PHILOSOPHICAL TRANSACTIONS B

royalsocietypublishing.org/journal/rstb

Research



Cite this article: Dantzer B *et al.* 2019 The development of individual differences in cooperative behaviour: maternal glucocorticoid hormones alter helping behaviour of offspring in wild meerkats. *Phil. Trans. R. Soc. B* **374**: 20180117.

http://dx.doi.org/10.1098/rstb.2018.0117

Accepted: 8 November 2018

One contribution of 18 to a theme issue 'Developing differences: early-life effects and evolutionary medicine'.

Subject Areas:

evolution, ecology, behaviour

Keywords:

cooperation, early life adversity, glucocorticoids, growth, maternal stress

Author for correspondence:

Ben Dantzer

e-mail: dantzer@umich.edu

The development of individual differences in cooperative behaviour: maternal glucocorticoid hormones alter helping behaviour of offspring in wild meerkats

Ben Dantzer^{1,2,3,4}, Constance Dubuc^{1,2}, Ines Braga Goncalves^{2,5}, Dominic L. Cram^{1,2}, Nigel C. Bennett⁶, Andre Ganswindt^{6,7}, Michael Heistermann⁸, Chris Duncan^{1,2}, David Gaynor^{2,6} and Tim H. Clutton-Brock^{1,2,6}

D BD, 0000-0002-3058-265X; DLC, 0000-0002-8790-8294; NCB, 0000-0001-9748-2947; THC-B, 0000-0001-8110-8969

The phenotype of parents can have long-lasting effects on the development of offspring as well as on their behaviour, physiology and morphology as adults. In some cases, these changes may increase offspring fitness but, in others, they can elevate parental fitness at a cost to the fitness of their offspring. We show that in Kalahari meerkats (Suricata suricatta), the circulating glucocorticoid (GC) hormones of pregnant females affect the growth and cooperative behaviour of their offspring. We performed a 3-year experiment in wild meerkats to test the hypothesis that GC-mediated maternal effects reduce the potential for offspring to reproduce directly and therefore cause them to exhibit more cooperative behaviour. Daughters (but not sons) born to mothers treated with cortisol during pregnancy grew more slowly early in life and exhibited significantly more of two types of cooperative behaviour (pup rearing and feeding) once they were adults compared to offspring from control mothers. They also had lower measures of GCs as they aged, which could explain the observed increases in cooperative behaviour. Because early life growth is a crucial determinant of fitness in female meerkats, our results indicate that GC-mediated maternal effects may reduce the fitness of offspring, but may elevate parental fitness as a consequence of increasing the cooperative behaviour of their daughters.

This article is part of the theme issue 'Developing differences: early-life effects and evolutionary medicine'.

1. Introduction

Parental effects are a mechanism of trans-generational phenotypic plasticity that occurs when the parental phenotype or parental environment modifies offspring characteristics [1]. Parental effects can increase the survival or reproduction of offspring, thereby elevating the direct fitness of both offspring and parents [2–6]. Alternatively, parental effects can increase parental fitness, but at some

¹Department of Zoology, University of Cambridge, Cambridge, CB2 3EJ, UK

²Kalahari Meerkat Project, Kuruman River Reserve, Northern Cape, South Africa

³Department of Psychology, University of Michigan, Ann Arbor, MI 48109, USA

⁴Department of Ecology and Evolutionary Biology, University of Michigan, Ann Arbor, MI 48109, USA

⁵School of Biological Sciences, University of Bristol, Bristol, BS8 1TQ, UK

⁶Department of Zoology and Entomology, Mammal Research Institute, University of Pretoria, 0002 Pretoria, South Africa

⁷Endocrine Research Laboratory, Department of Anatomy and Physiology, University of Pretoria, Onderstepoort 0110, South Africa

⁸Endocrinology Laboratory, German Primate Center, Leibniz Institute for Primate Research, Kellnerweg 4, 37077 Göttingen, Germany

cost to the fitness of their offspring [7,8]—a process regarded as a type of parental manipulation [9–12] or 'selfish parental effect' [13]. For example, in mammals, the optimal birth weight or litter size often differs between mothers and offspring [14] and pregnant females experiencing stressful environments may reallocate resources away from offspring and towards themselves, so that their offspring are smaller or grow more slowly before weaning [15]. Despite these observations, it has been suggested that selfish parental effects may be rare and unstable because selection would be expected to favour the evolution of resistance mechanisms in offspring [7,11,13,16,17].

Selfish parental effects may in fact be more likely in cooperatively breeding species where philopatric offspring (subordinates) help to rear the subsequent offspring of their parents or other close relatives. This could be especially likely under low food or high stress conditions as parents may gain substantial direct fitness benefits from delaying the development of their offspring if this causes them to invest in alloparental care directed at the parent's subsequent offspring [9,10]. In addition, the costs of selfish parental effects to offspring could be reduced in these circumstances, as offspring will gain indirect fitness benefits by contributing to raising the subsequent offspring of their parents [18]. For example, laboratory studies of eusocial insects suggest the possibility that selection will favour the evolution of alleles that enable mothers to increase the helping behaviour of their offspring while simultaneously reducing their capabilities of reproducing on their own ([19,20]; but see [21]).

To date, empirical field tests of how parental effects shape the helping behaviour of offspring are rare [22] and studies of selfish parental effects have mostly focused on non-social species [13,23]. Here, we report the results of experiments designed to test the hypothesis that elevated maternal glucocorticoid levels (GCs) reduce the potential for offspring to have direct reproductive opportunities and cause them to exhibit more cooperative behaviour. In a 3-year field study, we experimentally elevated maternal GCs by treating pregnant dominant female meerkats with cortisol and tracking the growth, stress physiology and cooperative behaviour of their offspring from birth until approximately 18 months of age, compared to those from control litters. We manipulated maternal GCs because they are known to cause mothers to reallocate energy away from offspring and towards themselves [15], indicating that they may function as a mediator of selfish maternal effects. Changes in maternal GCs have also previously been shown to delay the dispersal of offspring as well as influence the parental care behaviour of offspring [24,25], both traits that are important in cooperative breeders where philopatric offspring exhibit alloparental care behaviour towards non-descendent relatives.

To identify if the exposure of mothers to heightened GCs reduced the reproductive success of their offspring, we examined whether offspring from mothers treated with cortisol during pregnancy grew more slowly early in life. In meerkats, the rate of early life growth and body mass is closely linked to future direct fitness through its effects on survival, foraging success and adult body mass [26–28], as well as the probability of acquiring dominance [29,30] and other direct reproductive opportunities [31]. As elevated exposure to maternal GCs in some mammals may reduce offspring size and growth early in life [15], we predicted that offspring from mothers treated with cortisol during pregnancy would

be smaller or grow more slowly early in life. Because the rate of early life growth is predictive of future direct fitness in meerkats [26–31], we predicted that if offspring from mothers treated with cortisol did grow more slowly, they would consequently invest more in indirect fitness opportunities by contributing more to cooperative activities than controls

Second, we determined whether offspring from mothers treated with cortisol during pregnancy subsequently increased their contributions to two types of cooperative behaviours: pup rearing ('babysitting' [32]) and food provisioning during the period when the pups are foraging with their natal group, but are not yet nutritionally independent ('pup feeding' [33]). We chose these two behaviours as they appear to be the most costly from an energetic perspective [34] and are most closely tied to the probability of parents successfully rearing offspring. If offspring from mothers treated with cortisol during pregnancy exhibit more of either of these two types of alloparental care, this should increase both parental direct fitness (the number of offspring that they subsequently produce) and the indirect fitness of offspring, because subsequent offspring that receive more alloparental care should grow faster or have higher early life survival [26,31,35,36]. Previous studies in meerkats do indeed show that offspring with more helpers or those that receive more alloparental care grow faster or have increased early life survival [26,31,35,36].

To assess the mechanism by which elevated exposure to maternal stress may affect the alloparental care behaviour of off-spring, we repeatedly measured plasma cortisol and faecal glucocorticoid metabolite (fGCM) concentrations of offspring from when they were approximately one to 18 months of age to identify how our manipulations affected their neuroendocrine stress axes (GC output). Elevated maternal GCs can cause long-term changes in the neuroendocrine stress axis of offspring [37] and elevated activity of the neuroendocrine stress axis in meer-kats can reduce their contributions to alloparental care [38]. We therefore predicted that if offspring born to mothers treated with cortisol during pregnancy exhibited more alloparental care behaviour compared to controls, they would also have reduced plasma cortisol and fGCM concentrations.

2. Material and methods

(a) Study site and basic data collection

We studied free-living meerkats at the Kuruman River Reserve (26°58′ S, 21°49′ E) in the Northern Cape, South Africa from 2014 to 2017. Individuals were marked uniquely with PIT tags (Identipet®, Johannesburg, South Africa) as well as dye marks so that they could be identified. Groups were visited for approximately 4-8 h per day approximately four to six times per week throughout each year of study, and sometimes more frequently, such as when there were pups being babysat. Groups were visited at sunrise before meerkats emerged from their sleeping burrow. After all the meerkats had emerged, but before they started going foraging, we counted the total number of meerkats in the group (to get estimates of group size) and recorded which individuals were present (using their unique combinations of dye marks). We recorded their body mass on a portable balance each morning before foraging, 2-4 h after foraging was initiated, and immediately before the end of foraging [39]. These measures of body mass provided our estimates of growth, body mass and foraging success that are used in our analyses described below.

