

HHS Public Access

Author manuscript *Chembiochem.* Author manuscript; available in PMC 2020 April 01.

Published in final edited form as: *Chembiochem.* 2019 April 01; 20(7): 963–967. doi:10.1002/cbic.201800785.

Electronic and steric control of $n \rightarrow \pi^*$ interactions: stabilization of the α -helix conformation without a hydrogen bond

Nicole A. Wenzell, Himal K. Ganguly, Anil K. Pandey, Megh R. Bhatt, Glenn P. A. Yap, Neal J. Zondlo

Department of Chemistry and Biochemistry, University of Delaware, Newark, DE 19716, United States; zondlo@udel.edu, gpyap@udel.edu

Abstract

The preferred conformations of peptides and proteins are dependent on local interactions that bias the conformational ensemble. The $n \rightarrow \pi^*$ interaction between consecutive carbonyls promotes compact conformations, including a-helix and polyproline II helix. In order to further understand the $n \rightarrow \pi^*$ interaction and to develop methods to promote defined conformational preferences through acyl N-capping motifs, a series of peptides was synthesized in which the electronic and steric properties of the acyl group were modified. Using NMR spectroscopy, van't Hoff analysis of enthalpies, X-ray crystallography, and computational investigations, we observe that more electron-rich donor carbonyls (pivaloyl, *iso*-butyryl, propionyl) promote stronger $n \rightarrow \pi^*$ interactions and more compact conformations, compared to acetyl or to less electron-rich donor carbonyls (methoxyacetyl, fluoroacetyl, formyl). X-ray crystallography indicates a strong, electronically tunable preference for the α -helix conformation, as observed directly on the ϕ and ψ torsion angles. Electron-donating acyl groups promote the α -helical conformation, even in the absence of the hydrogen bonding that stabilizes the α -helix. In contrast, electron-withdrawing acyl groups exhibited more extended conformations. More sterically demanding groups can promote *trans* amide bonds independent of the electronic effect on $n \rightarrow \pi^*$ interactions. Chloroacetyl groups additionally promote $n \rightarrow \pi^*$ interactions via the interaction of the chlorine lone pair with the proximal carbonyl π^* . These data provide additional support for an important role of $n \rightarrow \pi^*$ interactions in the conformational ensemble of disordered or unfolded proteins. Moreover, this work suggests that readily incorporated acyl N-capping motifs that modulate $n \rightarrow \pi^*$ interactions may be employed rationally to promote conformational biases in peptides, with potential applications in molecular design and medicinal chemistry.

Graphical Abstract

How is the a-helix stabilized in the absence of hydrogen bonds? The strength of $n \rightarrow \pi^*$ interactions was systematically modulated via the identity of the peptide acyl N-cap, with strong donors promoting a-helix and polyproline II helix conformations, but weak donors resulting in more extended conformations. These results suggest the broader consideration of specific capping motifs for conformational control of peptides.

conformation

Keywords

amino acids; conformation analysis; hyperconjugation; peptides; stereoelectronic effects

electronics and sterics

The structure of peptides and proteins is dependent on the sum of a host of non-covalent interactions that are individually weak, but that collectively cause biomolecules to adopt a limited number of conformations, in the process solving the Levinthal paradox.^[1] The $n \rightarrow \pi^*$ interaction is a favorable interaction which is manifested locally in proteins between the lone pair (n) of one carbonyl and the π^* molecular orbital of the subsequent carbonyl (Figure 1).^[2] The $n \rightarrow \pi^*$ interaction has been identified as an important determinant of the backbone conformations, particularly in the α -helix and polyproline II helix (PPII) conformations.^[3] Importantly, the $n \rightarrow \pi^*$ interaction provides a significant basis for biases in the protein main chain conformation. Thus, the $n \rightarrow \pi^*$ interaction might especially impact structure in the disordered or unfolded states of proteins,^[3b, 4] due to the reduced long-range contacts and hydrogen bonding in disordered proteins.^[5]

