



HLA-B*07, HLA-DRB1*07, HLA-DRB1*12, and HLA-C*03:02 Strongly Associate With BMI: Data From 1.3 Million Healthy Chinese Adults

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Strong associations between HLA alleles and infectious and autoimmune diseases are well established. Although obesity is also associated with these diseases, the relationship between HLA and obesity has not been systematically investigated in a large cohort. In the current study, we analyzed the association of HLA alleles with BMI using data from 1.3 million healthy adult donors from the China Marrow Donor Program (CMDP). We found 23 HLA alleles, including 12 low-resolution and 11 high-resolution alleles, were significantly associated with BMI after correction for multiple testing. Alleles associated with high BMI were enriched in haplotypes that were common in both Chinese and European populations, whereas the alleles associated with low BMI were enriched in haplotypes common only in Asians. Alleles B*07, DRB1*07, DRB1*12, and C*03:02 provided the strongest associations with BMI ($P = 6.89 \times 10^{-10}$, 1.32×10^{-9} , 1.52×10^{-9} , and 4.45×10^{-8} , respectively), where B*07 and DRB1*07 also had evidence for sex-specific effects ($P_{\text{heterogeneity}} = 0.0067$ and 0.00058 , respectively). These results, which identify associations between alleles of HLA-B, DRB1, and

C with BMI in Chinese young adults, implicate a novel biological connection between HLA alleles and obesity.

Being overweight (BMI of 25–29.9 kg/m²) or obese (BMI ≥ 30 kg/m²) is well known to be a risk factor for several chronic diseases, including type 2 diabetes (T2D), cardiovascular disease, hypertension, and cancer. Because BMI is heritable, many groups have sought to identify its genetic determinants. Genome-wide association studies (GWAS) in individuals of European and Asian descent have identified >100 loci that associate with BMI or an obesity-related measure (1–6). However, commonly used GWAS platforms do not accurately capture genotypic variation within the HLA region due to its sequence complexity. In contrast, focused analyses of the HLA region have identified several alleles that have notable effects on disease susceptibility, including infectious diseases, autoimmune diseases, cancer, schizophrenia, and T2D (7–11). Despite obesity being a major

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risk factor for several of these diseases, no large-scale study published to date has interrogated the HLA region to determine whether HLA alleles associate with BMI. Among the few studies that have investigated HLA in relation to obesity in small cohorts, the results have been inconsistent (12–15).

The HLA region, also known as the MHC, encompasses ~3.6 Mb on chromosome 6p21.3 and includes five classic transplantation HLA genes: *HLA-A*, *HLA-C*, *HLA-B*, *HLA-DRB1*, and *HLA-DQB1*. There are two classes of MHC proteins, MHC class I (*HLA-A*, *-B*, and *-C*), which present peptides from inside the cell to T cells (endogenous antigens), and MHC class II (*HLA-DP*, *-DM*, *-DOA*, *-DOB*, *-DQ*, and *-DR*), which present antigens from outside of the cell (exogenous antigens) (16). HLA alleles affect the response to many infectious and autoimmune diseases due to their essential role in activation of both natural killer cells and T cells (17,18). Response to pathogens, such as human and nonhuman viruses, scrapie agents, bacteria, and gut microflora, has been reported to cause or exacerbate obesity in various experimental models (19). For example, the gut microbiota has been linked to multiple human diseases, including obesity (20,21), and human adenoviruses (Ad) 36, Ad-37, and Ad-5 have been shown to stimulate enzymes and transcription factors that cause accumulation of triglycerides and affect the differentiation of preadipocytes into mature adipocytes (22,23). Therefore, the diversity in HLA alleles may differentially alter the gut microbiota as well as differentially affect response to pathogens or infection, which in turn could have a subtle effect on obesity.

To determine whether a relationship exists between HLA alleles and BMI, we used the statistical power of data obtained

from 1.3 million healthy donors aged 18–50 years. Data were derived from the Data Bank of Chinese Hematopoietic Stem Cell Donors, also known as the Chinese Marrow Donor Program (CMDP), which has information on low-resolution and high-resolution HLA genotypes and anthropometric measures on >1.8 million donors through 31 branch registries.

RESEARCH DESIGN AND METHODS

Study Participants and Data Collection

The CMDP began in the year 2001 and is operated under the guidance of the Red Cross Society of China. Through this program, anthropometric data on >1.8 million donors from 31 branch registries have been collected. These branches cover the major geographic areas of all 31 provinces, autonomous regions, and municipalities in mainland China (24).

In the current study, basic information on the donor (age, sex, ethnicity/race) and anthropometric measures (weight, height) of 1,816,443 subjects with unique CMDP identification numbers (from dates 2 October 2001 to 2 October 2014) were obtained from the CMDP. Excluded were individuals with missing data on weight, height, age, or sex ($n = 600,352$) and individuals who were outliers for body weight (>216 kg or <30.8 kg) or height (>194.1 cm or <132.2 cm), as defined by the Survey on the Status of Nutrition and Health of the Chinese People conducted in 2002 (25) ($n = 11,871$). Also excluded were individuals aged <18 years or >50 years ($n = 12,338$) because relatively few individuals were in these age-groups in this database. All individuals examined from 2001 to 2003 ($n = 23,847$) were further excluded because of the small sample size and uneven distribution across different

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provinces. After these exclusions, data for 1,377,047 adults were available for analysis.

Individuals from the different provinces were grouped into seven geographical regions and analyzed separately. The seven regions are as follows: 1) Northeast China (Heilongjiang, Jilin, and Liaoning); 2) North China (Beijing, Tianjin, Hebei, Shanxi, and Inner Mongolia); 3) Northwest China (Xinjiang, Qinghai, Gansu, Ningxia, and Shaanxi); 4) East China (Shanghai, Shandong, Jiangsu, Anhui, Zhejiang, Jiangxi, and Fujian); 5) Central China (Henan, Hubei, Hunan); 6) South China (Guangdong, Guangxi, Hainan); and 7) Southwest China (Chongqing, Tibet, Yunnan, Sichuan, Guizhou). The locations of the seven geographical regions and the sample size from each region are shown in Fig. 1A and B. Taiwan, Hong Kong, and Macau were not included in this study.