Table 1. Summary of effects of dominant female treatments on litter characteristics and offspring survival. Number of pups emerged correspond to those that emerged from the natal burrow and in some cases these pups died before their sex could be determined (shown as 'Unk'). Three of the 13 litters treated with cortisol and one of the 7 control (fed) litters were aborted prior to birth. Two cortisol-treated mothers lost their litters before the pups first emerged from the burrow. Mean and SD shown.

treatment	total no. litters and females treated	total no. pups emerged (F, M, Unk)	avg. pups emerged	avg. pups surviving to three months	avg. pups surviving to six months	avg. pups surviving to 12 months
untreated	52 (21 females)	49 F, 84 M, 52 Unk	3.56 ± 1.24	2.68 ± 1.62	2.10 ± 1.61	1.62 ± 1.53
control	7 (6 females)	12 F, 13 M	4.17 ± 0.98	3.83 ± 1.17	3.83 <u>+</u> 1.17	2.83 ± 1.94
cortisol	13 (10 females)	13 F, 18 M	3.87 ± 0.83	3.25 ± 1.03	2.75 <u>+</u> 1.28	1.75 ± 1.75

(b) Experimental manipulations of dominant females

Dominant females in each group were identified via behavioural observations [40]. The pregnancy status of dominant females was determined visually (distended abdomen) as well as by noting a constant increase in their body mass. Dominant females were treated with either a cortisol solution or a control oil vehicle when they were pregnant by feeding them food containing one of these two treatments. We initially offered experimental animals hard boiled eggs with added cortisol but found that they rejected all foods that contained added cortisol with the exception of scorpions. We consequently fed experimental females with cortisol (10 mg kg⁻¹ of hydrocortisone, Sigma H4126) that was dissolved in 100 μl of 100% coconut oil and injected into a dead scorpion (Opistophthalmus spp.). Control females were fed a dead scorpion that was injected with 100 μl of 100% coconut oil. A previous study using the same protocol showed that meerkats that were fed cortisol had significantly higher plasma cortisol and fGCM concentrations than control animals and these increases were within a physiologically relevant range [38]. This indicated that our treatment causes the exogenous GCs that we feed the meerkats to enter their bloodstream and leads to sustained increases in their circulating GC concentrations.

Females were randomly allocated to the treatments. Across the 3 years of this study, we produced a total of 13 cortisol-treated litters from 10 females and 7 control litters produced by 6 females (table 1). Three of the females experienced both the control and cortisol treatments at different time points of the experiment, whereas one female experienced the control treatment once and the cortisol treatments twice. For these latter females treated multiple times, the order of treatments was randomly selected. We conducted these experiments over the course of three years: 13 litters from 10 females in 2014 (April-December 2014, 3 litters aborted), 5 litters from 5 females in 2015 (February-July 2015, 1 litter aborted) and 3 litters from 3 females in 2016 (July 2016). Three cortisol-treated mothers and one control mother aborted their offspring prior to birth and were excluded from any analyses except for assessing differences in the frequency of abortion between control and cortisol-treated mothers (table 2). Two cortisol-treated mothers lost their litters before the pups first emerged from the burrow, providing final sample sizes of 31 pups from 8 litters from 7 cortisol-treated females and 25 pups from 6 litters from 6 control females.

We aimed to experimentally increase the GC concentrations of pregnant dominant females from when they were first confirmed to be pregnant (second half of gestation) until parturition. Gestation in meerkats is approximately 70 days so we aimed to treat them with GCs from approximately 35 to 70 days during gestation. In reality, females that successfully produced a litter where pups emerged from the natal burrow were treated with cortisol for 12-36 days prior to birth (n=10 litters from 9 females, mean = 23.7 days, median = 23.5 days), whereas controls were fed for 12-58 days prior to birth (n=6 litters from 6 females, mean = 30 days, median = 20.5 days). Although controls were

treated for slightly longer, there was no significant difference in treatment duration between control and cortisol-treated females (general linear model, t = 1.05, p = 0.31).

To provide an additional comparison group to investigate how our treatments (fed during pregnancy or fed cortisol during pregnancy) affected offspring survival, growth and cooperative behaviour, we also monitored these traits in offspring produced by dominant females that were untreated during pregnancy (n = 52 litters from 21 dominant females, table 1). These females were not fed or treated with cortisol (hereafter, 'untreated mothers'). For our analyses of how the treatments affected offspring survival and growth, the untreated offspring were those from litters produced by dominant females in other meerkat groups in our same study area and were born during our study. We assessed the contributions of offspring from mothers treated with cortisol during pregnancy to two cooperative behaviours (babysitting and pup feeding) compared to those from control mothers, but also to other group members from untreated mothers. We did not have data from offspring from untreated mothers when we assessed how our treatments affected their plasma cortisol or fGCM concentrations.

(c) Quantifying early life growth of offspring

Meerkat pups typically first emerge from their natal burrow approximately 21–30 days after birth. Meerkat groups and dominant females were monitored daily around the estimated date of parturition and birth dates were estimated according to the change in the physical appearance of the dominant female, a large drop in body mass overnight, and group members exhibiting babysitting behaviour at the sleeping burrow. Burrows containing pups were monitored each day and, when pups emerged, they were uniquely marked by trimming small sections of hair before permanent PIT tags could be applied. Pups were weighed each time we visited the groups on a portable balance in the morning after group members emerged from their sleeping burrow (as above).

(d) Quantifying cooperative behaviour of offspring

We measured the babysitting (controls: 195–655 days; cortisol: 184–655 days; untreated: 155–655 days) and pup feeding (controls: 220–635 days; cortisol: 184–655 days; untreated: 155–626 days) contributions of offspring from cortisol-treated and control, and untreated mothers when they were more than six months of age until death or disappearance. We visited sleeping burrows containing pups every day in the morning and recorded the identity of the attending babysitters. As we have done previously [32,35,38,41], we calculated relative babysitting contributions of each individual meerkat for each litter by dividing the total number of days an individual babysat a litter by the total number of days that this specific litter had a babysitter. Pup feeding behaviour for each pup produced by the dominant females in the different treatment groups was estimated using *ad libitum*

Table 2. Effects of dominant female treatments (cortisol or control) on litter characteristics and pup survival. Results are from a linear mixed-effects model (LMM; no. pups emerged) or generalized linear mixed-effects models (GLMMs, all other response variables) that each contained random intercept terms for dominant female identity and year. No GLMM was overdispersed as indicated by the goodness of fit tests (R package aods3, p-values from Pearson χ^2 tests ranged from 0.13 to 1.00). The number of litters aborted by untreated females was not known so we only assessed the effects of cortisol versus control treatments on the number of litters aborted. Litter sex ratio is the proportion of males in the litter. Reference value for treatment (in the intercept) was cortisol-treated mothers. Data other than no. litters aborted are based upon an initial sample size of offspring from untreated (185 pups from 52 litters produced by 21 dominant females), control (25 pups from 6 litters produced by 6 females), or cortisol-treated (31 pups from 8 litters produced by 7 females) litters that produced pups that emerged from the burrow.

response variable	fixed effect	b	s.e.	t or z	<i>p</i> -value
no. litters aborted					
	intercept	— 1.21	0.66	— 1.83	0.07
	birthdate	-0.17	0.59	-0.29	0.77
	treatment				
	control	-0.59	1.27	- 0.46	0.64
no. pups emerged					
	intercept	3.84	0.41	9.31	< 0.000
	birthdate	0.27	0.15	1.78	0.08
	treatment				
	control	0.37	0.63	0.59	0.56
	none	-0.28	0.44	-0.63	0.53
litter sex ratio					
	intercept	-0.55	0.30	— 1.85	0.064
	birthdate	0.03	0.12	0.26	0.79
	treatment				
	control	-0.1	0.45	-0.23	0.82
	none	0.09	0.33	0.27	0.78
proportion litter surviving to three months					
	intercept	— 0.16	0.27	 0.62	0.53
	birthdate	— 0.12	0.11	— 1.09	0.27
	treatment				
	control	0.06	0.39	0.16	0.87
	none	— 0.19	0.30	 0.64	0.52
proportion litter surviving to six months					
	intercept	-0.32	0.28	— 1.16	0.24
	birthdate	— 0.19	0.11	— 1.65	0.099
	treatment				
	control	0.21	0.40	0.52	0.60
	none	-0.28	0.31	 0.91	0.36
proportion litter surviving to 12 months					
	intercept	— 0.91	0.46	— 1.999	0.046
	birthdate	— 0.22	0.14	— 1.58	0.11
	treatment				
	control	0.36	0.46	0.79	0.43
	none	0.096	0.38	0.25	0.80

sampling [33,38]. When the social group contained pups (up to 90 days of age), we recorded all pup feeding events from all that were visually and acoustically conspicuous to observers [42]. We then used these data to estimate the proportion of pup feeding events exhibited by an individual relative to all others in the group (i.e. relative pup feeding). Because the total amount of time devoted to the *ad libitum* recording sessions varied, we corrected for variation in observation time (see below).