The strength of the $n \rightarrow \pi^*$ interaction is impacted by the identities of both the donor and acceptor carbonyls.^[3a, 6] For example, thioamides function better than amides as $n \rightarrow \pi^*$ electron donors due to the greater nucleophilicity of sulfur and the higher energies of the lone pair orbitals of sulfur compared to oxygen.^[7] Similarly, the strength of the $n \rightarrow \pi^*$ interaction may be modulated by changing the electronic properties of the acceptor carbonyl: esters are better acceptors than amides, with the acceptor strength further enhanced by the incorporation of electron-withdrawing groups on the ester, which render the carbonyl more electrophilic and lower the energy of the π^* orbital.^[8]

We sought to further investigate the nature of the $n \rightarrow \pi^*$ interaction and the ability to electronically tune peptide structure via the systematic modulation of the electronic properties of the donor carbonyl. The incorporation of electron-donating acyl capping groups on the N-terminus should strengthen an $n \rightarrow \pi^*$ interaction, and thus promote α -helix and polyproline II helix conformations. In contrast, the introduction of electron-withdrawing substituents would be expected to weaken the $n \rightarrow \pi^*$ interaction and yield relatively more extended conformations. The N-terminus of peptides is readily modified as the last step of solid-phase peptide synthesis prior to cleavage from resin/side chain deprotection. As such, this approach to control the strength of the $n \rightarrow \pi^*$ interaction via the identity of the acyl Ncapping^[9] group could be used to change the conformational preferences of peptides in a general, predictable, and highly accessible manner.

In order to investigate the effects of modification of N-terminal electronics on peptide conformation, we synthesized a series of peptides based on the 4-nitrobenzoate ester of 4*R*-hydroxyproline (Hyp(4-NO₂-Bz)) (Figure 1). This proline derivative strongly promotes an *exo* ring pucker^[10] and readily crystallizes due to the nitrobenzoate group.^[11] Peptides (Figure 2) were synthesized with pivaloyl, *iso*-butyryl, and propionyl N-caps, which are more electron-donating than the acetyl group typically employed at the N-terminus of peptides. In addition, a series of N-caps was incorporated with more electron-deficient substituents than acetyl, including methoxyacetyl, fluoroacetyl, and formyl groups, as well as dichloro-, trichloro-, difluoro-, and trifluoro- acetyl groups.

All peptides were analyzed by NMR spectroscopy, to quantify how the identity of the acyl N-cap changes the preference for a *trans* amide bond, which provides one measure of the strength of the $n \rightarrow \pi^*$ interaction, with a stronger $n \rightarrow \pi^*$ interaction resulting in a higher population of *trans* amide bond (larger $K_{\text{trans/cis}}$) (Table 1), if all other factors are equal. For monosubstituted acetyl groups, other than chlorine substitution, an electron-donating substituent increased the population of *trans* amide bond, while electron-withdrawing substituents increased the population of *cis* amide bond, relative to acetyl (for N-caps C(O)-CH₂-X, X = -CH₃ > -H > -OMe > -F).

The most strongly *trans*-promoting N-cap was the pivaloyl group, which exhibited no evidence of *cis* amide bond by NMR. Proline *cis-trans* isomerism importantly also depends on steric effects, with larger groups destabilizing the *cis* amide conformation. The pivaloyl group thus was effective in promoting *trans* amide via both electronic and steric effects.^[3h] Similarly, the most *cis*-promoting N-cap was the formyl group, which is both less electron-donating than an acetyl group and is smaller, with both effects leading to a reduced^[3a] preference for a *trans* over a *cis* amide bond.

