

Cochrane Database of Systematic Reviews

Molecular assays for the diagnosis of sepsis in neonates (Review)

Pammi M, Flores A, Versalovic J, Leeflang MMG

Pammi M, Flores A, Versalovic J, Leeflang MMG. Molecular assays for the diagnosis of sepsis in neonates. *Cochrane Database of Systematic Reviews* 2017, Issue 2. Art. No.: CD011926. DOI: 10.1002/14651858.CD011926.pub2.

www.cochranelibrary.com



TABLE OF CONTENTS

PLAIN LANGUAGE SUMMARY 2 SUMMARY OF FINDINGS 3 BACKGROUND 5 OBJECTIVES 6 METHODS 6 RESULTS 8 Figure 1. 9 Figure 2. 10 Figure 3. 10 Figure 4. 11 Figure 5. 13 Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 76 MHAT'S NEW 99 DCLARATIONS OF INTEREST 99 DELARATIONS OF INTEREST 99 DELERENCES DET SUPPORT 99 DIFFERENCES DET SUPPORT 99	ABSTRACT	1
BACKGROUND 5 OBJECTIVES 6 METHODS 6 RESULTS 8 Figure 1. 9 Figure 2. 10 Figure 3. 10 Figure 4. 11 Figure 5. 13 Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 20 AUTHORS' CONCLUSIONS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 75 Test 3. Molecular tests. 76 MHAT'S NEW 99 CONTRIBUTIONS OF AUTHORS 99 DECLARATIONS OF INTEREST 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW 100	PLAIN LANGUAGE SUMMARY	2
OBJECTIVES 6 METHODS 6 RESULTS 8 Figure 1. 9 Figure 2. 10 Figure 3. 10 Figure 4. 11 Figure 5. 13 Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 22 ACKNOWLEDGEMENTS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARCTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 76 Test 3. Molecular tests. 76 MCHARTERS 76 WHAT'S NEW 99 CONTRIBUTIONS OF AUTHORS 99 DECLARATIONS OF INTEREST 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW<	SUMMARY OF FINDINGS	3
METHODS 6 RESULTS 8 Figure 1. 9 Figure 2. 10 Figure 3. 10 Figure 4. 11 Figure 6. 13 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Strigure 13. 22 AUTHORS' CONCLUSIONS 22 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 76 APPENDICES 76 ONTA 76 OPENDICES 76 PRENDICES 76 PRENDICES 76 PORCLARATIONS OF AUTHORS 99 DECLARATIONS OF INTEREST 99 DUFFERENCES BETWEEN PROTOCOL AND REVIEW 99	BACKGROUND	5
RESULTS 8 Figure 1. 9 Figure 2. 10 Figure 3. 10 Figure 4. 11 Figure 5. 13 Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 22 ACKNOWLEDGEMENTS 23 ACKNOWLEDGEMENTS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 75 Test 3. Molecular tests with good methodologic quality. 76 APPENDICES 76 WHAT'S NEW 99 COLCLARTIONS OF AUTHORS 99 DECLARATIONS OF INTEREST 99 DUFFERENCES BETWEEN PROTOCOL AND REVIEW 100 </td <td>OBJECTIVES</td> <td>6</td>	OBJECTIVES	6
Figure 1. 9 Figure 2. 10 Figure 3. 10 Figure 4. 11 Figure 5. 13 Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 76 MHAT'S NEW 99 CONTIBUTIONS OF AUTHORS 99 DECLARATIONS OF AUTHORS 99 DECLARATIONS OF INTEREST 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW 100	METHODS	6
Figure 2. 10 Figure 3. 10 Figure 3. 10 Figure 4. 11 Figure 5. 13 Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 22 AUTHORS' CONCLUSIONS 24 ACKNOWLEDGEMENTS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 75 Test 3. Molecular tests with good methodologic quality. 76 APPENDICES 76 WHAT'S NEW 99 SOURCES OF SUPPORT 99 DECLARATIONS OF AUTHORS 99 DECLARATIONS OF NUTREEST 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW <td>RESULTS</td> <td>8</td>	RESULTS	8
Figure 3. 10 Figure 4. 11 Figure 5. 13 Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 23 ACKNOWLEDGEMENTS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 76 APPENDICES 76 MPHENDICES 76 MORT'S NEW 99 COLLARATIONS OF AUTHORS 99 DECLARATIONS OF AUTHORS 99 DECLARATIONS OF INTEREST 99 DIFFERENCES OF SUPPORT 99 DIFFERENCES OF SUPPORT 99 DIFFERENCES OF SUPPORT 99 DIFFERENCES OF SUPPORT 99	Figure 1	9
Figure 4. 11 Figure 5. 13 Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 22 AUTHORS' CONCLUSIONS 23 ACKNOWLEDGEMENTS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 24 Test 1. All molecular tests. 74 Test 2. Molecular tests. 74 Test 2. Molecular tests. 75 Test 3. Molecular tests with good methodologic quality. 76 APPENDICES 76 WHAT'S NEW 99 ONTRIBUTIONS OF AUTHORS 99 DECLARATIONS OF INTEREST 99 SOURCES OF SUPPORT 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW 100	Figure 2	10
Figure 5. 13 Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 75 Test 3. Molecular tests with good methodologic quality. 76 APPENDICES 76 WHAT'S NEW 99 OONTRIBUTIONS OF INTEREST 99 DECLARATIONS OF INTEREST 99 DUFFERENCES BETWEEN PROTOCOL AND REVIEW 100	Figure 3	10
Figure 6. 14 Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests with good methodologic quality. 76 APPENDICES 76 WHAT'S NEW 99 DCONTRIBUTIONS OF AUTHORS 99 SOURCES OF SUPPORT 99 DICLARATIONS OF INTEREST 99 DICLARATIONS OF INTEREST 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW 100	Figure 4	11
Further Figure 7. 15 Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 22 AUTHORS' CONCLUSIONS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests biod samples only. 75 Test 3. Molecular tests with good methodologic quality. 76 APPENDICES 79 OUNTRIBUTIONS OF AUTHORS 99 DOLTRES OF SUPPORT 99 DICLARATIONS OF INTEREST 99 DIFERENCES BETWEEN PROTOCOL AND REVIEW 100	Figure 5	13
Figure 8. 16 Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 20 AUTHORS' CONCLUSIONS 22 ACKNOWLEDGEMENTS 22 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests: 74 Test 2. Molecular tests: 75 Test 3. Molecular tests: 76 APPENDICES 76 MHAT'S NEW 99 DONTRIBUTIONS OF AUTHORS 99 DECLARATIONS OF INTEREST 99 SOURCES OF SUPPORT 99 DIFERENCES BETWEEN PROTOCOL AND REVIEW 100	Figure 6	14
Figure 9. 17 Figure 10. 18 Figure 11. 19 Figure 12. 20 DISCUSSION 20 Figure 13. 22 AUTHORS' CONCLUSIONS 23 ACKNOWLEDGEMENTS 24 REFERENCES 25 CHARACTERISTICS OF STUDIES 29 DATA 73 Test 1. All molecular tests. 74 Test 2. Molecular tests. 76 APPENDICES 76 WHAT'S NEW 99 CONTRIBUTIONS OF AUTHORS 99 DOUTRIBUTIONS OF AUTHORS 99 SOURCES OF SUPPORT 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW 100	Figure 7	15
Figure 10.18Figure 11.19Figure 12.20DISCUSSION20Figure 13.22AUTHORS' CONCLUSIONS23ACKNOWLEDGEMENTS24REFERENCES25CHARACTERISTICS OF STUDIES29DATA73Test 1. All molecular tests.74Test 2. Molecular tests.75Test 3. Molecular tests.76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	Figure 8	16
G19Figure 11.19Figure 12.20DISCUSSION20Figure 13.22AUTHORS' CONCLUSIONS23ACKNOWLEDGEMENTS24REFERENCES25CHARACTERISTICS OF STUDIES29DATA73Test 1. All molecular tests.74Test 2. Molecular tests iblood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	Figure 9.	17
Figure 12.20DISCUSSION20Figure 13.22AUTHORS' CONCLUSIONS23ACKNOWLEDGEMENTS24REFERENCES25CHARACTERISTICS OF STUDIES29DATA73Test 1. All molecular tests.74Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	Figure 10	18
DISCUSSION20Figure 13.22AUTHORS' CONCLUSIONS23ACKNOWLEDGEMENTS24REFERENCES25CHARACTERISTICS OF STUDIES29DATA73Test 1. All molecular tests.74Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	Figure 11	19
Figure 13.22AUTHORS' CONCLUSIONS23ACKNOWLEDGEMENTS24REFERENCES25CHARACTERISTICS OF STUDIES29DATA73Test 1. All molecular tests.74Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	Figure 12	20
AUTHORS' CONCLUSIONS23ACKNOWLEDGEMENTS24REFERENCES25CHARACTERISTICS OF STUDIES29DATA73Test 1. All molecular tests.74Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	DISCUSSION	20
ACKNOWLEDGEMENTS24REFERENCES25CHARACTERISTICS OF STUDIES29DATA73Test 1. All molecular tests.74Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	Figure 13	22
REFERENCES25CHARACTERISTICS OF STUDIES29DATA73Test 1. All molecular tests.74Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	AUTHORS' CONCLUSIONS	23
CHARACTERISTICS OF STUDIES29DATA73Test 1. All molecular tests.74Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	ACKNOWLEDGEMENTS	24
DATA73Test 1. All molecular tests.74Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	REFERENCES	25
Test 1. All molecular tests.74Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	CHARACTERISTICS OF STUDIES	29
Test 2. Molecular tests: blood samples only.75Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	DATA	73
Test 3. Molecular tests with good methodologic quality.76APPENDICES76WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	Test 1. All molecular tests.	74
APPENDICES 76 WHAT'S NEW 99 CONTRIBUTIONS OF AUTHORS 99 DECLARATIONS OF INTEREST 99 SOURCES OF SUPPORT 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW 100	Test 2. Molecular tests: blood samples only.	75
WHAT'S NEW99CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	Test 3. Molecular tests with good methodologic quality.	76
CONTRIBUTIONS OF AUTHORS99DECLARATIONS OF INTEREST99SOURCES OF SUPPORT99DIFFERENCES BETWEEN PROTOCOL AND REVIEW100	APPENDICES	76
DECLARATIONS OF INTEREST 99 SOURCES OF SUPPORT 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW 100	WHAT'S NEW	99
SOURCES OF SUPPORT 99 DIFFERENCES BETWEEN PROTOCOL AND REVIEW 100	CONTRIBUTIONS OF AUTHORS	99
DIFFERENCES BETWEEN PROTOCOL AND REVIEW	DECLARATIONS OF INTEREST	99
	SOURCES OF SUPPORT	99
INDEX TERMS	DIFFERENCES BETWEEN PROTOCOL AND REVIEW	100
	INDEX TERMS	100



[Diagnostic Test Accuracy Review]

Molecular assays for the diagnosis of sepsis in neonates

Mohan Pammi¹, Angela Flores², James Versalovic³, Mariska MG Leeflang⁴

¹Section of Neonatology, Department of Pediatrics, Baylor College of Medicine, Houston, Texas, USA. ²Pediatrix Medical Group - NW Houston Practice, The Woodlands, Texas, USA. ³Pathology, Texas Children's Hospital and Baylor College of Medicine, Houston, Texas, USA. ⁴Department of Clinical Epidemiology, Biostatistics and Bioinformatics, Academic Medical Center, University of Amsterdam, Amsterdam, Netherlands

Contact: Mohan Pammi, Section of Neonatology, Department of Pediatrics, Baylor College of Medicine, 6621, Fannin, MC.WT 6-104, Houston, Texas, 77030, USA. mohanv@bcm.tmc.edu.

Editorial group: Cochrane Neonatal Group. **Publication status and date:** New, published in Issue 2, 2017.

Citation: Pammi M, Flores A, Versalovic J, Leeflang MMG. Molecular assays for the diagnosis of sepsis in neonates. *Cochrane Database of Systematic Reviews* 2017, Issue 2. Art. No.: CD011926. DOI: 10.1002/14651858.CD011926.pub2.

Copyright © 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

ABSTRACT

Background

Microbial cultures for diagnosis of neonatal sepsis have low sensitivity and reporting delay. Advances in molecular microbiology have fostered new molecular assays that are rapid and may improve neonatal outcomes.

Objectives

To assess the diagnostic accuracy of various molecular methods for the diagnosis of culture-positive bacterial and fungal sepsis in neonates and to explore heterogeneity among studies by analyzing subgroups classified by gestational age and type of sepsis onset and compare molecular tests with one another.

Search methods

We performed the systematic review as recommended by the Cochrane Diagnostic Test Accuracy Working Group. On 19 January 2016, we searched electronic bibliographic databases (the Cochrane Library, PubMed (from 1966), Embase (from 1982), and CINAHL (from 1982)), conference proceedings of the Pediatric Academic Societies annual conference (from 1990), clinical trial registries (ClinicalTrials.gov, International Standard Randomised Controlled Trial Number (ISRCTN) registry, and World Health Organization (WHO) International Clinical Trials Platform (ICTRP) Search portal), and Science Citation Index. We contacted experts in the field for studies.

Selection criteria

We included studies that were prospective or retrospective, cohort or cross-sectional design, which evaluated molecular assays (index test) in neonates with suspected sepsis (participants) in comparison with microbial cultures (reference standard).

Data collection and analysis

Two review authors independently assessed the methodologic quality of the studies and extracted data. We performed meta-analyses using the bivariate and hierarchical summary receiver operating characteristic (HSROC) models and entered data into Review Manager 5.

Main results

Thirty-five studies were eligible for inclusion and the summary estimate of sensitivity was 0.90 (95% confidence interval (CI) 0.82 to 0.95) and of specificity was 0.93 (95% CI 0.89 to 0.96) (moderate quality evidence). We explored heterogeneity by subgroup analyses of type of test, gestational age, type of sepsis onset, and prevalence of sepsis and we did not find sufficient explanations for the heterogeneity (moderate to very low quality evidence). Sensitivity analyses by including studies that analyzed blood samples and by good methodology revealed similar results (moderate quality evidence).



Authors' conclusions

Molecular assays have the advantage of producing rapid results and may perform well as 'add-on' tests.

PLAIN LANGUAGE SUMMARY

Molecular tests to detect infections in newborn babies

Review question: Do molecular tests detect infection better than the standard culture methods for detecting infection in newborn babies?

Background

The current method of detecting infection (illness caused by germs) in newborn babies is to obtain blood or other body fluids (or both) and culture (grow) the bacteria (germs) in a laboratory. However, culture methods may miss some infections and take a long time to produce results (48 to 72 hours). Newer methods of detecting infection are based on detecting DNA (a molecule that carries the genetic instructions used in growth, development, functioning, and reproduction) from bacteria and other organisms that cause infections. Advances in microbiology have introduced new molecular tests for detecting infections. Molecular tests are rapid and may detect more infections compared to the traditional culture methods.

Study characteristics

We searched for evidence for the use of the molecular methods to detect infection in newborn babies. We found 35 studies that compared the new molecular methods to culture methods of the blood and spinal fluid to diagnose infection.

Study funding sources

None.

Key results

We found that the molecular methods may be very helpful additional tests because they provide rapid results.

Quality of evidence

Although there were some issues with selection of newborn babies for this review, overall the methods used by the studies were adequate. We rated the quality of the evidence as moderate to low.

SUMMARY OF FINDINGS

Summary of findings 1. Summary of findings table

	Groups	Number of studies	Sensitivity	Specificity	Quality of evidence using GRADE
			(95% CI)	(95% CI)	
All studies	-	35	0.90 (0.82 to 0.95)	0.93 (0.89 to 0.96)	Moderate quality evidence*
Type of test	Broad-range PCR	9	0.97 (0.86 to 1.00)	0.93 (0.77 to 0.98)	Moderate quality evidence*
	Real-time PCR	9	0.86 (0.59 to 0.96)	0.94 (0.90 to 0.97)	Moderate quality evidence*
	Post-PCR processing	5	0.97 (0.40 to 1.00)	0.96 (0.93 to 0.98)	Low quality evidence**
	Multiplex PCR	6	0.76 (0.60 to 0.88)	0.81 (0.70 to 0.89)	Low quality evidence**
	Staphylococcal PCR*	2	-	-	Low quality evidence**
	Fungal PCR*	4	-	-	Low quality evidence**
Type of sepsis	EOS*	2	-	-	Low quality evidence**
	LOS	10	0.79 (0.69 to 0.86)	0.94 (0.85 to 0.98)	Low quality evidence**
	Mixed EOS and LOS	23	0.94 (0.84 to 0.98)	0.92 (0.87 to 0.95)	Moderate quality evidence*
Gestational age	Preterm	5	0.89 (0.75 to 0.96)	0.87 (0.71 to 0.94)	Low quality evidence**
	Mixed term and preterm	30	0.90 (0.80 to 0.96)	0.94 (0.90 to 0.96)	Moderate quality evidence*
Prevalence	< 15%	20	0.94 (0.80 to 0.99)	0.95 (0.92 to 0.97)	Moderate quality evidence*
	15% to 30%	8	0.85 (0.67 to 0.94)	0.88 (0.79 to 0.94)	Low quality evidence**
	>30%	7	0.87 (0.75 to 0.93)	0.93 (0.64 to 0.99)	Low quality evidence**
Specimen	Blood only	32	0.92 (0.84 to 0.96)	0.93 (0.89 to 0.95)	Low quality evidence**
	Blood and CSF*	3	-	-	Moderate quality evidence*

Cochrane Library

Trusted evidence. Informed decisions. Better health.

Quality	Good methodologic studies only	22	0.90 (0.78 to 0.96)	0.93 (0.88 to 0.96)	Moderate quality evidence*	
Summary estimat for the subgroups GRADE rating of e * Evidence downg	tes of sensitivity and specific are presented, where numb	tity were derived ber of studies ≥ 4 rading quality of stency of evidence	I. *Summary estimates of sensitivit Fevidence (Gopalakrishna 2014) Se.	riate random-effects model u	ion. sing statistical software STATA. Summary estimates e calculated using STATA if number of studies ≤ 4.	EIDIGI Y Better health.
						Cochrane Database of Systematic R

4



BACKGROUND

Sepsis is a frequent life-threatening event among neonates, particularly in very low birth weight infants (VLBW) (birth weight less than 1500 g) and is responsible for significant mortality and morbidity (Adams-Chapman 2006; Stoll 2002; Stoll 2004). Early diagnosis of infections in newborns may improve clinical outcomes. Microbial cultures of blood or other sterile body fluids are the gold standard in the diagnosis of neonatal bacterial and fungal sepsis. Blood cultures are generally assumed to have low sensitivity in neonates for the following reasons: low degree of neonatal bacteremia or fungemia, small inoculation volumes in culture bottles, and the use of intrapartum antibiotics (Chiesa 2004; Schelonka 1996). In addition, results of the microbial culture are not available for at least 24 to 72 hours. Diagnostic capabilities of blood culture systems have improved since the early 2000s with the advent of automated continuous blood culture monitoring systems but still, subcultures for specific assays (e.g. biochemical) are ultimately needed for pathogen identification. New molecular methods for detection of infection may provide results earlier and improve neonatal outcomes.

Target condition being diagnosed

Neonatal bacterial and fungal sepsis is the target condition to be diagnosed and often described based on the age of the infant at the onset of infection. Early-onset bacterial or fungal sepsis (sepsis in 72 hours of life or less) occurs in 1.5% to 1.9% of VLBW infants and late-onset bacterial or fungal sepsis (sepsis onset after 72 hours of life) in about 20% of VLBW infants (Stoll 2002). Neonatal mortality in late-onset sepsis (LOS) is approximately 18%, and in Gram-negative infections as high as 36%. The incidence of LOS in neonates less than 33 weeks' postmenstrual age (PMA) in the Canadian neonatal network was 10% but varied from 0.61% to 14% in other studies (Canadian Neonatal Network 2014; Dong 2015). Sepsis increases neonatal morbidities including patent ductus arteriosus, need for intravascular access, need for parenteral nutrition, bronchopulmonary dysplasia, necrotizing enterocolitis and length of hospital stay. In addition, sepsis significantly impairs long-term neurodevelopmental outcomes either by direct infection of the central nervous system or as a result of inflammatory injury (Adams-Chapman 2006). In one large cohort study of more than 6000 extremely low birth weight infants (birth weight 1000 g or less), infected infants had a significantly higher incidence of adverse developmental outcomes at follow-up, including cerebral palsy, lower Bayley's scores of infant development and visual impairment when compared to uninfected infants (Stoll 2004). Clinical signs and symptoms of neonatal sepsis are often nonspecific and early diagnosis and treatment may be critical to improve neonatal outcomes. Overdiagnosis of neonatal sepsis can lead to inappropriate antibiotic use that may foster antibiotic resistance.

Index test(s)

Advances in molecular microbiology have provided new molecular assays for the detection of infection. Molecular assays can be completed in less than 12 hours and may have better sensitivity than microbial cultures. In addition, the significant increase in workload related to bloodstream infections for the clinical microbiologic laboratory could potentially be offset by highthroughput molecular assays coupled with automation (RodriguezCreixems 2008). However, molecular assays do not provide information on antibiotic susceptibility.

Molecular pathogen detection methods are based on hybridization or amplification of pathogen DNA. Hybridization based methods (e.g. fluorescence in situ hybridization) have not yet been evaluated in the diagnosis of neonatal sepsis. However, neonatal studies have been conducted using amplification methods (e.g. polymerase chain reaction (PCR)) that amplify specific target regions in the microbial genome. Broad-range PCR targets the 16S ribosomal ribonucleic acid (rRNA) gene, a ubiquitous gene that is preserved in all bacteria and comprises both conserved and variable regions (Woese 1987). The conserved regions are targeted by universal primers for identifying bacterial infection, and the variable regions by genus or species-specific assays (Isaacman 1996; Relman 1999). Fungal PCRs target specific regions of the fungal genome (most commonly internal transcribed spacer regions of the rRNA). Amplified target regions may then be subjected to downstream applications such as sequencing or microarray/probe hybridization.

Amplification methods that have been evaluated in neonates for the diagnosis of sepsis can be grouped as follows.

- 1. Broad-range conventional PCR assays: PCR amplification strategies targeting conserved regions such as 16S rRNA in bacteria.
- 2. Real-time PCR, where amplification of the template is monitored in real time.
- 3. PCR followed by post-PCR processing, such as sequencing or hybridization.
- 4. Multiplex-PCR, where amplification is directed against multiple organisms in the same assay.
- 5. Species- or genus-specific assays: staphylococcal, fungal PCR assays or other organism-specific assays.

Clinical pathway

Neonates with clinical signs and symptoms of sepsis including lethargy, apnea, hypotension and oliguria are investigated for sepsis with a blood, cerebrospinal fluid (CSF) and urine cultures with or without markers of inflammation such as a white blood cell count, C-reactive protein (CRP) or others. However, to prove an infection beyond doubt, cultures should be positive, which takes more than 24 hours and usually 48 hours. Also, the sensitivity of cultures has been questioned. An ideal diagnostic test for neonatal bacterial or fungal sepsis should be rapid, sensitive, specific, detect all organisms relevant to neonatal sepsis and not be affected by maternal antibiotics. The test should have high sensitivity so that infections are not missed and a negative test should reliably exclude sepsis so that no neonate is unnecessarily treated with antibiotics.

Alternative test(s)

Traditionally sepsis diagnosis is aided by abnormal white blood cell count (white blood cell less than 5000 cells/ μ L, sensitivity 0.2, specificity 0.96; white blood cell less than 1000 cells/ μ L, sensitivity 0.3 specificity 1.0), altered white cell indices, differential white cell count, elevation of immature white cells (I:T ratio greater than 0.20, sensitivity 0.55 and specificity 0.74) and low platelet count (less than 50 × 10⁹/L, sensitivity 0.8 and specificity 0.99) (Hornik 2012). Serum biomarkers of infection consisting of acute-

Cochrane Library

Trusted evidence. Informed decisions. Better health.

phase proteins namely CRP (sensitivity 0.6 to 0.84, specificity 0.84 to 1.00), procalcitonin (sensitivity 0.77, specificity 0.62) or elevation of inflammatory cytokines; tumor necrosis factor (TNF)- α (sensitivity 0.6 to 0.82, specificity 0.86 to 0.93) and interleukin (IL)-6 (sensitivity 0.58 to 0.89, specificity 0.84 to 0.96) have also been used (Blommendahl 2002; Ng 1997; Ng 2012; Verboon-Maciolek 2006). All sensitivities and specificities were calculated with culture as the reference standard. White cell indices and other serum biomarkers may aid in the diagnosis but not necessarily confirm infection.

Rationale

Blood cultures are generally assumed to have a relatively low sensitivity for the diagnosis of neonatal bacterial or fungal sepsis and results of the microbial culture are not available for at least 24 to 72 hours. Also, some cases of sepsis may be missed by cultures and a more sensitive diagnostic test such as a molecular test may be useful. Rapid advances in technology have led to molecular methods with rapid turnaround times, that may be more sensitive than culture and which may have an impact on current clinical practice. We will not be able to show that the molecular tests are more sensitive than culture, as culture is our reference standard. Still, culture is used in practice as a confirmation test (100% specificity) and thus knowing the relative performance of molecular tests compared to culture is very relevant. If a test misses too many culture-positive samples, the test will not be implemented in practice. Alternative tests such as evaluation of acute phase reactants or cytokines are often used in conjunction with blood cultures but do not have sufficient diagnostic accuracy to replace microbial cultures as the reference standard. We have previously systematically reviewed molecular assays in the diagnosis of neonatal sepsis (Pammi 2011), but this is a rapidly advancing field. Optimization of the older molecular methods and development of newer methods may change the diagnostic accuracy of these tests and may change our clinical practice. In our view, a Cochrane Review is justified as new literature has accumulated since our last published review and will allow for updates as new studies are performed. We are not aware of any other systematic review on this topic in neonates although there are narrative reviews.