(e) Quantifying plasma cortisol concentrations from offspring

We obtained plasma samples from offspring from cortisol-treated and control mothers approximately every three months from the first emergence from the burrow (approx. one month) until approximately 18 months of age (controls: 20–548 days; cortisol-treated: 25–559 days). Capture and blood processing

procedures are described elsewhere [43,44]. The amount of time it took to obtain the blood samples varied (median = 10.6 min, s.d. = 7.2 min), but we included covariates for sampling time and sampling time² to control for effects of sampling time (described in [44]). We measured total plasma cortisol concentrations using a previously validated assay (Coat-a-Count, Siemens Diagnostic Products Corporation, Los Angeles, USA: validation described in [43]). The sensitivity of the assay was 1.9 ng ml⁻¹ and cross-reactivity to other hormones was 76% with prednisolone, 11.4% with 11-deoxycortisol, 2.3% with prednisone and less than 1% with aldosterone, corticosterone, cortisone, oestriol, estrone and pregnenolone. Intra-assay coefficient of variation (CV) was 7% (n = 20 samples). Inter-assay CV for a low control (78.5 \pm 6.3 ng ml⁻¹ n = 5 assays) was 8% and 2.8% for a high control (187 \pm 5.3 ng ml⁻¹, n = 5 assays).

(f) Quantifying fGCM concentrations from offspring

We collected faecal samples from offspring of cortisol-treated and control mothers opportunistically during behavioural observations over the course of the study (controls: 25-356 days; cortisol-treated: 32-326 days). Faecal samples were processed as described previously using a methanol solution to extract fGCMs for analysis [45,46]. Immunoreactive fGCM concentrations were determined using a group-specific enzyme immunoassay measuring cortisol metabolites with a 5β - 3α , 11β -diol-structure (11β-hydroxyetiocholanolone), already validated and established for monitoring fGCM alterations in meerkats [46]. Faecal GCMs that were measured reflect average adrenal cortisol production over the previous approximately 24 to 48 h period [46]. Detailed assay characteristics, including full descriptions of the assay components and cross-reactivities, are found elsewhere [47]. The sensitivity of the assay was 1.2 ng g^{-1} dry weight, intra-assay CVs determined by repeated measurements of high- and low-value quality controls were 6.9% and 7.4% and inter-assay CV values were 11.5% and 15.9% (n = 29 assays), respectively.

(g) Statistical analyses

We used generalized (binomial errors) or linear mixed-effects models (LMMs) to examine how our treatments affected the probability that the litter was aborted, litter size and sex ratio at emergence from the burrow, and the proportion of the litter that survived to emergence from the burrow, independence (approx. 90 days of age [28]), and six or 12 months of age. We focused on addressing whether the offspring from cortisol-treated mothers differed from control or untreated mothers. These models included a fixed effect for date of birth of the litter and random intercept terms for dominant female identity and year (as the experiments were conducted over three years). None of the generalized linear mixed-effects models (GLMMs) were overdispersed (table 2).

We used an LMM to investigate how the maternal treatments affected offspring growth from the first emergence from their natal burrow (approx. one month) to three months of age when the pups are typically foraging independently [28,33]. Morning body mass (in grams) was the response variable, with the fixed effects of maternal treatment (cortisol-treated, control or untreated), pup sex, pup age, litter size at burrow emergence, first measure of body mass when the pups first emerged from the burrow (to control for possible differences in age or development when they entered our study population), group size, group size², total rainfall in the previous 60 d, two measures of seasonality (sine and co-sine functions of day of weight measure: see [39]) and two three-way interactions between sex, treatment, age or age². Group size was defined as the average number of subordinate meerkats more than six months of age in the group during the entire period of offspring growth. Random intercept terms for year and the identity of the individual nested in litter, nested in dominant female identity, nested in

group were also included in this model. Fixed and random effects included in these models were based upon previous studies investigating meerkat body mass and/or growth from one to three months [27,30,39]. To prevent any issues associated with selective disappearance of specific individuals, only individuals that survived to 90 days were included in these analyses.

We assessed how the treatments affected the relative babysitting and pup feeding contributions of subordinates when they were more than six months old (as they rarely do alloparental care behaviour when less than six months old [35]) from cortisol-treated, control and untreated mothers. Relative babysitting and pup feeding contributions are defined as the proportion of babysitting or pup feeding contributions exhibited by a specific individual compared to the total number of babysitting or pup feeding contributions for that litter exhibited by all individuals in the group that were more than six months of age at the time of the birth of the litter [35,38,41]. In these GLMMs (binomial errors), we included a three-way interaction between treatment, sex and age of the subordinate to assess if the effects of the treatments on babysitting or pup feeding varied according to the sex or age of the subordinate, as contributions to cooperative behaviour in meerkats are known to vary according to subordinate sex and age [35]. To account for differences in observation time, we included a covariate for the number of days the litter was babysat (babysitting length) and the number of days the subordinate was observed in the group during babysitting as well as the total time spent observing the group during pup feeding (observation time). We included a range of covariates (see tables 4-6) that have been previously documented to affect relative contributions to babysitting and pup feeding, including age, foraging success, body mass and group size [33-35,41,48,49]. Group size was defined as the average number of subordinate meerkats more than six months of age in the group while the litter was being babysat or pup fed. Foraging success was defined as the average weight gained per hour estimated as the change in body mass from morning weight to evening weight over the total number of hours that had elapsed since those two weights [44]. Relatedness between the subordinate and the litter being babysat was not included as it has not been shown to impact babysitting or pup feeding contributions [26,41] and nearly all of the litters in our dataset were produced by the mother or full sibling of the subordinate. Random intercept terms for year and the identity of the individual, and litter being babysat or pup fed were nested within the group where the litter was being babysat or pup fed. Overdispersion was not an issue for our GLMM for babysitting, as indicated by the goodness of fit test (Pearson $\chi^2 = 147.1$, d.f. = 154, p = 0.64, using package aods3 [50]) but our GLMM for pup feeding was initially overdispersed (Pearson $\chi^2 = 310$, d.f. = 165, p < 0.0001) so we included an observation level random intercept term.

We used two separate LMMs to assess how our manipulations affected plasma cortisol and fGCMs in offspring from cortisol-treated and control mothers (we did not have these data from offspring from untreated mothers). Each model included fixed effects for maternal treatment, pup sex and age, time of day and year that the sample was acquired (2014 or 2015), and random intercept terms for the individual's identity nested in their birth litter and group. In the model for plasma cortisol concentrations, we also included a linear and second order fixed effect for the time it took to acquire the blood sample to control for any variation in plasma cortisol concentrations owing to restraint stress [44]. Year was included as a fixed effect because we only had samples from two separate years. We included covariates associated with the individual meerkat and weather or social group characteristics that are known to affect plasma cortisol [44] or fGCM [46] concentrations (see tables 6 and 7). This included a principal component axis (PC1) that described precipitation and temperature (weather) over the 30 days prior to sample acquisition where high PC1 scores correspond to warmer and wetter conditions (described in [44]).

Table 3. Effect of dominant female treatments on offspring growth from emergence to nutritional independence (one to three months of age). Data are from a LMM where the response variable was morning body mass that contained random intercept terms for individual identity nested in birth litter nested in mother nested in the natal group ($\sigma^2 = 116.7$) and year ($\sigma^2 = 0$). If fixed effects by themselves were involved in significant higher-order interactions with other variables, only parameter estimates are shown. Data used in these analyses were 4676 measures of body mass from 195 meerkats produced by 21 dominant females across 53 different litters in 16 different social groups in three different years. Only offspring that survived to 90 days of age were included in these analyses.

fixed effect	b	s.e.	t	d.f.	<i>p</i> -value
intercept					
females	184.6	9.75	18.93	53	< 0.0001
males	189.1	9.5	19.8	49	< 0.0001
litter size	-5.31	3.9	— 1.37	48	0.18
first weight	15.96	2.52	6.344	166	< 0.0001
sex (M)	4.54	4.32			
age	53.3	1.06			
age ²	-4.76	1.15			
rainfall	2.81	0.61	4.57	4550	< 0.0001
season (sine)	-67.6	3.3	— 20.6	4238	< 0.0001
season (co-sine)	66.22	3.4	19.2	4191	< 0.0001
group size	2.3	2.66	0.86	109	0.39
group size ²	0.09	2.21	0.04	178	0.97
treatment (control)					
females	22.6	15.6			
males	12.9	15.5			
treatment (none)					
females	23.6	10.6			
males	22.9	10.4			
age $ imes$ sex (M)	1.64	1.37	1.19	4567	0.23
$age^2 imes sex$ (M)	1.02	1.53	0.67	4543	0.5
sex (M) $ imes$ treatment (control)	−9.73	7.7	— 1.27	164	0.21
sex (M) $ imes$ treatment (none)	-0.63	5.1	-0.12	172	0.9
age $ imes$ treatment (control)					
females	7.6	1.8	4.17	4595	<0.0001
males	-2.3	1.5	— 1.48	4548	0.14
age $ imes$ treatment (none)					
females	0.79	1.2	0.65	4574	0.51
males	-0.53	1.01	-0.52	4580	0.60
$age^2 imes treatment$ (control)					
females	−0.13	1.93	-0.07	4521	0.94
males	— 1.05	1.6	-0.65	4547	0.52
$age^2 imes treatment (none)$					
females	-2.6	1.3	— 1.96	4534	0.051
males	-6.9	1.1	−6.23	4556	< 0.0001
age $ imes$ treatment (control) $ imes$ sex (M)	-9.9	2.3	-4.23	4574	< 0.0001
age $ imes$ treatment (none) $ imes$ sex (M)	-1.3	1.6	-0.84	4581	0.4
${\sf age}^2 imes {\sf treatment}$ (control) $ imes$ sex (M)	-0.93	2.5	-0.37	4552	0.71
$age^2 \times treatment (none) \times sex (M)$	-4.3	1.7	– 2.5	4552	0.01