The effect of steric *versus* electronic effects in impacting *cis-trans* isomerism was most observable in the halogen series, with dihalogenation and trihalogenation increasing the population of *trans* amide bond, despite the expectation of weaker $n \rightarrow \pi^*$ interactions with increasing F or Cl substitution. Notably, fluorine α -substitution on acyl groups has been previously identified to have effects on conformation that are dependent on the F-C-C-O torsion angle, via the balance of attractive and repulsive interactions between the fluorine and the carbonyl.^[6, 12]

In order to further understand how electronic factors impact the $n \rightarrow \pi^*$ interaction, a subset of peptides was examined by temperature-dependent NMR, to determine the effect of substitution on proline *cis-trans* isomerism enthalpies ($H_{trans/cis}$) via van't Hoff analysis (Figure 3, Table 1). These data indicate that more electron-donating substituents on the donor carbonyl increased the enthalpic preference for a *trans* amide bond. In contrast, electron-withdrawing substituents or formyl substitution on the donor carbonyl significantly reduced this preference (CH(CH₃)₂ > CH₂CH₃ > CH₃ ~ CH₂Cl >> CH₂OMe > H > CH₂F). Again, chloromethyl substitution exhibited anomalous behavior if only electronic effects are considered.

X-ray crystal structures were solved for nearly all derivatives (Figure 4, Table 2), with most structures exhibiting two molecules in each asymmetric section of their unit cell, providing the ability to examine electronic effects on structure in detail. The pivaloyl derivative adopted a polyproline II helix conformation, with a very close 2.68 Å $O_i \dots C_{i+1}$ intercarbonyl distance, indicating a particularly favorable $n \rightarrow \pi^*$ interaction for the most electron-rich donor carbonyl studied. Notably, the iso-butyryl, propionyl, chloroacetyl, acetyl, and methoxyacetyl derivatives all adopted an α -helical conformation in the crystalline form, indicating the ability of the $n \rightarrow \pi^*$ interaction to promote the α -helix independent of hydrogen bonding. The *iso*-butyryl N-cap exhibited quite close $O_{i}...C_{i+1}$ intercarbonyl distances (2.69, 2.74 Å) as well as surprisingly compact values of ϕ (-45°, -47°), which are consistent with particularly favorable $n \rightarrow \pi^*$ interactions. Among -CH₂-X N-caps, the propionyl (2.83, 2.88 Å) and chloroacetyl (2.85, 2.88 Å) groups exhibited the closest intercarbonyl distances, indicating the most favorable $n \rightarrow \pi^*$ interactions, consistent with the observations that these derivatives had the largest enthalpies and free energies favoring *trans* amide bonds. In contrast, the methoxyacetyl N-cap exhibited longer $n \rightarrow \pi^*$ interaction distances (2.89, 3.05 Å), consistent with its less favorable enthalpy and a lower K_{trans/cis}.

The peptide with the fluoroacetyl N-cap exhibited two structures: one was in an α -helical conformation, though with a longer (3.05 Å) intercarbonyl distance and a more extended value of ϕ (-69°, compared to -57° for propionyl); the other structure exhibited an extended conformation ((ϕ, ψ) = -83°, -173°). The trifluoroacetyl and formyl derivatives also exhibited longer intercarbonyl distances and extended conformations, consistent with weak $n \rightarrow \pi^*$ interactions in these more electron-poor derivatives. The observation of extended conformations is noteworthy given the inherent preference of the 4*R*-nitrobenzoate ester of Hyp for an *exo* ring pucker, and thus for more compact conformations. While crystal packing obviously impacts the specific details of the structures observed, collectively these data are consistent with a substantial ability of donor carbonyl electronic properties to modulate the strength of the $n \rightarrow \pi^*$ interaction, and thus the preference for a compact versus extended backbone conformation.

