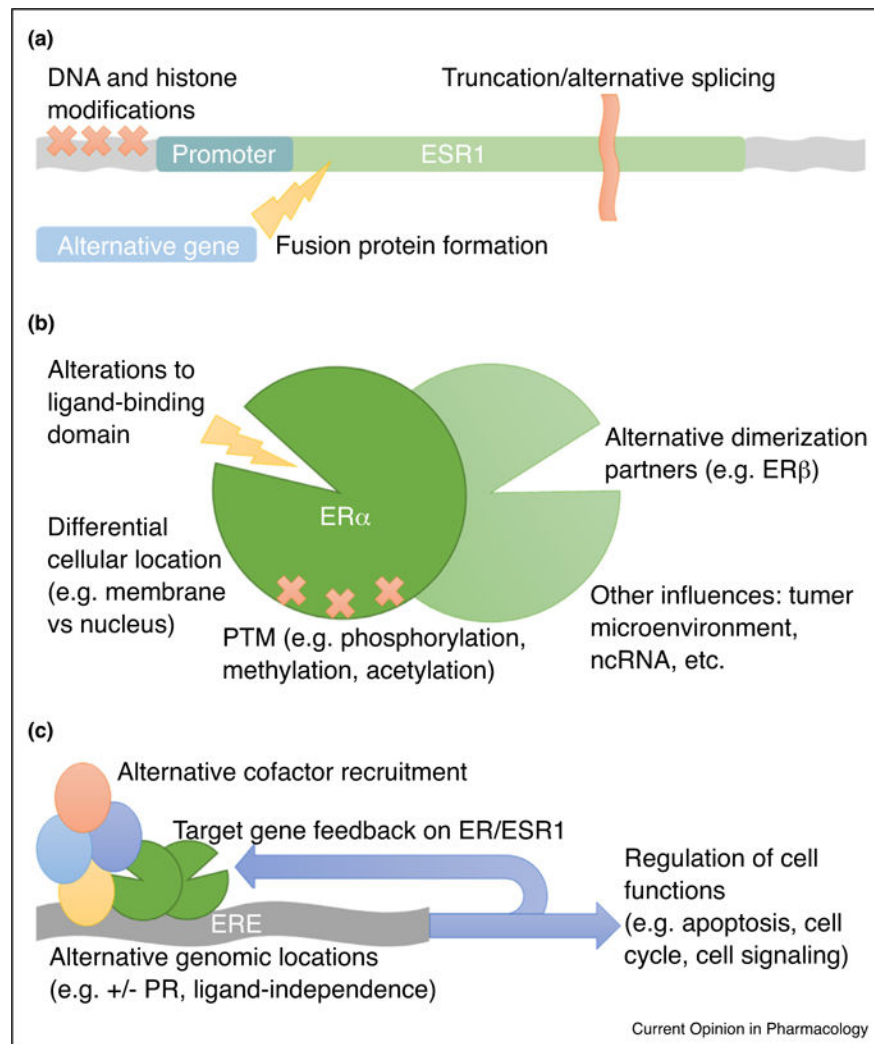


Figure 1.

Diagrammatic representation of known mechanisms of resistance to endocrine therapy. Mechanisms of resistance to endocrine therapy are known to occur at several levels. (a) At the level of ESR1, modifications include epigenetic modification of histone proteins and DNA to alter ESR1 transcription, truncation, alternative splicing events, and fusion events with alternative DNA sequences to produce variations of the ER protein. (b) The events outlined above can result in alterations to the ligand-binding domain and differential cellular localization of the receptor, altering function. Additionally, homo-dimerization or heterodimerization can occur as well as various post-translational modifications (PTM), dependent upon alterations to the proteome of tumor cells and possibly other factors, such as tumor microenvironment and non-coding RNA molecules, resulting in modifications to ER function and target gene selection. (c) Activated ER binds estrogen response elements (ERE) to dictate target genes for transcriptional activation, however alternate recruitment sites are well documented and largely dependent upon recruitment of specific cofactors and tethering proteins, such as progesterone receptor (PR). Differential recruitment of cofactors can also lead to repression of transcription rather than activation, modifying the downstream outcome

of ER activation. Target genes can create feedback loops to modify ESR1/ER expression and behavior, including cross-talk with other signaling pathways in the cell. Finally, all of the events outlined above converge on regulation of essential cellular functions, therefore identifying the many opportunities for development of endocrine therapy resistance has profound effects on clinical control of tumorigenesis.