We used R (v. 3.4.3 [51]) for all of our statistical analyses. R package lme4 (v. 1.1–14 [52]) was used for LMMs and p-values were estimated using lmerTest (version 2.0–33 [53]). A graphical approach was used to confirm normality and

homoscedasticity of residuals and to confirm there were no observations with high leverage [54]. Collinearity among predictor variables included in our models was assessed by calculating variance inflation factors (VIFs) [54] or generalized VIFs

Table 4. Effect of dominant female treatments on relative babysitting contributions. Data are from a GLMM where the response variable is the proportion of babysitting exhibited by the subordinate meerkat relative to the total babysitting contributions the litter received. The model contained random intercept terms for individual ($\sigma^2 = 0.12$), litter nested within group ($\sigma^2 = < 0.0001$) and year ($\sigma^2 = 0.000$). If fixed effects by themselves were involved in significant higher-order interactions with other variables, only parameter estimates are shown. Data used in these analyses were 182 observations of relative babysitting contributions to 28 litters produced in 9 groups across three years recorded from 105 subordinate meerkats.

fixed effect	b	s.e.	Z	<i>p</i> -value
intercept				
females	-2.1	0.22	-9.37	<0.0001
males	— 2.14	0.19	— 11.01	<0.0001
babysitting length	0.24	0.08	2.77	0.0056
observation time	- 0.28	0.08	-3.4	0.0007
litter size	0.015	0.05	0.32	0.75
mixed litter?	 0.04	0.12	−0.31	0.76
sex (M)	- 0.03	0.27	- 0.13	0.9
age				
females	0.13	0.17		
males	0.40	0.19		
foraging success	- 0.05	0.05	- 0.99	0.32
mass				
females	-0.32	0.12		
males	- 0.098	0.14		
group size				
females	-0.35	0.09	— 3.97	<0.0001
males	- 0.27	0.08	-3.28	0.001
treatment (control)				
females	- 0.67	0.31		
males	-0.12	0.28		
treatment (none)				
females	-0.06	0.23		
males	-0.16	0.18		
foraging success $ imes$ mass	0.036	0.06	0.57	0.57
age $ imes$ mass				
females	- 0.47	0.09	-5.08	<0.0001
males	- 0.29	0.08	−3.79	0.0001
age $ imes$ sex (M)	0.28	0.23	1.18	0.24
mass $ imes$ sex (M)	0.22	0.17	1.26	0.21
group size $ imes$ sex (M)	0.07	0.10	0.77	0.44
treatment (control) \times sex (M)	0.55	0.42	1.31	0.19
treatment (none) \times sex (M)	-0.10	0.28	-0.36	0.72
age $ imes$ treatment (control)				
females	-0.62	0.21	— 2.89	0.0039
males	- 0.52	0.27	— 1.92	0.055
age $ imes$ treatment (none)				
females	0.34	0.18	1.88	0.059
males	- 0.006	0.16	-0.03	0.97
age $ imes$ mass $ imes$ sex	0.18	0.12	1.50	0.13
age $ imes$ treatment (control) $ imes$ sex (M)	0.09	0.34	0.28	0.78
age \times treatment (none) \times sex (M)	-0.34	0.24	— 1.45	0.14

Table 5. Effect of dominant female treatments on relative pup feeding contributions. Data are from a GLMM where the response variable is the proportion of pup feeds exhibited by the subordinate meerkat relative to the total pup feeds the litter received. The model contained random intercept terms for individual ($\sigma^2 = 0.000$), litter nested within group ($\sigma^2 = 0.2$), year ($\sigma^2 = 0.08$) and an observational level random intercept term to control for overdispersion ($\sigma^2 = 0.19$). If fixed effects by themselves were involved in significant higher-order interactions with other variables, only parameter estimates are shown. Data used in these analyses were 192 observations of relative pup feeding contributions to 26 litters produced in 7 groups across 3 years recorded from 101 subordinate meerkats.

fixed effect	b	s.e.	Z	<i>p</i> -value
intercept				
females	— 2.27	0.28	-8.2	< 0.0001
males	— 2.66	0.25	— 10.45	< 0.0001
observation time	0.59	0.06	9.41	<0.0001
litter size	— 0.055	0.10	— 0.55	0.58
mixed litter (Y)	- 0.03	0.29	— 0.10	0.92
sex (M)	- 0.39	0.20		
age				
females	-0.37	0.16		
males	0.29	0.14		
foraging success	0.14	0.06	2.12	0.033
mass				
females	0.10	0.11		
males	- 0.37	0.11		
group size				
females	- 0.48	0.13	-3.57	0.0003
males	-0.32	0.13	− 2.46	0.014
treatment (control)				
females	-0.73	0.23	-3.12	0.0018
males	- 0.24	0.21	— 1.14	0.25
treatment (none)				
females	- 0.64	0.18	−3.49	0.0005
males	— 0.14	0.13	— 1.03	0.3
foraging success $ imes$ mass	0.06	0.06	0.97	0.33
age $ imes$ mass				
females	- 0.04	0.07	−0.62	0.54
males	— 0.19	0.07	— 2.68	0.007
age $ imes$ sex (M)	0.66	0.19	3.49	0.00048
$ ext{mass} imes ext{sex (M)}$	- 0.47	0.13	-3.63	0.0003
group size $ imes$ sex (M)	0.16	0.08	1.89	0.059
treatment (control) $ imes$ sex (M)	0.49	0.31	1.58	0.11
treatment (none) $ imes$ sex (M)	0.50	0.22	2.23	0.023
age $ imes$ treatment (control)				
females	−0.11	0.22	— 0.5	0.61
males	0.03	0.29	0.11	0.91
age $ imes$ treatment (none)				
females	0.17	0.16	1.03	0.3
males	- 0.02	0.14	-0.12	0.9
age \times mass \times sex	- 0.14	0.09	— 1.51	0.13
age $ imes$ treatment (control) $ imes$ sex (M)	0.14	0.34	0.42	0.67
age \times treatment (none) \times sex (M)	- 0.19	0.21	– 0.91	0.36

Table 6. Effect of dominant female treatments on plasma cortisol concentrations. Data are from a LMM where the response variable is plasma cortisol concentrations (In transformed) of the subordinate meerkat. The model contained random intercept terms for individual nested within their birth litter ($\sigma^2 = 0.034$) and capture group ($\sigma^2 = 0.000$). If fixed effects by themselves were involved in significant higher-order interactions with other variables, only parameter estimates are shown. Data used in these analyses were 299 measures of plasma cortisol concentrations from 49 subordinate meerkats produced in 14 litters from 10 different groups. Reference levels (intercept) for 'sex' was female, 'pups in group' was yes, and for relatedness was 'no parent was dominant'.

fixed effect	b	s.e.	t	d.f.	<i>p</i> -value
intercept					
females	3.18	0.29	11.31	128	<0.0001
males	3.37	0.25	13.37	113	< 0.0001
sampling time	1.16	0.10	11.95	259	<0.0001
sampling time ²	 0.24	0.03	−7.24	269	<0.0001
time of day	— 0.20	0.14	— 1.38	276	0.17
sample year (2015)	0.41	0.24	1.72	228	0.09
sex (M)	0.20	0.20	0.97	39	0.34
age					
females	— 0.24	0.18			
males	- 0.01	0.14			
foraging success					
females	-0.13	0.11	— 1.22	231	0.22
males	— 0.15	0.10	— 1.42	269	0.16
group size	0.07	0.11	0.62	242	0.54
group size ²	0.05	0.08	0.63	211	0.53
pups in group	- 0.31	0.18	— 1.79	276	0.075
group sex ratio	0.2	0.08	2.53	180	0.01
relatedness	- 0.09	0.18	— 0.50	40	0.62
weather (PC1)	- 0.08	0.10	— 0.75	276	0.45
treatment (cortisol)					
females	- 0.02	0.24			
males	-0.32	0.19			
sex (M) $ imes$ age	0.22	0.22	1.04	272	0.3
sex (M) $ imes$ foraging success	- 0.013	0.14	- 0.09	257	0.93
sex (M) $ imes$ treatment (cortisol)	-0.3	0.3	— 1.01	35	0.32
age \times treatment (cortisol)					
females	- 0.42	0.24	— 1.76	276	0.08
males	- 0.5	0.19	- 2.68	274	0.008
group size \times pups present (yes)	- 0.05	0.15	- 0.36	216	0.72
group size $^2 \times$ pups present (yes)	0.07	0.12	0.6	263	0.55
group size $ imes$ weather (PC1)	-0.02	0.08	- 0.30	273	0.76
age $ imes$ sex (M) $ imes$ treatment (cortisol)	-0.08	0.31	- 0.26	275	0.8

(for variables that had a second-order term or those included in an interaction [55]). Collinearity was not a problem as indicated by our VIFs, as VIFs or generalized VIFs were less than approximately 4 for all variables. In our model for how our treatments affected offspring growth (table 3), the generalized VIF for the two measures of seasonality (sine and co-sine functions of day of weight measure) was less than 6 but these two variables were included *a priori* given their previously documented effects on body mass and growth in meerkats [39]. All continuous variables were standardized to a mean of 0 and SD of 1.