In order to obtain further insights into the effect of N-cap identity on the $n \rightarrow \pi^*$ interaction, all derivatives were examined computationally, using DFT (M06–2X) and MP2 methods.^[13] Structures were examined as X-Pro-OMe and X-Flp-OMe peptides, with an *exo* ring pucker. Initial models were developed using crystallographically observed structures and then subjected to geometry optimization. 4*R*-Fluoroproline (Flp) and 4*R*-hydroxyproline nitrobenzoate have similar effects on conformation.^[10] Therefore, Flp was used in modeling, due to its much lower computational cost, which allowed the use of larger basis sets and analysis using the MP2 method, which better addresses electron correlation than DFT methods. Peptides were analyzed with an *exo* ring pucker in both the α -helix and PPII conformations, in order to specifically examine effects on both secondary structures stabilized by $n \rightarrow \pi^*$ interactions.

Computational results corroborated observations by NMR spectroscopy and X-ray crystallography (Table 2, Figure 5). Using the distance between the donor carbonyl O (O_{*i*}) and the acceptor carbonyl C (C_{*i*+1}) as a reporter of $n \rightarrow \pi^*$ interaction strength, the pivaloyl

group exhibited the closest $O_i...C_{i+1}$ distances, while the formyl group exhibited the longest $O_i...C_{i+1}$ distances, with the distances $Piv > iBut > CH_2CH_3 > CH_3 > CH_2Cl \sim CH_2OMe \sim CH_2F > CHF_2 \sim CF_3 > H$. In addition, a stronger donor resulted in a more compact geometry in a given conformation (e.g. PPII conformation, by DFT: propionyl (ϕ, ψ) = -58°, +143°, versus formyl (ϕ, ψ) = -64°, +151°).

The computational results also provided insights into the basis for the higher-than-expected preference of the chloroacetyl group for a *trans* amide bond and a close $n \rightarrow \pi^*$ interaction. This derivative exhibited a Cl-C-C-O torsion angle of ~ -90° both crystallographically and computationally. A torsion angle scan confirmed that this conformation was preferred.^[14] NBO analysis^[15] revealed that this conformation allows for favorable orbital overlap between one chlorine lone pair (n) and the donor carbonyl π^* orbital, as an $n \rightarrow \pi^*$ interaction with an NBO interaction energy of 2.5 kcal mol⁻¹. These results suggest that the chloroacetyl group enhances the carbonyl/carbonyl $n \rightarrow \pi^*$ interaction via two serial $n \rightarrow \pi^*$ interactions, whereby one $n \rightarrow \pi^*$ interaction (here, Cl...C=O) makes that acceptor carbonyl a better donor for a subsequent $n \rightarrow \pi^*$ interaction. These serial effects have been proposed to stabilize both the α -helix and polyproline helix conformations in proteins.^[3b, 3d, 3e, 4]

We have described the application of peptide acyl N-capping motifs to modulate peptide structure, via the tunable control of the strength of $n \rightarrow \pi^*$ interactions. We have observed crystallographically that more electron-donating N-cap motifs promote closer $n \rightarrow \pi^*$ interactions and more compact conformations, including α -helix and polyproline II helix, whereas more electron-withdrawing N-cap motifs relatively promote more extended conformations. Notably, electronic modulation of the $n \rightarrow \pi^*$ donor carbonyl was observed to *directly* impact the ϕ and ψ main chain torsion angles, as expected based on prior, more indirect analyses that employed readout of the ω torsion angle. Collectively, these results provide further support for the importance of $n \rightarrow \pi^*$ interactions in protein structure, with particular relevance in understanding disordered states of proteins. The unfolded states of proteins remain not well understood, yet are central to understanding protein folding and recognition. In addition, these results should have significant applications in peptide design and in medicinal chemistry, whereby the identity of the acyl N-capping motif could be used to predictably promote defined conformations, for optimized structure, function, and molecular recognition.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

We thank NSF (CHE-1412978) for support. Instrumentation support was provided by NIH (GM110758) and NSF (CHE-1229234).