The Institutional Review Board of the First Affiliated Hospital within Nanjing Medical University and the CMDP Ethics Review Board approved the study protocol.

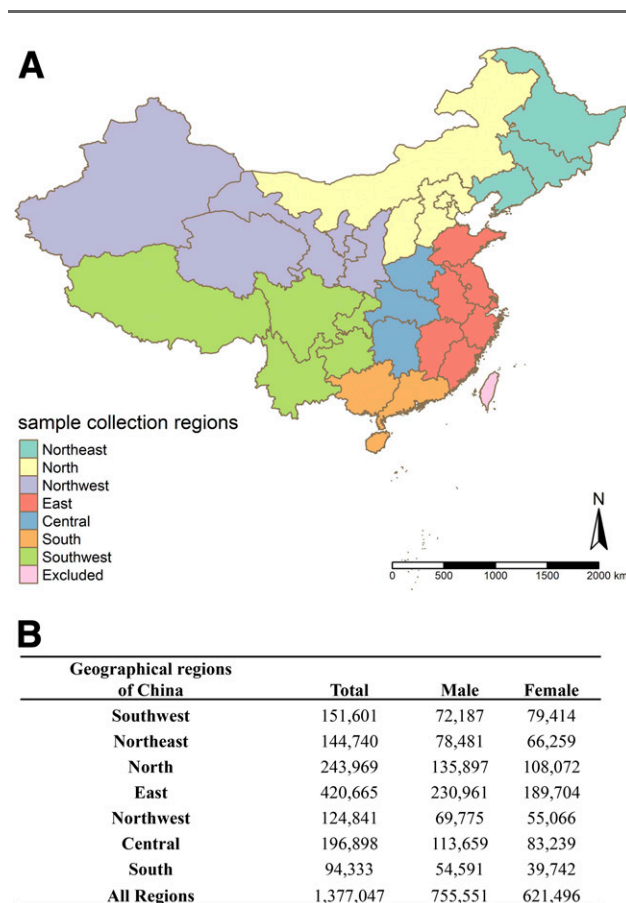


Figure 1—Seven geographical regions in China map (A) and sample size from each region (B). Individuals from the different provinces were grouped into seven geographical regions and analyzed separately. The seven regions are as follows: 1) Northeast China (Heilongjiang, Jilin, and Liaoning); 2) North China (Beijing, Tianjin, Hebei, Shanxi, and Inner Mongolia); 3) Northwest China (Xinjiang, Qinghai, Gansu, Ningxia, and Shaanxi); 4) East China (Shanghai, Shandong, Jiangsu, Anhui, Zhejiang, Jiangxi, and Fujian); 5) Central China (Henan, Hubei, Hunan); 6) South China (Guangdong, Guangxi, Hainan); and 7) Southwest China (Chongqing, Tibet, Yunnan, Sichuan, Guizhou). Taiwan, Hong Kong, and Macau were not included in this study.

Study Outcome Definitions

Anthropometric measurements were made by trained technicians in mobile examination centers at each of the 31 CMDP branch registries. Height (cm) and weight (kg) were measured using metal weighing scales with height rods. Subjects were required to stand straight on the instrument, barefoot, and at ease. BMI was calculated as weight in kilograms divided by height in meters squared and rounded to the nearest tenth. Birth date and sex were obtained from the individual’s identification card, and age was calculated using the sampling date and birth date.

HLA Genotyping

HLA typing was performed at 31 CMDP-accredited HLA typing laboratories. HLA typing was done by serological methods from 2001 to 2005. In 2005, high-resolution HLA genotyping was performed using three methods: PCR–reverse sequence-specific oligonucleotide probes method (26), Sanger sequencing-based typing, and next-generation high-throughput sequence-based typing (27). HLA data, including *HLA-A*, *-B*, *-C*, *-DRB1*, and *-DQB1*, were submitted to the CMDP database annually by each HLA typing laboratory. HLA high-resolution genotypes are given a four-digit code representing protein level assignment, and HLA low-resolution genotypes are given a two-digit code.

Statistical Analysis

For each HLA locus, multiple alleles were expanded as multiple biallelic markers for each testing allele by custom python scripts (<https://github.com/zhenyisong/guo.project>). Homozygotes for the testing allele (T/T), heterozygotes (T/O), and homozygotes for the other allele combined (O/O) were coded as a continuous numeric variable for genotype (2, 1, and 0 copies of the testing allele). Association analysis with BMI was performed using multiple linear regression with adjustment for age, sex, residential province, and sampling year in each of the seven geographical regions separately, and then meta-analyzed using the inverse variance method, which is based on β values or the odds ratio (OR) and SE from each geographical region. Cochran’s Q statistics were used to assess consistency of effect and to quantify heterogeneity among the seven regions. The regression analysis and meta-analysis were performed in Golden Helix SNP & Variation Suite 8.1 software (Bozeman, MT). A Bonferroni multiple testing correction was used to account for the number of common low-resolution or high-resolution HLA alleles.

Multiple linear regression, as described above, was repeated for men and women separately with correction for age, residential province, and sampling year in each of the seven geographical regions separately. A meta-analysis was performed to combine a sex-differentiated test of association and a test of heterogeneity in allelic effects between men and women implemented in GWAMA (Genome-Wide Association Meta-Analysis) software (28).

The variance explained by a single HLA allele was calculated by $2 \times MAF \times (1 - MAF) \times \beta^2$, where *MAF*

is the minor allele frequency, and β is the HLA allele effect estimate, assuming an additive model, computed by meta-analysis (29,30).