3. Results

(a) Effects of treatments on litter characteristics and offspring survival

There was no evidence that the treatment of pregnant females with cortisol affected their ability to maintain litters to term or the survival of their pups prior to emergence from the natal burrow (tables 1 and 2). The number of pups surviving to emergence from the natal burrow or three, six or

Table 7. Effect of dominant female treatments on fGCM concentrations. Data are from a LMM where the response variable was fGCM concentrations ($\ln + 1$ transformed) of the subordinate meerkat. The model contained random intercept terms for individual nested within their birth litter ($\sigma^2 = 0.094$) and collection group ($\sigma^2 = 0.000$). If fixed effects by themselves were involved in significant higher-order interactions with other variables, only parameter estimates are shown. Data used in these analyses were 542 faecal samples (n = 355 from controls, n = 187 from cortisol-treated litters) from 34 subordinate meerkats (control: n = 12 females, n = 11 males; cortisol-treated: n = 5 females, n = 6 males) produced in 10 litters from 7 different groups. Reference levels (intercept) for 'sex' was female, 'pups in group' was yes and for relatedness was 'no parent was dominant'.

fixed effect	b	s.e.	t	d.f.	<i>p</i> -value
intercept					
females	5.40	0.25	21.36	50	< 0.0001
males	5.33	0.23	23.4	23	< 0.0001
time of day	— 0.14	0.04	−3.29	516	0.001
sample year (2015)	0.07	0.17	0.43	120	0.67
sex (M)	- 0.06	0.20	- 0.31	22	0.76
age					
females	0.48	0.16			
males	0.06	0.10			
foraging success					
females	0.16	0.13	1.29	355	0.2
males	0.04	0.06	0.81	520	0.42
group size	0.41	0.12	3.52	96	0.0007
group size ²	— 0.05	0.11	— 0.78	267	0.43
pups in group	— 0.13	0.16	— 0.78	267	0.43
group sex ratio	0.16	0.07	2.27	158	0.024
relatedness	0.28	0.25	1.10	14	0.29
weather (PC1)	0.09	0.07	1.40	203	0.16
treatment (cortisol)					
females	— 0.12	0.29			
males	- 0.03	0.24			
sex (M) $ imes$ age	— 0.42	0.17	— 2.47	221	0.014
sex (M) $ imes$ foraging success	— 0.12	0.13	— 0.87	375	0.38
sex (M) $ imes$ treatment (cortisol)	0.10	0.33	0.29	18	0.77
age $ imes$ treatment (cortisol)					
females	-0.63	0.22	— 2.9	447	0.004
males	— 0.10	0.14	— 0.69	491	0.49
group size $ imes$ pups present (yes)	— 0.17	0.12	— 1.4 6	216	0.14
group size $^2 imes$ pups present (yes)	0.11	0.12	456	0.97	0.33
group size $ imes$ weather (PC1)	0.04	0.05	0.69	129	0.49
age $ imes$ sex (M) $ imes$ treatment (cortisol)	0.53	0.25	2.15	413	0.032

12 months of age and the litter sex ratio were not different among litters from cortisol-treated, control or untreated females (tables 1 and 2).

(b) Effects of treatments on offspring early life growth

The effects of the treatments on offspring growth from initial emergence to nutritional independence (one to three months) differed between daughters and sons, as reflected in the significant three-way interaction between treatment, sex and age (table 3). Daughters (but not sons) from cortisol-treated mothers grew more slowly from one to three months compared with those from control (fed) mothers (daughters: age \times treatment, t=-4.17, p<0.0001; sons: t=-1.48, p=0.14), but

exhibited similar growth compared to those from untreated (unfed) mothers (daughters: age \times treatment, t = 0.65, p = 0.51; sons: t = -0.52, p = 0.6, table 3, figure 1). Daughters, but not sons, from control (fed) mothers grew faster than those from untreated mothers (daughters: age \times treatment, t = -4.24, p < 0.0001; sons: t = 1.35, p = 0.18).

(c) Effects of treatments on offspring cooperative behaviour

The effects of the maternal treatments on babysitting behaviour of offspring depended upon the age and sex of the offspring (table 4). Babysitting contributions in daughters

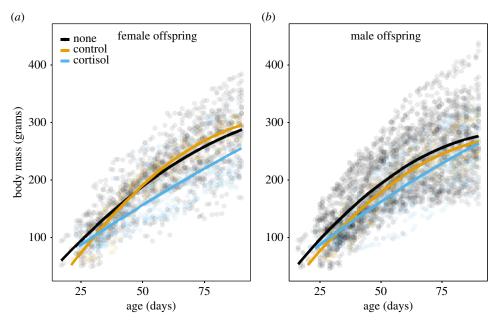


Figure 1. (*a*) Daughters but not (*b*) sons from mothers treated with cortisol during pregnancy were significantly smaller from initial emergence from their natal burrow to nutritional independence (approx. one to three months) compared with those from control litters (daughters: age \times treatment, t = -4.17, p < 0.0001; sons: t = -1.48, p = 0.14), but not untreated ('none') litters (daughters: age \times treatment, t = 0.65, p = 0.51; sons: t = -0.52, p = 0.6, table 3). Data are body mass measures from offspring from cortisol-treated (females: n = 373 estimates; males: n = 488), control (females: n = 215; males: n = 241) and untreated litters (females: n = 1121; males: n = 2238). Raw data and regression lines are shown (full results in table 3).

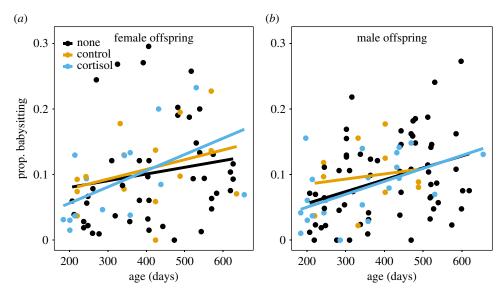


Figure 2. Babysitting contributions of (a) daughters and (b) sons from mothers treated with cortisol during pregnancy increased with age at a faster rate than those from control (females: z = -2.89, p = 0.0039; males: z = -1.92, p = 0.055), but not untreated (none) litters (females: z = 1.88, p = 0.06; males: z = -0.03, p = 0.97, table 4). Data are relative babysitting contributions from offspring from cortisol-treated (females: n = 15 estimates; males: n = 24), control (females: n = 15; males: n = 10) and untreated litters (females: n = 49; males: n = 69). Raw data and regression lines are shown (full results in table 4).

from mothers treated with cortisol during pregnancy were slightly, but significantly higher with increasing age of the babysitter compared with those from control mothers (age \times treatment, z=-2.89, p=0.0039) but not untreated mothers (age \times treatment, z=1.88, p=0.059; table 4, figure 2). Babysitting contributions in sons from mothers treated with cortisol during pregnancy showed a similar tendency to slightly increase with age compared to those from control mothers, but this difference was not significant (age \times treatment, z=-1.92, p=0.055). Further, age-related increases in babysitting contributions between males from mothers treated with cortisol during pregnancy and

untreated mothers did not differ (age×treatment, z = -0.03, p = 0.97; table 4, figure 2). Comparisons of the magnitude of effect sizes showed that the interaction between age and maternal treatment had a larger effect on babysitting contributions in daughters but not sons than other variables known to impact babysitting contributions, such as foraging success, age-related body mass, or group size (table 4).

The effects of the maternal treatments on pup feeding depended upon the sex of the offspring, but not their age (table 5). Daughters, but not sons from mothers treated with cortisol during pregnancy exhibited significantly more

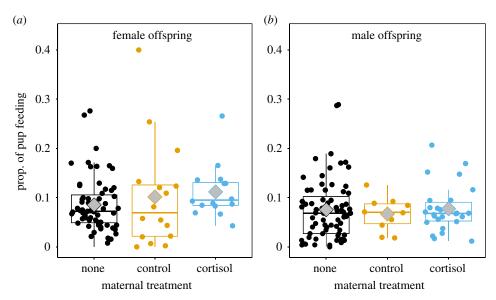


Figure 3. Pup feeding contributions of (a) daughters, but not (b) sons, from mothers treated with cortisol during pregnancy were significantly higher compared with from control (females: z = -3.12, p = 0.0018; males: z = -1.14, p = 0.25) or untreated (none) litters (females: z = -3.49, p = 0.0005; males: z = -1.03, p = 0.3, table 5). Data are relative pup feeding contributions from offspring from cortisol-treated (females: n = 16 estimates; males: n = 26), control (females: n = 16; males: n = 10) and untreated litters (females: n = 64; males: n = 71). Raw data are shown (full results in table 5). Boxplots show median (solid horizontal line), mean (grey diamonds) and first (25%) and third (75%) quartiles.

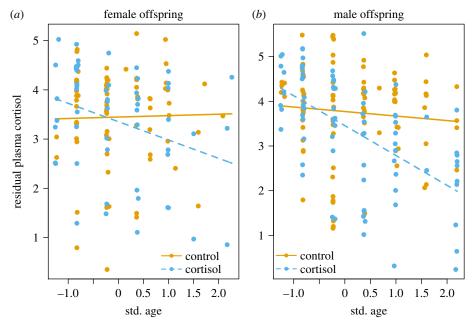


Figure 4. (*a*) Daughters and (*b*) sons from mothers treated with cortisol during pregnancy had lower plasma cortisol concentrations as they became older compared to those from control litters, though the difference was only significant in males (daughters: age \times treatment, t = -1.76, p = 0.08; sons: age \times treatment t = -2.68, p = 0.008, table 6). Data are residual plasma cortisol concentrations from offspring from cortisol-treated (females: n = 64 samples; males: n = 92) and control (females: n = 89; males: n = 104) litters. Residuals from a LMM (table 6) are shown on *y*-axis.

pup feeding contributions than those from control mothers (females: z=-3.12, p=0.00018; males: z=-1.14, p=0.25) or untreated mothers (females: z=-3.49, p=0.0005, sons: z=-1.03, p=0.3, table 5, figure 3). Notably, the magnitude of the effect size of maternal treatment for daughters was much larger than other variables known to impact baby-sitting contributions such as foraging success, age-related body mass and group size (table 5).