References

- [1]. Levinthal C, J. Chim. Phys. Physico-Chimie Biol. 1968, 65, 44-45.
- [2]. a Bretscher LE, Jenkins CL, Taylor KM, DeRider ML, Raines RT, J. Am. Chem. Soc. 2001, 123, 777–778; [PubMed: 11456609] b Newberry RW, Raines RT, Acc. Chem. Res. 2017, 50, 1838–

1846; [PubMed: 28735540] c Singh SK, Das A, Phys. Chem. Chem. Phys. 2015, 17, 9596–9612; [PubMed: 25776003] d Singh SK, Mishra KK, Sharma N, Das A, Angew. Chem., Int. Ed. 2016, 55, 7801–7805; e Deb P, Jin GY, Singh SK, Moon J, Kwon H, Das A, Bagchi S, Kim YS, J. Phys. Chem. Lett. 2018, 9, 5425–5429. [PubMed: 30149713]

- [3]. a Hinderaker MP, Raines RT, Protein Sci. 2003, 12, 1188–1194; [PubMed: 12761389] b Bartlett GJ, Choudhary A, Raines RT, Woolfson DN, Nat. Chem. Biol. 2010, 6, 615–620; [PubMed: 20622857] c Sonntag LS, Schweizer S, Ochsenfeld C, Wennemers H, J. Am. Chem. Soc. 2006, 128, 14697–14703; [PubMed: 17090057] d Brister MA, Pandey AK, Bielska AA, Zondlo NJ, J. Am. Chem. Soc. 2014, 136, 3803–3816; [PubMed: 24559475] e Elbaum MB, Zondlo NJ, Biochemistry 2014, 53, 2242–2260; [PubMed: 24641765] f Wilhelm P, Lewandowski B, Trapp N, Wennemers H, J. Am. Chem. Soc. 2014, 136, 15829–15832; [PubMed: 25368901] g Rahim A, Saha P, Jha KK, Sukumar N, Sarma BK, Nat. Commun. 2017, 8;h D. N. Reddy, E. N. Prabhakaran 2014, 101, 66–77.
- [4]. Zondlo NJ, Nat. Chem. Biol. 2010, 6, 567–568. [PubMed: 20644541]
- [5]. a Uversky VN, Oldfield CJ, Dunker AK, J. Mol. Recognit. 2005, 18, 343–384; [PubMed: 16094605] b Shi Z, Chen K, Li Z, Kallenbach NR Chem. Rev. 2006, 106, 1877–1897; [PubMed: 16683759] c Tompa P, Schad E, Tantos A, Kalmar L, Curr. Opin. Struct. Biol. 2015, 35, 49–59; [PubMed: 26402567] d Wright PE, Dyson HJ, Nat. Rev. Mol. Cell. Biol. 2015, 16, 18–29. [PubMed: 25531225]
- [6]. Choudhary A, Fry CG, Raines RT, ARKIVOC 2010, 8, 251-262.
- [7]. Newberry RW, VanVeller B, Guzei IA, Raines RT, J. Am. Chem. Soc. 2013, 135, 7843–7846.[PubMed: 23663100]
- [8]. Hodges JA, Raines RT, Org. Lett. 2006, 8, 4695–4697. [PubMed: 17020280]
- [9]. a Shoemaker KR, Kim PS, York EJ, Stewart JM, Baldwin RL, Nature 1987, 326, 563–567;
 [PubMed: 3561498] b Chakrabartty A, Doig AJ, Baldwin RL, Proc. Natl. Acad. Sci. USA 1993, 90, 11332–11336; [PubMed: 8248248] c Doig AJ, Baldwin RL, Protein Sci. 1995, 4, 1325–1336;
 [PubMed: 7670375] d Aurora R, Rose GD, Protein Sci. 1998, 7, 21–38. [PubMed: 9514257]
- [10]. a Thomas KM, Naduthambi D, Tririya G, Zondlo NJ, Org. Lett. 2005, 7, 2397–2400; [PubMed: 15932207] b Naduthambi D, Zondlo NJ, J. Am. Chem. Soc. 2006, 128, 12430–12431; [PubMed: 16984189] c Pandey AK, Naduthambi D, Thomas KM, Zondlo NJ, J. Am. Chem. Soc. 2013, 135, 4333–4363. [PubMed: 23402492]
- [11]. Pandey AK, Yap GPA, Zondlo NJ, J. Org. Chem. 2014, 79, 4174–4179. [PubMed: 24720652]
- [12]. a Holl MG, Struble MD, Siegler MA, Lectka T, J. Fluorine Chem. 2016, 188, 126–130;b Hof F, Scofield DM, Schweizer WB, Diederich F, Angew. Chem., Int. Ed. 2004, 43, 5056–5059;c Paulini R, Muller K, Diederich F, Angew. Chem., Int. Ed. 2005, 44, 1788–1805;d Fischer FR, Schweizer WB, Diederich F, Angew. Chem., Int. Ed. 2007, 46, 8270–8273.
- [13]. a Leverentz HR, Truhlar DG, J. Phys. Chem. A 2008, 112, 6009–6016; [PubMed: 18540587] b
 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. Montgomery, J. A., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski, D. J. Fox, Gaussian, Inc., Wallingford, CT, 2013.
- [14]. See the Supporting Information for details.
- [15]. Glendening CR, Landis CR, Weinhold F, WIREs Comput. Mol. Sci. 2012, 2, 1-42.