Multilocus haplotypes were constructed, and their frequencies were calculated using the expectation-maximization algorithm based on a subset of the samples in this study using the Arlequin software (31).

The estimated allele frequencies in Chinese from the current study were compared with the corresponding allele frequencies of Europeans obtained from the Allele Frequency Net Database (<http://www.allelefrequencies.net/>). The allele count calculated from frequency and total number of the samples was used for χ^2 or the Fisher exact test. Testing the difference in the proportions of alleles associated with high BMI between alleles that have significantly higher allele frequencies in European and those in Chinese was performed by the Fisher exact test. The significance level of variable difference used a value of $P < 0.05$.

RESULTS

Characteristics of the Participants

A total of 1,377,047 subjects (755,551 men and 621,496 women) aged 18–50 years were included in the association

analysis for BMI. The age distribution for men and women was asymmetric due to an overrepresentation of younger people who typically participate in the CMDP. The mean age of subjects included in the final analysis was 27.2 for men and 25.6 years for women (Supplementary Fig. 1).

Association Between HLA Alleles and BMI

Approximately 99% of the individuals had low-resolution HLA genotypic data, where 125 different alleles were detected, and ~67% of the individuals had both low- and high-resolution HLA genotypic data, where 1,455 different alleles were detected (allele frequencies reported in Supplementary Table 1). To preserve statistical power and reduce the multiple testing burden, we initially focused on 72 low-resolution alleles (57% of total) with a minor allele frequency >0.001 and performed a multiple linear regression analysis for BMI with adjustment for appropriate covariates in each geographical region separately. We then performed a meta-analysis for the seven geographical regions. Significance for the low-resolution alleles was defined as a P value of ≤ 0.0006 ($0.05/72$) based on a stringent Bonferroni correction, which assumes 72 independent tests, although linkage disequilibrium (LD) within the HLA region has been well documented. We then performed multiple linear regression

Table 1—Meta-analysis results of 23 HLA alleles that associated with BMI in CMDP

Allele	Gene	Frequency	P value	$\beta \pm SE$	Explained variance (%)
A*02	<i>HLA-A</i>	0.306	4.07E-04	0.015 \pm 0.004	0.010
B*07	<i>HLA-B</i>	0.030	6.89E-10	0.071 \pm 0.011	0.029
B*08	<i>HLA-B</i>	0.009	1.53E-04	0.079 \pm 0.021	0.011
B*13	<i>HLA-B</i>	0.110	4.37E-04	0.022 \pm 0.006	0.009
B*46	<i>HLA-B</i>	0.100	1.10E-06	0.042 \pm 0.009	0.032
B*58	<i>HLA-B</i>	0.057	3.75E-05	0.086 \pm 0.021	0.080
C*07	<i>HLA-C</i>	0.176	5.09E-04	0.038 \pm 0.011	0.042
DRB1*03	<i>HLA-DRB1</i>	0.048	1.06E-07	0.048 \pm 0.009	0.021
DRB1*04	<i>HLA-DRB1</i>	0.113	6.15E-06	0.028 \pm 0.006	0.016
DRB1*07	<i>HLA-DRB1</i>	0.093	1.32E-08	0.041 \pm 0.007	0.028
DRB1*09	<i>HLA-DRB1</i>	0.145	9.76E-05	0.024 \pm 0.006	0.014
DRB1*12	<i>HLA-DRB1</i>	0.121	1.52E-09	0.047 \pm 0.008	0.047
A*02:01	<i>HLA-A</i>	0.125	3.75E-06	0.035 \pm 0.007	0.027
B*07:02	<i>HLA-B</i>	0.022	1.12E-07	0.089 \pm 0.017	0.034
B*27:04	<i>HLA-B</i>	0.010	4.15E-05	0.113 \pm 0.028	0.025
B*40:01	<i>HLA-B</i>	0.099	5.78E-05	0.038 \pm 0.009	0.026
C*01:03	<i>HLA-C</i>	0.006	1.00E-04	0.182 \pm 0.047	0.042
C*03:02	<i>HLA-C</i>	0.059	4.45E-08	0.085 \pm 0.016	0.081
DQB1*03:02	<i>HLA-DQB1</i>	0.057	1.94E-04	0.066 \pm 0.018	0.047
DQB1*06:02	<i>HLA-DQB1</i>	0.076	9.55E-06	0.122 \pm 0.028	0.210
DRB1*03:01	<i>HLA-DRB1</i>	0.048	1.11E-05	0.049 \pm 0.011	0.022
DRB1*07:01	<i>HLA-DRB1</i>	0.093	6.49E-06	0.040 \pm 0.009	0.027
DRB1*12:02	<i>HLA-DRB1</i>	0.082	5.13E-05	0.047 \pm 0.012	0.033

Bold font indicates the HLA alleles with the strongest associations ($P < 5.0 \times 10^{-8}$). Significance is defined as $P \leq 0.0006$ ($0.05/72$) for low-resolution alleles and $P \leq 0.0003$ ($0.05/163$) for high-resolution HLA alleles based on a stringent Bonferroni correction. Explained variance (%) was estimated based on the effect sizes (β) and allele frequencies in CMDP. $\beta \pm SE$ is the regression slope and its SE.

analysis and meta-analysis for BMI on the 163 high-resolution alleles (11% of total) with a minor allele frequency >0.001. Significance for the high resolution alleles was defined as a *P* value of ≤0.0003 (0.05/163).