OBJECTIVES

To assess the diagnostic accuracy of various molecular methods for the diagnosis of culture-positive bacterial and fungal sepsis in neonates and to explore heterogeneity among studies by analyzing subgroups classified by gestational age and type of sepsis onset and compare molecular tests with one another.

METHODS

Criteria for considering studies for this review

Types of studies

We included prospective or retrospective, cohort or cross-sectional studies that assessed the diagnostic accuracy of a molecular test in the clinical context of diagnosis of neonatal bacterial or fungal sepsis. We excluded studies that assessed the diagnostic accuracy of the test using only positive samples or healthy controls and not in the clinical context of suspected neonatal bacterial or fungal sepsis.

Participants

Neonates with clinically suspected bacterial or fungal sepsis. Clinical signs and symptoms of sepsis in neonates can be nonspecific and hence a high index of clinical suspicion is required for the diagnosis. Neonates are defined as a newborn of 28 days of age or less. We defined gestational age subgroups of preterm and term infants as:

- 1. preterm: neonates born at less than 37 completed weeks of gestation;
- 2. term: neonates born at 37 completed weeks of gestation or greater.

We made a post-hoc decision to include data from studies that included infants aged more than 28 days if more than 50% of the study participants were under 28 days of age.

Index tests

We defined molecular assays as any assay that involves extraction and evaluation of nucleic acid from bacteria or fungi, and performed for the diagnosis of neonatal sepsis. The results of the index test were dichotomous; positive or negative. We assessed the results of the index test with the reference standard done at approximately same time. In the event of the index test identifying a different organism compared to the reference standard or identifying an organism when the reference standard was negative, we discussed among our author group as to whether we should discard or include as a false positive based on whether it was a contaminant or not. We analyzed subgroups of type of molecular assay namely broad-range conventional PCR, real-time PCR, PCR followed by post-PCR processing, multiplex PCR, staphylococcal PCR and fungal PCR. New tests/methodology may arrive in the future as the technology advances and we will address this by subgroup analyses and using year of publication as a covariate in future meta-analyses. We excluded molecular methods assessing infections other than those caused by bacteria or fungi (e.g. viruses or protozoa).

Target conditions

Neonatal bacterial or fungal sepsis, defined as a neonate with a positive culture of bacteria or fungi from the blood or CSF, or both. We analyzed subgroups of type of sepsis onset namely early-onset sepsis (EOS) (72 hours of age or less) and LOS (greater than 72 hours of age).

Reference standards

The reference standard for the diagnosis of sepsis was microbial culture of blood or CSF, or both, for bacteria or fungi, or both. Microbial cultures are generally assumed to have low sensitivity but this decreased sensitivity has not been quantified. The low sensitivity of cultures in neonates may be due to the low degree of neonatal bacteremia or fungemia, small inoculation volumes in culture bottles and the use of intrapartum antibiotics. We documented the participant characteristics, risk factors and outcomes of people who were index test positive and reference standard negative to gain insight into the sensitivity of the reference standard. Alternative tests, such as evaluation of acute phase reactants or cytokines, are often used in conjunction with blood cultures but do not have sufficient diagnostic accuracy to replace microbial cultures as the reference standard.

Molecular assays for the diagnosis of sepsis in neonates (Review) Copyright © 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

Search methods for identification of studies

We used the standard search methods recommended by the Cochrane Neonatal Group and searched the literature on 19 January 2016. We applied no language restrictions in our search methods.

Electronic searches

- 1. Bibliographic databases: the Cochrane Library (2016, Issue 1), PubMed (from 1966), Embase (from 1982) and CINAHL (from 1982) using the search engines at Texas Medical Center library.
- 2. Abstract of conferences: proceedings of meetings of American Pediatric Society, Society for Pediatric Research and European Society for Pediatric Research (from 1990).
- 3. ClinicalTrials.gov (clinicaltrials.gov/), International Standard Randomised Controlled Trial Number (ISRCTN) registry (www.isrctn.com/), and the World Health Organization (WHO) International Clinical Trials Platform (ICTRP) Search portal (apps.who.int/trialsearch/).
- 4. Science Citation Index, Web of Science using subject search.

Our search strategies for PubMed and other databases including the platforms are outlined in Appendix 1. The search strategy was developed by discussion between the review author team, librarians and the Cochrane Neonatal Group's Trials Search Coordinator.

Searching other resources

We screened reference lists of identified studies, relevant review articles and other publications held in our personal files. We also searched for ongoing and unpublished studies by contacting experts in this field.

Data collection and analysis

Selection of studies

Two review authors (MP, AF) screened all titles and abstracts identified by our search strategy for relevance to the inclusion criteria as detailed in Criteria for considering studies for this review. We retrieved full-text articles of all identified articles that were deemed relevant to the review and evaluated them against our inclusion eligibility. We resolved disagreements by mutual discussion.

Data extraction and management

Two review authors (MP, AF) independently extracted the following data.

- 1. Author, year of publication and name of journal.
- 2. Study design including sample size, type of study (prospective or retrospective, cohort or cross-sectional).
- 3. Study population characteristics and the clinical context in which the test was evaluated (e.g. suspected sepsis), and type of participant sample tested.
- 4. Type of reference standard, performance of the reference standard and whether evaluated manually or automated.
- 5. Index tests, performance of the index tests, type of assay, manufacturer, positivity thresholds, time between the performance of index and reference tests.

- 6. Information regarding quality assessment items of the Quality Assessment of Diagnostic Accuracy Studies-2 (QUADAS-2) tool (Assessment of methodological quality).
- 7. Data in two by two tables for calculation of diagnostic accuracy parameters.

Studies report number of neonates or episodes of sepsis as the unit of analysis. Some studies included neonates with more than one episode of sepsis. As the comparison here was between two tests, cultures versus molecular tests, we included the number of samples wherever possible for our analysis and most studies reported only one sample per participant which we analyzed as such. We compared the extracted data, and resolved discrepancies found upon comparison by mutual discussion. Data extracted from included studies are presented in Appendix 2.

Assessment of methodological quality

We assessed methodologic quality of each included study following guidance from the Cochrane Diagnostic Test Accuracy Working Group, which is adapted from the QUADAS-2 tool (Whiting 2011). The four domains assessed for risk of bias are participant selection, index test, reference test, and flow and timing. Applicability concerns were assessed in the first three domains (participant selection, index test, reference test). In each domain, we answered the questions with 'Yes', 'No' or 'Unclear' and for each domain judged the risk of bias as 'Low', 'High' or 'Unclear' risk (Appendix 3).

Sources of bias in diagnostic accuracy studies that we assessed include those related to participants (spectrum bias and selection bias), the index test (information bias), the reference standard (misclassification bias, partial verification bias, differential verification bias, incorporation bias, disease progression bias and information bias) and data analysis (excluded data bias) (Appendix 3).

In addition, we decided post-hoc to present quality of evidence using GRADE methodology recommended for diagnostic tests (Gopalakrishna 2014).

Statistical analysis and data synthesis

In our included studies, the reference standard and the index tests have dichotomous outcomes. We constructed two by two tables for all included studies and enumerated true positives, false positives, false negatives and true negatives. Any positive blood or CSF culture was considered a positive for the reference standard. Nine studies reported data from episodes of sepsis and hence more than one sample from some infants and other studies reported one episode of sepsis from one infant. We have meta-analyzed data from both studies that reported as episodes of sepsis or as number of infants in this review with advice from our statistician.

As the results of the index tests were dichotomous without an explicit threshold, we used a bivariate random-effects approach to estimate summary sensitivity and specificity for each index test type separately (Macaskill 2010; Reitsma 2009). The bivariate random-effects approach enabled us to calculate the summary estimates of sensitivity and specificity, while dealing with the imprecision by which sensitivity and specificity have been measured within each study, variation beyond chance in sensitivity and specificity between studies and any correlation that may exist between sensitivity and specificity. We calculated summary estimates of sensitivity and specificity using 'xtmelogit' in the

Molecular assays for the diagnosis of sepsis in neonates (Review)



STATA software (Stata 2011) (Harbord 2007; Harbord 2008; Harbord 2009).

We generated forest plots with 95% confidence intervals (CIs) for sensitivity and specificity for each study using Review Manager 5 (RevMan 2014). We entered the relevant 'xtmelogit' STATA output in Review Manager 5 (RevMan 2014) for the creation of receiver operating characteristic (ROC) space, including summary estimates with 95% CIs and the summary curve.

Investigations of heterogeneity

Sepsis prevalence is higher in premature infants than in term infants because of their relative immunodeficiency, compromise in mucosal and skin integrity, need for intensive care and exposure to invasive procedures. The diagnostic accuracy parameters are likely to be influenced by prevalence of sepsis in term and preterm infants. Therefore, we investigated the effect of prevalence by including it as a covariate in the bivariate model. The same will be true for the onset of sepsis: prevalence rates and spectrum of organisms are different in late-onset and early-onset disease and may account for variation among studies. Therefore, we also included sepsis onset as a covariate in the models.

We compared the accuracy of different test types by comparing their summary estimates of sensitivity and specificity and the respective CIs. We did not report P values because the results are prone to confounding due to variations in participant characteristics and study methodology.

We used statistical tests using the 'xtmelogit' command in the statistical software STATA (Stata 2011) for evaluation of heterogeneity by subgroup and sensitivity analysis. We reported summary sensitivity and specificity for each subgroup in the subgroup analyses.

Sensitivity analyses

After performing analyses with data of all included studies, we performed sensitivity analysis to assess test accuracy in studies that evaluated blood samples only as well as studies that evaluated both blood and CSF samples, to test if inhibitors of PCRs in blood samples might influence our results. Furthermore, we investigated the effect of the potential sources of bias by removing biased studies from the total set of studies and re-analyzing this new set.

Assessment of reporting bias

We used the Deeks' test to assess publication or reporting bias in this diagnostic test accuracy review (Reitsma 2009; Van Enst 2014).

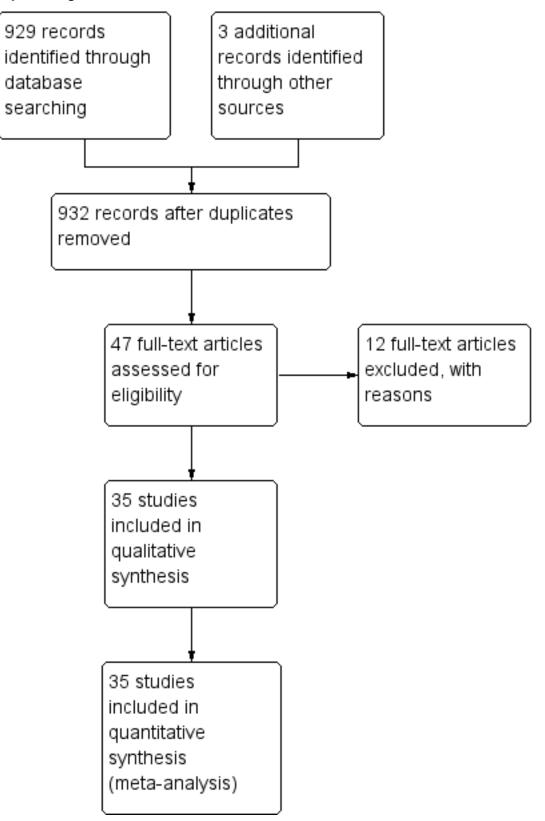
RESULTS

Results of the search

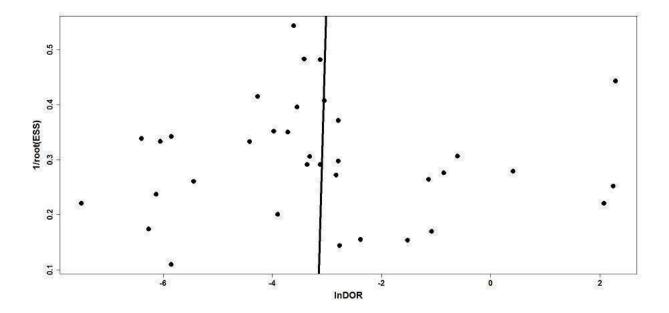
Our comprehensive search identified 932 studies of which we selected 47 relevant articles based on the title and abstract. We obtained the full publications whenever possible for the 47 relevant articles. Twelve articles were irrelevant to this review and discarded. Thirty-five studies met the inclusion criteria assessing the diagnostic accuracy of molecular assays in neonatal sepsis. The inclusion process is detailed in the PRISMA flow diagram (Figure 1). Some studies did not include an upper limit for age and hence some infants were greater than 28 days of age (Chan 2009; Enomoto 2009; Esparcia 2011; Fujimori 2010; Jordan 2000; Lima 2007; Makhoul 2005; Makhoul 2006; Ohlin 2008; Ohlin 2012; Tirodker 2003; Torres-Martos 2013). We made a post-hoc decision that we would include studies where an upper age limit was not specified but more than 50% of the sample were from newborn to less than 28 days of age. Our decision was supported by the reasoning that LOS extends up to three months of age and participant characteristics are similar in the first two to three months of age. The included studies and their risk of bias are presented in Characteristics of included studies table and 12 excluded studies with reasons for exclusion are presented in the Characteristics of excluded studies table. We found no publication bias. Funnel plots were created with In(DOR) on the x-axis and the reciprocal of the effective sample size (ESS) on the y-axis where $1/ESS = (1/(FP + TN) + 1/(TP + FN))^{1/2}$ (Figure 2). Then Deeks' test for publication bias was applied by computing Spearman's rank correlation (r_s) for the association between ln(DOR) and 1/ESS. Asymmetry is not evident in the funnel plot, and Deeks' test did not indicate the presence of publication bias (r_s = 0.012, *p* = 0.944).



Figure 1. Study flow diagram.



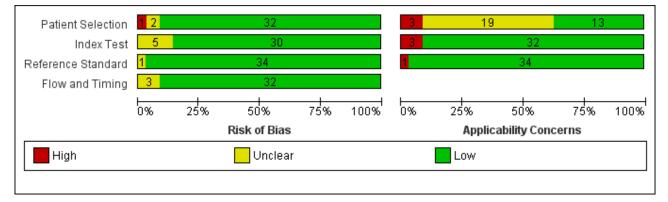




Methodological quality of included studies

The results of the methodologic assessment of the studies included in the meta-analyses are presented in Figure 3; Figure 4. Major risks for bias pertained to participant selection and blinding of index test. Applicability concerns pertained to selection of participants and blinding of the index test and blinding of the reference standard. All studies used an acceptable reference standard, avoided partial and differential verification, and avoided incorporation of the reference standard. Uninterpretable results and withdrawals were explained where applicable.

Figure 3. Risk of bias and applicability concerns graph: review authors' judgments about each domain presented as percentages across included studies.

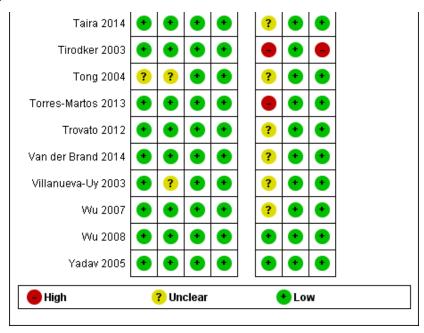




								
	- 1	Risk o	of Bias	6	Applicability Concerns			
	Patient Selection	Index Test	Reference Standard	Flow and Timing	Patient Selection	Index Test	Reference Standard	
Briones 2003	?	?	•	•	?	•	•	
Chan 2009	•	•	•	•	•	•	•	
Chen 2009	•	•	•	•	?	•	•	
Draz 2013	•	•	•	•	?	•	•	
Dutta 2009	•	•	•	•	•	•	•	
Enomoto 2009	•	•	•	•	•	•	•	
Esparcia 2011	•	•	•	•	•	•	•	
Fujimori 2010	•	•	•	•	•	•	•	
Garcia-Elorriaga 2012	•	•	•	•	•	•	•	
Ibarra 2015	•	•	•	•	•	•	•	
Jordan 2000	•	•	•	?	?	•	•	
Jordan 2005a	•	•	•	?	?	•	•	
Jordan 2006	•	•	•	?	•	•	•	
Kasper 2013	•	•	•	•	?	•	•	
Laforgia 1997	•	•	•	•	•	•	•	
Lima 2007	•	?	•	•	?	•	•	
Liu 2014	•	•	•	•	•	•	•	
Makhoul 2005	•	•	•	•	?	•	•	
Makhoul 2006	•	•	•	•	?	•	•	
Ohlin 2008	•	•	•	•	•	•	•	
Ohlin 2012	•	•	•	•	•	•	•	
Paolucci 2009	•	•	•	•	?	•	•	
Reier-Nilsen 2009	•	•	•	•	?	•	•	
Shaat 2013	•	?	?	•	?	•	•	
Shang 2005	•	•	•	•	?	•	•	
Taira 2014	•	•	•	•	?	•	•	

Figure 4. Risk of bias and applicability concerns summary: review authors' judgments about each domain for each included study.

Figure 4. (Continued)



Findings

Summary estimates of mean sensitivity for the 35 included studies were 0.90 (95% CI 0.82 to 0.95), while the mean specificity was 0.93 (95% CI 0.89 to 0.96) (moderate quality evidence) (Summary of findings 1). Forest plot (Figure 5) shows that sensitivity across

studies ranged for 0.38 to 1.0 and specificity from 0.32 to 1.0. We also plotted the included studies in the ROC space to give a sense of distribution of sensitivity and specificity of the studies (Figure 6). Each study is represented by an oval symbol, with the width proportional to the inverse standard error of the specificity and the height to the inverse standard error of sensitivity.

Figure 5. Forest plot of 1 All molecular tests. CI: confidence interval; FN: false negative; FP: false positive; TN: true negative; TP: true positive.

Study	TP	FP			Sensitivity (95% CI)		Sensitivity (95% CI)	Specificity (95% CI)
Draz 2013	20	15	8	7	0.71 [0.51, 0.87]	0.32 [0.14, 0.55]		
Garcia-Elorriaga 2012	9	42	0	47	1.00 [0.66, 1.00]	0.53 [0.42, 0.63]		
Ibarra 2015	9	25	4	48	0.69 [0.39, 0.91]	0.66 [0.54, 0.76]		
Kasper 2013	15	9	0	22	1.00 [0.78, 1.00]	0.71 [0.52, 0.86]		
Fujimori 2010	6	9	0	24	1.00 [0.54, 1.00]	0.73 [0.54, 0.87]		
Torres-Martos 2013	12	6	5	19	0.71 [0.44, 0.90]	0.76 [0.55, 0.91]		
Tirodker 2003	10	13	3	44	0.77 [0.46, 0.95]	0.77 [0.64, 0.87]		
Shaat 2013	17	- 7	0	26	1.00 [0.80, 1.00]	0.79 [0.61, 0.91]		
Taira 2014	3	3	0	18	1.00 [0.29, 1.00]	0.86 [0.64, 0.97]		
Reier-Nilsen 2009	4	6	2	36	0.67 [0.22, 0.96]	0.86 [0.71, 0.95]		
Paolucci 2009	3	4	1	26	0.75 [0.19, 0.99]	0.87 [0.69, 0.96]		
Lima 2007	3	10	5	75	0.38 [0.09, 0.76]	0.88 [0.79, 0.94]		
Trovato 2012	7	8	1	70	0.88 [0.47, 1.00]	0.90 [0.81, 0.95]		-
Ohlin 2012	44	31	12	281	0.79 [0.66, 0.88]	0.90 [0.86, 0.93]		•
Enomoto 2009	3	6	3	65	0.50 [0.12, 0.88]	0.92 [0.83, 0.97]		
Laforgia 1997	4	2	0	27	1.00 [0.40, 1.00]	0.93 [0.77, 0.99]		
Chen 2009	15	10	0	170	1.00 [0.78, 1.00]	0.94 [0.90, 0.97]		-
Shang 2005	8	9	0	155	1.00 [0.63, 1.00]	0.95 [0.90, 0.97]		-
Makhoul 2006	8	- 7	6	125	0.57 [0.29, 0.82]	0.95 [0.89, 0.98]		-
Briones 2003	20	2	1	38	0.95 [0.76, 1.00]	0.95 [0.83, 0.99]		
Ohlin 2008	21	12	29	233	0.42 [0.28, 0.57]	0.95 [0.92, 0.97]		
Liu 2014	95	28	0	583	1.00 [0.96, 1.00]	0.95 [0.93, 0.97]	•	
Yadav 2005	9	4	0	87	1.00 [0.66, 1.00]	0.96 [0.89, 0.99]		-
Esparcia 2011	3	3	4	73	0.43 [0.10, 0.82]	0.96 [0.89, 0.99]		-
Dutta 2009	50	- 7	2	183	0.96 [0.87, 1.00]	0.96 [0.93, 0.99]		•
Tong 2004	8	9	0	268	1.00 [0.63, 1.00]	0.97 [0.94, 0.99]		•
Chan 2009	33	5	9	171	0.79 [0.63, 0.90]	0.97 [0.93, 0.99]		•
Wu 2007	20	23	0	787	1.00 [0.83, 1.00]	0.97 [0.96, 0.98]		
Wu 2008	34	16	0	550	1.00 [0.90, 1.00]	0.97 [0.95, 0.98]		
Jordan 2006	7	30	10	1186	0.41 [0.18, 0.67]	0.98 [0.96, 0.98]		
Jordan 2000	24	3	1	520	0.96 [0.80, 1.00]	0.99 [0.98, 1.00]		
Makhoul 2005	9	0	4	202	0.69 [0.39, 0.91]	1.00 [0.98, 1.00]		
Van der Brand 2014	10	0	3	7	0.77 [0.46, 0.95]	1.00 [0.59, 1.00]		
Villanueva-Uy 2003	23	0	6	32	0.79 [0.60, 0.92]	1.00 [0.89, 1.00]		
Jordan 2005a	51	0	2	32	0.96 [0.87, 1.00]	1.00 [0.89, 1.00]		· · · · · · · · · · · · · · · · · · ·
							0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8 1



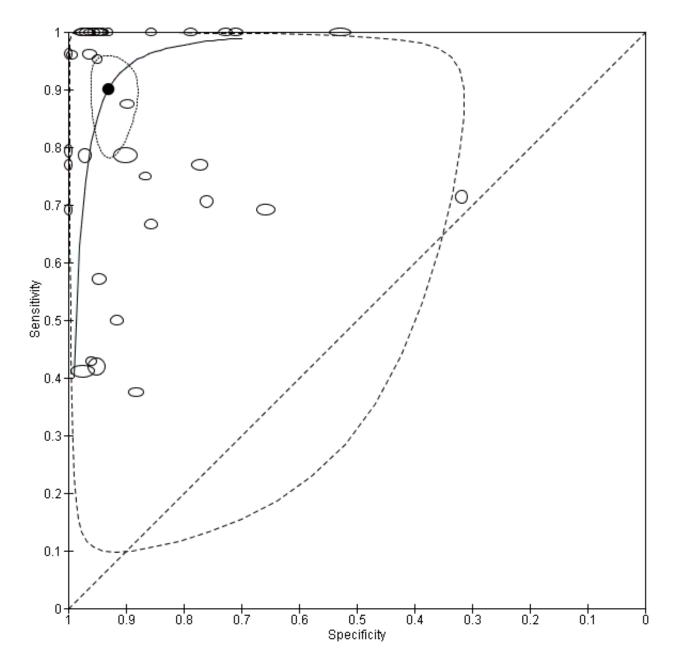


Figure 6. Summary receiver operating characteristic plot of all molecular tests.

We explored heterogeneity by differentiating studies based on the type of molecular assay, onset of sepsis, gestational age and prevalence, and plotted the subgroups of studies in the ROC space (moderate to low quality evidence). Figure 7 represents the studies differentiated by the type of molecular assay in the ROC space. Summary estimates for real-time PCR assays were sensitivity 0.86 (95% CI 0.59 to 0.96) and specificity 0.94 (95% CI 0.90 to 0.97). Broad-range conventional PCR performed with sensitivity 0.97 (95% CI 0.86 to 1.00), specificity 0.93 (95% CI 0.77 to 0.98), tests with post-PCR processing, sensitivity 0.97 (95% CI 0.40 to 1.00) and specificity 0.96 (95% CI 0.93 to 0.98) and multiplex PCR, sensitivity 0.76 (95% CI 0.60 to 0.88), specificity 0.81 (95% CI 0.70 to 0.89) (Summary of findings 1). Summary estimates of sensitivity and specificity for Staphylococcal PCR and fungal PCR were not possible as there four or fewer studies.

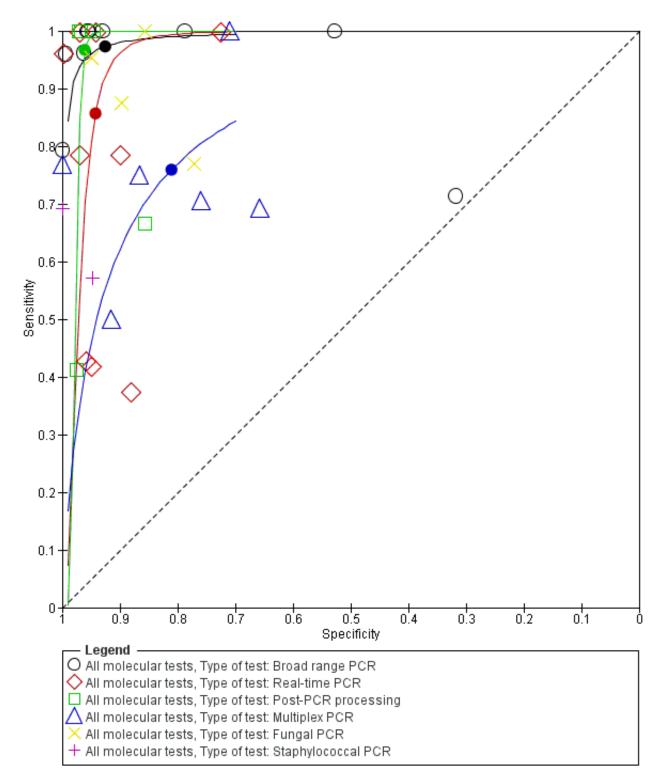


Figure 7. Summary receiver operating characteristic plot by type of molecular test. PCR: polymerase chain reaction.