(d) Effects of treatments on offspring stress physiology Daughters from mothers treated with cortisol during pregnancy had lower plasma cortisol concentrations (age ×

treatment, t = -1.76, p = 0.08, table 6, figure 4a) and lower fGCM concentrations (age × treatment, t = -2.9, p = 0.004, table 7, figure 5a) as they became older compared with those from control mothers but these differences were only significant for fGCM concentrations. Sons from mothers treated with cortisol during pregnancy had significantly lower plasma cortisol concentrations as they became older compared with those from control mothers (age × treatment t = -2.68, p = 0.008, table 6, figure 4b) but similar fGCM concentrations compared with those from control mothers as they became older (age×treatment, t = -0.1, p = 0.49, table 7, figure 5b).

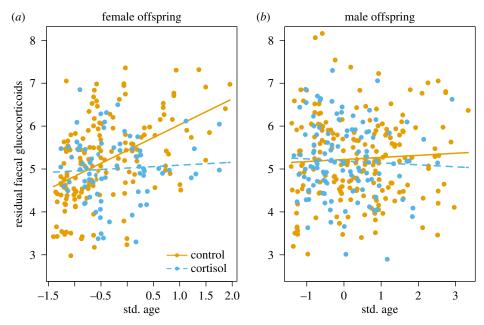


Figure 5. Faecal glucocorticoid metabolite (fGCM) concentrations in (a) daughters but not (b) sons from mothers treated with cortisol during pregnancy were significantly lower than those from control litters as they became older (daughters: age \times treatment, t=-2.9, p=0.004; sons: age \times treatment, t=-0.1, p=0.49, table 7). Data are residual fGCM concentrations from offspring from cortisol-treated (females: n=79 samples; males: n=118) and control (females: n=154; males: n=201) litters. Residuals from a LMM (table 7) are shown on y-axis.

4. Discussion

We found some support for our hypothesis that elevated maternal GCs would reduce the potential for offspring to have direct reproductive opportunities and would therefore shift them towards exhibiting more cooperative behaviour that could increase their indirect fitness. Daughters, but not sons, from mothers treated with cortisol during pregnancy grew more slowly early in life and exhibited more babysitting and pup feeding behaviour as they became older compared with controls. Other than offspring survival (table 2), we were unable to quantify the direct and indirect fitness of offspring from control or cortisol-treated mothers, but early life growth or body mass (which we measured here) is closely linked to direct fitness opportunities in daughters [26-31]. Previous studies in meerkats show that female, but not male, offspring that grow faster from one to three months are more likely to acquire the dominant breeding position [30], perhaps because offspring that grow faster in their first three months of life are heavier later in life [31,56,57], and heavier females are more likely to acquire a vacant dominant breeding position [29,31]. As such, daughters, but not sons, from mothers treated with cortisol during pregnancy should have reduced future direct fitness opportunities and therefore increase their investment in behaviours that elevate their indirect fitness. Our results are consistent with studies in other taxa that suggest that individuals adjust their contributions to cooperative behaviour according to their future reproductive potential. For example, in cooperatively breeding birds, when the chances of direct reproduction are elevated, subordinates often stop helping at the nest [58]. Studies of social wasps show that individuals whose probability of acquiring the dominant breeding position was experimentally increased exhibited significantly less helping behaviour [59,60]. Finally, in cooperatively breeding fish, subordinates will reduce their helping investment immediately prior to dispersal from their natal group where they attempt to

reproduce on their own rather than stay in their natal group and queue for dominance [61].

Our results show that increases in maternal GCs can increase the cooperative behaviour of daughters, which should lead to substantial direct fitness benefits to mothers. Daughters from mothers treated with cortisol during pregnancy exhibited more alloparental care compared with controls, such that subsequent offspring produced in groups with offspring from cortisol-treated mothers should have received more alloparental care. Because offspring that receive more alloparental care grow faster early in life or are larger later in life [31,56], the presence of offspring from cortisol-treated mothers should increase the direct fitness of dominant breeders and the indirect fitness of the offspring from cortisol-treated mothers. Taken together, our results suggest that this GC-mediated maternal effect reduced the direct fitness opportunities of daughters by reducing their early life growth, but they compensated by increasing their investment in indirect fitness opportunities (helping to rear non-descendent offspring). This is in line with theoretical predictions that parental manipulation of the cooperative behaviour of offspring can evolve if the costs of resisting the parental effect are high and inclusive fitness benefits of helping rear subsequent offspring are increased [18], as is the case in cooperative breeders.

Control females that were fed during pregnancy produced daughters that grew faster during early development (one to three months) compared with daughters from cortisol-treated or untreated mothers. Although mothers that were treated with cortisol during pregnancy received the same amount of supplemental food as controls, daughters and sons from mothers fed cortisol during pregnancy did not differ in early life growth compared with those from untreated mothers. This indicates that the additional food provided to dominant females during pregnancy had the potential to increase growth, but the added cortisol prevented those gains in body mass. This has implications for

understanding the fitness consequences of maternal stress on offspring growth trajectories [15,62] because our results show that elevated circulating GC levels in pregnant females in the absence of obvious energetic constraints induced reductions in the early life growth of offspring. This supports the hypothesis that maternal GC levels during offspring development act as a cue that induces plasticity in offspring growth rather than simply mediating the effects of energetic constraints. Alternatively, elevated maternal GCs could alter patterns of maternal investment in offspring. Identifying whether offspring or mothers are driving these effects is a major challenge in studies of maternal stress effects in wild animals.

The reductions in the activity of the neuroendocrine stress axis of daughters may have potentiated the increased alloparental care behaviour that we observed. Compared to daughters from control mothers, daughters from mothers treated with cortisol during pregnancy exhibited more babysitting as they became older, more overall pup feeding, and they also had lower plasma cortisol and fGCM concentrations as they got older. Males from mothers treated with cortisol during pregnancy had significantly lower plasma cortisol concentrations, but not fGCM concentrations as they got older and also tended to exhibit more babysitting as they aged. The activity of the neuroendocrine stress axis is closely linked to an array of social behaviours [63] and our recent work shows that elevated activity of the neuroendocrine stress axis reduces babysitting in both females and males and decreases pup feeding in females [38]. Together, this supports the hypothesis that the mechanism by which early life stress increases the cooperative behaviour of daughters is by dampening the activity of their neuroendocrine stress axis.

Our results show that the effects of maternal GCs on offspring growth, physiology and behaviour were greater in daughters than in sons, which adds to biomedical [64,65] and ecological [66-68] studies that highlight how early life conditions or maternal GC levels can have sex-specific consequences for offspring. In meerkats, there may be added benefits for the dominant female of altering the cooperative behaviour of daughters compared to sons; daughters exhibit more cooperative behaviour than sons [35] and are more responsive to the begging calls of subsequent offspring that they provision with food [69]. More broadly, sex-differences in natal dispersal may cause these differential responses to parental effects. In meerkats, subordinate males voluntarily disperse from their natal group to look for receptive females but can return to their natal group, whereas subordinate females rarely voluntarily disperse from their natal group [70]. In our case and in others [62], the more philopatric sex (females) is more sensitive to early life conditions, which may be owing to differential costs of parental modification between the philopatric and dispersing sex. If parental effects have long-term consequences on offspring characteristics, as we show here, there may be an increased degree of mismatch between the phenotype of the dispersing sex and the postnatal environment where individuals eventually settle. If this mismatch has fitness costs, this should select for individuals from the dispersing sex to be less responsive to cues from the parental phenotype or environment.

Our results provide some support for the hypothesis that parents may alter the cooperative tendencies of their offspring by manipulating the characteristics of their offspring [9,10], though we note that it is uncertain whether the

transfer of maternal GCs to offspring was passive or active. Explanations regarding the evolutionary origins of cooperative behaviour involve nepotism or kin selection [71], mutualisms or reciprocity [72], but few studies have tested the 'parental manipulation' hypothesis proposed by Alexander [9]. Some studies show that alleles that increase maternal fitness at the expense of the direct fitness of offspring can evolve [19] and that cooperative breeders may bias investment towards offspring that exhibit more cooperative behaviour [73]. Our study supports the hypothesis that environmental stressors may induce a parental effect that can modify the cooperative tendencies of their offspring.

Finally, our results have two implications for theoretical models examining the evolution of parental effects. First, given the sex-specificity of parental effects, our results challenge the conclusions of models examining the evolution of parental effects that assume that all offspring are equally sensitive to the parental effect [16], or those that assume that the benefits of exhibiting the phenotype resulting from the parental effect are equal for all offspring [18]. Second, selfish parental effects are thought to be relatively rare [8,13] and theory [16,17] and empirical studies showing sex-specific responses to early life stress [64,65] indicate that offspring can become resistant to such selfish parental effects. However, some models indicate that the evolution of selfish parental effects may be dependent upon the social environment [23], especially if the selfish parental effect influences the expression of alloparental care behaviour of offspring and therefore increases the indirect fitness of offspring. Our results provide an example whereby a GC-mediated maternal effect should decrease the direct fitness of daughters (by reducing their early life growth), but should increase the direct fitness of mothers and indirect fitness of daughters by elevating their cooperative behaviour.