Meta-analysis of the results from the seven geographical regions showed that 12 low-resolution alleles (*A*02*, *B*07*, *B*08*, *B*13*, *B*46*, *B*58*, *C*07*, *DRB1*03*, *DRB1*04*, *DRB1*07*, *DRB1*09*, and *DRB1*12*) and 11 high-resolution alleles (*A*02:01*, *B*07:02*, *B*27:04*, *B*40:01*, *C*01:03*, *C*03:02*, *DQB1*03:02*, *DQB1*06:02*, *DRB1*03:01*, *DRB1*07:01*, and *DRB1*12:02*) were significantly associated with BMI (Table 1). Performance of a genotype pairwise correlation analysis among these alleles identified strong correlations ($r^2 > 0.3$) between high-resolution alleles (*A*02:01*, *B*07:02*, *DRB1*03:01*, *DRB1*07:01*, and *DRB1*12:02*) and their respective low resolution alleles (*A*02*, *B*07*, *DRB1*03*, *DRB1*07*, and *DRB1*12*) (Fig. 5A and Supplementary Table 4), suggesting

these high-resolution alleles are almost identical to their respective low-resolution alleles. Therefore, 19 unique HLA alleles were found to be associated BMI.

Three low-resolution alleles and one high-resolution allele had the strongest evidence for association with BMI ($P < 5 \times 10^{-8}$) (Fig. 2). These alleles are *B*07* ($P = 6.89 \times 10^{-10}$) from the *HLA-B* locus, *C*03:02* ($P = 4.45 \times 10^{-8}$) from the *HLA-C* locus, and *DRB1*07* ($P = 1.32 \times 10^{-8}$) and *DRB1*12* ($P = 1.52 \times 10^{-9}$) from the *HLA-DRB1* locus. Heterogeneity in the direction of the effect was not detected among the different geographical regions (heterogeneity $P > 0.05$) (forest plots shown in Fig. 3). These alleles were also assessed for their association with categories of BMI, namely, overweight (BMI 25.0–30.0 kg/m²; $n = \sim 236,000$ case subjects and $\sim 850,000$ control subjects) and obese (BMI ≥30 kg/m²; $n = \sim 40,000$ case subjects and $\sim 850,000$ control subjects) (Supplementary Table 3). Notably, *B*07*, *DRB1*07*, and

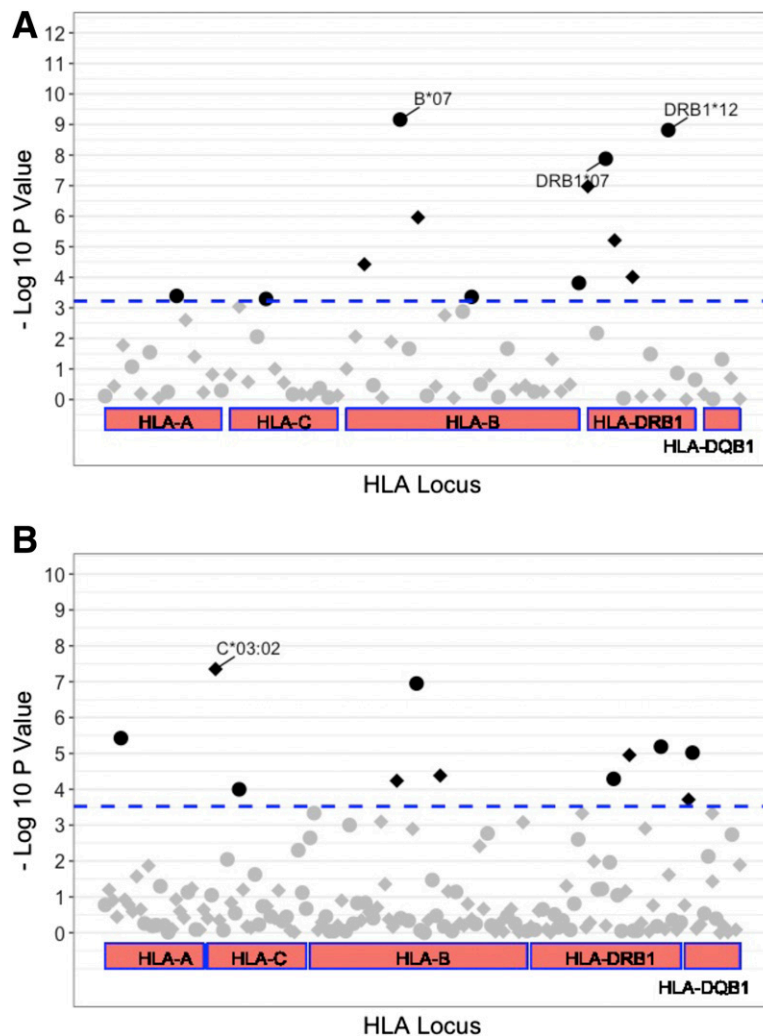


Figure 2—Scatter plots of association results of low-resolution (A) and high-resolution (B) HLA alleles with BMI in CMDP. The $-\log_{10} P$ values were plotted against their positions in each gene. HLA alleles associated with higher BMI are indicated by circles and HLA alleles associated with lower BMI are indicated by diamonds. The HLA alleles with the strongest association ($P < 0.00000005$) are labeled with their allele name. The horizontal dashed line represents the threshold for the statistical significance after Bonferroni correction ($P = 0.0006$ for low-resolution allele and $P = 0.0003$ for high-resolution alleles).