Two studies reported on EOS, 10 on only LOS and 23 studies on both. Summary estimates for the molecular tests in the diagnosis of LOS were sensitivity 0.79 (95% CI 0.69 to 0.86), specificity 0.94

(95% CI 0.85 to 0.98) and mixed EOS and LOS were sensitivity 0.94 (95% CI 0.84 to 0.98), specificity 0.92 (95% CI 0.87 to 0.95) (Figure 8; Summary of findings 1).



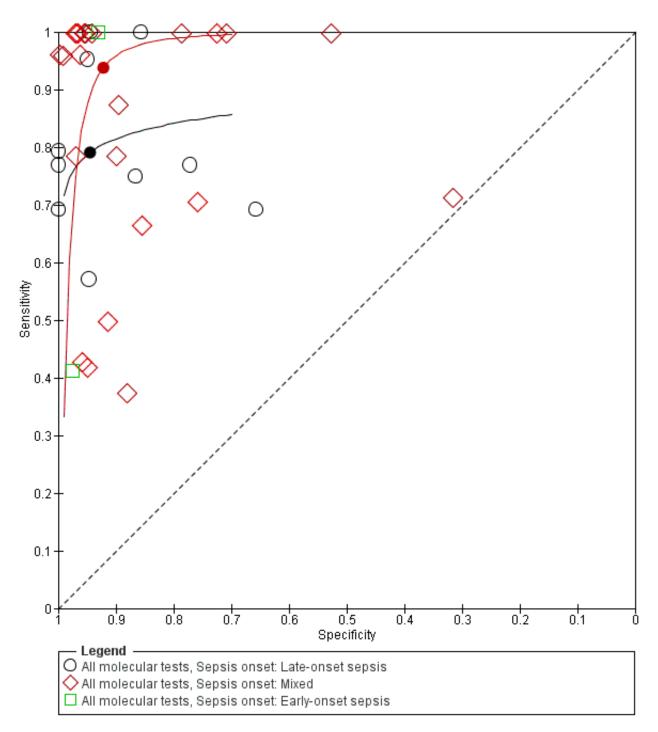
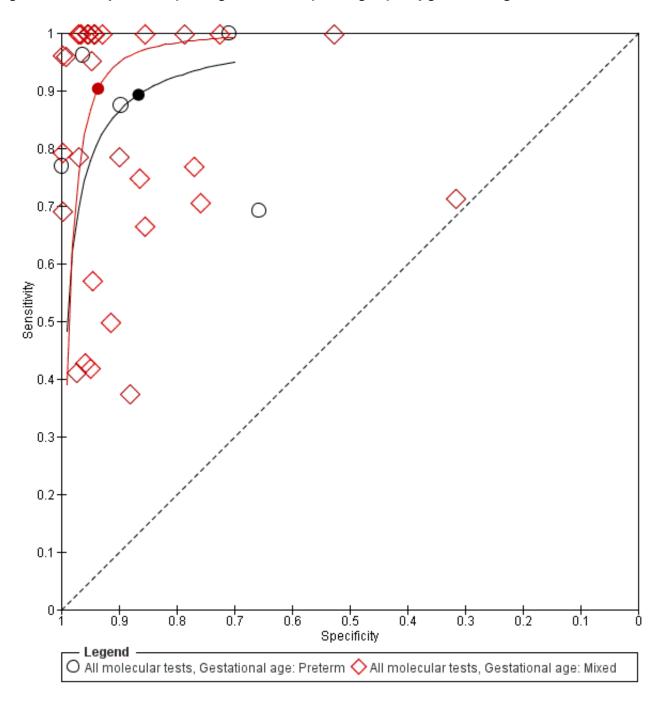


Figure 8. Summary receiver operating characteristic plot subgrouped by sepsis onset.

Five studies reported on testing on preterm infants only and 30 studies on a combination of preterm and term infants. Summary estimates for studies reporting on only preterm infants were sensitivity 0.89 (95% Cl 0.75 to 0.96), specificity 0.87 (95% Cl 0.71 to 1.25% cl 0.71% cl 0.75% cl 0.71% cl 0.75% cl 0.71% cl 0.71% cl 0.75% cl 0.71% cl 0.71% cl 0.75% cl 0.71% cl 0.75% cl 0.71% cl 0.71% cl 0.75% cl 0.71% cl 0.75% cl 0.

0.94) and those for mixed term and preterm infants were sensitivity 0.90 (95% CI 0.80 to 0.96), specificity 0.94 (0.90 to 0.96) (Figure 9; Summary of findings 1).





We categorized studies into three groups based on prevalence less than 15%, 15 % to 30% and greater than 30%. Summary estimates for 20 studies with a prevalence of less than 15% were sensitivity 0.94 (95% CI 0.80 to 0.99), specificity 0.95 (95% CI 0.92 to 0.97), with prevalence 15% to 30% were sensitivity 0.85 (95% CI 0.67 to 0.94),

specificity 0.88 (95% CI 0.79 to 0.94) and those for studies with a sepsis prevalence greater than 30% were sensitivity 0.87 (95% CI 0.75 to 0.93), specificity 0.93 (95% CI 0.64 to 0.99) (moderate to low quality evidence) (Summary of findings 1; Figure 10; Figure 11).



Cochrane

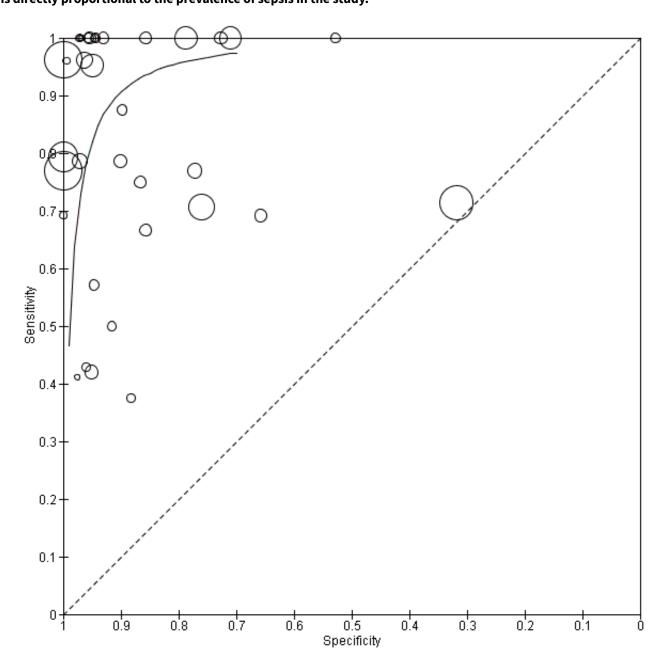
Library

Figure 10. Forest plot of all molecular tests sorted in order of prevalence. CI: confidence interval; FN: false negative; FP: false positive; TN: true negative; TP: true positive.

Study	ΤР	FP	FN	TN	Prevalence	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Jordan 2006	7	30	10	1186	1.38	0.41 [0.18, 0.67]	0.98 [0.96, 0.98]		
Wu 2007	20	23	0	787	2.41	1.00 [0.83, 1.00]	0.97 [0.96, 0.98]		
Tong 2004	8	9	0	268	2.81	1.00 [0.63, 1.00]	0.97 [0.94, 0.99]		•
Jordan 2000	24	3	1	520	4.56	0.96 [0.80, 1.00]	0.99 [0.98, 1.00]		•
Shang 2005	8	9	0	155	4.65	1.00 [0.63, 1.00]	0.95 [0.90, 0.97]		-
Wu 2008	34	16	0	550	5.67	1.00 [0.90, 1.00]	0.97 [0.95, 0.98]		•
Makhoul 2005	9	0	4	202	6.05	0.69 [0.39, 0.91]	1.00 [0.98, 1.00]	_	•
Chen 2009	15	10	0	170	7.69	1.00 [0.78, 1.00]	0.94 [0.90, 0.97]		
Enomoto 2009	3	6	3	65	7.79	0.50 [0.12, 0.88]	0.92 [0.83, 0.97]		-
Esparcia 2011	3	3	4	73	8.43	0.43 [0.10, 0.82]	0.96 [0.89, 0.99]		
Lima 2007	3	10	5	75	8.6	0.38 [0.09, 0.76]	0.88 [0.79, 0.94]		-
Yadav 2005	9	- 4	0	87	9.0	1.00 [0.66, 1.00]	0.96 [0.89, 0.99]		-=
Garcia-Elorriaga 2012	9	42	0	47	9.18	1.00 [0.66, 1.00]	0.53 [0.42, 0.63]		
Trovato 2012	- 7	8	1	70	9.3	0.88 [0.47, 1.00]	0.90 [0.81, 0.95]		
Makhoul 2006	8	- 7	6	125	9.59	0.57 [0.29, 0.82]	0.95 [0.89, 0.98]		-
Paolucci 2009	3	4	1	26	11.76	0.75 [0.19, 0.99]	0.87 [0.69, 0.96]		
Laforgia 1997	4	2	0	27	12.12	1.00 [0.40, 1.00]	0.93 [0.77, 0.99]		
Reier-Nilsen 2009	4	6	2	36	12.5	0.67 [0.22, 0.96]	0.86 [0.71, 0.95]		
Taira 2014	3	3	0	18	12.5	1.00 [0.29, 1.00]	0.86 [0.64, 0.97]		
Liu 2014	95	28	0	583	13.46	1.00 [0.96, 1.00]	0.95 [0.93, 0.97]	-	•
Ibarra 2015	9	25	4	48	15.12	0.69 [0.39, 0.91]	0.66 [0.54, 0.76]	_	
Ohlin 2012	44	31	12	281	15.22	0.79 [0.66, 0.88]	0.90 [0.86, 0.93]		•
Fujimori 2010	6	9	0	24	15.38	1.00 [0.54, 1.00]	0.73 [0.54, 0.87]		
Ohlin 2008	21	12	29	233	16.95	0.42 [0.28, 0.57]	0.95 [0.92, 0.97]		•
Tirodker 2003	10	13	3	44	18.57	0.77 [0.46, 0.95]	0.77 [0.64, 0.87]		
Chan 2009	33	- 5	9	171	19.27	0.79 [0.63, 0.90]	0.97 [0.93, 0.99]		•
Dutta 2009	50	- 7	2	183	21.49	0.96 [0.87, 1.00]	0.96 [0.93, 0.99]		•
Kasper 2013	15	9	0	22	32.61	1.00 [0.78, 1.00]	0.71 [0.52, 0.86]		
Shaat 2013	17	- 7	0	26	34.0	1.00 [0.80, 1.00]	0.79 [0.61, 0.91]		
Briones 2003	20	2	1	38	34.43	0.95 [0.76, 1.00]	0.95 [0.83, 0.99]		
Torres-Martos 2013	12	6	5	19	40.48	0.71 [0.44, 0.90]	0.76 [0.55, 0.91]		
Villanueva-Uy 2003	23	0	6	32	47.54	0.79 [0.60, 0.92]	1.00 [0.89, 1.00]		
Draz 2013	20	15	8	7	56.0	0.71 [0.51, 0.87]	0.32 [0.14, 0.55]	_	
Jordan 2005a	51	0	2	32	62.35	0.96 [0.87, 1.00]	1.00 [0.89, 1.00]		
Van der Brand 2014	10	0	3	7	65.0	0.77 [0.46, 0.95]	1.00 [0.59, 1.00]		
								0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8 1



Figure 11. Summary receiver operating characteristic plot of all molecular tests where the size of the study symbol is directly proportional to the prevalence of sepsis in the study.



We performed sensitivity analyses using data from studies evaluating blood samples alone (not CSF) excluding three studies; the summary sensitivity was 0.92 (95% CI 0.84 to 0.96), specificity 0.93 (95% CI 0.89 to 0.95) (Figure 12; Summary of findings 1) (moderate quality evidence). Furthermore, we investigated the effect of the potential sources of bias by removing studies with

unclear or high risk of bias or applicability concerns (13 studies) from the total set of studies and re-analyzing this new set (22 studies) and found no differences in summary estimates; the summary sensitivity was 0.90 (95% CI 0.78 to 0.96), specificity 0.93 (95% CI 0.88 to 0.96) (moderate quality evidence) (Summary of findings 1).

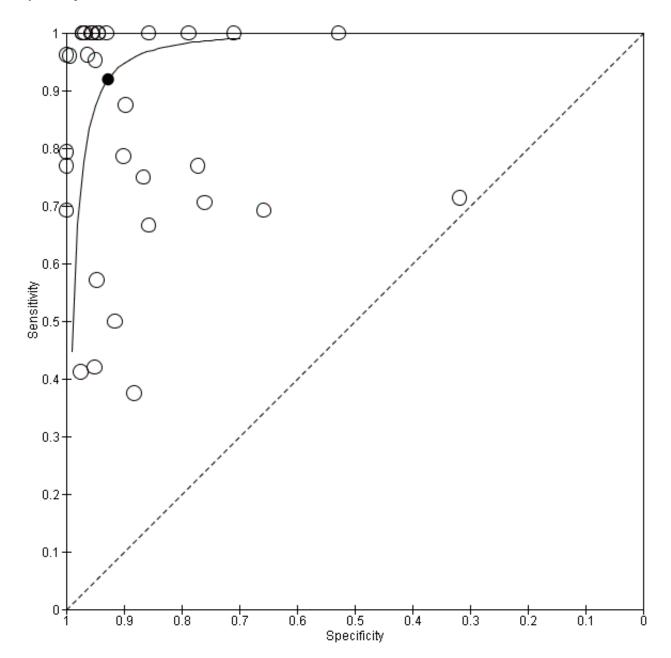


Figure 12. Summary receiver operating characteristic plot of studies that performed molecular tests on blood samples only.

DISCUSSION

Summary of main results

Our search strategy identified 35 eligible studies and mean sensitivity of molecular tests in the diagnosis of neonatal sepsis was 0.90 (95% CI 0.82 to 0.95) and specificity was 0.93 (95% CI 0.89 to 0.96) and evidence was of moderate quality. We explored heterogeneity by subgroup analyses based on type of test, gestational age, type of sepsis onset and prevalence of neonatal sepsis (moderate to low quality evidence). We also performed sensitivity analysis by excluding studies which used both blood and CSF samples and excluding studies with high or uncertain risk of bias and applicability concerns.

Low sensitivity (less than 0.7 in nine studies) in some of the studies may be explained by the technicalities of the multiplex PCR assay, use of stored blood samples that were drawn by heel stick at a different time to the blood culture sample, participant characteristics and to Staphylococcus-specific PCR. Jordon and colleagues commented that presence of white blood cells in the samples and hence human genomic DNA interference may have inhibited the PCR assay accounting for low sensitivity (Jordan 2006). However, 13 studies reported a sensitivity of 1.00 that did not conform to any particular type of test or participant population. In contrast, specificity was consistently higher than sensitivity and all except three of the included studies had specificity more than 0.70 (Draz 2013; Garcia-Elorriaga 2012; Ibarra 2015). Primers used in the

tests and differences in participant characteristics may accounted for low specificity. Four studies reported a specificity of 1.00 but with varying sensitivities and type of molecular assays (Jordan 2005a; Makhoul 2005; Van der Brand 2014; Villanueva-Uy 2003).

We explored sources of heterogeneity by subgroup analyses based on type of test, gestational age, type of sepsis onset and prevalence. We found that studies evaluating molecular tests with post-PCR processing, real-time PCR and broad-range conventional PCR plotted in the left upper corner of the ROC space and had higher sensitivity and specificity than multiplex PCR assay. Summary sensitivities from subgroups based on gestational age were similar with overlapping CIs and summary specificity was higher in studies that evaluated both preterm and term infants. In 10 studies that evaluated only LOS, the sensitivity was lower than the summary estimate for mixed EOS and LOS (0.79 (95% CI 0.66 to 0.87) versus 0.94 (95% CI 0.84 to 0.98)) but had higher specificity (0.94 (95% CI 0.85 to 0.98) versus 0.92 (95% CI 0.87 to 0.95)). But the wide 95% CIs precluded any delineation based on these subgroup analyses. We categorized studies into three groups based on sepsis prevalence less than 15%, 15% to 30% and greater than 30%. Studies that evaluated molecular tests in a population with a sepsis prevalence less than 15% had higher sensitivity and specificity (sensitivity 0.94 (95% CI 0.80 to 0.99), specificity 0.95 (95% CI 0.92 to 0.97)) compared with studies in a higher sepsis prevalence population (Summary of findings 1). Variations in participant characteristics or test methodology may account for some of these differences.

We performed sensitivity analyses by type of samples used (blood or both blood and CSF, because inhibitors of PCRs may be present only in blood samples) and for studies evaluating blood samples alone (not CSF), the summary sensitivity was 0.92 (95% CI 0.84 to 0.96) and specificity 0.93 (95% CI 0.89 to 0.95) (Figure 12; Summary of findings 1). We also investigated the effect of the potential sources of bias by removing studies with unclear or high risk of bias or applicability concerns (13 studies) from the total set of studies and re-analyzing this new set (22 studies) and found no differences (summary sensitivity 0.90 (95% CI 0.78 to 0.96), specificity 0.93 (95% CI 0.88 to 0.96)) (Figure 13; Summary of findings 1).



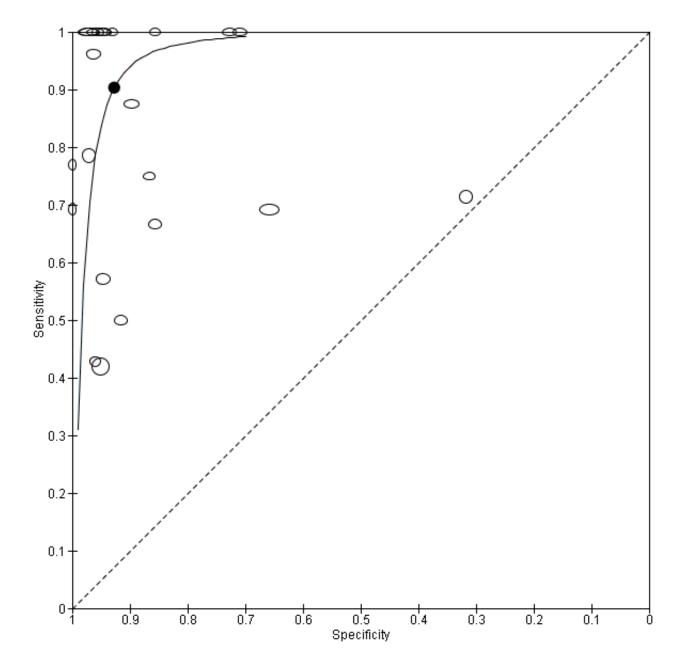


Figure 13. Summary receiver operating characteristic plot of molecular tests with good methodologic quality.

Other sources for variation of diagnostic test accuracy among studies evaluating molecular tests may be due to methods of DNA extraction or preprocessing the sample before DNA extraction (e.g. preincubation of the blood culture media before DNA extraction). Studies using whole blood DNA extraction had low sensitivity and preincubation of sample for five hours in tryptic soy broth increased sensitivity significantly. However, the methodologies of DNA extraction, samples from which DNA were extracted, varied considerably among the studies to make any meaningful comparisons.

New diagnostic tests can assume the following roles in a diagnostic pathway: replacement of the existing test, triage or 'add on' to the existing test (Bossuyt 2006). Our meta-analysis estimated

a mean sensitivity of 0.90 (95% CI 0.82 to 0.95) and a mean specificity 0.93 (95% CI 0.89 to 0.96) for molecular assays. The mean estimated sensitivity of molecular assays are better than other alternative tests used to diagnose sepsis such as platelet count, CRP, procalcitonin, TNF and IL-6 while mean specificity was similar to these tests (Blommendahl 2002; Hornik 2012; Ng 1997; Ng 2012; Verboon-Maciolek 2006). Theoretically, in 1000 VLBW neonates screened for EOS, where the prevalence was 2% (using the summary estimates of this review), we would miss two cases of sepsis and overtreat 69 neonates without sepsis. Similarly, in 1000 VLBW neonates screened for LOS (prevalence 10%), we would miss 10 culture-positive cases and overtreat 63 neonates without sepsis. Thus, currently available molecular assays may not have sufficient diagnostic accuracy to replace microbial cultures.

Cochrane Library

Trusted evidence. Informed decisions. Better health.

However, advancing technologies in molecular microbiology may bring forth newer assays with higher sensitivity and specificity, sufficient to replace microbial cultures in the diagnosis of neonatal sepsis.

In addition to test accuracy, it is important to consider management strategies for neonatal sepsis where molecular tests may be useful. Evidence to decision frameworks are recommended to assess how test results affect participant outcomes (Schünemann 2016; Trenti 2016). In the context of neonatal sepsis, molecular assays are unlikely to be used as a triage test that will select neonates who would undergo cultures. An unwanted delay in performing blood cultures may ensue and may postpone treatment. False negatives on the molecular tests will compromise neonatal safety. However, molecular assays have a faster turnaround time and may perform well as 'add-on' tests where molecular assays may be performed concurrently with the gold standard (cultures). Results of molecular assays are available in six to eight hours and may help in optimizing clinical therapy. If the molecular test is negative, antibiotics may be discontinued if the test assay has high specificity and high negative predictive value. Decrease in antibiotic doses and decreased length of stay are potential advantages of such a strategy (Brozanski 2006). If the molecular test assay is positive (and if the assay has high sensitivity) then a case could be made for continuation of antibiotics. Molecular assays may theoretically diagnose sepsis in neonates exposed to antibiotics including maternal exposure to antibiotics in EOS, where cultures are negative and potentially decrease resource utilization. Combination of blood cultures with an 'add-on' molecular test may improve sensitivity at the cost of specificity. Newer molecular assays that can identify the organism or detect antibiotic resistance can guide antibiotic therapy.

Jordan and colleagues and our group reviewed the methodology of molecular assays used in the diagnosis of neonatal sepsis without synthesizing data using meta-analyses (Jordan 2010; Venkatesh 2010). Our group published one systematic review with metaanalysis of 23 studies evaluating molecular assays in the diagnosis of neonatal sepsis (Pammi 2011). Overall, the summary estimates of sensitivity and specificity were similar with larger CIs and slightly higher specificity (sensitivity 0.90 (95% CI 0.78 to 0.95), specificity 0.96 (95% CI 0.94 to 0.97)). In our previous review, we were unable to analyze reasons for heterogeneity as data were not available, which we were able to do in this review.

Strengths and weaknesses of the review

Strengths: our systematic review was based on methodology recommended by the Cochrane Diagnostic Test Accuracy Working Group (Leeflang 2008). We performed a comprehensive search for all eligible studies using clinically relevant inclusion criteria. We used the bivariate random-effects model for meta-analyses of the included studies. We strived to explain the sources of heterogeneity by subgroup analyses using test type, gestational age of participants, type of sepsis onset and prevalence.

Weaknesses: evolution in methodology in the included studies over time (1997 to 2016) may account for variations in the diagnostic accuracy among studies. Unlike meta-analyses of randomized controlled trials, heterogeneity is a well-recognized problem in reviews of diagnostic test accuracy (Reitsma 2009). Despite our extensive search strategy, we may have missed potential studies, as diagnostic accuracy studies are poorly tagged in electronic databases. Publication bias in studies reporting diagnostic test accuracy has been poorly studied (Leeflang 2008). Poor reporting of study design, method of enrollment and participant characteristics may hamper methodologic assessment and external validity of the studies. Another limitation of our review might be that the reference standard (microbial cultures) is thought to be far from perfect. Interpretation of the accuracy of molecular assays is challenging given the assumed low sensitivity of the blood cultures. However, as our summary sensitivity of the molecular assays was poor (0.90) and the proportion of false positives was low, it does not seem to be the case.

Applicability of findings to the review question

Molecular assays have significant advantages when performed in conjunction with microbial cultures as an 'add-on' test. The high specificity of molecular assay in LOS evaluation (0.94 (95% CI 0.85 to 0.98)) has the potential of decreasing antibiotic exposure by aiding physicians to make earlier decisions about discontinuation of antibiotics. Molecular assays, including PCR and hybridization methods, are feasible in neonates and have rapid detection times compared to blood cultures (six to eight hours versus 20 to 36 hours). Detection of pathogen DNA in the absence of viable organisms by culture and false-negative results due to the presence of inhibitors may require careful interpretation. Molecular assays may have a significant impact on early diagnosis and treatment of neonatal sepsis. However, current molecular assays do not provide antibiotic susceptibility that may be important clinically. Microbiologic cultures detect most organisms causing neonatal sepsis, whereas molecular assays focused on fungi or a specific organism (Staphylococcus- or fungus-specific PCR) do not. Costs, availability of equipment and technical skills in the microbiologic laboratory are important considerations that will impact applicability.

AUTHORS' CONCLUSIONS

Implications for practice

The mean sensitivity of molecular assays in the diagnosis of clinically suspected neonatal sepsis was 0.90 (95% CI 0.82 to 0.95) and mean specificity was 0.93 (95% CI 0.89 to 0.96) (moderate quality evidence) and the diagnostic accuracy was variable among reported studies. Molecular tests for the diagnosis of sepsis may be useful 'add-on' tests as they give rapid information that may aid clinical decisions regarding treatment. Our recommendations are based on moderate to low quality evidence. Optimization of existing assays or the development of new molecular assays in the future may improve diagnostic accuracy. Future molecular tests that may identify the pathogen and evaluate pathogen virulence and antibiotic susceptibility, in addition to diagnosis of sepsis may aid clinical management tremendously.

Implications for research

Investigators evaluating current as well as future molecular tests should design their studies satisfying the items expounded in the QUADAS-2 evaluation system, so that studies are of high methodologic quality and bias is minimal. Studies reporting diagnostic test accuracy should explicitly state the method of enrollment (prospective or retrospective), characteristics of the population assessed (such as gestational age, chronologic age range, birth weight, comorbidity), blinding of reference standard and index tests, and explanation of withdrawals. Details of the clinical setting and participant characteristics will help clinicians decide whether a diagnostic test is applicable in their population. Costs of the molecular assays need to be balanced with their ability to impact clinical outcomes before widespread acceptance in clinical practice.