Ethics. All protocols used in our experiments were approved by the Animal Ethics Committee at the University of Pretoria (Pretoria, South Africa: no. EC031-13, no. EC047-16) and the Northern Cape Department of Environment and Nature Conservation Research (South Africa: FAUNA 050/2013, FAUNA 192/2014, FAUNA 1020/2016).

Data accessibility. All data are available in the Dryad Digital Repository: https://doi.org/10.5061/dryad.260t0rm [74]. All code for statistical analyses is available from the first author.

Authors' contributions. B.D. and T.H.C.-B. planned the study, B.D. designed the experiments, B.D., Co.D, D.L.C., T.H.C.-B., D.G., I.B.G., N.C.B., M.H., A.G. and Ch.D. coordinated and collected data, B.D. conducted analyses and produced figures, B.D. and T.H.C.-B. authored the manuscript with contributions from all authors.

Competing interests. We have no competing interests.

Funding. Research was supported by grants from National Environment Research Council (RG53472) and the European Research Council (294494) to T.H.C.-B. I.B.G. was funded by the Swiss National Science Foundation (grant no. 31003A_13676 to M. Manser). N.C.B. was supported by a DST-NRF SARCHI chair of Mammal Behavioural Ecology and Physiology. The Kalahari Meerkat Project (KMP) has also been financed by the University of Cambridge (T.H.C.-B.) and University of Zurich (M. Manser).

Acknowledgements. We thank M. Manser for her contributions to maintain the long-term KMP and her contributions to this specific project. Thanks to P. Roth, S. Bischoff-Mattson, L. Howell, J. Samson, N. Thavarajah and numerous KMP volunteers who contributed to data collection. Thanks to M. Manser and S. Guindre-Parker for helpful comments. Thanks to the Northern Cape Conservation Authority for permission to conduct research and the Kotze family for allowing us to study meerkats on their property.

References

- Wolf JB, Wade MJ. 2009 What are maternal effects (and what are they not)? *Phil. Trans. R. Soc. B* 364, 1107 – 1115. (doi:10.1098/rstb.2008.0238)
- Kirkpatrick M, Lande R. 1989 The evolution of maternal characters. *Evolution* 43, 485-503. (doi:10.1111/j.1558-5646.1989.tb04247.x)
- Mousseau TA, Fox CW (eds). 1998 Maternal effects as adaptations. Oxford, UK: Oxford University Press.
- Agrawal AA, Laforsch C, Tollrian R. 1999
 Transgenerational induction of defences in animals and plants. *Nature* 401, 60–63. (doi:10.1038/43425)
- Dantzer B, Newman AE, Boonstra R, Palme R, Boutin S, Humphries MM, McAdam AG. 2013 Density triggers maternal hormones that increase adaptive offspring growth in a wild mammal. Science 340, 1215–1217. (doi:10.1126/science. 1235765)
- Duckworth RA, Belloni V, Anderson SR. 2015 Cycles of species replacement emerge from locally induced maternal effects on offspring behavior in a passerine bird. Science 347, 875 – 877. (doi:10. 1126/science.1260154)
- Uller T. 2008 Developmental plasticity and the evolution of parental effects. *Trends Ecol. Evol.* 23, 432–438. (doi:10.1016/j.tree.2008.04.005)
- Uller T, Nakagawa S, English S. 2013 Weak evidence for anticipatory parental effects in plants and animals. J. Evol. Biol. 26, 2161 – 2170. (doi:10.1111/ ieb.12212)
- Alexander RD. 1974 The evolution of social behavior. *Annu. Rev. Ecol. Syst.* 5, 325–383. (doi:10.1146/annurev.es.05.110174.001545)
- Michener CD, Brothers DJ. 1974 Were workers of eusocial Hymenoptera initially altruistic or oppressed? *Proc. Natl Acad. Sci. USA* 71, 671 – 674. (doi:10.1073/pnas.71.3.671)
- Müller W, Lessells CKM, Korsten P, Engelhardt NV. 2007 Manipulative signals in family conflict? On the function of maternal yolk hormones in birds. *Am. Nat.* **169**, E84–E96. (doi:10.1086/511962)
- Schwarzkopf L, Andrews RM. 2012 Are moms manipulative or just selfish? Evaluating the 'maternal manipulation hypothesis' and implications for life-history studies of reptiles. *Herpetologica* 68, 147 – 159. (doi:10.1655/HERPETOLOGICA-D-11-00009.1)
- Marshall DJ, Uller T. 2007 When is a maternal effect adaptive? *Oikos* 116, 1957 – 1963. (doi:10.1111/j. 2007.0030-1299.16203.x)
- Wilson AJ, Pilkington JG, Pemberton JM, Coltman DW, Overall AD, Byrne KA, Kruuk LEB. 2005 Selection on mothers and offspring: whose phenotype is it and does it matter? *Evolution* 59, 451–463. (doi:10. 1111/j.0014-3820.2005.tb01003.x)
- Berghänel A, Heistermann M, Schülke O, Ostner J. 2017 Prenatal stress accelerates offspring growth to compensate for reduced maternal investment across mammals. Proc. Natl Acad. Sci. USA 114, E10658—E10666. (doi:10.1073/pnas.1707152114)

- Uller T, Pen I. 2011 A theoretical model of the evolution of maternal effects under parent – offspring conflict. *Evolution* 65, 2075 – 2084. (doi:10.1111/j.1558-5646.2011.01282.x)
- 17. Kuijper B, Johnstone RA. 2018 Maternal effects and parent offspring conflict. *Evolution* **72**, 220 233. (doi:10.1111/evo.13403)
- González-Forero M, Gavrilets S. 2013 Evolution of manipulated behavior. *Am. Nat.* 182, 439–451. (doi:10.1086/671932)
- Wade MJ. 2001 Maternal effect genes and the evolution of sociality in haplo-diploid organisms. Evolution 55, 453 – 458. (doi:10.1554/0014-3820(2001)055[0453:MEGATE]2.0.C0;2)
- Linksvayer TA, Wade MJ. 2005 The evolutionary origin and elaboration of sociality in the aculeate Hymenoptera: maternal effects, sib-social effects, and heterochrony. Q Rev. Biol. 80, 317–336. (doi:10.1086/432266)
- 21. Bourke AF, Franks NR. 1995 *Social evolution in ants*. Princeton, NJ: Princeton University Press.
- 22. Russell A F, Lummaa V. 2009 Maternal effects in cooperative breeders: from hymenopterans to humans. *Phil. Trans. R. Soc. B* **364**, 1143–1167. (doi:10.1098/rstb.2008.0298)
- 23. Taborsky B. 2017 Developmental plasticity: preparing for life in a complex world. *Adv. Study Behav.* **49**, 49–99. (doi:10.1016/bs.asb.2016. 12.002)
- Carter CS, Roberts RL. 1997 The psychobiological basis of cooperative breeding in rodents. In Cooperative breeding in mammals (eds NG Solomon, JA French), pp. 231–266. Cambridge, UK: Cambridge University Press.
- 25. De Fraipont M, Clobert J, Alder HJ, Meylan S. 2000 Increased pre-natal maternal corticosterone promotes philopatry of offspring in common lizards *Lacerta vivipara*. *J. Anim. Ecol.* **69**, 404–413. (doi:10.1046/j.1365-2656.2000.00405.x)
- Clutton-Brock TH, Brotherton PNM, O'Riain MJ, Griffin AS, Gaynor D, Kansky R, Sharpe L, McIlrath GM. 2001 Contributions to cooperative rearing in meerkats. *Anim. Behav.* 61, 705 – 710. (doi:10.1006/ anbe.2000.1631)
- Russell AF, Clutton-Brock TH, Brotherton PN, Sharpe LL, Mcilrath G, Dalerum FD, Cameron EZ, Barnard JA. 2002 Factors affecting pup growth and survival in co-operatively breeding meerkats *Suricata* suricatta. J. Anim. Ecol. 71, 700 – 709. (doi:10.1046/ i.1365-2656.2002.00636.x)
- 28. Thornton A. 2008 Social learning about novel foods in young meerkats. *Anim. Behav.* **76**, 1411–1421. (doi:10.1016/j.anbehav.2008.07.007)
- Hodge SJ, Manica A, Flower TP, Clutton-Brock TH. 2008 Determinants of reproductive success in dominant female meerkats. J. Anim. Ecol. 77, 92 – 102. (doi:10.1111/j.1365-2656.2007.01318.x)
- English S, Huchard E, Nielsen JF, Clutton-Brock TH.
 2013 Early growth, dominance acquisition and lifetime reproductive success in male and female