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Table 1

Summary of ongoing clinical trials involving PI3K/AKT/mTOR in ER+ breast cancer

Target	Compound	NCT number	Population
PI3K	AZD5363, Paclitaxel	NCT01625286	ER+ MBC
	BKM120 or BYL719 and Capecitabine	NCT01300962	
	BYL719, Letrozole or Exemestane	NCT01870505	
	BYL719, Letrozole	NCT01791478	
	GDC-0032, Letrozole or Fulvestrant	NCT01296555	
	Taselisib, Fulvestrant	NCT02340221	
	Alpelisib, Fulvestrant	NCT02437318	
	Alpelisib, hormone therapy	NCT03056755	PIK3CA-mutant
	BYL719, Fulvestrant	NCT01219699	
	AZD8186, Docetaxel	NCT03218826	PTEN or PIK3CB mutant
mTOR	Taselisib, Enzalutamide	NCT02457910	Androgen Receptor+ MBC
	Everolimus to adjuvant hormone therapy	NCT01805271	Adjuvant ER+
	Everolimus, standard hormone therapy	NCT01805271	
	TAK-228 then Letrozole	NCT02619669	
	TAK-228, Tamoxifen	NCT02988986	ER+ MBC
	Everolimus, Letrozole	NCT01698918	
	AZD2014, Fulvestrant	NCT02216786	
	Everolimus, Exemestane	NCT01783444	
	Everolimus, Tamoxifen	NCT01298713	
	Ridaforolimus, Dalotuzumab, Exemestane	NCT01605396	ER+ MBC
AKT	AZD5363	NCT03310541	AKT-mutant
	AZD5363, Fulvestrant	NCT01992952	ER+ MBC
	AZD5363	NCT01226316	MBC
AKT, PD-L1, MEK, VEGF	MSC2363318A, with Trastuzumab or Tamoxifen	NCT01971515	
	Ipatasertib, Atezolizumab, Cobimetinib, Bevacizumab	NCT03395899	Neoadjuvant ER+
PI3K, mTOR, CDK	Ribociclib, Everolimus, Exemestane	NCT01857193	ER+ MBC
	Gedatolisib, Fulvestrant, Palbociclib, Goserelin	NCT02626507	
PI3K, CDK	Ribociclib, Fulvestrant and BYL719 or BKM120	NCT02088684	

Target	Compound	NCT number	Population
	Ribociclib, Letrozole, BYL719	NCT01872260	
	Ribociclib, Tamoxifen, Letrozole, Anastrozole, Goserelin	NCT02278120	
mTOR, CDK	Ribociclib, Everolimus, Exemestane	NCT02732119	
	Hormone therapy, hormone therapy with Everolimus, or hormone therapy with CDK4/6 inhibition	NCT02753686	

MBC, metastatic breast cancer; IGF1R, Insulin Growth Factor 1 Receptor; PD-L1, programmed death-ligand 1; MEK, Mitogen-activated protein kinase kinase. Table updated from clinicaltrials.gov on March 11, 2018.

Table 2

Ongoing clinical trials involving CDK inhibitors in ER+ breast cancer

Target	Compound	NCT number	Population
CDK	Ribociclib, Hormone therapy	NCT03285412	Adjuvant ER+
	Ribociclib, Hormone therapy	NCT03078751	
	Ribociclib, Hormone therapy	NCT03081234	
	Palbociclib, Anastrozole, Goserelin	NCT01723774	
	Abemaciclib	NCT02831530	
	Chemotherapy or Ribociclib and Letrozole	NCT03283384	Neoadjuvant ER+
	Fulvestrant with or without Palbociclib	NCT03447132	
	Paclitaxel, Tamoxifen + Palbociclib, Aromatase Inhibitor	NCT02603679	
	+ Palbociclib, Goserelin + Aromatase Inhibitor + Palbociclib		
	Palbociclib, Hormone therapy	NCT01864746	ER+ post-neoadjuvant
Ribociclib, Letrozole	NCT03439046	ER+ MBC	
Ribociclib, Letrozole	NCT01958021		
Ribociclib, Letrozole, Goserelin	NCT02941926		
Ribociclib, Fulvestrant	NCT02422615		
Endocrine therapy plus CDK 4/6	NCT03425838		
Ribociclib, Letrozole, Goserelin	NCT03096847		
Ribociclib or Palbociclib, Fulvestrant	NCT02632045		
Palbociclib, Letrozole	NCT02499146		
Palbociclib, hormone therapy	NCT02040857		
GI38 with Fulvestrant	NCT02983071		
Letrozole alone or with Palbociclib	NCT00721409		
Palbociclib, hormone therapy	NCT03184090		
Palbociclib, Letrozole	NCT01740427		
Palbociclib, Exemestane, Goserelin	NCT02917005		
Palbociclib, Letrozole	NCT02297438		
Palbociclib	NCT03159195		
Palbociclib, Tamoxifen	NCT02668666		
Aromatase inhibitor, Palbociclib	NCT03439735		

Target	Compound	NCT number	Population
	Palbociclib and aromatase inhibitor or Palbociclib and Fulvestrant	NCT03220178	
	Exemestane plus Goserelin with Palbociclib versus Capecitabine	NCT02592746	
	Abemaciclib, Fulvestrant	NCT02107703	
	Abemaciclib	NCT02763566	
	Palbociclib, Letrozole	NCT01684215	
CDK, HER2	Palbociclib, Letrozole, Trastuzumab, Goserelin	NCT02907918	Neoadjuvant ER+/HER2+
	Ribociclib with Trastuzumab or Trastuzumab emtansine	NCT02657343	ER+/HER2+ MBC
	Tucatinib in combination with Palbociclib and Letrozole	NCT03054363	
CDK, PD-1	Ribociclib, PDR001, Fulvestrant	NCT03294694	ER+ MBC
CDK, IGF	Xentuzumab, Abemaciclib, Fulvestrant	NCT03099174	ER+ MBC refractory to CDK4/6
CDK, FGFR	Erdafitinib, Palbociclib, Fulvestrant	NCT03238196	FGFR-amplified ER+

MBC, metastatic breast cancer; PD-1, programmed cell death protein 1; IGF, Insulin Growth Factor; FGFR, Fibroblast Growth Factor Receptor. Table updated from clinicaltrials.gov on March 11, 2018.