ACKNOWLEDGEMENTS

The Cochrane Neonatal Group has been funded in part with federal funds from the Eunice Kennedy Shriver National Institute of Child Health and Human Development National Institutes of Health, Department of Health and Human Services, USA, under Contract No. HHSN267200603418C. We acknowledge the help of Nha Huynh, librarian at the Texas Medical Center library in Houston, TX, USA and Milagros De Jesus, librarian at Texas Technical University Health Sciences Center, Lubbock, TX, USA for invaluable help with search strategy.

We acknowledge the help of Joe Hagan, statistician, Section of Neonatology at Baylor College of Medicine in creating the Deeks' funnel plot and checking the statistics for the hierarchical summary receiver operating characteristic (HSROC) and the bivariate model.



REFERENCES

References to studies included in this review

Briones 2003 {published data only}

Briones CR, Villanueva-Uy ME, Uy HGTI. The use of polymerase chain reaction in neonatal candidemia. *Pediatric Research* 2003;**53**:396A.

Chan 2009 {published data only}

Chan KY, Lam HS, Cheung HM, Chan AK, Li K, Fok TF, et al. Rapid identification and differentiation of Gram-negative and Gram-positive bacterial bloodstream infections by quantitative polymerase chain reaction in preterm infants. *Critical Care Medicine* 2009;**37**:2441-7. [PUBMED: 19531943]

Chen 2009 {published data only}

Chen LH, Duan QJ, Cai MT, Wu YD, Shang SQ. Rapid diagnosis of sepsis and bacterial meningitis in children with realtime fluorescent quantitative polymerase chain reaction amplification in the bacterial 16S rRNA gene. *Clinical Pediatrics* 2009;**48**(6):641-7. [PUBMED: 19407210]

Draz 2013 {published data only}

Draz NI, Taha SE, Abou Shady NM, Abdel Ghany YS. Comparison of broad range 16S rDNA PCR to conventional blood culture for diagnosis of sepsis in the newborn. *Egyptian Journal of Human Medical Genetics* 2013;**14**:403-11. [PUBMED: 19152691]

Dutta 2009 {published data only}

Dutta S, Narang A, Chakraborty A, Ray P. Diagnosis of neonatal sepsis using universal primer polymerase chain reaction before and after starting antibiotic drug therapy. *Archives of Pediatrics and Adolescent Medicine* 2009;**163**(1):6-11. [PUBMED: 19124696]

Enomoto 2009 {published data only}

Enomoto M, Morioka I, Morisawa T, Yokoyama N, Matsuo M. A novel diagnostic tool for detecting neonatal infections using multiplex polymerase chain reaction. *Neonatology* 2009;**96**(2):102-8. [PUBMED: 19279393]

Esparcia 2011 {published data only}

Esparcia O, Montemayor M, Ginovart G, Pomar V, Soriano G, Pericas R, et al. Diagnostic accuracy of a 16S ribosomal DNA gene-based molecular technique (RT-PCR, microarray, and sequencing) for bacterial meningitis, early-onset neonatal sepsis, and spontaneous bacterial peritonitis. *Diagnostic Microbiology and Infectious Disease* 2011;**69**(2):153-60. [PUBMED: 21251558]

Fujimori 2010 {published data only}

Fujimori M, Hisata K, Nagata S, Matsunaga N, Komatsu M, Shoji H, et al. Efficacy of bacterial ribosomal RNA-targeted reverse transcription-quantitative PCR for detecting neonatal sepsis: a case control study. *BMC Pediatrics* 2010;**10**:53. [PUBMED: 20667142]

Garcia-Elorriaga 2012 {published data only}

Garcia-Elorriaga G, Cortes-Torres N, Ballesteros-Del-Olmo JC, Del Rey-Pineda G, Gonzaez-Bonilla C. The usefulness of the buffy coat smear and panbacterial polymerase chain reaction in early diagnosis of neonatal sepsis. *Revista de Investigacion Clinica* 2012;**64**(3):275-83. [PUBMED: 23045950]

Ibarra 2015 {published data only}

Ibarra JO, Valdez PT, Mendez EV, Rojas AL, Flores GL, Bocanegra AC, et al. Evaluation of the Light-Cycler® SeptiFast Test in newborns with suspicion of nosocomial sepsis. *Iran Journal of Pediatrics* 2015;**25**(1):e253. [PUBMED: 26199693]

Jordan 2000 {published data only}

Jordan JA, Durso MB. Comparison of 16S rRNA gene PCR and BACTEC 9240 for detection of neonatal bacteremia. *Journal of Clinical Microbiology* 2000;**38**(7):2574-8. [PUBMED: 10878046]

Jordan 2005a {published data only}

Jordan JA, Durso MB. Real-time polymerase chain reaction for detecting bacterial DNA directly from blood of neonates being evaluated for sepsis. *Journal of Molecular Diagnostics* 2005;**7**(5):575-81. [PUBMED: 16258155]

Jordan 2006 {published data only}

Jordan JA, Durso MB, Butchko AR, Jones JG, Brozanski BS. Evaluating the near-term infant for early onset sepsis: progress and challenges to consider with 16S rDNA polymerase chain reaction testing. *Journal of Molecular Diagnostics* 2006;**8**(3):357-63. [PUBMED: 16825509]

Kasper 2013 {published data only}

Kasper DC, Altiok I, Mechtler TP, Bohm J, Straub J, Langgartner M, et al. Molecular detection of late-onset neonatal sepsis in premature infants using small blood volumes: proofof-concept. *Neonatology* 2013;**103**(4):268-73. [PUBMED: 23485823]

Laforgia 1997 {published data only}

Laforgia N, Coppola B, Carbone R, Grassi A, Mautone A, Lolascon A. Rapid detection of neonatal sepsis using polymerase chain reaction. *Acta Paediatrica* 1997;**86**(10):1097-9. [PUBMED: 9350892]

Lima 2007 {published data only}

Lima V, Alpuche A, Noyola D, Soria R, Nieto K. Polymerase chain reaction technique in the diagnosis of neonatal sepsis: future gold standard?. Pediatric Academic Societies Annual Meeting; 2007 May 5-8; Toronto (ON) 2007.

Liu 2014 {published data only}

Liu CL, Ai HW, Wang WP, Chen L, Hu HB, Ye T, et al. Comparison of 16S rRNA gene PCR and blood culture for diagnosis of neonatal sepsis. *Archives of Pediatrics* 2014;**21**(2):162-9. [PUBMED: 24388336]

Makhoul 2005 {published data only}

Makhoul I, Smolkin RT, Sujov P, Kassis I, Tamir A, Shalginov R, et al. PCR-based diagnosis of neonatal staphylococcal bacteremias. *Journal of Clinical Microbiology* 2005;**43**(9):4823-5. [PUBMED: 16145149]



Makhoul 2006 {published data only}

Makhoul IR, Yacoub A, Smolkin T, Sujov P, Kassis I, Sprecher H. Values of C-reactive protein, procalcitonin, and Staphylococcusspecific PCR in neonatal late-onset sepsis. *Acta Paediatrica* 2006;**95**(10):1218-23. [PUBMED: 16982493]

Ohlin 2008 {published data only}

Ohlin A, Backman A, Bjorkqvist M, Molling P, Jurstrand M, Schollin J. Real-time PCR of the 16S-rRNA gene in the diagnosis of neonatal bacteraemia. *Acta Paediatrica* 2008;**97**(10):1376-80. [PUBMED: 18624992]

Ohlin 2012 {published data only}

Ohlin A, Backman A, Ewald U, Schollin J, Bjorkqvist M. Diagnosis of neonatal sepsis by broad-range 16S real-time polymerase chain reaction. *Neonatology* 2012;**10**(4):241-6. [PUBMED: 22205207]

Paolucci 2009 {published data only}

Paolucci M, Capretti MG, Dal Monte P, Corvaglia L, Landini MP, Varani S, et al. Laboratory diagnosis of late-onset sepsis in newborns by multiplex real-time PCR. *Journal of Medical Microbiology* 2009;**58**(Pt 4):533-4. [PUBMED: 19273654]

Reier-Nilsen 2009 {published data only}

Reier-Nilsen T, Farstad T, Nakstad B, Lauvrak V, Steinbakk M. Comparison of broad range 16S rDNA PCR and conventional blood culture for diagnosis of sepsis in the newborn: a case control study. *BMC Pediatrics* 2009;**9**:5. [PUBMED: 19152691]

Shaat 2013 {published data only}

Shaat SS, El Shazly SA, Badr Eldin MM, Barakat SS, Hashish MH. Role of polymerase chain reaction as an early diagnostic tool for neonatal bacterial sepsis. *Journal of Egypt Public Health Association* 2013;**88**(3):160-4. [PUBMED: 24374951]

Shang 2005 {published data only}

Shang S, Chen G, Wu Y, Du L, Zhao Z. Rapid diagnosis of bacterial sepsis with PCR amplification and microarray hybridization in 16S rRNA gene. *Pediatric Research* 2005;**58**(1):143-8. [PUBMED: 15985688]

Taira 2014 {published data only}

Taira CL, Okay TS, Delgado AF, Ceccon ME, de Almeida MT, Del Negro GM. A multiplex nested PCR for the detection and identification of Candida species in blood samples of critically ill paediatric patients. *BMC Infectious Diseases* 2014;**14**:406. [PUBMED: 25047415]

Tirodker 2003 {published data only}

Tirodker UH, Nataro JP, Smith S, LasCasas L, Fairchild KD. Detection of fungemia by polymerase chain reaction in critically ill neonates and children. *Journal of Perinatology* 2003;**23**(2):117-22. [PUBMED: 12673260]

Tong 2004 {published data only}

Tong MQ, Shang SQ, Wu YD, Zhao ZY. Rapid diagnosis of neonatal sepsis by 16SrRNA genes PCR amplification and genechip hybridization. *Zhonghua Er Ke Za Zhi* 2004;**42**(9):663-7. [PUBMED: 15482666]

Torres-Martos 2013 {published data only}

Torres-Martos E, Perez-Ruiz M, Pedrosa-Corral I, Pena-Caballero M, Jimenez-Valera MM, Perez-Ramirez MD, et al. Evaluation of the LightCycler[®] SeptiFast test in newborns and infants with clinical suspicion of sepsis. *Enfermedades Infecciosas Microbiologia Clinica* 2013;**31**(6):375-9. [PUBMED: 23137657]

Trovato 2012 {published data only}

Trovato L, Betta P, Romeo MG, Oliveri S. Detection of fungal DNA in lysis-centrifugation blood culture for the diagnosis of invasive candidiasis in neonatal patients. *Clinical Microbiology and Infection* 2012;**18**(3):E63-5. [PUBMED: 22192484]

Van der Brand 2014 {published data only}

Van den Brand M, Peters RP, Catsburg A, Rubenjan A, Broeke FJ, van den Dungen FA, et al. Development of a multiplex real-time PCR assay for the rapid diagnosis of neonatal late onset sepsis. *Journal of Microbiological Methods* 2014;**106**:8-15. [PUBMED: 25102109]

Villanueva-Uy 2003 {published data only}

Villanueva-Uy ME, Briones CR, Uy HG. Application of polymerase chain reaction in late-onset neonatal sepsis. *Pediatric Research* 2003;**53**:313A.

Wu 2007 {published data only}

Wu YD, Shang SQ, Li JP, Yang ZQ, Zheng ZB, Du LZ, et al. A broad-range 16S rRNA gene real-time PCR assay for the diagnosis of neonatal septicemia. *Zhonghua Er Ke Za Zhi* 2007;**45**(6):446-9. [PUBMED: 17880793]

Wu 2008 {published data only}

Wu YD, Chen LH, Wu XJ, Shang SQ, Lou JT, Du LZ, et al. Gram stain-specific-probe-based real-time PCR for diagnosis and discrimination of bacterial neonatal sepsis. *Journal of Clinical Microbiology* 2008;**46**(8):2613-9. [PUBMED: 18550744]

Yadav 2005 {published data only}

Yadav AK, Wilson CG, Prasad PL, Menon PK. Polymerase chain reaction in rapid diagnosis of neonatal sepsis. *Indian Pediatrics* 2005;**42**(7):681-5. [PUBMED: 16085969]

References to studies excluded from this review

Chiba 2009 {published data only}

Chiba N, Murayama SY, Morozumi M, Nakayama E, Okada T, Iwata S, et al. Rapid detection of eight causative pathogens for the diagnosis of bacterial meningitis by real-time PCR. *Journal* of Infection and Chemotherapy 2009;**15**(2):92-8. [PUBMED: 19396518]

Das 2015 {published data only}

Das BK, Suri S, Nath G, Prasad R. Urine nested polymerase chain reaction in neonatal septicemia. *Journal of Tropical Pediatrics* 2015;**61**(4):295-300. [PUBMED: 26130622]

de Zoysa 2012 {published data only}

de Zoysa A, Edwards K, Gharbia S, Underwood A, Charlett A, Efstratiou A. Non-culture detection of *Streptococcus agalactiae*



(Lancefield group B Streptococcus) in clinical samples by real-time PCR. *Journal of Medical Microbiology* 2012;**61**(Pt 8):1086-90. [PUBMED: 22740612]

Golden 2004 {published data only}

Golden SM, Stamilio DM, Faux BM, dela Cruz WP, Shoemaker CT, Blackmon CL, et al. Evaluation of a real-time fluorescent PCR assay for rapid detection of Group B Streptococci in neonatal blood. *Diagnostic Microbiology and Infectious Disease* 2004;**50**(1):7-13. [PUBMED: 15380273]

Jones 2010 {published data only}

Jones V, Wilks M, Johnson G, Warwick S, Hennessey E, Kempley S, et al. The use of molecular techniques for bacterial detection in the analysis of gastric aspirates collected from infants on the first day of life. *Early Human Development* 2010;**86**(3):167-70. [PUBMED: 20223606]

Jordan 2005b {published data only}

Jordan JA, Butchko, AR, Durso MB. Use of pyrosequencing of 16S rRNA fragments to differentiate between bacteria responsible for neonatal sepsis. *Journal of Molecular Diagnostics* 2005;**7**(1):105-10. [PUBMED: 15681481]

Jordan 2009 {published data only}

Jordan JA, Jones-Laughner J, Durso MB. Utility of pyrosequencing in identifying bacteria directly from positive blood culture bottles. *Journal of Clinical Microbiology* 2009;**47**(2):368-72. [PUBMED: 19091813]

Lucignano 2011 {published data only}

Lucignano B, Ranno S, Liesenfeld O, Pizzorno B, Putignani L, Bernaschi P, et al. Multiplex PCR allows rapid and accurate diagnosis of bloodstream infections in newborns and children with suspected sepsis. *Journal of Clinical Microbiology* 2011;**49**(6):2252-8. [PUBMED: 21471340]

Makhoul 2007 {published data only}

Makhoul IR, Sprecher H, Smolkin T, Sawaid R, Ben-David S, Sujov P, et al. Approach to term neonates born after maternal intrapartum fever and unknown maternal group B Streptococcus status: value of serum C-reactive protein and 16S rRNA gene PCR amplification. *Pediatric Infectious Disease Journal* 2007;**26**(11):1064-6. [PUBMED: 17984819]

Shang 2001 {published data only}

Shang S, Chen Z, Yu X. Detection of bacterial DNA by PCR and reverse hybridization in the 16S rRNA gene with particular reference to neonatal septicemia. *Acta Paediatrica* 2001;**90**(2):179-83. [PUBMED: 11236048]

Shen 2004 {published data only}

Shen DX, Du J, Feng ZC. Rapid diagnosis of common pathogenic bacteria infection in newborn infants by 16SrDNA oligonucleotide array. *Zhonghua Er Ke Za Zhi* 2004;**42**(9):668-72. [PUBMED: 15482667]

Tschiedel 2012 {published data only}

Tschiedel E, Steinmann J, Buer J, Onnebrink JG, Felderhoff-Muser U, Rath PM, et al. Results and relevance of molecular detection of pathogens by SeptiFast - a retrospective analysis Cochrane Database of Systematic Reviews

in 75 critically ill children. *Klinische Padiatrie* 2012;**224**(1):12-6. [PUBMED: 22258624]

Additional references

Adams-Chapman 2006

Adams-Chapman I, Stoll BJ. Neonatal infection and long-term neurodevelopmental outcome in the preterm infant. *Current Opinion in Infectious Diseases* 2006;**19**(3):290-7. [PUBMED: 16645492]

Blommendahl 2002

Blommendahl J, Janas M, Laine S, Miettinen A, Ashorn P. Comparison of procalcitonin with CRP and differential white blood cell count for diagnosis of culture-proven neonatal sepsis. *Scandinavian Journal of Infectious Diseases* 2002;**34**(8):620-2. [PUBMED: 12238581]

Bossuyt 2006

Bossuyt PM, Irwig L, Craig J, Glasziou P. Comparative accuracy: assessing new tests against existing diagnostic pathways. *BMJ* 2006;**332**(7549):1089-92. [PUBMED: 16675820]

Brozanski 2006

Brozanski BS, Jones JG, Krohn MJ, Jordan JA. Use of polymerase chain reaction as a diagnostic tool for neonatal sepsis can result in a decrease in use of antibiotics and total neonatal intensive care unit length of stay. *Journal of Perinatology* 2006;**26(11)**:688-692.

Canadian Neonatal Network 2014

Shah P, Yoon EW, Chan P, Members of the Annual Report Review Committee. Annual report of the Canadian Neonatal Network, 2004. www.canadianneonatalnetwork.org/Portal/ LinkClick.aspx?fileticket=eGgxmMubxjk%3d&tabid=39 (accessed 18 February 2017).

Chiesa 2004

Chiesa C, Panero A, Osborn JF, Simonetti AF, Pacifico L. Diagnosis of neonatal sepsis: a clinical and laboratory challenge. *Clinical Chemistry* 2004;**50**(2):279-87. [PUBMED: 14752012]

Dong 2015

Dong Y, Speer CP. Late-onset neonatal sepsis: recent developments. *Archives of Disease in Childhood. Fetal and Neonatal Edition* 2015;**100**(3):F257-63.

Gopalakrishna 2014

Gopalakrishna G, Mustafa RA, Davenport C, Scholten RJ, Hyde C, Brozek J, et al. Applying Grading of Recommendations Assessment, Development and Evaluation (GRADE) to diagnostic tests was challenging but doable. *Journal of Clinical Epidemiology* 2014;**67**(7):760-8.

Harbord 2007

Harbord RM, Deeks JJ, Egger M, Whiting P, Sterne JA. A unification of models for meta-analysis of diagnostic accuracy studies. *Biostatistics* 2007;**8**(2):239-51. [PUBMED: 16698768]



Harbord 2008

Harbord RM, Whiting P, Sterne JA, Egger M, Deeks JJ, Shang A, et al. An empirical comparison of methods for meta-analysis of diagnostic accuracy showed hierarchical models are necessary. *Journal of Clinical Epidemiology* 2008;**61**(11):1095-103. [PUBMED: 19208372]

Harbord 2009

Harbord RM, Whiting P. Metandi: meta-analysis of diagnostic accuracy using hierarchical logistic regression. *Stata Journal* 2009;**9**(2):211-29.

Hornik 2012

Hornik CP, Benjamin DK, Benjamin DKJ, Li J, Clark RH, Cohen-Wolkowiez M, et al. Use of the complete blood cell count in late-onset neonatal sepsis. *Pediatric Infectious Disease Journal* 2012;**31**(8):803-7. [PUBMED: 22531232]

Isaacman 1996

Isaacman DJ, Karasic RB, Reynolds EA, Kost SI. Effect of number of blood cultures and volume of blood on detection of bacteremia in children. *Pediatrics* 1996;**128**(2):190-5. [PUBMED: 8636810]

Jordan 2010

Jordan JA. Molecular diagnosis of neonatal sepsis. *Clinics in Perinatology* 2010;**37**(2):411-9. [PUBMED: 20569815]

Leeflang 2008

Leeflang MM, Deeks JJ, Gatsonis C, Bossuyt PM. Systematic reviews of diagnostic test accuracy. *Annals of Internal Medicine* 2008;**149**(12):889-97. [PUBMED: 19075208]

Macaskill 2010

Macaskill P, Gatsonis C, Deeks JJ, Harbord RM, Takwoingi Y. Chapter 10: Analysing and presenting results. In: Cochrane Handbook for Systematic Reviews of Diagnostic Test Accuracy Version 1.0.0. The Cochrane Collaboration, 2010. Available from srdta.cochrane.org/.

Ng 1997

Ng PC, Cheng SH, Chui KM, Fok TF, Wong MY, Wong W, et al. Diagnosis of late onset neonatal sepsis with cytokines, adhesion molecule, and C-reactive protein in preterm very low birthweight infants. *Archives of Disease in Childhood Fetal and Neonatal Edition* 1997;**77**(3):F221-7. [PUBMED: 9462194]

Ng 2012

Ng PC, Lam HS. Biomarkers in neonatology: the next generation of tests. *Neonatology* 2012;**102**(2):145-51. [PUBMED: 22759988]

Pammi 2011

Pammi M, Flores A, Leeflang M, Versalovic J. Molecular assays in the diagnosis of neonatal sepsis: a systematic review and metaanalysis. *Pediatrics* 2011;**128**(4):e973-85. [PUBMED: 21949139]

Reitsma 2009

Reitsma JB, Whiting P, Vlassov VV, Leeflang MMG, Deeks JJ. Chapter 9: Assessing methodological quality. In: Cochrane Handbook for Systematic Reviews of Diagnostic Test Accuracy Version 1.0.0. The Cochrane Collaboration, 2009. Available from srdta.cochrane.org/.

Relman 1999

Relman DA. The search for unrecognized pathogens. *Science* 1999;**284**(5418):1308-10. [PUBMED: 10334977]

RevMan 2014 [Computer program]

The Nordic Cochrane Centre, The Cochrane Collaboration. Review Manager (RevMan). Version 5.3. Copenhagen: The Nordic Cochrane Centre, The Cochrane Collaboration, 2014.

Rodriguez-Creixems 2008

Rodriguez-Creixems M, Alcala L, Munoz P, Cercenado E, Vicente T, Bouza E. Bloodstream infections: evolution and trends in the microbiology workload, incidence, and etiology, 1985-2006. *Medicine* 2008;**87**(4):234-49. [PUBMED: 18626306]

Schelonka 1996

Schelonka RL, Chai MK, Yoder BA, Hensley D, Brockett RM, Ascher DP. Volume of blood required to detect common neonatal pathogens. *Journal of Pediatrics* 1996;**129**(2):275-8. [PUBMED: 8765627]

Schünemann 2016

Schünemann HJ, Mustafa R, Brozek J, Santesso N, Alonso-Coello P, Guyatt G, et al. GRADE Working Group. GRADE Guidelines: 16. GRADE evidence to decision frameworks for tests in clinical practice and public health. *Journal of Clinical Epidemiology* 2016;**76**:89-98.

Stata 2011 [Computer program]

StataCorp. LP. Stata Statistical Software: Release 11. College Station (TX): StataCorp. LP, 2011.

Stoll 2002

Stoll BJ, Hansen N, Fanaroff AA, Wright LL, Carlo WA, Ehrenkranz RA, et al. Late-onset sepsis in very low birth weight neonates: the experience of the NICHD Neonatal Research Network. *Pediatrics* 2002;**110**(2 Pt 1):285-91. [PUBMED: 12165580]

Stoll 2004

Stoll BJ, Hansen NI, Adams-Chapman I, Fanaroff AA, Hintz SR, Vohr B, et al. Neurodevelopmental and growth impairment among extremely low-birth-weight infants with neonatal infection. *JAMA* 2004;**292**(19):2357-65. [PUBMED: 15547163]

Trenti 2016

Trenti T, Schunemann HJ, Plebani M. Developing GRADE outcome-based recommendations about diagnostic tests: a key role in laboratory medicine policies. *Clinical Chemistry and Laboratory Medicine* 2016;**54**(4):535-43.

Van Enst 2014

Van Enst WA, Ochodo E, Scholten RJPM, Hooft L, Leeflang MA. Investigation of publication bias in meta-analyses of diagnostic test accuracy: a meta-epidemiologic study. *BMC Medical Research Methodology* 2014;**14**:70. [PUBMED: 24884381]



Venkatesh 2010

Venkatesh M, Flores A, Luna RA, Versalovic J. Molecular microbiological methods in the diagnosis of neonatal sepsis. *Expert Reviews in Anti Infective Therapy* 2010;**8**(9):1037-48. [PUBMED: 20818947]

Verboon-Maciolek 2006

Verboon-Maciolek MA, Thijsen SF, Hemels MA, Menses M, van Loon AM, Krediet TG, et al. Inflammatory mediators for the diagnosis and treatment of sepsis in early infancy. *Pediatric Research* 2006;**59**(3):457-61. [PUBMED: 16492989]

CHARACTERISTICS OF STUDIES

Characteristics of included studies [ordered by study ID]

Whiting 2011

Whiting PF, Rutjes AW, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Annals of Internal Medicine* 2011;**155**(8):529-36. [PUBMED: 22007046]

Woese 1987

Woese CR. Bacterial evolution. *Microbiology Reviews* 1987;**51**(2):221-71. [PUBMED: 2439888]

Briones 2003

Study characteristics					
Patient sampling	Participant samplin	ig not clearly describ	ed.		
Patient characteristics and setting	Newborns > 3 days old with suspected sepsis. No information or participant demographics or study period.				
Index tests	PCR using universa	l candida DNA seque	nce.		
Target condition and reference standard(s)	Neonatal sepsis and	d blood culture.			
Flow and timing	Blood samples drav	vn at the same time.			
Comparative					
Notes	Data from conferen demographics or st		information on participant		
Methodological quality					
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns		
DOMAIN 1: Patient Selection					
Was a consecutive or random sample of patients enrolled?	Unclear				
Was a case-control design avoided?	Yes				
Did the study avoid inappropriate exclusions?	Yes				
		Unclear	Unclear		
DOMAIN 2: Index Test All tests					
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear				
If a threshold was used, was it pre-specified?	Unclear				

Molecular assays for the diagnosis of sepsis in neonates (Review)



		Unclear	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

Chan 2009

Study characteristics

Item	Authors' judgement	Risk of bias	Applicability con- cerns					
Methodological quality								
Notes	15 samples were excluded due to insufficient amount of sample (n = 9) and mistakenly left in the refrigerator for > 72 hours (n = 6). Excluded samples not included in the analysis. Cycle threshold cut-off values for positive PCR were defined. Interquartile range of age reported suggests some infants may have been > 28 days of age.							
Comparative								
Flow and timing	Index test and the refer	ence standard perfo	rmed at the same time.					
Target condition and reference standard(s)	Neonatal sepsis and blood, peritoneal fluid and CSF cultures.							
Index tests	Real-time PCR using universal primers and Gram-specific probes.							
Patient characteristics and setting	Preterm infants < 37 weeks and > 72 hours old with signs and symptoms of sepsis requiring antibiotic treatment. Interquartile range of age re- ported suggests some infants may have been > 28 days of age. Study period: March 2006 to June 2008 (28 months).							
Patient sampling	Participants were recru	ited consecutively.						