- cooperative meerkats. *Ecol. Evol.* **3**, 4401 4407. (doi:10.1002/ece3.820)
- Russell AF, Young AJ, Spong G, Jordan NR, Clutton-Brock TH. 2007 Helpers increase the reproductive potential of offspring in cooperative meerkats. *Proc. R. Soc. B* 274, 513 – 520. (doi:10.1098/rspb.2006.3698)
- 32. Clutton-Brock TH *et al.* 1998 Costs of cooperative behaviour in suricates (*Suricata suricatta*). *Proc. R. Soc. Lond. B* **265**, 185 190. (doi:10.1098/rspb.1998.0281)
- Brotherton PNM, Clutton-Brock TH, O'Riain MJ, Gaynor D, Sharpe L, Kansky R, McIlrath GM. 2001 Offspring food allocation by parents and helpers in a cooperative mammal. *Behav. Ecol.* 12, 590 – 599. (doi:10.1093/beheco/12.5.590)
- Russell AF, Sharpe LL, Brotherton PNM, Clutton-Brock TH. 2003 Cost minimization by helpers in cooperative vertebrates. *Proc. Natl Acad. Sci. USA* 100, 3333-3338. (doi:10.1073/pnas.0636503100)
- Clutton-Brock TH, Russell AF, Sharpe LL, Young AJ, Balmforth Z, McIlrath GM. 2002 Evolution and development of sex differences in cooperative behavior in meerkats. Science 297, 253 – 256. (doi:10.1126/science.1071412)
- Bell MBV, Cant MA, Borgeaud C, Thavarajah N, Samson J, Clutton-Brock TH. 2014 Suppressing subordinate reproduction provides benefits to dominants in cooperative societies of meerkats. *Nat. Commun.* 5, 4499. (doi:10.1038/ncomms5499)
- 37. Harris A, Seckl J. 2011 Glucocorticoids, prenatal stress and the programming of disease. *Horm. Behav.* **59**, 279–289. (doi:10.1016/j.yhbeh.2010.06.007)
- Dantzer B et al. 2017 The influence of stress hormones and aggression on cooperative behaviour in subordinate meerkats. Proc. R. Soc. B 284, 20171248. (doi:10.1098/rspb.2017.1248)
- 39. English S, Bateman AW, Mares R, Ozgul A, Clutton-Brock TH. 2014 Maternal, social and abiotic environmental effects on growth vary across life stages in a cooperative mammal. *J. Anim. Ecol.* **83**, 332–342. (doi:10.1111/1365-2656.12149)
- Kutsukake N, Clutton-Brock TH. 2006 Aggression and submission reflect reproductive conflict between females in cooperatively breeding meerkats Suricata suricatta. Behav. Ecol. Sociobiol. 59, 541–548. (doi:10.1007/s00265-005-0079-7)
- Clutton-Brock TH, Brotherton PNM, O'Riain MJ, Griffin AS, Gaynor D, Sharpe L, Kansky R, Manser MB, McIlrath GM. 2000 Individual contributions to babysitting in a cooperative mongoose, Suricata suricatta. Proc. R. Soc. B 267, 301–305. (doi:10. 1098/rspb.2000.1000)
- Kunc HP, Madden JR, Manser MB. 2007 Begging signals in a mobile feeding system: the evolution of different call types. *Am. Nat.* 170, 617–624. (doi:10.1086/521233)
- Carlson AA, Young AJ, Russell AF, Bennett NC, McNeilly AS, Clutton-Brock T. 2004 Hormonal correlates of dominance in meerkats (*Suricata suricatta*). Horm. Behav. 46, 141–150. (doi:10. 1016/j.yhbeh.2004.01.009)

- 44. Dantzer B, Bennett NC, Clutton-Brock TH. 2017 Social conflict and costs of cooperation in meerkats are reflected in measures of stress hormones. Behav. Ecol. 28, 1131-1141. (doi:10. 1093/beheco/arx077)
- 45. Heistermann M, Finke M, Hodges JK. 1995 Assessment of female reproductive status in captivehoused Hanuman langurs (Presbytis entellus) by measurement of urinary and fecal steroid excretion patterns. Am. J. Primatol. 37, 275-284. (doi:10. 1002/ajp.1350370402)
- 46. Braga Goncalves I, Heistermann M, Santema P, Dantzer B, Mausbach J, Ganswindt A, Manser MB. 2016 Validation of a fecal glucocorticoid assay to assess adrenocortical activity in meerkats using physiological and biological stimuli. PLoS ONE 11, e0153161. (doi:10.1371/journal.pone.0153161)
- 47. Heistermann M, Palme R, Ganswindt A. 2006 Comparison of different enzyme immunoassays for assessment of adrenocortical activity in primates based on fecal analysis. Am. J. Primatol. 68, 257 – 273. (doi:10.1002/ajp.20222)
- 48. Clutton-Brock TH, Russell AF, Sharpe LL. 2003 Meerkat helpers do not specialize in particular activities. Anim. Behav. 66, 531-540. (doi:10.1006/ anbe.2003.2209)
- 49. Carlson AA, Manser MB, Young AJ, Russell AF, Jordan NR, McNeilly AS, Clutton-Brock T. 2006 Cortisol levels are positively associated with pupfeeding rates in male meerkats. Proc. R. Soc. B 273, 571 – 577. (doi:10.1098/rspb.2005.3087)
- 50. Lesnoff M, Lancelot R. 2013 aods3: analysis of overdispersed data using S3 methods. aods3 package version 0.4-1.
- 51. R Development Core Team. 2013 R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. See www.r-project.org/.
- 52. Bates D, Maechler M, Bolker B, Walker S. 2014 lme4: linear mixed-effects models using Eigen and S4. R Package Version 1, 1-23.
- 53. Kuznetsova A, Brockhoff PB, Christensen RHB. 2016 ImerTest: tests in linear mixed effects models. R

- package version 2.0-30. See http://CRAN.R-project. org/package=ImerTest.
- 54. Zuur AF, Ieno EN, Elphick CS. 2010 A protocol for data exploration to avoid common statistical problems. *Meth. Ecol. Evol.* **1**, 3 – 14. (doi:10.1111/j. 2041-210X.2009.00001.x)
- 55. Fox J, Monette G. 1992 Generalized collinearity diagnostics. J. Am. Stat. Assoc. 87, 178-183. (doi:10.1080/01621459.1992.10475190)
- 56. Clutton-Brock TH, Russell AF, Sharpe LL, Brotherton PNM, McIlrath GM, White S, Cameron EZ. 2001 Effects of helpers on juvenile development and survival in meerkats. Science 293, 2446-2449. (doi:10.1126/science.1061274)
- 57. Huchard E, English S, Bell MB, Thavarajah N, Clutton-Brock T. 2016 Competitive growth in a cooperative mammal. Nature 533, 532. (doi:10. 1038/nature17986)
- 58. Hatchwell BJ, Komdeur J. 2000 Ecological constraints, life history traits and the evolution of cooperative breeding. Anim. Behav. 59, 1079 - 1086. (doi:10.1006/anbe.2000.1394)
- 59. Cant MA, Field J. 2001 Helping effort and future fitness in cooperative animal societies. Proc. R. Soc. B 268, 1959-1964. (doi:10.1098/ rspb.2001.1754)
- 60. Field J, Cronin A, Bridge C. 2006 Future fitness and helping in social queues. Nature 441, 214. (doi:10. 1038/nature04560)
- 61. Zöttl M, Chapuis L, Freiburghaus M, Taborsky M. 2013 Strategic reduction of help before dispersal in a cooperative breeder. Biol. Lett. 9, 20120878. (doi:10.1098/rsbl.2012.0878)
- 62. Sheriff MJ et al. 2017 Integrating ecological and evolutionary context in the study of maternal stress. Integr. Comp. Biol. **57**, 437 – 449. (doi:10.1093/icb/ icx105)
- 63. Raulo A, Dantzer B. 2018 Associations between glucocorticoids and sociality across a continuum of vertebrate social behaviour. Ecol. Evol. 8, 7697 – 7716. (doi:10.1002/ece3.4059)
- 64. Weinstock M. 2007 Gender differences in the effects of prenatal stress on brain development and

- behaviour. Neurochem. Res. 32, 1730-1740. (doi:10.1007/s11064-007-9339-4)
- 65. Mueller BR, Bale TL. 2008 Sex-specific programming of offspring emotionality after stress early in pregnancy. J. Neurosci. 28, 9055-9065. (doi:10. 1523/JNEUROSCI.1424-08.2008)
- 66. Wilkin TA, Sheldon BC. 2009 Sex differences in the persistence of natal environmental effects on life histories. Curr. Biol. 19, 1998 – 2002. (doi:10.1016/j. cub.2009.09.065)
- 67. Metzger DC, Schulte PM. 2016 Maternal stress has divergent effects on gene expression patterns in the brains of male and female threespine stickleback. Proc. R. Soc. B 283, 20161734. (doi:10.1098/rspb. 2016.1734)
- 68. Marshall HH et al. 2017 Lifetime fitness consequences of early-life ecological hardship in a wild mammal population. Ecol. Evol. 7, 1712 – 1724. (doi:10.1002/ece3.2747)
- 69. English S, Kunc HP, Madden JR, Clutton-Brock TH. 2008 Sex differences in responsiveness to begging in a cooperative mammal. Biol. Lett. 4, 334-337. (doi:10.1098/rsbl.2008.0200)
- 70. Clutton-Brock TH, Manser M. 2016 Meerkats: cooperative breeding in the Kalahari. In Cooperative breeding in vertebrates (eds WD Koenig, JL Dickinson), pp. 294-317. Cambridge, UK: Cambridge University Press.
- 71. Hamilton WD. 1964 The genetical evolution of social behaviour. II. J. Theor. Biol. 7, 17-52. (doi:10.1016/0022-5193(64)90039-6)
- 72. Trivers RL. 1971 The evolution of reciprocal altruism. Q. Rev. Biol. 46, 35-57. (doi:10.1086/
- 73. Griffin AS, Sheldon BC, West SA. 2005 Cooperative breeders adjust offspring sex ratios to produce helpful helpers. Am. Nat. **166**, 628 – 632. (doi:10.1086/491662)
- 74. Dantzer B et al. 2019 Data from: The development of individual differences in cooperative behaviour: maternal glucocorticoid hormones alter helping behaviour of offspring in wild meerkats. Dryad Digital Repository. (https://doi.org/10.5061/dryad.