Figure 1.

Proline *cis-trans* isomerism to probe $n \rightarrow \pi^*$ interactions. (a) Proline *trans* and *cis* amide bonds (blue) are in slow exchange on the NMR timescale. Only the *trans* amide bond may be stabilized by an $n \rightarrow \pi^*$ interaction. Thus, molecular properties that promote an $n \rightarrow \pi^*$ interaction lead to a larger $K_{\text{trans/cis}}$. (b) Synthesis of peptides with different acyl N-caps from the common intermediate **5**. (c) Overlap of the donor (red) carbonyl O_i lone pair (n) with the acceptor carbonyl $C_{i+1}=O_{i+1}$ (blue) π^* molecular orbital leads to electron delocalization. The extent of orbital overlap is associated with the $O_{j}...C_{i+1}$ distance (purple), with $n \rightarrow \pi^*$ interactions exhibiting distances significantly below the 3.22 Å sum of the van der Waals radii of O and C. (d) Crystal structure of **5**, which exhibits an *exo* ring pucker due to the strong stereoelectronic effect of the nitrobenzoate ester.

ÔMe



Figure 2.

Modulation of the electronic and steric properties of acyl N-capping motifs (N-caps). R = 4-NO₂-benzoate ester.



Figure 3.

van't Hoff analysis of the temperature dependence of $K_{\text{trans/cis}}$ for the peptides R–C(O)– Hyp(4-NO₂-Bz)-OMe. From top to bottom, R = –CH(CH₃)₂ (black squares),–CH₂CH₃ (open red squares), –CH₂Cl (open blue circles), –CH₃ (orange circles), –CH₂OCH₃ (green inverted triangles), –CH₂F (cyan diamonds), –H (purple triangles).



Figure 4.

X-ray crystal structures of derivatives, with the $O_{i}...C_{i+1}$ intercarbonyl distance *d* and the peptide main chain (ϕ, ψ) torsion angles in each molecule indicated. Nbz = 4-NO₂-benzoate ester.



Figure 5.

NBO analysis of $n \rightarrow \pi^*$ interactions in the limiting cases of pivaloyl and formyl N-caps. The extent of orbital overlap between the p-like O_i orbital and the C=O_{i+1} π^* orbital is significantly greater with the geometry observed in the pivaloyl N-cap than that in the formyl N-cap. The former also exhibits overlap between the s-like O_i and π^* orbitals. Bottom right: the chloroacetyl N-cap exhibits an $n \rightarrow \pi^*$ interaction between the chlorine lone pair and the adjacent carbonyl, resulting in serial $n \rightarrow \pi^*$ interactions. Calculations were conducted at the MP2 level with the 6–311++G(2d,2p) basis set in implicit water.