Molecular assays for the diagnosis of sepsis in neonates (Review)



Chan 2009 (Continued)			
Was a consecutive or random sample of patients en- rolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Low
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

Chen 2009

Study characteristics	
Patient sampling	Study did not classify whether participants were enrolled randomly or consecutively. Negative controls (n = 30) were not included in the analysis.
Patient characteristics and setting	Infants with suspected sepsis, admitted to the neonatal department and the intensive care unit of the Zhejiang University Children's Uni- versity in China. It was unclear how many infants were < 28 days old as no participant demographics are available. Study period: Septem- ber 2007 to June 2008.
Index tests	Broad-range 16S rRNA-based real-time fluorescent PCR.

Molecular assays for the diagnosis of sepsis in neonates (Review)



Chen 2009 (Continued)						
Target condition and reference standard(s)	Suspected sepsis and the reference standard were cultures of bloo and CSF. Both index test and reference standard samples were drawn simula neously.					
Flow and timing						
Comparative						
Notes	No participant demographics available and unclear if some infants were > 28 days of age.					
Methodological quality						
Item	Authors' judgement	Risk of bias	Applicability con- cerns			
DOMAIN 1: Patient Selection						
Was a consecutive or random sample of patients enrolled?	Unclear					
Was a case-control design avoided?	Yes					
Did the study avoid inappropriate exclusions?	Yes					
		Low	Unclear			
DOMAIN 2: Index Test All tests						
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear					
If a threshold was used, was it pre-specified?	Yes					
		Low	Low			
DOMAIN 3: Reference Standard						
Is the reference standards likely to correctly classify the tar- get condition?	Yes					
Were the reference standard results interpreted without knowledge of the results of the index tests?	Unclear					
		Low	Low			
DOMAIN 4: Flow and Timing						
Was there an appropriate interval between index test and reference standard?	Yes					
Did all patients receive the same reference standard?	Yes					
Were all patients included in the analysis?	Yes					
		Low				

Molecular assays for the diagnosis of sepsis in neonates (Review)



Draz 2013 **Study characteristics** Patient sampling All neonates with suspected sepsis admitted during the period of May 2012 to August 2012 were enrolled. Patient characteristics and setting Neonates with suspected sepsis admitted to the NICU of Ain Shams University Hospitals. Study period: May 2012 to August 2012. Age range reported was 0 to 50 days. Index tests Broad-range 16S rDNA PCR. Target condition and reference standard(s) Neonatal sepsis and blood culture. Flow and timing Blood sample for culture and PCR were collected concurrently using standard sterile procedures. Comparative Participants were referred to as neonates although the age range Notes reported was 0 to 50 days. Participants included both preterm and full-term infants. Methodological quality Applicability con-Item Authors' judge-**Risk of bias** ment cerns **DOMAIN 1: Patient Selection** Was a consecutive or random sample of patients enrolled? Yes Was a case-control design avoided? Yes Did the study avoid inappropriate exclusions? Yes Low Unclear **DOMAIN 2: Index Test All tests** Were the index test results interpreted without knowledge of Yes the results of the reference standard? If a threshold was used, was it pre-specified? Yes Low Low **DOMAIN 3: Reference Standard** Is the reference standards likely to correctly classify the target Yes condition? Were the reference standard results interpreted without knowl-Yes edge of the results of the index tests? Low Low

Molecular assays for the diagnosis of sepsis in neonates (Review)



Draz 2013 (Continued)

DOMAIN 4: Flow and Timing	
Was there an appropriate interval between index test and refer- ence standard?	Yes
Did all patients receive the same reference standard?	Yes
Were all patients included in the analysis?	Yes
	Low

Dutta 2009

Study characteristics		
Patient sampling	Not clearly reported.	
Patient characteristics and setting	Neonates with suspected sepsis admitted to Level III NICU. Study period not mentioned.	
Index tests	Broad-range conventional PCR after 5-hour preamplification cul- ture.	
Target condition and reference standard(s)	Neonatal sepsis and blood culture.	
Flow and timing	Blood samples for culture and PCR were drawn simultaneously. Reason for exclusion of participants were reported.	
Comparative		
Notes	Of the 64 participants that were excluded, 34 had malformations, 15 had < 12-hour life expectancy and the remaining 15 had conta- minated blood cultures. Study period not mentioned.	

Methodological quality

Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Low
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		



Low

Low

D

_

-

_

Dutta 2009 (Continued)		
If a threshold was used, was it pre-specified?	Yes	
		Low
DOMAIN 3: Reference Standard		
Is the reference standards likely to correctly classify the target condition?	Yes	
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear	
		Low
DOMAIN 4: Flow and Timing		
Was there an appropriate interval between index test and refer-	Ves	

	Low
Were all patients included in the analysis?	Yes
Did all patients receive the same reference standard?	Yes
ence standard?	res

Enomoto 2009

Infants were enrolled if they met the inclusion criteria during the study period. Controls (n = 50) were not included in the analysis.
Newborn participants with signs and history suggestive of sepsis mitted in the NICU at Kobe Hospital University from June 2005 to September 2006.
Multiplex PCR targeting 8 common pathogens.
Neonatal sepsis and bacterial culture of blood, skin, bronchoalve lavage, mucus, CSF, urine and ascitic fluid.
Only 77 samples with paired specimen culture and PCR were inclued of the set
Of the 6 specimens that were positive for PCR but negative for cul ture, 1 culture was positive for normal flora and was considered n ative.
Authors' judgement Risk of bias Applicability co cerns

Enomoto 2009 (Continued)

DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Low
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the tar- get condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	
Esparcia 2011			
Study characteristics			
Patient sampling	Infants were enrollec study period.	if they met the inclusio	on criteria during the
Patient characteristics and setting		d with suspected sepsi ing hospital from Nover	
Index tests	RT-PCR targeting the	16S rRNA.	

Molecular assays for the diagnosis of sepsis in neonates (Review)



Esparcia 2011 (Continued)			
Target condition and reference standard(s)	Suspected early-onset neonatal sepsis and blood and CSF cul- tures.		
Flow and timing	Sample for PCR and culture were drawn concurrently. Samples for PCR were stored until DNA extraction.		
Comparative			
Notes	Analyzed only EOS i	n neonates and inclue	ded 83 neonates.
Methodological quality			
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Low
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Unclear		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	



Fujimori 2010

Study characteristics				
Patient sampling	Neonates were enro study period.	olled if they met inclu	sion criteria during the	
Patient characteristics and setting	Jutendo Shizuoka H (SD) gestational age	Neonates admitted to the NICU of Jutendo University Hospital or Jutendo Shizuoka Hospital from February to August 2009. Mean (SD) gestational age was 34.8 ± 5.8 weeks. There were 36 partici- pants with 39 episodes of sepsis.		
Index tests	RT-PCR targeting 16	S rRNA.		
Target condition and reference standard(s)	Neonatal sepsis and	Neonatal sepsis and blood culture.		
Flow and timing	Whole blood collect	ted concurrently for F	PCR and culture.	
Comparative				
Notes				
Methodological quality				
ltem	Authors' judge- ment	Risk of bias	Applicability con cerns	
DOMAIN 1: Patient Selection				
Was a consecutive or random sample of patients enrolled?	Unclear			
Was a case-control design avoided?	Yes			
Did the study avoid inappropriate exclusions?	Yes			
		Low	Low	
DOMAIN 2: Index Test All tests				
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear			
If a threshold was used, was it pre-specified?	Yes			
		Low	Low	
DOMAIN 3: Reference Standard				
Is the reference standards likely to correctly classify the target condition?	Yes			
	l- Unclear			
Were the reference standard results interpreted without knowl edge of the results of the index tests?				

Fujimori 2010 (Continued)

Was there an appropriate interval between index test and refer- ence standard?	Yes
Did all patients receive the same reference standard?	Yes
Were all patients included in the analysis?	Yes
	Low

Garcia-Elorriaga 2012

Study characteristics				
Patient sampling	Neonates were enrolled if they met inclusion criteria during the study period.			
Patient characteristics and setting	Neonates up to 28 days old admitted to the NICU from August 2005 to July 2006.			
Index tests	Broad-range PCR.			
Target condition and reference standard(s)	Neonatal sepsis and	blood culture.		
Flow and timing	Index test and refere ously.	ence standard sampl	ing performed simultane-	
Comparative				
Notes	Only blood culture-p	positive samples wer	e included in the analysis.	
Methodological quality				
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns	
DOMAIN 1: Patient Selection				
Was a consecutive or random sample of patients enrolled?	Unclear			
Was a case-control design avoided?	No			
Did the study avoid inappropriate exclusions?	Yes			
		High	Low	
DOMAIN 2: Index Test All tests				
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes			
If a threshold was used, was it pre-specified?	Unclear			

Low

Low

Molecular assays for the diagnosis of sepsis in neonates (Review)

Garcia-Elorriaga 2012 (Continued)

DOMAIN 3: Reference Standard	
Is the reference standards likely to correctly classify the target condition?	Yes
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Yes
	Low Low
DOMAIN 4: Flow and Timing	
Was there an appropriate interval between index test and refer- ence standard?	Yes
Did all patients receive the same reference standard?	Yes
Were all patients included in the analysis?	Yes
	Low
Ibarra 2015	
Study characteristics	
Patient sampling	Participants who met the inclusion criteria were enrolled prospec- tively.
Patient characteristics and setting	Neonates with suspected clinical sepsis admitted to the Central South Hospital of Petroleos Mexicanos, the Gynecological-Obstet- rics Hospital number 4 of the Mexican Institute of Social Securi- ty, the Dalinde Hospital and the Monterrey Nuevo Leon University Hospital and National Institute of Perinatology. Study period not mentioned.
Index tests	LightCycler SeptiFast Test.
Target condition and reference standard(s)	Suspected neonatal sepsis and blood culture.
Flow and timing	Samples for blood culture and LightCycler SeptiFast were drawn concurrently.
Comparative	
Notes	Study period not mentioned in the report.
Methodological quality	

DOMAIN 1: Patient Selection

Item

Was a consecutive or random sample of patients enrolled?

Yes

ment

Authors' judge-

Risk of bias

Molecular assays for the diagnosis of sepsis in neonates (Review)

Copyright @ 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

Applicability con-

cerns



Ibarra 2015 (Continued)			
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Low
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Yes		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	
Jordan 2000			
Study characteristics			
Patient sampling	All infants admitted to the NICU for sepsis evaluation.		
Patient characteristics and setting	All infants admitted to the NICU for sepsis evaluation. No participant demographics available.		
Index tests	Broad-range co	onventional PCR and D	NA dot-blot hybridization.
Target condition and reference standard(s)	Neonatal seps	s and blood culture.	
Flow and timing	Index test and ly.	reference standard we	ere performed simultaneous-

Comparative



Jordan 2000 (Continued)

Notes

This was a feasibility study and blood sample for PCR was from discarded or unused sample sent to evaluate CBCs. It was not clear whether blood drawn for CBC was also done with the same aseptic technique as blood culture. Study period not mentioned.

Methodological quality

Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	High
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Unclear	

Jordan 2005a

Study characteristics



ordan 2005a (Continued) Patient sampling	Infants were enrolled if	they met inclusion of	criteria.	
	Infants were enrolled if they met inclusion criteria.			
Patient characteristics and setting	Infant admitted to the NICU for sepsis evaluation that included at least blood culture and CBC. No demographic information or study period details available.			
Index tests	Real-time 16S rRNA PCF	۶.		
Target condition and reference standard(s)	Neonatal sepsis and blo	ood culture.		
Flow and timing		BC. Unclear whethe	rded or unused samples r blood drawn for CBC was	
Comparative				
Notes	eliminate tryptic soy br ventional PCR assay to here is real-time PCR f	oth pre-enrichment a real-time PCR plat rom whole blood w	ation protocol that would step and to convert con- form. The methodology vithout enrichment. So a paper and overlap is very	
Methodological quality				
Item	Authors' judgement	Risk of bias	Applicability con- cerns	
DOMAIN 1: Patient Selection				
Was a consecutive or random sample of patients enrolled?	Unclear			
Was a case-control design avoided?	Yes			
Did the study avoid inappropriate exclusions?	Yes			
		Low	Unclear	
DOMAIN 2: Index Test All tests				
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear			
If a threshold was used, was it pre-specified?	Yes			
		Low	High	
DOMAIN 3: Reference Standard				
Is the reference standards likely to correctly classify the tar- get condition?	Yes			
Were the reference standard results interpreted without knowledge of the results of the index tests?	Unclear			
		Low	Low	

Molecular assays for the diagnosis of sepsis in neonates (Review)



Jordan 2005a (Continued)

DOMAIN 4: Flow and Timing	
Was there an appropriate interval between index test and reference standard?	Yes
Did all patients receive the same reference standard?	Yes
Were all patients included in the analysis?	Yes
	Unclear

Jordan 2006

Study characteristics			
Patient sampling	All NICU admissions dur ty.	ing the period of stud	y were screened for eligibili-
Patient characteristics and setting	Infants > 34 weeks adm September 2000 to 1 Ap		uspected EOS from 1
Index tests	Broad-range conventior	nal PCR followed by py	rosequencing.
Target condition and reference standard(s)	EOS in near-term infant	s and blood culture.	
Flow and timing	•	aluated from sample s	dard were collected simulta- sent for CBC. Concerns about
Comparative			
Notes	evaluate CBC and were was conventional PCR PCR just like the paper to be from September	collected by venous, a with enrichment wit Jordan 2000 . The stu 2000. Jordan 2000 pa the title page of the a	tion of the sample sent to arteria or heel stick. The PCR th Trypticose soy before udy period here was stated aper was submitted for pub- article and hence overlap of
Methodological quality			
Item	Authors' judgement	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients en- rolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		

Low

Molecular assays for the diagnosis of sepsis in neonates (Review)

Copyright @ 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

Low



Jordan 2006 (Continued)

DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowl- edge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	High
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted with- out knowledge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Unclear	

Kasper 2013

Study characteristics	
Patient sampling	Neonates who met inclusion criteria were enrolled on admission.
Patient characteristics and setting	VLBW infants > 72 hours old. Participant demographics or study period not available.
Index tests	Multiplex real-time PCR using Roche LightCycler SeptiFast MGRADE system.
Target condition and reference standard(s)	Neonates with suspected LOS and blood culture.
Flow and timing	Blood sample for PCR was collected during routine sepsis work-up and before antibiotics.
Comparative	
Notes	Participant demographics or study period not available.
Methodological quality	



Kasper 2013 (Continued)

Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	
Laforgia 1997			
Study characteristics			
Patient sampling	Neonates were enrolled if they met inclusion criteria during the study period.		
Patient characteristics and setting			September 1996. Prede- to classify participants

Molecular assays for the diagnosis of sepsis in neonates (Review)



Laforgia 1997 (Continued)			
Index tests	Broad-range conventional PCR		
Target condition and reference standard(s)	Neonatal EOS and blood culture.		
Flow and timing	Blood samples for analyses were drawn concurrently.		
Comparative			
Notes			
Methodological quality			
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Low
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	



Lima 2007

Flow and timing Blood samples for PCR and cultu Comparative Recalculated sensitivity, specifici Notes Recalculated sensitivity, specifici Methodological quality Item Item Authors' judge- ment Risk of bi DOMAIN 1: Patient Selection Unclear Was a consecutive or random sample of patients enrolled? Unclear Did the study avoid inappropriate exclusions? Yes DOMAIN 2: Index Test All tests Unclear Were the index test results interpreted without knowledge of the results of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Yes DOMAIN 3: Reference Standard Yes Us the reference standards likely to correctly classify the target condition? Yes Were the reference standard results interpreted without knowledge of Unclear	et inclusion criteria during the
Target condition and reference standard(s) Neonatal sepsis and blood cultur Flow and timing Blood samples for PCR and cultur Comparative Recalculated sensitivity, specifici itive for PCR were also positive for DNA. Participant demographics of DNA. Participant demographics of Methodological quality Item Authors' judge-ment Methodological quality Risk of bi ment DOMAIN 1: Patient Selection Unclear Was a consecutive or random sample of patients enrolled? Unclear Did the study avoid inappropriate exclusions? Yes DOMAIN 2: Index Test All tests Low DOMAIN 2: Index test results interpreted without knowledge of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Yes DOMAIN 3: Reference Standard Yes Were the reference standards likely to correctly classify the target condition? Yes	
Flow and timing Blood samples for PCR and cultu Comparative Recalculated sensitivity, specifici Notes Recalculated sensitivity, specifici Methodological quality DNA. Participant demographics r Methodological quality Authors' judge- ment Risk of bi DOMAIN 1: Patient Selection Unclear Vas a consecutive or random sample of patients enrolled? Unclear Was a case-control design avoided? Yes Ves Ves Did the study avoid inappropriate exclusions? Yes Low DOMAIN 2: Index Test All tests Unclear Unclear Ware the index test results interpreted without knowledge of the reference standard? Unclear Unclear DOMAIN 3: Reference Standard Unclear Unclear Unclear DOMAIN 3: Reference Standard Yes Unclear Unclear Were the reference standards likely to correctly classify the target condition? Yes Unclear	mers.
Comparative Notes Recalculated sensitivity, specificitive for PCR were also positive for DNA. Participant demographics of DNA. Participant demographics of ment Methodological quality Authors' judge-ment Item Authors' judge-ment DOMAIN 1: Patient Selection Was a consecutive or random sample of patients enrolled? Was a case-control design avoided? Yes Did the study avoid inappropriate exclusions? Yes DOMAIN 2: Index Test All tests Low DOMAIN 2: Index test results interpreted without knowledge of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Yes DOMAIN 3: Reference standards likely to correctly classify the target condition? Yes	2.
Notes Recalculated sensitivity, specific itive for PCR were also positive fo DNA. Participant demographics r Methodological quality Authors' judge- ment Risk of bi Item Authors' judge- ment Risk of bi DOMAIN 1: Patient Selection Unclear Was a consecutive or random sample of patients enrolled? Unclear Was a case-control design avoided? Yes Did the study avoid inappropriate exclusions? Yes DOMAIN 2: Index Test All tests Low DOMAIN 2: Index test results interpreted without knowledge of the results of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Yes DOMAIN 3: Reference standard results interpreted without knowl- uoriear Yes	e were drawn concurrently.
itive for PCR were also positive for DNA. Participant demographics r Methodological quality Item Authors' judge-ment Nas a consecutive or random sample of patients enrolled? Unclear Was a consecutive or random sample of patients enrolled? Unclear Was a case-control design avoided? Yes Did the study avoid inappropriate exclusions? Yes Low DOMAIN 2: Index Test All tests Were the index test results interpreted without knowledge of the results of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Patients Is the reference standards likely to correctly classify the target condition? Yes	
Item Authors' judge-ment Risk of biment DOMAIN 1: Patient Selection	r human DNA and not bacterial
ment DOMAIN 1: Patient Selection Was a consecutive or random sample of patients enrolled? Unclear Was a case-control design avoided? Yes Did the study avoid inappropriate exclusions? Yes Did the study avoid inappropriate exclusions? Yes DOMAIN 2: Index Test All tests Low DOMAIN 2: Index Test All tests Unclear Were the index test results interpreted without knowledge of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Unclear Is the reference standards likely to correctly classify the target condition? Yes Were the reference standard results interpreted without knowl- Unclear	
Was a consecutive or random sample of patients enrolled? Unclear Was a case-control design avoided? Yes Did the study avoid inappropriate exclusions? Yes Domain 2: Index Test All tests Low DOMAIN 2: Index Test All tests Unclear Were the index test results interpreted without knowledge of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Unclear Is the reference standards likely to correctly classify the target condition? Yes Were the reference standard results interpreted without knowl- Unclear	as Applicability con- cerns
Was a case-control design avoided? Yes Did the study avoid inappropriate exclusions? Yes Low DOMAIN 2: Index Test All tests Were the index test results interpreted without knowledge of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Unclear Is the reference standards likely to correctly classify the target condition? Yes Were the reference standard results interpreted without knowl- Unclear	
Did the study avoid inappropriate exclusions? Yes Low DOMAIN 2: Index Test All tests Were the index test results interpreted without knowledge of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Unclear Is the reference standards likely to correctly classify the target condition? Yes Were the reference standard results interpreted without knowl- Unclear	
Low DOMAIN 2: Index Test All tests Were the index test results interpreted without knowledge of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Unclear Is the reference standards likely to correctly classify the target condition? Yes Were the reference standard results interpreted without knowl- Unclear	
DOMAIN 2: Index Test All tests Were the index test results interpreted without knowledge of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear DOMAIN 3: Reference Standard Unclear Is the reference standards likely to correctly classify the target condition? Yes Were the reference standard results interpreted without knowl- Unclear	
Were the index test results interpreted without knowledge of the reference standard? Unclear If a threshold was used, was it pre-specified? Unclear Unclear Unclear DOMAIN 3: Reference Standard Is the reference standards likely to correctly classify the target condition? Were the reference standard results interpreted without knowl- Unclear	Unclear
the results of the reference standard? If a threshold was used, was it pre-specified? Unclear Unclear DOMAIN 3: Reference Standard Is the reference standards likely to correctly classify the target Yes condition? Were the reference standard results interpreted without knowl- Unclear	
Unclear DOMAIN 3: Reference Standard Is the reference standards likely to correctly classify the target condition? Were the reference standard results interpreted without knowl- Unclear	
DOMAIN 3: Reference Standard Is the reference standards likely to correctly classify the target condition? Were the reference standard results interpreted without knowl- Unclear	
Is the reference standards likely to correctly classify the target Yes condition? Were the reference standard results interpreted without knowl- Unclear	Low
condition? Were the reference standard results interpreted without knowl- Unclear	
edge of the results of the index tests?	
Low	Low



Lima 2007 (Continued)

Was there an appropriate interval between index test and refer- ence standard?	Yes
Did all patients receive the same reference standard?	Yes
Were all patients included in the analysis?	Yes

Low

Liu 2014

Study characteristics	
Patient sampling	All neonates with suspected sepsis and had blood samples drawn for concomitant culture, CBC and CRP assay were included in the study.
Patient characteristics and setting	Neonates with suspected sepsis admitted to the NICU of the Women and Children's Hospital, the Children's Hospital and Tongji Hospital in Hubei Province from 1 September 2011 to 31 De- cember 2011. Participants were from 4 hour to 28 days old.
Index tests	16S rRNA gene PCR.
Target condition and reference standard(s)	Neonatal sepsis and blood culture.
Flow and timing	Additional 0.5 mL to 1 mL EDTA blood sample was collected for PCR at the time of sepsis workup.
Comparative	
Notes	

Methodological quality

Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Low
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		

Molecular assays for the diagnosis of sepsis in neonates (Review)



		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Yes		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	
Makhoul 2005			
Study characteristics			
Patient sampling	Prospective enrollmening a 12-month period	nt of infants that met in I.	clusion criteria dur-
Patient characteristics and setting	Neonates aged > 3 days, admitted to the NICU with suspected LOS. Gestational age range 24 to 42 weeks and range of age at en- rollment was 4 to 96 days. Study period not mentioned although reported over 12 months.		
Index tests	Staphylococcal 16S rR coagulase-negative St	RNA PCR (both <i>Staphylo</i> caphylococcus).	coccus aureus and
Target condition and reference standard(s)	Neonatal LOS and blo	od culture.	
Flow and timing	Blood samples for PCR and culture were drawn concurrently.		
Comparative			
Notes	There were 32 culture-positive samples for bacteria and fungi but only 13 were positive for staphylococci and this was incorporated into the analysis.		
Methodological quality			
ltem	Authors' judge- ment	Risk of bias	Applicability con- cerns

DOMAIN 1: Patient Selection

Molecular assays for the diagnosis of sepsis in neonates (Review)

Makhoul 2005 (Continued)			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

Makhoul 2006

Study characteristics	
Patient sampling	Prospective enrollment of neonates that met the criteria for suspected LOS.
Patient characteristics and setting	Neonates aged > 3 days with suspected LOS. The age range of in- fants included were 4 to 105 days. Study period not available.
Index tests	Staphylococcal 16S rRNA PCR (both <i>Staphylococcus aureus</i> and coagulase-negative Staphylococci).
Target condition and reference standard(s)	Neonates with suspected LOS and blood culture.
Flow and timing	Blood samples for PCR and culture were drawn concurrently.