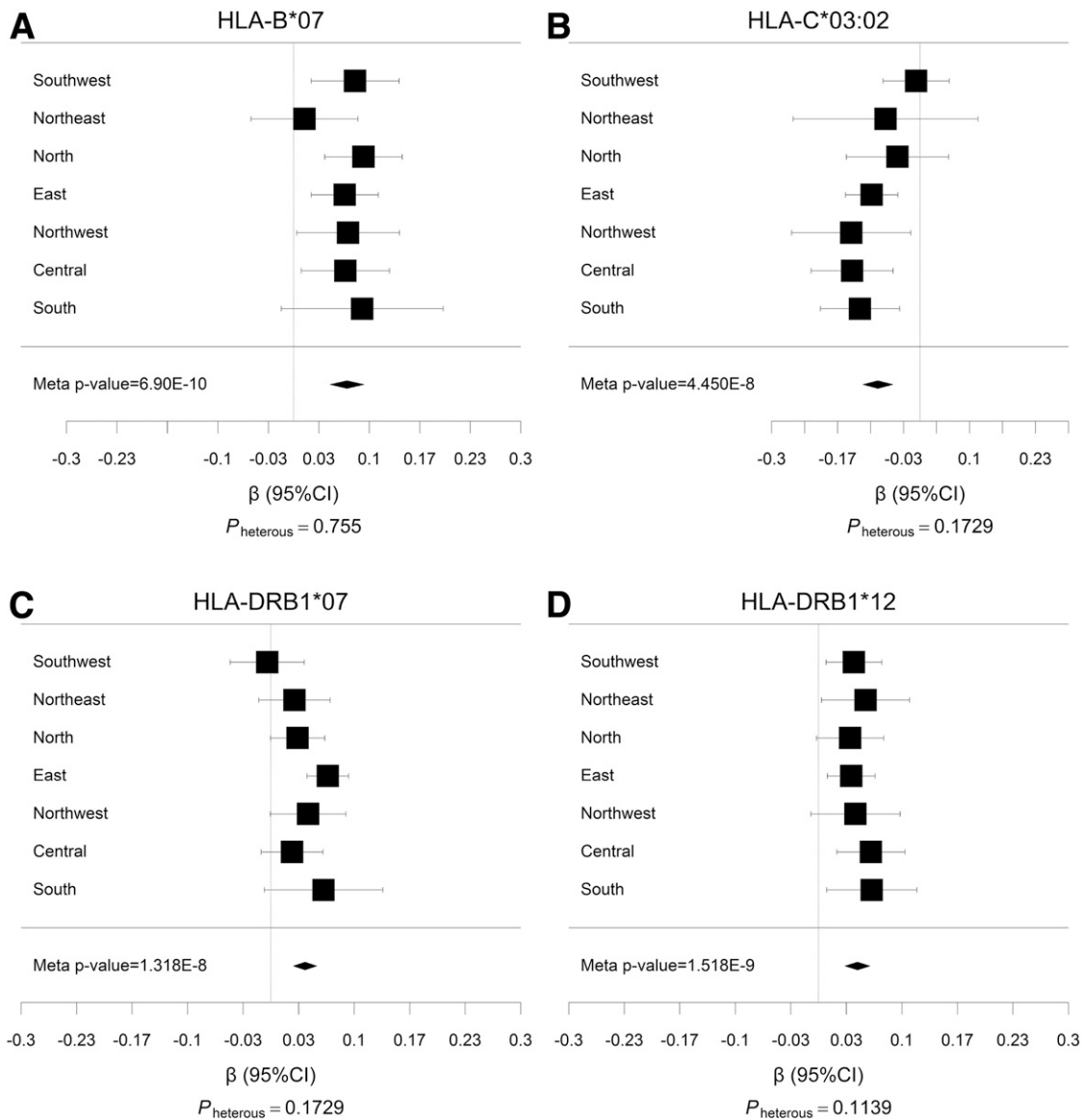


Figure 3—Meta-analysis forest plots for HLA-B*07 (A), HLA-C*03:02 (B), HLA-DRB1*07 (C), and HLA-DRB1*12 (D), HLA alleles with the strongest associations with BMI in Chinese adults. Plots show the study-specific association estimates (β) and 95% CI for the seven geographical regions presented as bars. The scale is the β value. The association estimate and CI for the meta-analysis combining the seven geographical regions is shown as a diamond. $P_{heteros}$ is the P value for heterogeneity among seven regions.

C*03:02, which had strong evidence for association with BMI as a quantitative trait, also associated with overweight ($P = 7.80 \times 10^{-5}$, 4.70×10^{-12} , and 2.08×10^{-5} , respectively) and obesity ($P = 4.79 \times 10^{-4}$, 0.03, and 4.05×10^{-3} , respectively) (Supplementary Table 2).

Sex-Stratified Analyses

We performed sex-specific meta-analyses to test for an association of each allele with BMI in men and women separately, which identified nominal evidence for heterogeneity ($P < 0.05$) for the DRB1*07 and B*07 alleles. The sex-specific effects of DRB1*07 ($P_{heterogeneity} = 0.00058$) met Bonferroni significance (significance requires $P_{heterogeneity} \leq 0.002$; 0.05/23 alleles tested), where the association was

much stronger in men ($\beta = 0.066$, $P = 1.54 \times 10^{-10}$) than in women ($\beta = 0.018$, $P = 0.056$) (Fig. 4). The association between B*07 and BMI was also stronger in men ($\beta = 0.099$, $P = 1.50 \times 10^{-9}$) than in women ($\beta = 0.039$, $P = 0.009$), although the evidence for heterogeneity for this allele was statistically nominal ($P_{heterogeneity} = 0.0067$).

Comparison of HLA Allele Frequencies in Chinese and Europeans

Among the 19 unique HLA alleles that associated with BMI in our Chinese cohort, 18 had significantly different allele frequencies between Chinese and European populations (χ^2 or Fisher exact test, $P < 0.05$). Of the eight HLA alleles with higher frequency in Europeans, five associated with a higher