Table 1.

Effects of acyl N-cap substitution on the thermodynamic properties of peptides in $CDCl_3$. – = not determined. Errors are in the Supporting Information.

X-(C=O)- Hyp(Nbz)-OMe X=	K _{trans/cis} 298 K	K _{trans/cis} 263 K	G _{298 K,} kcal mol ⁻¹	<i>H</i> , kcal mol ⁻¹	S, cal mol ⁻¹ K ⁻¹	
C(CH ₃) ₃	>20	-	_	_	_	
CH(CH ₃) ₂	7.6	10.8	-1.20	-1.88	-2.4	
CH ₂ CH ₃	5.7	8.1	-1.03	-1.62	-2.0	
CH ₃	4.4	6.2	-0.88	-1.50	-2.1	
CH ₂ CI	6.0	8.0	-1.06	-1.39	-1.2	
CH ₂ OCH ₃	3.2	4.0	-0.69	-0.87	-0.5	
CH ₂ F	2.6	2.9	-0.57	-0.54	-0.1	
Н	1.8	2.0	-0.35	-0.67	-1.1	
CHCI ₂	11.1	—	-1.43	—	—	
CHF ₂	5.1	_	-0.96	—	—	
CCI ₃	>20	—	—	—	—	
CF ₃	7.8	—	-1.22	—	—	

Table 2.

Summary^{*a*} of results from X-ray crystallography and calculations^{*b*}

X-(C=O)-	<u>X-ray c</u>	<u>graphy</u>		calculations. Flp							
Hyp(Nbz)-OMe X=	molecule 1		molecule 2		<u>a-helix</u>		PPII				
	¢, ⊻ °	d, Å	¢, ₩°°	d, Å	d, Å	NBO E ^c	d, Å				
C(CH ₃) ₃	-56, +138	2.681	_	-	2.723	2.27 ^d	2.726				
CH(CH ₃) ₂	—45, —44	2.687	-47,-40	2.738	2.785	1.97	2.781				
CH ₂ CH ₃	-57,-38	2.828	-61,-34	2.883	2.807	1.87	2.816				
CH ₃	-65,-34	2.991	-69, -29	3.049	2.812	1.85	2.816				
CH ₂ CI	-56, -36	2.847	-61,-32	2.879	2.837	1.68	2.834				
CH ₂ OCH ₃	-59, -41	2.893	-69, -44	3.046	2.841	1.70	2.844				
CH ₂ F	-69,-32	3.053	-83, -173	3.320	2.838	1.71	2.837				
Н	-76, +163	3.197	-84, +173	3.336	2.961	1.11	2.963				
CHF ₂	-56, -36	2.858	-63,-30	2.921	2.861	1.40	2.864				
CF ₃	-74, +167	3.099	_	_	2.854	1.41	2.859				

^{*a*}Additional details are in the Supporting Information. d = distance between the donor O (O_{*i*}) and the acceptor carbon (C_{*i*+1}).

^bCalculations on X-C(O)-Flp-OMe at the MP2 level of theory and 6–311++G(2d,2p) basis set in implicit water, on peptides with an *exo* ring pucker and the indicated general conformation. ϕ and ψ torsion angles are tabulated in the Supporting Information, as are calculations using the M06–2X DFT method and similar calculations on X-C(O)-Pro-OMe.

^{*c*}NBO energies for orbital overlap between the O_{*i*} p-like orbital and the C=O_{*i*+1} π^* orbital in kcal mol⁻¹.

^dAn interaction energy of 0.66 kcal mol⁻¹ was in addition calculated between the O_j s-like orbital and the C=O_{j+1} π^* orbital.