Molecular assays for the diagnosis of sepsis in neonates (Review)



Makhoul 2006 (Continued)

Comparative			
Notes	The article mentioned 148 events of LOS but on further scrutiny there were on 146 events which were incorporated into the analy sis.		
Methodological quality			
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	No		
		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Unclear		
		Low	

Ohlin 2008

Study characteristics



ohlin 2008 (Continued)				
Patient sampling		t met inclusion criter ing the period of 1999	ia for EOS and LOS admit) to 2005.	
Patient characteristics and setting		Newborn infants < 28 days old with suspected EOS or LOS admit- ted to Öbrero University from 1999 to 2005.		
Index tests	Real-time PCR targe	eting 16S rRNA.		
Target condition and reference standard(s)	Neonates with susp	Neonates with suspected EOS or LOS and blood culture.		
Flow and timing	Blood samples for PCR and culture were drawn simultaneously.			
Comparative				
Notes			ive for culture and PCR esult showed double se-	
Methodological quality				
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns	
DOMAIN 1: Patient Selection				
Was a consecutive or random sample of patients enrolled?	Yes			
Was a case-control design avoided?	Yes			
Did the study avoid inappropriate exclusions?	Yes			
		Low	Low	
DOMAIN 2: Index Test All tests				
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes			
If a threshold was used, was it pre-specified?	Yes			
		Low	Low	
DOMAIN 3: Reference Standard				
Is the reference standards likely to correctly classify the target condition?	Yes			
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear			
		Low	Low	
DOMAIN 4: Flow and Timing				
Was there an appropriate interval between index test and reference standard?	Yes			



Ohlin 2008 (Continued)		
Did all patients receive the same reference standard?	Yes	
Were all patients included in the analysis?	Yes	
		Low

Study characteristics			
Patient sampling	All infants that met	inclusion criteria wer	e enrolled prospectively
Patient characteristics and setting	admitted to the NIC October 2007 and N	CU at 2 Swedish Unive	ent sepsis evaluation and rsity Hospitals between e participants enrolled ir
Index tests	Broad-range 16S re	al-time PCR.	
Target condition and reference standard(s)	Suspected sepsis a	nd blood culture.	
Flow and timing	Blood samples for PCR and culture were drawn simultaneously.		
Comparative			
Notes	older than 3 month PCR and culture we	s and 10 participants	k of consent, 7 for being whose blood sample for ently. Excluded partici-
Methodological quality			
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		

DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low

Molecular assays for the diagnosis of sepsis in neonates (Review)

nun 2012 (Continued)			
DOMAIN 3: Reference Standard			
s the reference standards likely to correctly classify the target condition?	Yes		
Nere the reference standard results interpreted without knowledge of the results of the index tests?	Unclear		
	Low Low		
DOMAIN 4: Flow and Timing			
Nas there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Nere all patients included in the analysis?	Yes		
	Low		
nolucci 2009			
Study characteristics			
Patient sampling	34 newborns with LOS were enrolled in the study.		
Patient characteristics and setting	Newborns > 3 days old with suspected LOS. Age of participants at enrollment and study period not available.		
ndex tests	Commercial real-time PCR using LightCycler SeptiFast system (multiplex PCR).		
Farget condition and reference standard(s)	Neonatal LOS and blood culture.		
Flow and timing	Blood samples for LightCycler SeptiFast and culture were simulta neously.		
Comparative			
Notes	Age of participants at enrollment and study period not available.		
Methodological quality			
tem	Authors' judge- Risk of bias Applicability con- ment cerns		
DOMAIN 1: Patient Selection			
	Unclear		
Nas a consecutive or random sample of patients enrolled?			
Nas a consecutive or random sample of patients enrolled? Nas a case-control design avoided?	Yes		

Molecular assays for the diagnosis of sepsis in neonates (Review)



Paolucci 2009 (Continued)

Trusted evidence. Informed decisions. Better health.

		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

Reier-Nilsen	2009
--------------	------

Study characteristics	
Patient sampling	Prospective, non-randomized enrollment of participants that met inclusion criteria.
Patient characteristics and setting	Infants with birth weight > 1000 g admitted to the NICU at Akershus University Hospital with suspected sepsis during the first week of life. Age at study enrollment and study period not mentioned.
Index tests	Broad-range 16S rRNA PCR followed by sequencing.
Target condition and reference standard(s)	Suspected neonatal sepsis and blood culture.
Flow and timing	Blood samples for PCR and culture were drawn concurrently.
Comparative	
Notes	PCR samples were stored until analysis. 4 infants were excluded from the study with 3 having incomplete registration and 1 with missing sample. 1 infant in the final analysis ended up with a diag-

Molecular assays for the diagnosis of sepsis in neonates (Review)



Reier-Nilsen 2009 (Continued)

nosis of asphyxia rather than sepsis. Age at study enrollment and study period not mentioned.

Methodological quality	Meth	odolo	ogical	qua	lity
------------------------	------	-------	--------	-----	------

Item	Authors' judgement	Risk of bias	Applicability con cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Yes		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

Shaat 2013

Study characteristics

Patient sampling

Neonates with clinically suspected sepsis.

Molecular assays for the diagnosis of sepsis in neonates (Review)



Trusted evidence. Informed decisions. Better health.

Shaat 2013 (Continued)			
Patient characteristics and setting	Neonates with suspected sepsis. The gestational age ranged from 26 to 39 weeks but age at enrollment not mentioned. Study peri- od: October 2010 to December 2012.		
Index tests	16S rDNA PCR.		
Target condition and reference standard(s)	Neonatal sepsis and	d blood culture.	
Flow and timing	Blood samples for bously.	plood culture and PCF	R were done simultane-
Comparative			
Notes	Age at enrollment r	not available.	
Methodological quality			
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Unclear		
		Unclear	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Unclear	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		

Did all patients receive the same reference standard?

Yes



Shaat 2013 (Continued)

Were all patients included in the analysis?

Yes

Low

Study characteristics			
Patient sampling		inclusion criteria duri excluded from analy	ng a specified period of sis.
Patient characteristics and setting	who developed clin	ical signs of LOS duri	neonatal ward or NICU ng the period of 1 January demographics not avail-
Index tests	Broad-range 16S rR	NA PCR followed by n	nicroarray hybridization.
Target condition and reference standard(s)	Suspected neonata	l LOS and blood cultu	re.
Flow and timing	Unclear whether blo drawn simultaneou		and blood culture were
Comparative			
Notes	Participant demogr	aphics not available.	
Methodological quality			
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		



Shang 2005 (Continued)

Is the reference standards likely to correctly classify the target Yes condition?

Were the reference standard results interpreted without knowl- Unclear edge of the results of the index tests?

		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Unclear		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

Taira 2014

Study characteristics				
Patient sampling	Consecutive enrollment of infants (24 were neonates) with signs of systemic inflammatory response syndrome and risk factors for candidemia.			
Patient characteristics and setting	Infants who were admitted to the ICU of 2 pediatric hospital in Sac Paulo State, Brazil over an 18-month period. Study period (month and year) or participant demographics not available. Author pro- vided results for the 24 neonates.			
Index tests	Multiplex nested PC <i>Candida</i> species	Multiplex nested PCR with specific primers designed to identify 7 <i>Candida</i> species		
Target condition and reference standard(s)	Candidemia and blood culture.			
Flow and timing	Blood sample for both culture and PCR were done concurrently.			
Comparative				
Notes	Data based on ema	Data based on email communication with Dr. Del Negro.		
Methodological quality				
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns	
DOMAIN 1: Patient Selection				
Was a consecutive or random sample of patients enrolled?	Yes			
Was a case-control design avoided?	Yes			
was a case-control design avoided?	Yes			

Molecular assays for the diagnosis of sepsis in neonates (Review)



Taira 2014 (Continued)

Did the study avoid inappropriate exclusions?

		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Yes		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

Yes

Tirodker 2003

Study characteristics	
Patient sampling	All infants with suspected sepsis in the NICU and PICU during the study period were considered for inclusion in the study.
Patient characteristics and setting	Infants admitted in the NICU (n = 46) and PICU (n = 17) with suspected sepsis during the period from November 1999 to November 2000. PCR and blood culture data separately for neonates not available.
Index tests	Fungal conventional PCR targeting 18S rRNA.
Target condition and reference standard(s)	Suspected sepsis and blood culture.
Flow and timing	Excess blood used for culture was used for PCR.
Comparative	
Notes	PCR and blood culture data separately for neonates not available. It was unclear how many of the infants admitted in the PICU were

Molecular assays for the diagnosis of sepsis in neonates (Review)



Tirodker 2003 (Continued)

neonates hence, not all infants may have met the target condition of neonatal sepsis defined in this study. PCR products were analyzed by 2 independent observers blinded to blood culture results and participant information.

Item	Authors' judgement	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	High
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the tar- get condition?	Yes		
Were the reference standard results interpreted without knowledge of the results of the index tests?	Unclear		
		Low	High
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and reference standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

Tong 2004

Study characteristics



ong 2004 (Continued)				
Patient sampling	Study data derived from conference abstract only and hence limited.			
Patient characteristics and setting	Neonates with suspected sepsis. No participant demographics or study period details available.			
Index tests	16S rRNA-based PCR followed by hybridization to chips with 18 probes.			
Target condition and reference standard(s)	Infants with suspec	ted sepsis and blood	culture.	
Flow and timing	Possible simultaned dard.	ous sampling for inde	ex test and reference star	
Comparative				
Notes	Limited information or study period deta		articipant demographics	
Methodological quality				
Item	Authors' judge- ment	Risk of bias	Applicability con cerns	
DOMAIN 1: Patient Selection				
Was a consecutive or random sample of patients enrolled?	Unclear			
Was a case-control design avoided?	Yes			
Did the study avoid inappropriate exclusions?	Unclear			
		Unclear	Unclear	
DOMAIN 2: Index Test All tests				
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear			
If a threshold was used, was it pre-specified?	Unclear			
		Unclear	Low	
DOMAIN 3: Reference Standard				
Is the reference standards likely to correctly classify the target condition?	Yes			
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear			
		Low	Low	
DOMAIN 4: Flow and Timing				
Was there an appropriate interval between index test and refer- ence standard?	Unclear			



Tong 2004 (Continued)			
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	
Torres-Martos 2013			
Study characteristics			
Patient sampling	Participants who m tively.	et inclusion criteria we	ere admitted consecu-
Patient characteristics and setting	tal Universitario Virg April 2009. Participa	gen de las Nieves. Stud ints enrolled in the stu owever, age of particip	the NICU at the Hospi- dy period: April 2007 to Idy were both preterm pants at the time of en-
Index tests	LightCycler SeptiFast Assay.		
Target condition and reference standard(s)	Neonatal sepsis and blood culture.		
Flow and timing	Sample for blood culture and LightCycler SeptiFast assay were collected at the same time.		
Comparative			
Notes		of participants at the	oth preterm and term in- time of enrollment range
Methodological quality			
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	High
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		

If a threshold was used, was it pre-specified?

Unclear

Molecular assays for the diagnosis of sepsis in neonates (Review)



Torres-Martos 2013 (Continued)

DOMAIN 3: Reference Standard				
Is the reference standards likely to correctly classify the target condition?	Yes			
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Yes			
		Low	Low	
DOMAIN 4: Flow and Timing				
Was there an appropriate interval between index test and refer- ence standard?	Yes			
Did all patients receive the same reference standard?	Yes			
Were all patients included in the analysis?	Yes			
		Low		
rovato 2012				
rovato 2012				
Study characteristics				
	Only participants wit study.	h probable can	didiasis were included	in the
Study characteristics	study. 	for invasive ca	didiasis were included ndidiasis from Jan 2009 demographics availab	9 to Dec
Study characteristics Patient sampling	study. Neonates at high risk 2010. No information Detection of fungal D culture. Fungus-spec	for invasive can on participant NA directly fron ific universal pr	ndidiasis from Jan 2009	9 to Dec le. blood re used
Study characteristics Patient sampling Patient characteristics and setting	study. Neonates at high risk 2010. No information Detection of fungal D culture. Fungus-spec to amplify 18S rDNA,	for invasive can on participant DNA directly fror ific universal pr the adjacent IT	ndidiasis from Jan 2009 demographics availab m lysis-centrifugation b rimer ITS1 and ITS2 wer S1 and a small portion	9 to Dec le. blood re used
Study characteristics Patient sampling Patient characteristics and setting Index tests	study. Neonates at high risk 2010. No information Detection of fungal D culture. Fungus-spec to amplify 18S rDNA, 28S rDNA region. Suspected neonatal	for invasive can on participant ONA directly fror ific universal pr the adjacent IT candidiasis and	ndidiasis from Jan 2009 demographics availab m lysis-centrifugation b rimer ITS1 and ITS2 wer S1 and a small portion	9 to Dec le. Dlood re used of the
Study characteristics Patient sampling Patient characteristics and setting Index tests Target condition and reference standard(s)	study. Neonates at high risk 2010. No information Detection of fungal D culture. Fungus-spec to amplify 18S rDNA, 28S rDNA region. Suspected neonatal Blood samples for PC	for invasive can on participant ONA directly fror ific universal pr the adjacent IT candidiasis and	ndidiasis from Jan 2009 demographics availab n lysis-centrifugation b rimer ITS1 and ITS2 wer S1 and a small portion	9 to Dec le. Dlood re used of the
Study characteristics Patient sampling Patient characteristics and setting Index tests Target condition and reference standard(s) Flow and timing	study. Neonates at high risk 2010. No information Detection of fungal D culture. Fungus-spec to amplify 18S rDNA, 28S rDNA region. Suspected neonatal Blood samples for PC	for invasive can on participant NA directly from ific universal pr the adjacent IT candidiasis and CR and culture c	ndidiasis from Jan 2009 demographics availab m lysis-centrifugation b rimer ITS1 and ITS2 wer S1 and a small portion blood culture.	9 to Dec le. Dlood re used of the
Study characteristics Patient sampling Patient characteristics and setting Index tests Target condition and reference standard(s) Flow and timing Comparative	study. Neonates at high risk 2010. No information Detection of fungal D culture. Fungus-spec to amplify 18S rDNA, 28S rDNA region. Suspected neonatal Blood samples for PC 1.5 microbial tubes.	for invasive can on participant NA directly from ific universal pr the adjacent IT candidiasis and CR and culture c	ndidiasis from Jan 2009 demographics availab m lysis-centrifugation b rimer ITS1 and ITS2 wer S1 and a small portion blood culture.	9 to Dec le. blood re used of the

DOMAIN 1: Patient Selection

Unclear
Low
Low

Van der Brand 2014

Study characteristics	
Patient sampling	Consecutive enrollment of preterm infants with suspected LOS.
Patient characteristics and setting	Preterm infants with suspected LOS admitted to the NICU. Participant demographics or study period not mentioned.
Index tests	Multiplex real-time PCR assay.
Target condition and reference standard(s)	LOS in neonates and blood culture.
Flow and timing	Blood samples for culture and PCR were drawn concurrently.
Comparative	

Molecular assays for the diagnosis of sepsis in neonates (Review)



Van der Brand 2014 (Continued)

Notes

Participant demographics or study period not available.

Methodological quality

Item	Authors' judge- ment	Risk of bias	Applicability con- cerns
DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Yes		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Unclear
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Yes		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	
Villanueva-Uy 2003			

Study characteristics

Patient sampling

Limited information from abstract.

Molecular assays for the diagnosis of sepsis in neonates (Review)

Trusted evidence. Informed decisions. Better health.

Villanueva-Uy 2003 (Continued)

Patient characteristics and setting	Newborns aged > 3 days with suspected LOS. Participant demo- graphics or study period data not available.			
Index tests	Broad-range 16S rRNA conventional PCR.			
Target condition and reference standard(s)	Neonatal LOS and blood culture.			
Flow and timing	Blood samples for PCR and culture were drawn concurrently.			
Comparative				
Notes	Study data derived from abstract only. Participant demographics or study period data not available.			
Methodological quality				
ltem	Authors' judge- ment	Risk of bias	Applicability con- cerns	
DOMAIN 1: Patient Selection				
Was a consecutive or random sample of patients enrolled?	Unclear			
Was a case-control design avoided?	Yes			
Did the study avoid inappropriate exclusions?	Yes			
		Low	Unclear	
DOMAIN 2: Index Test All tests				
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear			
If a threshold was used, was it pre-specified?	Unclear			
		Unclear	Low	
DOMAIN 3: Reference Standard				
Is the reference standards likely to correctly classify the target condition?	Yes			
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear			
		Low	Low	
DOMAIN 4: Flow and Timing				
Was there an appropriate interval between index test and refer- ence standard?	Yes			
Did all patients receive the same reference standard?	Yes			
Were all patients included in the analysis?	Yes			

Molecular assays for the diagnosis of sepsis in neonates (Review)



Villanueva-Uy 2003 (Continued)

Low

Study characteristics				
Patient sampling	Limited information from abstract. Controls not included in the analysis.			
Patient characteristics and setting	Newborns with suspected sepsis admitted to the neonatal ward or NICU. Participant demographics or study period data not avail- able.			
Index tests	Real-time PCR targeting 16S rRNA.			
Target condition and reference standard(s)	Neonatal sepsis and blood culture.			
Flow and timing	Blood samples were tested for routine culture and PCR separately There was no mention if blood sample was drawn simultaneously.			
Comparative				
Notes	Abstract only. Participant demographics or study period data not available.			
Methodological quality				
Item	Authors' judge- ment	Risk of bias	Applicability con- cerns	
DOMAIN 1: Patient Selection				
Was a consecutive or random sample of patients enrolled?	Unclear			
Was a case-control design avoided?	Yes			
Did the study avoid inappropriate exclusions?	Yes			
		Low	Unclear	
DOMAIN 2: Index Test All tests				
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear			
If a threshold was used, was it pre-specified?	Yes			
		Low	Low	
DOMAIN 3: Reference Standard				
Is the reference standards likely to correctly classify the target condition?	Yes			



Wu 2007 (Continued)

Were the reference standard results interpreted without knowl- Unclear edge of the results of the index tests?

		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Unclear		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

Wu 2008 **Study characteristics** Patient sampling Neonates who met inclusion criteria during the study period were enrolled. Controls were not included in the analysis. Patient characteristics and setting Neonates aged 1 to 28 days with suspected sepsis admitted to the neonatal ward and NICU of Zhejiang University Children's Hospital from January 2005 to January 2007. 108 of the participants were preterm infants. Index tests Real-time PCR with Gram-specific probes followed by sequencing. Target condition and reference standard(s) Suspected neonatal EOS and LOS and blood culture. Flow and timing PCR and culture were done simultaneously. Unclear if samples were concurrently. Comparative Notes **Methodological quality** Item Authors' judge-**Risk of bias** Applicability conment cerns

DOMAIN 1: Patient Selecti	on
---------------------------	----

Was a consecutive or random sample of patients enrolled?	Unclear			
Was a case-control design avoided?	Yes			
Did the study avoid inappropriate exclusions?	Yes			
		Low	Low	

Molecular assays for the diagnosis of sepsis in neonates (Review)



Wu 2008 (Continued)

DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Yes		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Yes		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Unclear		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	
Yadav 2005 Study characteristics			
Patient sampling	Infants were enrolled	if they met inclusion cr	iteria.
Patient characteristics and setting	Infants < 7 days old w NICU. Study period d	vith suspected sepsis ad etails not available.	mitted to a level II
Index tests	Broad-range 16S rRN	A PCR.	
Target condition and reference standard(s)	Suspected neonatal	sepsis and blood culture	2.
Flow and timing	Blood samples for PC	R and culture were draw	wn concurrently.

Comparative

Notes

Methodological quality

ltem	Authors' judge- ment	Risk of bias	Applicability con- cerns

Study period details not available.

Molecular assays for the diagnosis of sepsis in neonates (Review)

Copyright @ 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

Yadav 2005 (Continued)

DOMAIN 1: Patient Selection			
Was a consecutive or random sample of patients enrolled?	Unclear		
Was a case-control design avoided?	Yes		
Did the study avoid inappropriate exclusions?	Yes		
		Low	Low
DOMAIN 2: Index Test All tests			
Were the index test results interpreted without knowledge of the results of the reference standard?	Unclear		
If a threshold was used, was it pre-specified?	Yes		
		Low	Low
DOMAIN 3: Reference Standard			
Is the reference standards likely to correctly classify the target condition?	Yes		
Were the reference standard results interpreted without knowl- edge of the results of the index tests?	Unclear		
		Low	Low
DOMAIN 4: Flow and Timing			
Was there an appropriate interval between index test and refer- ence standard?	Yes		
Did all patients receive the same reference standard?	Yes		
Were all patients included in the analysis?	Yes		
		Low	

CBC: complete blood count; CSF: cerebrospinal fluid; EDTA: ethylenediaminetetraacetic acid; EOS: early-onset sepsis; LOS: late-onset sepsis; n: number of participants; NICU: neonatal intensive care unit; NPV: negative predictive value; PCR: polymerase chain reaction; PICU: pediatric intensive care unit; PPV: positive predictive value; rDNA: ribosomal DNA; rRNA: ribosomal ribonucleic acid; RT-PCR: real-time polymerase chain reaction; SD: standard deviation; VLBW: very low birth weight.

Characteristics of excluded studies [ordered by study ID]

Study	Reason for exclusion
Chiba 2009	All samples (CSF) were positive by culture for bacterial meningitis and not in the context of suspected infection.
Das 2015	Urine instead of blood sample was used for broad-range 16S rDNA in detecting neonatal sep- ticemia.

Molecular assays for the diagnosis of sepsis in neonates (Review) Copyright © 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

Study	Reason for exclusion
de Zoysa 2012	All samples investigated were culture negative samples and not in the context of suspected infec- tion.
Golden 2004	GBS fluorescent PCR not compared with the reference standard (all were culture negative sam- ples).
Jones 2010	Analyzed gastric aspirates by molecular methods for DNA load followed by sequencing and cul- tures. Neonates were suspected of sepsis but no details of blood cultures to diagnose sepsis were available.
Jordan 2005b	Culture-positive specimens were examined for 16srRNA for PCR and sequencing. Not evaluated in the clinical context of suspected sepsis.
Jordan 2009	Pyrosequencing used to identify bacteria from positive blood culture bottles. Not evaluated in the clinical context of suspected sepsis.
Lucignano 2011	It is unclear how many participants included in the study were neonates. Attempt made to contact author for details.
Makhoul 2007	Term neonates had risk factors of sepsis (maternal fever, unknown maternal GBS) but not suspect- ed of having sepsis. Both blood cultures and PCR were negative in this cohort.
Shang 2001	Culture-positive specimens and healthy controls were evaluated and not in the clinical context of suspected sepsis.
Shen 2004	No clinical specimens from neonates with suspected sepsis. Spiked samples were used.
Tschiedel 2012	Non-neonatal population.

CSF: cerebrospinal fluid; GBS: group B streptococcus; PCR: polymerase chain reaction.

DATA

Presented below are all the data for all of the tests entered into the review.

Table Tests. Data tables by test

Test	No. of studies	No. of participants
1 All molecular tests	35	7339
2 Molecular tests: blood samples only	32	6999
3 Molecular tests with good methodologic quality	22	4150

Test 1. All molecular tests.

Review: Molecular assays for the diagnosis of sepsis in neonates Test: 1 All molecular tests

udy	ТΡ	FP	FN	TN	Sensitivity	Specificity	Sensitivity	Specificity
Briones 2003	20	2	1	38	0.95 [0.76, 1.00]	0.95 [0.83, 0.99]		·
Chan 2009	33	5	9	171	0.79[0.63,0.90]	0.97 [0.93, 0.99]	_	
Chen 2009	15	10	0	170	1.00 [0.78, 1.00]	0.94 [0.90, 0.97]		-
Draz 2013	20	15	8	7	0.71[0.51,0.87]	0.32 [0.14, 0.55]	_	
Dutta 2009	50	7	2	183	0.96 [0.87, 1.00]	0.96 [0.93, 0.99]		
Enomoto 2009	3	6	3	65	0.50[0.12,0.88]	0.92 [0.83, 0.97]		_
Esparcia 2011	3	3	4	73	0.43 [0.10, 0.82]	0.96 [0.89, 0.99]		-
Fujimori 2010	6	9	0	24	1.00 [0.54, 1.00]	0.73 [0.54, 0.87]		
Garcia-Elorriaga	20192	42	0	47	1.00 [0.66, 1.00]	0.53 [0.42, 0.63]		_ _
lbarra 2015	9	25	4	48	0.69[0.39,0.91]	0.66 [0.54, 0.76]	_	
Jordan 2000	24	3	1	520	0.96[0.80,1.00]	0.99 [0.98, 1.00]		
Jordan 2005a	51	0	2	32	0.96 [0.87, 1.00]	1.00 [0.89, 1.00]		-
Jordan 2006	7	30	10	1186	0.41[0.18,0.67]	0.98 [0.96, 0.98]		
Kasper 2013	15	9	0	22	1.00 [0.78, 1.00]	0.71[0.52,0.86]		_
Laforgia 1997	4	2	0	27	1.00 [0.40, 1.00]	0.93 [0.77, 0.99]		
Lima 2007	3	10	5	75	0.38[0.09,0.76]	0.88 [0.79, 0.94]	—	
Liu 2014	95	28	0	583	1.00 [0.96, 1.00]	0.95 [0.93, 0.97]	-	
Makhoul 2005	9	0	4	202	0.69[0.39,0.91]	1.00 [0.98, 1.00]	_	
Makhoul 2006	8	7	6	125	0.57 [0.29, 0.82]	0.95 [0.89, 0.98]		-
Ohlin 2008	21	12	29	233	0.42[0.28,0.57]	0.95 [0.92, 0.97]	_	
Ohlin 2012	44	31	12	281	0.79[0.66,0.88]	0.90 [0.86, 0.93]	_	-
Paolucci 2009	3	4	1	26	0.75 [0.19, 0.99]	0.87 [0.69, 0.96]		
Reier-Nilsen 200	94	6	2	36	0.67 [0.22, 0.96]	0.86 [0.71, 0.95]		_ _
Shaat 2013	17	7	0	26	1.00[0.80,1.00]	0.79[0.61,0.91]		-
Shang 2005	8	9	0	155	1.00[0.63,1.00]	0.95 [0.90, 0.97]		-
Taira 2014	3	3	0	18	1.00[0.29,1.00]	0.86 [0.64, 0.97]		- _
Tirodker 2003	10	13	3	44	0.77 [0.46, 0.95]	0.77 [0.64, 0.87]	_	_ _
Tong 2004	8	9	0	268	1.00[0.63,1.00]	0.97 [0.94, 0.99]		
Torres-Martos 20	1312	6	5	19	0.71[0.44,0.90]	0.76[0.55,0.91]		_
Trovato 2012	7	8	1	70	0.88[0.47,1.00]	0.90 [0.81, 0.95]		
Van der Brand 2	01410	0	3	7	0.77 [0.46, 0.95]	1.00 [0.59, 1.00]	_	
Villanueva-Uy 20	0323	0	6	32	0.79[0.60,0.92]	1.00 [0.89, 1.00]	_	
Wu 2007	20	23	0	787	1.00[0.83,1.00]	0.97 [0.96, 0.98]		
Wu 2008	34	16	0	550	1.00 [0.90, 1.00]	0.97 [0.95, 0.98]	_	
Yadav 2005	9	4	0	87	1.00 [0.66, 1.00]	0.96 [0.89, 0.99]		-
							0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8



Test 2. Molecular tests: blood samples only.