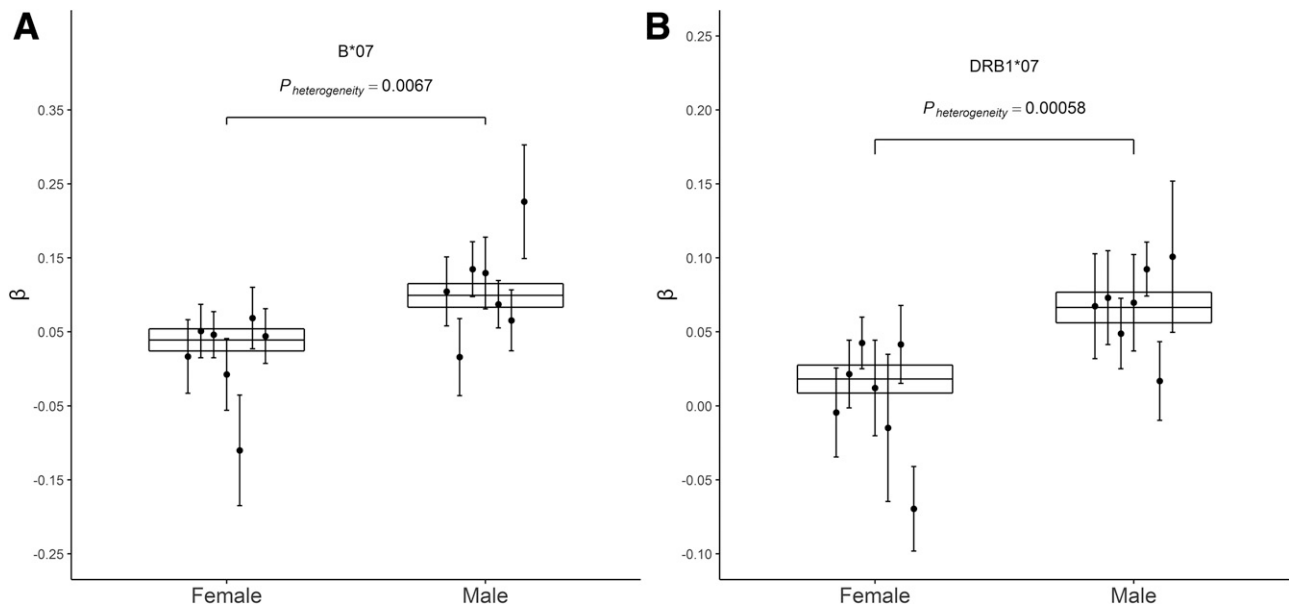


Figure 4—HLA alleles that associate with BMI in a sex-dependent manner. Each of the seven geographic study-specific association estimates (β ; represented by a circle) and SE (represented by a vertical line) are shown in males and females separately for B*07 (A) and DRB1*07 (B). The meta-analysis mean and SE (represented by the horizontal lines) for all seven studies was calculated using the GWAMA software. $P_{\text{heterogeneity}}$ is the P value for heterogeneity between male and female.

BMI (A*02:01, B*08, B*07:02, DRB1*07:01, and DQB1*06:02) and three associated with lower BMI (DRB1*04, DRB1*03:01, and DQB1*03:02) in our study of the Chinese cohort. Of the 10 HLA alleles with higher frequency in Chinese, 4 associated with a high BMI (B*13, C*01:03, C*07, and DRB1*12:02) and 6 associated with a lower BMI (B*27:04, B*58, B*46, B*40:01, C*03:02, and DRB1*09:01). Tests of the difference in the proportions of alleles associated with high BMI between populations that have significantly higher allele frequencies in Caucasians ($n = 8$) and those in Chinese ($n = 10$) show no statistical significance ($P > 0.05$). Interestingly, six (A*02:01, DQB1*06:02, DRB1*07:01, B*07:02, B*08, and C*07) of nine alleles associated with a higher BMI have an allele frequency >0.10 in Caucasians, whereas six (B*27:04, C*03:02, B*58, B*46, DRB1*09:01, and B*40:01) of nine alleles associated with lower BMI have an allele frequency <0.10 in Caucasians (Fig. 5B).

Assignment of BMI-Associated Alleles to Common Haplotypes

The extremely high LD within the HLA region can confound genetic association studies. Therefore, haplotypes were constructed based on 169,995 volunteers, which represents a subset of the samples in this study (27). The BMI-associated alleles were mapped onto 105 five-locus haplotypes with allele frequencies >0.001 . For the BMI-associated low-resolution alleles, we used r^2 to identify high-resolution alleles (B*46:01, B*58:01, and DRB1*04:03), which could be used to represent them.

Among alleles associated with high BMI, B*13:02 and DRB1*07:01 are strongly correlated ($r^2 = 0.31$) and were observed in multiple common haplotypes “CH” that contain

C*06:02-B*13:02-DRB1*07:01-DQB1*02:02 (Fig. 5C). These haplotypes are common in both Chinese and European populations (32). The A*02:01 allele was observed on haplotype “CH” (A*02:01-C*06:02-B*13:02-DRB1*07:01-DQB1*02:02) and also on other haplotypes containing C*07:02, DRB1*12:02, or DQB1*06:02. The three alleles C*07:02, B*07:02, and DQB1*06:02 were frequently carried on the same haplotypes. Most often, these are preceded by A*03:01 or A*24:02 (Fig. 5C).

Among alleles associated with lower BMI, B*46:01 correlated with DRB1*09:01 ($r^2 = 0.028$) and B*58:01 correlated with DRB1*03:01 ($r^2 = 0.051$). These alleles have been observed on two common four-locus haplotypes (“AH”), C*03:02-B*58:01-DRB1*03:01-DQB1*02:01 and C*01:02-B*46:01-DRB1*09:01-DQB1*03:03, which have higher frequencies in Asians compared with Africans or Caucasians (Fig. 5C) (32).