Review: Molecular assays for the diagnosis of sepsis in neonates Test: 2 Molecular tests: blood samples only

tudy 1	ГР	FP	FN	TN	Sensitivity	Specificity	Sensitivity	Specificity
Briones 2003	20	2	1	38	0.95 [0.76, 1.00]	0.95 [0.83, 0.99]		
Chen 2009	15	10	0	170	1.00[0.78,1.00]	0.94 [0.90, 0.97]		-
Draz 2013	20	15	8	7	0.71[0.51,0.87]	0.32 [0.14, 0.55]	_	
Dutta 2009	50	7	2	183	0.96[0.87,1.00]	0.96 [0.93, 0.99]		
Fujimori 2010	3	6	3	65	0.50[0.12,0.88]	0.92 [0.83, 0.97]		_
Garcia-Elorriaga	20122	42	0	47	1.00[0.66,1.00]	0.53 [0.42, 0.63]		_ _
Ibarra 2015	9	25	4	48	0.69[0.39,0.91]	0.66 [0.54, 0.76]		_
Jordan 2000	24	3	1	520	0.96[0.80,1.00]	0.99[0.98, 1.00]		
Jordan 2005a	51	0	2	32	0.96[0.87,1.00]	1.00 [0.89, 1.00]		-
Jordan 2006	7	30	10	1186	0.41[0.18,0.67]	0.98 [0.96, 0.98]		
Kasper 2013	15	9	0	22	1.00[0.78,1.00]	0.71[0.52,0.86]		_
Laforgia 1997	4	2	0	27	1.00[0.40,1.00]	0.93 [0.77, 0.99]		
Lima 2007	3	10	5	75	0.38[0.09,0.76]	0.88 [0.79, 0.94]		
Liu 2014	95	28	0	583	1.00 [0.96, 1.00]	0.95 [0.93, 0.97]		
Makhoul 2005	9	0	4	202	0.69[0.39,0.91]	1.00 [0.98, 1.00]		
Makhoul 2006	8	7	6	125	0.57 [0.29, 0.82]	0.95 [0.89, 0.98]	_	
Ohlin 2008	21	12	29	233	0.42[0.28,0.57]	0.95 [0.92, 0.97]		
Ohlin 2012	44	31	12	281	0.79[0.66,0.88]	0.90 [0.86, 0.93]	_ _	-
Paolucci 2009	3	4	1	26	0.75 [0.19, 0.99]	0.87 [0.69, 0.96]		
Reier-Nilsen 2009	4	6	2	36	0.67 [0.22, 0.96]	0.86 [0.71, 0.95]		_ _
Shaat 2013	17	7	0	26	1.00 [0.80, 1.00]	0.79[0.61,0.91]		_
Shang 2005	8	9	0	155	1.00 [0.63, 1.00]	0.95 [0.90, 0.97]		
Taira 2014	3	3	0	18	1.00 [0.29, 1.00]	0.86 [0.64, 0.97]		_
Tirodker 2003	10	13	3	44	0.77 [0.46, 0.95]	0.77 [0.64, 0.87]		_
Tong 2004	8	9	0	268	1.00 [0.63, 1.00]	0.97 [0.94, 0.99]		
Torres-Martos 20	1312	6	5	19	0.71[0.44,0.90]	0.76[0.55,0.91]	_	_
Trovato 2012	7	8	1	70	0.88[0.47,1.00]	0.90 [0.81, 0.95]		
Van der Brand 20	1410	0	3	7	0.77 [0.46, 0.95]	1.00 [0.59, 1.00]		
Villanueva-Uy 200	0323	0	6	32	0.79[0.60,0.92]	1.00 [0.89, 1.00]		
Wu 2007	20	23	0	787	1.00[0.83,1.00]	0.97 [0.96, 0.98]		
Wu 2008	34	16	0	550	1.00 [0.90, 1.00]	0.97 [0.95, 0.98]		
Yadav 2005	9	4	0	87	1.00 [0.66, 1.00]	0.96 [0.89, 0.99]		-
							0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8

Test 3. Molecular tests with good methodologic quality.

Review: Molecular assays for the diagnosis of sepsis in neonates Test: 3 Molecular tests with good methodologic quality

tudy T	P	FP	FN	TN	Sensitivity	Specificity	Sensitivity	Specificity
Chan 2009	33	5	9	171	0.79 [0.63, 0.90]	0.97 [0.93, 0.99]		-
Draz 2013	20	15	8	7	0.71[0.51,0.87]	0.32[0.14,0.55]	_	
Dutta 2009	50	7	2	183	0.96[0.87,1.00]	0.96 [0.93, 0.99]		-
Enomoto 2009	3	6	3	65	0.50[0.12,0.88]	0.92 [0.83, 0.97]		
Esparcia 2011	3	3	4	73	0.43[0.10,0.82]	0.96 [0.89, 0.99]		
Fujimori 2010	6	9	0	24	1.00[0.54,1.00]	0.73[0.54,0.87]		_
Ibarra 2015	9	25	4	48	0.69[0.39,0.91]	0.66 [0.54, 0.76]	_	_
Kasper 2013	15	9	0	22	1.00[0.78,1.00]	0.71[0.52,0.86]		_
Laforgia 1997	4	2	0	27	1.00 [0.40, 1.00]	0.93 [0.77, 0.99]		
Liu 2014	95	28	0	583	1.00 [0.96, 1.00]	0.95 [0.93, 0.97]		-
Makhoul 2005	9	0	4	202	0.69[0.39,0.91]	1.00 [0.98, 1.00]		
Makhoul 2006	8	7	6	125	0.57 [0.29, 0.82]	0.95 [0.89, 0.98]	_	-
Ohlin 2008	21	12	29	233	0.42[0.28,0.57]	0.95 [0.92, 0.97]		-
Paolucci 2009	3	4	1	26	0.75[0.19,0.99]	0.87 [0.69, 0.96]		_
Reier-Nilsen 2009	4	6	2	36	0.67 [0.22, 0.96]	0.86[0.71,0.95]		_
Shang 2005	8	9	0	155	1.00 [0.63, 1.00]	0.95 [0.90, 0.97]		-
Taira 2014	3	3	0	18	1.00 [0.29, 1.00]	0.86 [0.64, 0.97]		_
Trovato 2012	7	8	1	70	0.88[0.47,1.00]	0.90 [0.81, 0.95]		
Van der Brand 20	1410	0	3	7	0.77 [0.46, 0.95]	1.00 [0.59, 1.00]	_	
Wu 2007	20	23	0	787	1.00[0.83,1.00]	0.97 [0.96, 0.98]	4	
Wu 2008	34	16	0	550	1.00 [0.90, 1.00]	0.97 [0.95, 0.98]		
Yadav 2005	9	4	0	87	1.00[0.66,1.00]	0.96 [0.89, 0.99]		

APPENDICES

Appendix 1. Search strategy

1. Our search strategy for **PubMed** below was developed by discussion between the author team and the Cochrane Neonatal Group's Trials Search coordinator. We adapted it for use in other databases. www-ncbi-nlm-nih-gov.ezproxyhost.library.tmc.edu/pubmed? otool=hamtmc

Search (((((((((("Sepsis/blood"[Mesh] OR "Sepsis/cerebrospinal fluid"[Mesh] OR "Sepsis/diagnosis"[Mesh]))) OR neonatal sepsis) OR neonatal bacteremia) OR neonatal infections)) AND "Infant, Newborn"[Mesh])) AND (((((((((polymerase chain reaction) OR pcr) OR real time pcr) OR multiplex pcr) OR molecular probes) OR nucleic acid amplification) OR hybridization) OR sequencing) OR pyrosequencing) OR genechip))) AND (((((diagnosis) OR detection) OR identification) OR rapid identification) OR diagnostic tool)

2. EMBASE search strategy (provided by Elsevier through TMC library)

#1 sepsis

#2 Infection

#3 bacteremia

#4 #1 OR #2 OR #3

#5 neonate

#6 newborn

#7 #5 OR #6

#8 diagnosis OR detection OR identification OR diagnostic

#9 PCR

#10 molecular AND methods

#11 nucleic AND acid AND amplification

Molecular assays for the diagnosis of sepsis in neonates (Review) Copyright © 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.



- #12 hybridization
- #13 sequencing
- #14 polymerase AND chain AND reaction

#15 #9 OR #10 OR #11 OR #12 OR #13 OR #14

#16 Human

#17 #4 AND #7 AND #8 AND #15 AND #16

3. CINAHL search strategy (platform EBSCO host)

#1 sepsis

- #2 Infection
- #3 bacteremia
- #4 #1 OR #2 OR #3

#5 neonate

#6 newborn

#7 #5 OR #6

#8 diagnosis OR detection OR identification OR diagnostic

#9 PCR

#10 molecular AND methods

#11 nucleic AND acid AND amplification

- #12 hybridization
- #13 sequencing
- #14 polymerase AND chain AND reaction

#15 #9 OR #10 OR #11 OR #12 OR #13 OR #14

#16 Human

#17 #4 AND #7 AND #8 AND #15 AND #16

4. Cochrane library http://www.cochranelibrary.com.ezproxyhost.library.tmc.edu/

Using advanced search and selecting Cochrane Reviews, other reviews, trials and methods

studies. Using search words, molecular, neonate, newborn, PCR and sepsis

5. Science citation index, platform-Web of science

Searched using advanced search and subject search with search words, 'molecular', 'neonate',

'newborn', 'PCR', 'nucleic acid' 'diagnostic' and sepsis using BOOLEAN combination words.

Ref	Method	Data				ТР	FP	FN	TN	Sen- sitiv- ity (%)	Speci- ficity (%)	PPV (%)	NPV (%)	Participants	Study peri- od	Comments
			Refere Blood	nce std Cx												
			Posi- tive	Neg- ative	-											
Briones 2003	Fungal conven- tional	Posi- tive	20	2	22	20	2	1	38	95.24	95.00	90.91	97.44	Newborns > 3 days old suspected of sepsis.	Not men- tioned	Abstract. Same autho as Villanue-
	PCR tar- geting ITS3 and	Neg- ative	1	38	39	_								No information on demographics.	tioned.	va-Uy and same numbe of cases but
	ITS3 and ITS4 re- gions of the 5S rRNA.		21	40	61	_										using differ- ent primers (bacterial vs fungal).
			Refere	nce std												
			Blood	Сх												
			Posi- tive	Neg- ative	-											
Chan 2009	RT-PCR with uni- versal	Posi- tive	33	5	38	33	5	9	171	78.57	97.16	86.84	95.00	Preterm infants < 37 wk GA, > 72 hr of age with signs and	Over 28- month	-
	primers and Gram-	Neg- ative	9	171	180									symptoms of sys- temic infection re- quiring full sepsis	pe- riod from	
	specific probes		42	176	218	_								evaluation and an- tibiotic treatment. Interquartile range	Mar 2006 to	

Molecular assays for the diagnosis of sepsis in neonates (Review) Copyright © 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

78

Cochrane Database of Systematic Reviews

Cochrane Tru Library Bet

(Continued	⁷ Blood, peri- toneal fluid and urine.													of age as reported in results suggest- ed some infants > 28 days old.	Jun 2008.	
			Refere Blood CSF Cx													
			Posi- tive	Neg- ative	-											
Chen 2009	Broad- range 16S	Posi- tive	15	10	25	15	10	0	170	100.00	94.44	60.00	100.00	Neonates admitted to the neonatal de- partment and ICU	Sept 2007 to	Blood (n = 190) and CSF (n = 5) sam-
	rRNA- based real-time	Neg- ative	0	170	170	_								of the Children's Hospital at Zhe- jiang University in	Jun 2008.	ples. Each sample test- ed for Cx and
	FQ-PCR.		15	180	195	_								China with suspect- ed sepsis or menin- gitis. No information on demographics.		PCR. Not sure if blood drawn con- currently for Cx and PCR. Not blinded.
			Refere	nce std												
			Blood	Сх	_											
			Posi- tive	Neg- ative												
Draz 2013	Broad- range 16S rD-	Posi- tive	20	15	35	20	15	8	7	71.43	31.82	57.14	46.67	Neonates with clin- ical or lab findings suggestive of sep-	May 2012 to	The authors mentioned 6 samples
	NA PCR.	Neg- ative	8	7	15	_								sis.	Aug 2012.	were consid- ered contami- nated: 4 with
			28	22	50											Diphtheroid

Copyright © 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

																spp. and 2 with Candid. Appears the 6 were even- tually consic ered as nega tive blood C
			Refere	nce std												
			Blood	Сх												
			Posi- tive	Neg- ative	-											
Dutta 2009	Broad- range conven-	Posi- tive	50	7	57	50	7	2	183	96.15	96.32	87.72	98.92	Neonates who were clinically sus- pected to have an	Not men- tioned	Aseptically collected an concurrent
	tional PCR af- ter 5-hr	Neg- ative	2	183	185									episode of sepsis with onset of ≥ 72 hr after cessation	tioned.	blood draw for PCR and Cx. Not blind
	pream- plifica- tion Cx.		52	190	242									of antibiotics.		ed.
			Refere Cx	nce std												
			Posi- tive	Neg- ative	-											
Enomo to 2009	- Multi- plex PCR target-	Posi- tive	3	5	8	3	5	3	66	50.00	92.96	37.50	95.65	130 clinical sam- ples from 62 new- borns with any sus-	Jun 2005 to	In Table 2, number of positive PCF
2005	ing 8 pathogens	Neg- ative	3	66	69	_								picious infectious signs or infections and 50 cord bloods	Sept 2006.	was 9 not 8 as in Table 3. Number o
	Also in- cludes skin, BAL, mu-		6	71	77	_								and blood after birth from healthy term infants with-		samples wit no test was unless pha- ryngeal mu-

Cochrane Database of Systematic Reviews

Cochrane Library

(Continue	cus, CSF, urine and as- cites.													out signs or history of infection. Total of 77 paired samples.		cus was in- cluded. Those doing Cx were blinded but no mention of those doing PCR.
			Refere Blood	nce std Cx												
			Posi- tive	Neg- ative	-											
Es- par-	16S RT- PCR fol-	Posi- tive	3	3	6	3	3	4	73	42.86	96.05	50.00	94.81	Newborn < 7 days old with suspect-	Nov 2005	There were 105 samples from 83 new-
cia 2011	lowed by microar- ray and sequenc-	Neg- ative	4	73	77									ed sepsis or early meningitis.	to Jan 2007.	borns for EOS. In the paper, results re-
	Includes CSF samples where PCR and Cx were per- formed.		7	76	83											ferred to cas- es of EOS and not samples, hence n = 83.
			Refere	nce std												
			Blood	Сх	_											
			Posi- tive	Neg- ative	_											

$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	⁻ uji- nori 2010	RT-PCR.	Posi- tive	6	9	15	6	9	0	24	100.00	72.73	40.00	100.00	Neonates admit- ted to NICU with suspected sep-	Feb 2009 to	Concurrent blood draw. Repeated
$\begin{array}{c c c c c c c c c c c c c c c c c c c $				0	24	24									sis. Mean (SD) GA 34.8 ± 5.8 wk. 36	Aug	samples tak- en in same
Blood Cx Posi- tive Neg- ative Gar- cia-Elor-range pringa 2012 Posi- primer 9 38 47 9 38 0 2 100.00 5.00 19.15 100.00 Neonates aged ≤ 28 days admitted to NICU with clin- ical Dx of sepsis without mibiotic treatment or with maximum 48 hr an- tibiotic treatment or > 3 days' treat- ment but without response. Aug 2005 Calculation blood Cx of case only. To- tal positive to Jul 2006.				9	33	39									episodes of neona-		excluded. Not
Posi- tive Neg- ative Gar- cia-Elor-range riaga 2012 Posi- primer Note: au- thors' gold std was clin- ical Dx. Posi- Posi- Posi- Posi- Posi- Posi- Posi- Posi- 2012 9 38 47 9 38 0 2 100.00 5.00 19.15 100.00 Neonates aged ≤ 28 days admitted to NICU with clin- ical Dx of sepsis without antibiotic treatment or with maximum 48 hr an- tibiotic treatment or > 3 days' treat- ment but without response. Aug 2005 Calculation based on to Jul blood Cx of case only. To- tal positive CX on table 2				Refere	nce std												
Gar- cia-Elor- range primer 2012 Posi- primer Note: au- thors' 9 38 47 9 38 0 2 100.00 5.00 19.15 100.00 Neonates aged ≤ 28 days admitted to NICU with clin- ical Dx of sepsis without antibiotic treatment or with was clin- ical Dx. Aug 2012 Calculation based on to Jul 9 40 49				Blood	Сх												
cia-Elor-range riaga 2012tive28 days admitted to NICU with clin- ical Dx of sepsis without antibiotic thors' gold std was clin- ical Dx.tive2005 to Jul blood Cx of 2006.based on to Jul blood Cx of 2006.Note: au- thors' gold std was clin- ical Dx.94049493049Note: au- thors' gold std was clin- ical Dx.940494933 but Ta- tibiotic treatment or > 3 days' treat- ment but without response.33 but Ta- to add 2 posi- tive catheter as it is un- clear in table where PCR						-											
2012 primer Neg- ative 0 2 2 ical Dx of sepsis without antibiotic treatment or with 2006. case only. To- tal positive Note: au- thors' 9 40 49 maximum 48 hr an- tibiotic treatment was clin- ical Dx. = 33 but Ta- ble 4 = 23. Un- or > 3 days' treat- ment but without response. ble 4 = 23. Un- sure where treatment ble 4 = 23. Un- to add 2 posi- tive catheter as it is un- clear in table where PCR	cia-Eloi	r- range		9	38	47	9	38	0	2	100.00	5.00	19.15	100.00	28 days admitted	2005	based on
thors'94049maximum 48 hr an-= 33 but Ta-gold stdtibiotic treatmentble 4 = 23. Un-was clin-or > 3 days' treat-sure whereical Dx.to add 2 posi-response.tive catheteras it is un-clear in tablewhere PCR		primer	-	0	2	2	-								ical Dx of sepsis without antibiotic		case only. To- tal positive
		gold std was clin-		9	40	49	_								maximum 48 hr an- tibiotic treatment or > 3 days' treat- ment but without		= 33 but Ta- ble 4 = 23. Un- sure where to add 2 posi- tive catheter as it is un- clear in table where PCR
Reference std				Posi-	Neg-	-											
Blood Cx				tive	ative												

Copyright © 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

82

Reviews

Ibox	ed)	Deei	0	25	24	0	25	4	40	co 22		26.47	02.21	Neenetee with eve	Net	Consumment
lbar- ra 2015	LightCy- cler Sep- tiFast.	Posi- tive	9	25	34	9	25	4	48	69.23	65.75	26.47	92.31	Neonates with sus- pected clinical sepsis and those	Not men- tioned.	Concurrent samples for Cx and Light-
		Neg- ative	4	48	52									presenting > 8 on NOSEP-1 scale. 86 samples from 86		Cycler Septi- Fast. PPV and NPV reported
			13	73	86	-								neonates included.		were differ-
														Table 4 shows that neonates in the blood Cx group may be > 28 days old as it reported (mean ± SD) 23 ± 9.2 days.		ent (69% and 65%, respec- tively).
			Refere	nce std												
			Blood	Сх	_											
			Posi- tive	Neg- ative												
									500							
dan	Broad- range PCR and	Posi- tive	24	3	27	24	3	1	520	96.00	99.43	88.89	99.81	All infants admit- ted to NICU for sep- sis evaluation	Not men- tioned	Not blinded. Good tech- nique Flimi-
Jor- dan 2000			24	3	27 521	24 	3	1	520	96.00	99.43	88.89	99.81		men-	
dan	range PCR and DNA blot	tive Neg-				24 - -	3	1	520	96.00	99.43	88.89	99.81	ted to NICU for sep- sis evaluation. No information on	men-	Good tech- nique. Elimi- nated conta-
dan	range PCR and DNA blot	tive Neg-	1 25	520 523	521	24	3	1	520	96.00	99.43	88.89	99.81	ted to NICU for sep- sis evaluation. No information on	men-	Good tech- nique. Elimi- nated conta-
dan	range PCR and DNA blot	tive Neg-	1 25 Referen	520 523 nce std	521	24	3	1	520	96.00	99.43	88.89	99.81	ted to NICU for sep- sis evaluation. No information on	men-	Good tech- nique. Elimi- nated conta-
dan	range PCR and DNA blot	tive Neg-	1 25 Referen Blood (520 523 nce std Cx	521	24	3	1	520	96.00	99.43	88.89	99.81	ted to NICU for sep- sis evaluation. No information on	men-	Good tech- nique. Elimi- nated conta-
dan	range PCR and DNA blot	tive Neg-	1 25 Referen	520 523 nce std	521	24	3	1	520	96.00	99.43	88.89	99.81	ted to NICU for sep- sis evaluation. No information on	men-	Good tech- nique. Elimi- nated conta-

Molecular assays for the diagnosis of sepsis in neonates (Review) Copyright © 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

Cochrane Database of Systematic Reviews

Cochrane Library

			53			-										bers were de-
				32	85											rived from the paper that stated 53 were Cx posi- tive and of the 53, 51 were al- so PCR posi- tive and 2 that were PCR neg- ative. 32 sam- ples were Cx negative and PCR negative. No mention if blinded.
			Refere	nce std												
			Blood													
			Posi- tive	Neg- ative	-											
or- Conv an tion- 006 al PC	- t	Posi- tive	7	30	37	7	30	10	1186	41.18	97.53	18.92	99.16	Eligible infants had to be > 34 wk GA at time of birth,	1 Sept 2000	No mention if blinded.
base on 1	ed N .6S a	Neg- ative	10	1186	1196	-								admitted to NICU within a few hours	to 1 Apr	
sayf	ed by ose-		17	1216	1233	-								for EOS evaluation, and have both a blood Cx and CBC ordered. No details on demographics.	2004.	

Copyright @ 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

			Blood	Сх											
			Posi- tive	Neg- ative	-										
Kasper 2013	tiplex	Posi- tive	15	9	24	15	9	0	22	100.00	70.97	62.50	100.00	46 VLBW infants > 72 hr of life with	Not - men-
	RT-PCR (Light- Cycler) Sep-	Neg- ative	0	22	22	_								suspected LOS. Details on demo- graphic including day of life of sep-	tioned.
	tiFast MGRADE system for de- tection of LOS. Targeted Gram- positive and G- negative organ- isms be- tween 16S and 23S rRNA genes, and fun- gi by 18S and 5.8S rDNA.		15	31	46									sis evaluation were not mentioned.	
			Refere Blood												
			Posi- tive	Neg- ative	-										

Cochrane Library

> Trusted evidence. Informed decisions. Better health.