DISCUSSION

In this study, we examined 72 low-resolution HLA alleles and 163 high-resolution HLA alleles in 1.3 million individuals who had been HLA typed as part of the CMDP repository. We identified 19 unique HLA alleles that associated with BMI after correction for multiple testing. However, the effect size of these alleles (β is defined as change in kg/m^2 per allele) ranged from -0.024 to -0.11 for alleles associated with low BMI and from 0.01 to 0.12 for alleles associated with high BMI. These effects are smaller than those reported in prior GWAS of Caucasians (4) (β s ranged from 0.06 to 0.39 kg/m^2 per allele); therefore, the variance explained by a single HLA

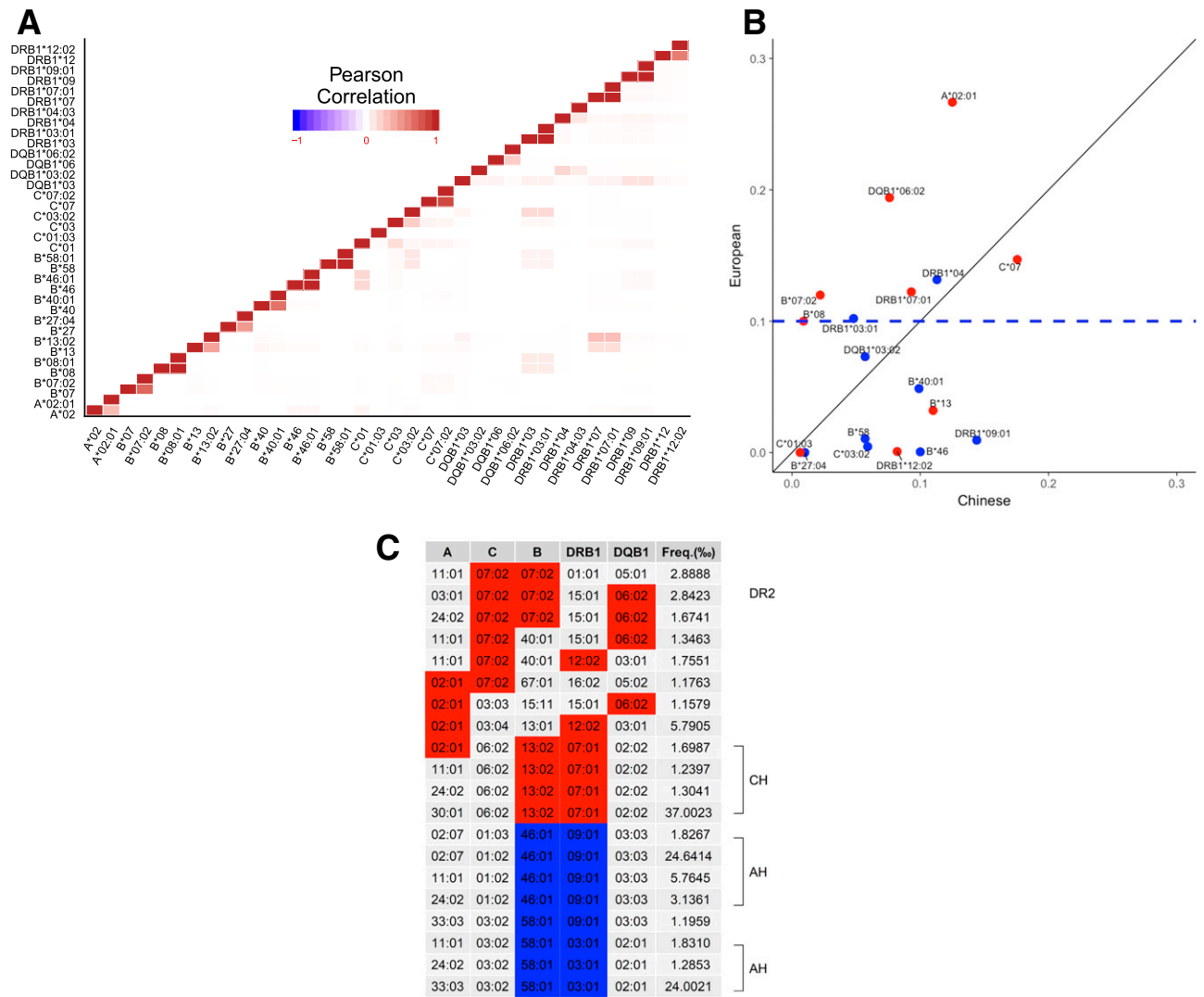


Figure 5—BMI-associated HLA alleles and haplotype. **A:** Pairwise correlation matrix heat map of BMI-associated HLA alleles. The heat map grid represents the square of the Pearson correlation coefficient (r^2). Three high-resolution alleles (DRB1*03:01, DRB1*07:01, and DRB1*09:01) are nearly perfectly correlated ($r^2 > 0.99$) with their respective low-resolution alleles (DRB1*03, DRB1*07, and DRB1*09). In addition, B*07:02 is strongly correlated with B*07 ($r^2 = 0.73$), and A*02:01 is strongly correlated with A*02 ($r^2 = 0.32$). Among alleles associated with higher BMI, DRB1*07:01 and B*13:02 are strongly correlated ($r^2 = 0.31$). Among alleles associated with lower BMI, B*46:01 is correlated with DRB1*09:01 ($r^2 = 0.028$), and B*58:01 is correlated with DRB1*03:01 ($r^2 = 0.051$). **B:** Allele frequency between European and Chinese. The allele frequencies in Europeans were derived from the Allele Frequency Net Database (<http://www.allelefrequencies.net>), and the frequencies in Chinese are from the current study. Eighteen had significantly different allele frequencies between Chinese and European populations (χ^2 or Fisher exact test $P < 0.05$). HLA alleles highlighted in red are the alleles associated with higher BMI, and the HLA alleles highlighted in blue are the alleles associated with lower BMI. The alleles around the diagonal line $y = x$ are similar in frequency between European and Chinese. The blue horizontal dashed line represents the allele frequency of 0.1 in Europeans. Six (A*02:01, DQB1*06:02, DRB1*07:01, B*07:02, B*08, and C*07) of nine alleles associated with a higher BMI have an allele frequency >0.10 in Caucasians, whereas six (B*27:04, C*03:02, B*58, B*46, DRB1*09:01, and B*40:01) of nine alleles associated with lower BMI have an allele frequency <0.10 in Caucasians. There is no significant difference in the proportions of alleles associated with high BMI between alleles that have significantly higher allele frequencies in Caucasian and those in Chinese (Fisher exact test $P > 0.05$). **C:** Assignment of BMI-associated HLA alleles into five locus haplotypes. Blue box: alleles associated with lower BMI; red box: alleles associated with high BMI. The first row contains the gene name, A = HLA-A, C = HLA-C, B = HLA-B, DRB1 = HLA-DRB1, DQB1 = HLA-DQB1. Each row contains one common haplotype with at least two alleles associated with lower or higher BMI. The last column (Freq. [%]) is the haplotype frequency reported in the Chinese population. Most of the 19 BMI-associated alleles mapped onto one of these common haplotypes and several alleles mapped to multiple haplotypes. Three alleles associated with high BMI (A*02:01, B*13:02, and DRB1*07:01) are in a four-locus haplotype (CH: C*06:02-B*13:02-DRB1*07:01-DQB1*02:02), two alleles associated with high BMI (B*07:02 and DQB1*06:02) are in the five-locus haplotype (DR2: A*03:01-C*07:02-B*07:02-DRB1*15:01-DQB1*06:02). Both of these haplotypes are common in Chinese and European populations. The latter is also called ancestral DR2 haplotype. A four-locus haplotype (AH) C*03:02-B*58:01-DRB1*03:01-DQB1*02:01 contains two alleles associated with lower BMI and has a higher frequency in Asians than Africans or Caucasians. Most of the haplotypes contained more than one risk or protective allele, suggesting high LD among these alleles.