Cochrane Database of Systematic Reviews

Copyright \circledast 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

85

afor- gia .997	Mul- tiplex PCR.	Posi- tive	4	2	6	4	2	0	27	100.00	93.10	66.67	100.00	33 newborns at risk for EOS.	Jan to Sept	-
		Neg- ative	0	27	27										1996.	
			4	29	33	_										
			Refere	nce std												
			Blood	Сх	_											
			Posi- tive	Neg- ative	_											
ima 007	RT-PCR using universal	Posi- tive	3	10	13	3	10	5	75	37.50	88.24	23.08	93.75	93 samples for neonates with sus- pected sepsis.	Dec 2004 to	Abstract. 93 blood sam- ples. 3 were
	primer.	Neg- ative	5	75	80	_								No information on demographics.	Jun 2005.	blood Cx and PCR-positive. 5 were blood
			8	85	93											Cx positive, 10 were posi- tive by molec- ular method. 4 samples not included as it was posi- tive for hu- man chromo- somes.
			Refere	nce std												
			Blood													
			Posi- tive	Neg- ative	-											

Copyright \circledast 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

Liu 2014	Broad- range 16S	Posi- tive	95	28	133	95	28	0	583	100.00	95.42	77.24	100.00	Neonates who had blood drawn for CBC and CRP. In-	1 Sept to 31	-
	rRNA gene PCR.	Neg- ative	0	583	583	_								fants were 4 hr to 28 days old.	Dec 2011.	
			95	911	706											
			Refere	nce std												
			Blood	Сх												
			Posi- tive	Neg- ative	-											
Makhou 2005	IStaphy- lococ- cal 16S	Posi- tive	9	0	9	9	0	4	202	69.23	100.00	100.00	98.06	Neonates hospi- talized in the NICU with clinical signs	12- month peri-	Mean (± SD) GA 33.5 ± 4.4 (range 24 to
	rRNA PCR (both	Neg- ative	4	202	206	_								suggestive of sep- sis after 3 days of life. 124 neonates	od.	42 wk), mean birth weight 1962 ± 874
	S. <i>au-</i> <i>reus</i> and CONS).		13	202	215									with 215 events. There was no men- tion of how many infants were > 28 days old.		g (range 560 g to 3939 g), mean age at onset of pre- sumed sep- sis was 15.4 ± 17.3 days (range 4 to 96 days).
																Not blinded.
			Refere	nce std												
			Blood													
			Posi- tive	Neg- ative	-											

Copyright @ 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

Makho 2006	ulStaphy- lococ-	Posi- tive	8	7	15	8	7	6	125	57.14	94.70	53.33	95.42	Neonates with clin- ically suspected	Not men-	Mean age (± SD) at onset
	cal 16S rRNA PCR (both	Neg- ative	6	125	131	-								LOS beyond 3 days of life. No mention how	tioned.	of presumed sepsis was 17.3 ± 18.7 down (congo 4
	<i>S. au-</i> <i>reus</i> and CONS).		14	132	146 (? 148)	-								many infants were > 28 days old.		days (range 4 to 105 days). Not men- tioned if blinded. Discrepan- cy with pub- lished num- ber and actual number (148
			Blood Posi-	Neg-												
Ohlin 2008	RT-PCR 16S RNA.	Posi- tive	tive 21	ative 12	33	21	12	29	233	42.00	95.10	63.64	88.93	Newborns < 28 days old admitted	1995 to	Not blinded.
		Neg- ative	29	233	262	-								to NICU. n = 295 refers to samples from 288 infants.	2005.	
			50	245	295											
			Refere	nce std												
			Refere Blood													

Copyright \circledast 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

(Continued							_ .			-					-	·
Ohlin 2012	Broad- range 16S RT-	Posi- tive	44	31	75	44 -	31	12	281	78.57	90.06	58.67	95.90	Infants < 3 months of age subjected to blood Cx. total of	Oct 2007 to	34 samples were collect- ed at postna-
	PCR.	Neg- ative	12	281	293									368 samples from 317 infants.	Nov 2009.	tal age from 29 days to 3 months; how
			56	312	368	-										ever, no spe- cific informa- tion on the blood Cx and PCR results of these sam- ples.
			Refere	nce std												
			Blood													
			Posi- tive	Neg- ative	-											
Paoluc- ci	mercial	Posi- tive	3	4	7	3	4	1	26	75.00	86.67	42.86	96.30	Newborns with suspected LOS.	Not men-	Not blinded.
2009	LightCy- cler Sep- tiFast System.	Neg- ative	1	26	27	-								Age of infant at time of Dx not men- tioned.	tioned.	
	System.		4	30	34											
			Refere	nce std												
			Blood	Сх												
			Posi- tive	Neg- ative	-											
Reier- Nilsen 2009	Broad- range 16S rRNA	Posi- tive	4	6	10	4	6	2	36	66.67	85.70	40.00	94.70	Infants with birth weight > 1000 g with suspected	Not men- tioned.	Prospective, non-RCT. Sterile tech- nique. Same

Copyright \circledast 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

Cochrane Database of Systematic Reviews

Cochrane Library

															ble was used in article. In- cluded all (n
															= 48) cases of suspected sepsis.
		Refere	nce std												
		Blood	Сх	_											
		Posi- tive	Neg- ative												
road- ange 6S rD-	Posi- tive	17	7	24	17	7	0	26	100.00	78.79	70.83	100.00	Newborns with clinically suspected	Oct 2010	GA ranged from 26 to 39 wk, mean (±
A PCR.	Neg- ative	0	26	26	-								seµsis.	Dec 2012.	SD) 32.44 ± 2.91 wk; how- ever, age at
		17	33	50											Dx not men- tioned.
		Refere	nce std												
		Blood	Сх												
		Posi- tive	Neg- ative	-											
road- ange CR with	Posi- tive	8	9	17	8	9	0	155	100.00	94.51	47.06	100.00	developed clinical	1 Jan to 30	Authors did not provide additional
nicroar- ay hy-	Neg- ative	0	155	155	_								sepsis after 3 days of life.	2004.	characteris- tics of infants
ar 6: A roar	nge S rD- A PCR.	nge tive S rD- NPCR. Neg- ative 	tive tive tive 0 ad- Posi- tive NPCR. Neg- 0 ative 17 Referent Blood Posi- tive 0 ad- Referent Posi- tive 0 0 0 0 0 0 0 0 0 0 0 0 0	tive ative tive ative tive ative tive ative tive ative tive ative 17 7 APCR. Neg- ative 17 33 17 33 Reference std Blood Cx Posi- Neg- ative Neg- ative Neg- ative 17 33	tiveativeoad- nge S rD- APCR.Posi- tive17724Neg- ative02626173350Reference std Blood CxPosi- tiveNeg- ativeoad- nge R with croar-Posi- tive8917Neg- tive0155155	tive ative tive ative tive ative tive ative restring SrD-APCR. Neg- ative restring SrD-APCR. Neg- ative restring SrD-APCR. Neg- ative restring SrD-APCR. Neg- ative restring SrD-APCR. Neg- ative restring SrD-APCR. Neg- ative restring SrD-APCR. Neg- ative restring SrD-APCR. Neg- ative restring SrD-APCR. restring SrD-	$\frac{\text{tive}}{\text{nge}} = \frac{17}{17} + \frac{7}{7} + \frac{24}{17} + \frac{17}{7} + \frac{7}{7} + \frac{7}{7} + \frac{17}{7} + \frac{7}{7} + \frac{17}{7} + \frac{7}{7} + \frac{17}{7} + \frac{7}{7} + \frac{17}{7} + \frac{7}{7} + \frac{17}{17} + \frac{7}{7} + \frac{17}{17} + \frac{7}{17} + \frac{17}{17} +$	$\frac{\text{tive}}{\text{nge}} = \frac{17}{17} + \frac{7}{7} + \frac{24}{17} + \frac{17}{7} + \frac{7}{7} + \frac{9}{7} + \frac{17}{7} + \frac{9}{7} + \frac{17}{7} + \frac{9}{7} + \frac{17}{7} + \frac{9}{7} + \frac{17}{7} + \frac{9}{7} + \frac{17}{17} + \frac{9}{7} + \frac{17}{17} + 17$	$\frac{\text{tive}}{\text{nge}} = \frac{17}{17} + \frac{7}{7} + \frac{24}{17} + \frac{17}{7} + \frac{7}{7} + \frac{26}{17} + \frac{17}{7} + \frac{7}{7} + \frac{26}{17} + \frac{17}{17} + \frac{7}{7} + \frac{26}{17} + \frac{17}{17} + \frac{7}{17} + \frac{17}{17} + \frac{17}{$	$\frac{\text{tive}}{\text{nge}} = \frac{17}{\text{tive}} = \frac{17}{7} = \frac{7}{7} = \frac{24}{17} = \frac{17}{7} = \frac{7}{0} = \frac{26}{100.00}$ $\frac{\text{Neg-}}{17} = \frac{0}{17} = \frac{26}{26} = \frac{26}{17} = \frac{17}{17} = \frac{33}{33} = 50$ $\frac{\text{Reference std}}{17} = \frac{17}{17} = \frac{17}{33} = \frac{17}{17} = \frac{100.00}{17} = \frac$	$\frac{\text{tive}}{\text{nge}} = \frac{17}{\text{tive}} = 17$ $\frac{\text{Posi-tive}}{17} = 17$ $\frac{7}{7} = 24$ $\frac{17}{7} = 7$ $\frac{7}{7} = 24$ $\frac{17}{7} = 7$ $\frac{7}{7} = 24$ $\frac{17}{7} = 7$ $\frac{7}{7} = 26$ $\frac{100.00}{78.79}$ $\frac{78.79}{78.79}$ $\frac{17}{17} = 33$ $\frac{7}{50}$ $\frac{17}{17} = 33$ $\frac{7}{50}$ $\frac{17}{17} = 8$ $\frac{17}{17} = 15$ $\frac{100.00}{155} = 100.00$ $\frac{155}{100.00} = 94.51$ $\frac{17}{17} = 15$ $\frac{17}{17} = 15$ $\frac{17}{100.00} = 94.51$ $\frac{17}{100.00} = 94.51$ $\frac{17}{100.00} = 94.51$ $\frac{17}{100.00} = 155$ $\frac{100.00}{155} = 155$	$\frac{\text{tive}}{\text{ative}} \frac{\text{ative}}{\text{tive}} = \frac{17}{7} + \frac{7}{7} + \frac{24}{26} + \frac{17}{7} + \frac{7}{7} + \frac{9}{7} + \frac{26}{7} + \frac{100.00}{78.79} + \frac{70.83}{70.83} + \frac{70.83}{70.83$	$\frac{\text{tive}}{\text{ative}} = \frac{\text{ative}}{\text{ative}}$ $\frac{\text{Posi-}}{\text{five}} \frac{17}{17}, \frac{7}{7}, \frac{24}{26}, \frac{17}{7}, \frac{7}{7}, \frac{24}{26}, \frac{17}{7}, \frac{7}{7}, \frac{26}{26}, \frac{100.00}{78.79}, \frac{78.79}{70.83}, \frac{100.00}{100.00}$ $\frac{\text{Neg-}}{17}, \frac{33}{33}, \frac{50}{50}$ $\frac{\text{Reference std}}{17}, \frac{\text{Reference std}}{17}, \frac{17}{33}, \frac{17}{50}, 1$	$\frac{\text{tive}}{\text{affine}} = \frac{\frac{\text{Posi-}}{\text{tive}}}{\frac{17}{\text{r}}} = \frac{7}{17} = \frac{24}{17} = \frac{17}{7} = \frac{7}{17} = \frac{24}{17} = \frac{17}{7} = \frac{7}{17} = \frac{9}{17} = \frac{26}{100.00} = \frac{100.00}{78.79} = \frac{70.83}{100.00} = \frac{100.00}{\text{Newborns with clinically suspected sepsis.}}$	$\frac{\text{tive} \text{ative}}{\text{reg}} \frac{\text{Posi-}}{\text{tive}} \frac{17}{17} \frac{7}{1} \frac{24}{26} \frac{17}{7} 7 0 26 100.00 78.79 70.83 100.00 \text{Newborns with} \\ \text{clinically suspected} \frac{\text{Oct}}{\text{sepsis.}} \frac{\text{Oct}}{17} 33 50 \frac{17}{17} 33 50 \frac{17}{17} \frac{33}{17} 50 \frac{17}{17} \frac{33}{17} \frac{17}{17} \frac{10}{17} \frac{100.00}{17} 100.0$

Cochrane Database of Systematic Reviews

Trusted evidence. Informed decisions. Better health.

Cochrane Library

Continued) bridiza- tion. Positive speci- mens subject- ed to mi- croar- ray hy- bridiza		8	164	172											included in the study. Sensitivity was 94.51% (155/164). Not sure how the authors came up with 97.85%.
bridiza- tion.			_								-				Not blinded.
		Refere	nce std												
		Blood													
		Posi- tive	Neg- ative	-											
Taira Mul- 2014 tiplex	Posi- tive	8	5	13	8	5	0	41	100.00	89.13	61.54	100.00	neonates was	18- month	-
nested PCR for detec- tion and	Neg- ative	0	41	41	_								based on corre- spondence with Dr Del Negro.	peri- od.	
identifi- cation of Candida species.		8	46	54	_										
		Refere	nce std												
		Blood	Сх												
		Posi- tive	Neg- ative	-											
	Posi-	10	13	23	10	13	3	44	76.92	77.19	43.48	93.62	70 samples from 63	Nov	Study infants

Copyright @ 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

(Continued	al PCR target- ing 18S	Neg- ative	3	44	47	_								PICU) with suspect- ed clinical sepsis.	Nov 2000.	PICU (17 in- fants). Neona tal specific
	rRNA fungi.		13	57	70											data on bloo Cx and PCR not available Aseptic and concurrent blood sam- pling. Blind- ed.
			Refere	nce std												
			Blood	Сх												
			Posi- tive	Neg- ative	-											
Tong 2004	16S rRNA- based	Posi- tive	8	9	17	8	9	0	268	100.00	96.75	47.06	100.00	Neonates with sus- pected sepsis.	Not men- tioned.	Abstract on- ly. No specific details pro
	PCR fol- lowed by hy-	Neg- ative	0	268	268											vided for de- mographics.
	bridiza- tion to chips with 18 probes.		8	277	285	_										
			Refere	nce std												
			Blood	Сх	-											
			Posi- tive	Neg- ative												
Tor- res-Mai	LightCy- r- cler Sep- tiFast.	Posi- tive	12	6	18	12	6	5	19	70.59	76.00	66.67	79.17	42 blood sam- ples from 35 in- fants with febrile	Apr 2007 to	Sensitivity, specificity, PPV and NP\

Cochrane Database of Systematic Reviews

Cochrane Library

(Continued) tos 2013		Neg- ative	5	9 25	24 42	_								episodes. Based on Table 1. Infants were 0 to 151 days.	Apr 2009.	values report- ed in paper were based on compari- son on Light- Cycler Septi- Fast with clin- ical Dx.
			Refere Blood	ence std Cx												
			Posi- tive	Neg- ative	-											
to § 2012 c 1 1 2 1 2 1 2 2 1 2 2 2 2 2 2 2 2 2 2	Fun- gus-spe- cific uni- versal primers ITS1	Posi- tive	7	8	15	7	8	1	70	87.50	89.74	46.67	98.59	Neonates at high risk for invasive candidiasis.	Jan 2009 to Dec 2010.	No detailed information on demo-
		Neg- ative	1 70	70	71											graphics.
	and ITS2 used to ampli- fy rDNA, the ad- jacent ITS1 and small portion of the 28S rD- NA.		8	78	86											
			Refere Blood	ence std Cx												
					-											

Trusted evidence

(Continued	d)		Posi- tive	Neg- ative												
Van Multi- der plex RT- Brand PCR. 2014	plex RT-	Posi- tive	10	0	10	10	0	3	7	76.92	100.00	100.00	70.00	Preterm infants ad- mitted to NICU and	Not men-	-
	PCR.	Neg- ative	3	7	10	_								suspected to have LOS. No details on age of infants dur- ing evaluation for	tioned.	
			13	7	20									LOS.		
			Refere	nce std												
			Blood	Cx												
			Posi- tive	Neg- ative	-											
lanue- rang va-Uy 16S 2003 rRNA conv	Broad- range 16S	Posi- 23 tive Neg- 6 ative	23	0	23	23	0	6	32	79.31	100.00	100.00	84.21	old with suspected	Not men- tioned.	Abstract.
	rRNA conven- tional		6	32	38											
	PCR.		29	32	61					_						
			Refere	nce std												
			Blood	Сх												
			Posi- tive	Neg- ative	-											
	RT-PCR 16S RNA.	Posi- tive	20	23	43	20	23	0	787	100.00	97.16	46.51	100.00	Blood samples from cases of sus- pected septicemia.	Not men- tioned	Abstract only.
		Neg- ative	0	787	787									pected septicemia. No mention of up- per age limit.	tioned.	

Copyright © 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

94

Cochrane Database of Systematic Reviews

Cochrane Library

(Continue	d)		20	810	830											
			Refere Blood	nce std Cx												
			Posi- tive	Neg- ative	-											
2008 with	RT-PCR with Gram-	Posi- tive	34	16	50 34 550	34	16	0	550 100.	100.00	97.17	68.00	100.00	Infants aged 1 to 28 days admitted to the neonatal ward or NICU for clin- ically suspected to have bacterial infection or to be susceptible to in- fection.	Jan 2005 to Jan 2007.	Not blinded but implied as Cx and PCR were done si- multaneously.
	specific probes followed	Neg- ative	0	550		_										
	by se- quenc- ing.		34	566	600											
			Refere	nce std												
			Blood	Cx												
			Posi- tive	Neg- ative	-											
dav i	Broad- range 16S	Posi- tive	9	4	13	9	4	0	87	100.00	95.60	69.23	100.00	Newborns with risk factor for sepsis.	Not men-	Not blinded.
2005	rRNA PCR.	Neg- ative	0	87	87	-									tioned.	
			9	91	100	_										

BAL: bronchoalveolar lavage; CBC: complete blood count; CONS: coagulase-negative staphylococci; CRP: C-reactive protein; CSF: cerebrospinal fluid; Cx: culture; Dx: diagnosis; EOS: early-onset sepsis; FP: false positive; FN: false negative; FQ-PCR: quantitative fluorescence polymerase chain reaction; GA: gestational age; hr: hour; ICU: intensive care unit; LOS: late-onset sepsis; NICU: neonatal intensive care unit; NPV: negative predictive value; PCR: polymerase chain reaction; PICU: pediatric intensive care unit; PPV: positive predictive value; RCT: randomized controlled trial; rRNA: ribosomal ribonucleic acid; RT-PCR: real-time polymerase chain reaction; SD: standard deviation; std: standard; TP: true positive; TN: true negative; wk: week. Cochrane Library





Appendix 3. QUADAS-2 methodologic assessment tool

QUADAS-2 is structured so that four key domains are each rated in terms of the risk of bias and the concern regarding applicability to the research question (as defined above). Each key domain has a set of signaling questions to help reach the judgments regarding bias and applicability.

Domain 1: Participant selection

A. Risk of bias

Was a consecutive or random sample of participants enrolled?

YES: if the articles clearly stated that a consecutive or random samples was enrolled; **NO:** if it was clear that this was not the case (e.g. if a study included participants 'at the discretion of the clinician'); **UNCLEAR:** in other cases where it was not clear if consecutive or random samples were enrolled.

Was a case-control design avoided?

YES: if the enrolled sample was a random or consecutive enrollment of neonates with suspected sepsis and not separate samples from sepsis-positive participants and healthy controls; **NO:** if the enrolled samples consisted of sepsis-confirmed cases and healthy controls; **UNCLEAR:** if the sampling regarding case-control design was not clear.

Did the study avoid inappropriate exclusions?

Inappropriate exclusions included neonates whose mothers were treated with antibiotics, neonates from mothers infected with the human immunodeficiency virus (HIV), etc. **YES:** if inappropriate exclusions were not found in the included study, **NO:** if reasons for inappropriate exclusion were found. **Unclear:** if there was no description of the inclusion and exclusion criteria and inappropriate exclusion could not be ascertained.

Could the selection of participants have introduced bias?

LOW RISK: if all questions were scored "YES", or a maximum of one question with unclear.

HIGH RISK: if at least one question was scored as "NO".

UNCLEAR RISK: if at least two questions were scored as "UNCLEAR" and one as "NO".

B. Concerns regarding applicability

Was there concern that the included participants did not match the review question?

LOW CONCERN: if all included participants were neonates according to our definition and if they were suspected of sepsis.

HIGH CONCERN: if at least 10% of the included participants were not neonates or not suspected of sepsis.

UNCLEAR CONCERN: if it is unclear whether the study fulfilled either the criteria for low concern or for high concern.

Domain 2: Index test(s)

Describe the index test and how it was conducted and interpreted. If more than one index test was used, please complete for each test.

A. Risk of bias

· Describe the index test and how it was conducted and interpreted

Were the index test results interpreted without knowledge of the results of the reference standard?

YES: if people performing the molecular assays were blinded to the results of blood or cerebrospinal fluid (or both) cultures or if the index test was performed and interpreted prior to the reference standard; **NO:** if people performing the molecular assays had knowledge of the results of blood or cerebrospinal fluid (or both) cultures; **UNCLEAR:** if the study did not explicitly describe how the index test was conducted and interpreted.

If a threshold was used, was it prespecified?

This signaling question is not applicable to the study as no thresholds were used in the conduct and interpretation of the index and the reference standards. Results of the tests were dichotomous and were reported as either positive or negative.

Could the conduct or interpretation of the index test have introduced bias?

LOW RISK: if the study was performed blinded to the results of the reference standard.

HIGH RISK: if there was prior knowledge of the results of the reference standard.

UNCLEAR RISK: if there was no clear description of how the tests were conducted and interpreted.

B. Concerns regarding applicability

Was there concern that the index test, its conduct, or interpretation differed from the review question?

LOW CONCERN: if the index test used for the diagnosis of sepsis was a molecular assay as defined in our protocol and if the index test was interpreted without the knowledge of the results of the reference standard.

HIGH CONCERN: if the index test used for the diagnosis of sepsis varied from what was defined in the protocol and if the index test was interpreted with knowledge of the results of the reference standard.

UNCLEAR CONCERN: if it was unclear whether the study fulfilled criteria for "low concern" or "high concern" or if the study provided limited information regarding the conduct and interpretation of the index test.

Domain 3: Reference standard

A. Risk of bias

· Describe the reference standard and how it was conducted and interpreted

Was the reference standard likely to correctly classify the target condition?

YES: if the reference standard used was microbial culture of blood or cerebrospinal fluid (or both) in the diagnosis of neonatal sepsis. Microbial culture is currently the "gold standard" used in clinical practice in the diagnosis of neonatal sepsis; **NO:** if the test used as reference standard was a test other than microbial culture; **UNCLEAR:** if there was no description of the reference standard or if microbial cultures were used in combination with an "add-on" test.

Were the reference standard results interpreted without knowledge of the results of the index test?

YES: if people evaluating the results of the microbial culture were blinded to the results of the molecular assays and if the reference standard was performed and interpreted prior to the index test; **NO:** if people evaluating the results of the microbial culture had knowledge of the results of the molecular assays; **UNCLEAR:** if the study did not explicitly describe how the reference standard was conducted and interpreted.

Could the reference standard, its conduct, or its interpretation have introduced bias?

LOW RISK: if the reference standard used met the definition described in the protocol, performed and evaluated without knowledge of the results of the index test.

HIGH RISK: if the reference standard did not meet the definition described in the protocol or was evaluated with the knowledge of the results of the index test.

UNCLEAR RISK: if there was no clear description of the reference standard used, how it was performed and interpreted in relation to the results of the index test.

B. Concerns regarding applicability

Was there concern that the target condition as defined by the reference standard did not match the review question?

LOW CONCERN: if the reference standard was microbial culture of blood or cerebrospinal fluid (or both) and if the target condition was suspected sepsis in a neonate as defined in our protocol.

HIGH CONCERN: if the reference standard was a test other than microbial culture of blood or cerebrospinal fluid (or both) and if the target condition included participants other than neonates or if the participants were not suspected of neonatal sepsis.

UNCLEAR CONCERN: if it was unclear whether the study fulfilled either the criteria for "low concern" or for "high concern".

Domain 4: Flow and timing

A. Risk of bias

- Describe any participants who did not receive the index test(s) or reference standard (or both) or who were excluded from the 2 × 2 table (refer to flow diagram).
- Describe the time interval and any interventions between index test(s) and reference standard.

Was there an appropriate interval between index test(s) and reference standard?

YES: if blood or cerebrospinal fluid (or both) samples used for both microbial culture and molecular assay were drawn concurrently at the same time during the workup for neonatal sepsis; NO: if blood or cerebrospinal fluid (or both) samples used for both microbial culture and

Molecular assays for the diagnosis of sepsis in neonates (Review)

Copyright ${\ensuremath{\mathbb C}}$ 2017 The Cochrane Collaboration. Published by John Wiley & Sons, Ltd.

molecular assay were drawn more than 6 hours apart for the workup of neonatal sepsis; **UNCLEAR:** if there was no description of how and when the samples for both the index text and the reference standard were collected.

Did all participants receive a reference standard?

YES: if all participants underwent microbial culture testing for their blood or cerebrospinal fluid (or both); **NO:** if at least 1 participant did not have the reference standard performed. **UNCLEAR:** if the study did not describe clearly which participants received the reference standard and which ones did not.

Did participants receive the same reference standard?

YES: if all participants underwent microbial culture testing for their blood or cerebrospinal fluid (or both); **NO:** if a different reference standard other than culture of blood or cerebrospinal fluid (or both) was used in at least 1 participant; **UNCLEAR:** if the study did not describe clearly what type of reference standard was used to diagnose a participant with neonatal sepsis.

Were all participants included in the analysis?

YES: if all enrolled participants with the target condition who underwent testing using the index test and reference standard were included in the analysis; **NO:** if all enrolled participants were not accounted in the analysis; **UNCLEAR:** if it was unclear from the study about the inclusion of all enrolled participants in the analysis.

Could the participant flow have introduced bias?

LOW CONCERN: if the answers to above questions were all "YES" which means that all participants enrolled in the study were subjected to the same reference standard and index test, clinical samples for testing were drawn concurrently from the same participant, and all participants were included in the final analysis.

HIGH CONCERN: if at least 2 questions had a "NO" answer.

UNCLEAR CONCERN: if at least 1 question had a "NO" answer or it was unclear whether the study fulfilled either the criteria for "low concern" or for "high concern".

WHAT'S NEW

Date	Event	Description
26 December 2016	Amended	Revised based on suggestions from reviewers

CONTRIBUTIONS OF AUTHORS

MP conceived the project, searched literature, extracted and analyzed data, and wrote the review.

AF participated in the design, searched literature, extracted data, performed the QUADAS evaluation of included studies and assisted in writing the review.

JV provided critical intellectual input and revised the review.

ML provided critical intellectual input and revised the review.

DECLARATIONS OF INTEREST

Mohan Pammi, Angela Flores, James Versalovic and Mariska MG Leeflang have no financial or other conflicts of interest to disclose.

SOURCES OF SUPPORT

Internal sources

• No sources of support supplied

External sources

• Eunice Kennedy Shriver National Institute of Child Health and Human Development National Institutes of Health, Department of Health and Human Services, USA.

Editorial support of the Cochrane Neonatal Review Group has been funded with Federal funds from the Eunice Kennedy Shriver National Institute of Child Health and Human Development National Institutes of Health, Department of Health and Human Services, USA, under Contract No. HHSN275201100016C

DIFFERENCES BETWEEN PROTOCOL AND REVIEW

1. We decided post-hoc to present quality of evidence using GRADE methodology recommended for diagnostic tests.

2. Some studies did not include an upper limit for age and hence some infants were over 28 days of age. We made a post-hoc decision that we would include studies where an upper age limit was not specified but where more than 50% of the samples were from newborn less than 28 days of age. Our decision was supported by the reasoning that LOS extends up to three months of age and participant characteristics are similar in the first two to three months of age.

INDEX TERMS

Medical Subject Headings (MeSH)

DNA, Bacterial [blood] [cerebrospinal fluid] [isolation & purification]; DNA, Fungal [blood] [cerebrospinal fluid] [isolation & purification]; Infant, Premature; Polymerase Chain Reaction [methods]; Sepsis [*diagnosis] [microbiology]

MeSH check words

Humans; Infant, Newborn