allele in our study of Chinese is very small (ranging from 0.009 to 0.21%).

The HLA alleles with the strongest evidence for association with BMI were B*07, DRB1*07, DRB1*12, and C*03:02, where B*07, DRB1*07, and DRB1*12 were associated with higher BMI, and C*03:02 was associated with lower BMI. Several of these alleles have been previously implicated in response to viral infection. For example, HLA-DRB1*07 has been associated with viral persistence in both hepatitis C and B virus infection (33–35) and hepatitis B vaccine failure (36–39). Chronic infection with hepatitis C or B virus is the most common risk factor for hepatocellular carcinoma. HLA-B*07 has been associated with increased risk of cervical cancer (40–42), which is often caused by human papillomavirus. Two of the HLA alleles, B*07 and DRB1*07, had evidence for a sex-specific effect on BMI in our study. Sex specificity in response to viral infection has been reported, where males mount more vigorous immune and behavioral responses to influenza viral infection than females (43).

Prior studies in mice and humans have also suggested an impact of classic HLA alleles on the gut microbiota composition (44–46), and the role of the gut microbiota on obesity is an area of current investigation. Therefore, further studies are needed to investigate the role of HLA-B*07, -DRB1*07, and -C*03:02 on gut microbiota in Chinese populations. Infectious pathogens are arguably among the strongest selective forces that act on human populations (47).

During migrations of modern humans, immune response genes, in particular the HLA system, have been under strong selective pressure to respond and adapt to pathogens. Many pathogens have been reported to cause or exacerbate obesity in animals by damaging the central nervous system (48). In our haplotype analysis, we found that three risk alleles are carried in the haplotype A*02:01-C*06:02-B*13:02-DRB1*07:01-DQB1*02:02, and three risk alleles are carried in the haplotype of A*03:01-C*07:02-B*07:02-DRB1*15:01-DQB1*06:02. These haplotypes are both common in Chinese and European populations. The later haplotype is the “ancestral DR2” haplotype, which is considered to be the longest of the widely distributed ancestral haplotypes (49). These common haplotypes, which could increase survival by providing immune and/or stored energy advantage, are thought to have occurred around the time of the “out of Africa” and expansion into Europe migration (50,51). Four HLA alleles, which we found were protective of obesity, are carried on the most common haplotypes in Asians. These haplotypes are much less common in Africans and Caucasians (52) and therefore are likely to have occurred after the “out of Africa” migration, allowing Asians to gain resistance for “new” pathogens in Asia (53,54).

Future studies are needed to investigate the role of these BMI-associated HLA alleles on T2D. The prevalence of T2D in China has escalated during the past two decades. In 2010, the prevalence of T2D was comparable in China and the U.S. (both ~11.3%), yet among the Chinese population, T2D occurred at a significantly lower BMI (mean BMI 23 kg/m²) compared with the U.S. population (mean BMI 28.7 kg/m²)

(55). A prior study of individuals living in southern China, a region that is experiencing the world’s fastest economic development, reported that increases in BMI and overall obesity have been leveling off, while increases in waist circumference and abdominal obesity have continued to rise (56). Waist circumference has been proven to be a better predictor of T2D than BMI among the Chinese adult population (57). Therefore, the increased prevalence of T2D in China may possibly be due to increases in abdominal obesity, even though HLA alleles predictive of lower BMI are more common in this population. In addition, the small effect sizes of HLA alleles on BMI suggest that the effect of HLA-protective alleles on T2D is minimal. Thus, environmental factors, gene-gene, and gene-environmental interaction likely also play important roles.

Our study has three major strengths. First, to our knowledge, this study is the largest genetic association study for BMI in the world, affording >80% power to detect variants with an allele frequency of 0.001. Second, HLA genotypes were not imputed (58,59), but rather directly typed. Third, phenotypic data were all derived from a single source.

A weakness of our study is that it only identifies associations, and the strong LD across the HLA region makes it difficult to define the exact location of etiological variants. For example, *TNFA*, which encodes tumor necrosis factor- α (TNF- α) is among the 200 genes in the HLA locus. TNF- α is a key regulator of the inflammatory response and can cause necrosis of tumors (60). Dysregulation of TNF- α production has been implicated in a variety of human diseases such as obesity, T2D, autoimmune diseases, and cancers (61–63). Therefore, variation in the *TNFA* gene or other genes in this locus could possibly account for some of the association between HLA and obesity observed in our study.

In summary, the results of this study suggest that specific HLA alleles are associated with higher or lower BMI in human populations and that these alleles may cause alterations in obesity risk through their roles in immune responses.

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