Letter

N‑Nitrosulfonamides as Carbonic Anhydrase Inhibitors: A Promising Chemotype for Targeting Chagas Disease and Leishmaniasis

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S Supporting Information

[ABSTRACT:](#page-4-0) Trypanosoma cruzi and Leishmania spp. are protozoa of the Trypanosomatidae family, respectively, responsible of the neglected tropical disorders (NTDs) Chagas disease and leishmaniasis. The present pharmacotherapy is often ineffective and exhibits serious side effects. The metalloenzyme carbonic anhydrases (CAs, EC 4.2.1.1) recently identified in these protozoans $(\alpha$ -TcCA and β -LdcCA) are novel promising targets for chemotherapeutic interventions. Herein, we report a series of N-nitrosulfonamides, as a novel chemotype to yield the target CA isoform selective inhibition over ubiquitous human isozymes. Two derivatives selected among the most active and selective ones for TcCA/LdcCA over off-target CAs were progressed as silver salts to in vitro studies with various developmental forms and spp of Trypanosoma cruzi and leishmania. Excellent values of parasites growth inhibition (IC_{50}) were observed, with some selectivity index (over cytotoxicity for macrophages and Vero cells) being comparable or better than reference drugs. These findings make N-nitrosulfonamides and their salts promising lead compounds for a rational optimization of innovative agents for the treatment of Chagas disease and leishmaniasis based on CA inhibition.

KEYWORDS: Chagas disease, Trypanosoma cruzi, leishmania, carbonic anhydrase, zinc-binding group, inhibition, silver, antiparasitic

World Health Organization (WHO) included Chagas disease (American trypanosomiasis) and leishmaniasis in the list of neglected tropical diseases (NTDs). Parasites of the kinetoplastidae family are responsible for these infections, both belonging to the vector-borne diseases affecting 20 million people and killing more than 50,000 every year.¹

Trypanosoma cruzi is naturally transmitted by kissing bugs (mainly belonging to the genera Triatoma and R[h](#page-4-0)odnius), which primarily diffuse in Latin America. The disease evolves producing potentially fatal lesions to organs in the cardiac, digestive, or neurological systems.¹

Leishmaniasis is transmitted by the bite of an infected phlebotomine and works out ski[n](#page-4-0) or visceral aches that could turn out to be fatal if untreated. Among the NTDs, leishmaniasis is the first-in-class in terms of mortality and morbidity.¹

Available pharmacological treatments for the majority of NTDs are limited in terms of cost and toxicity and in[e](#page-4-0)ffective, and resistance phenomena constantly increase throughout the world.^{2−4} Pharmaceutical industry shows poor interest in searching new effective drugs for the treatment of NTDs due to hig[h](#page-4-0) [co](#page-4-0)sts and expected low financial return. It is urgent to find new therapeutic targets for these parasitosis, which WHO classifies as priority infections.^{2,5} Novel targets have been identified driven by large-scale analysis on the completely known genome sequence of both prot[oz](#page-4-0)oans. Indeed, endeavors to enrich the therapeutic arsenal against Chagas disease and leishmaniasis based on enzymatic inhibition have been starting in many laboratories with synthetic drugs representing a valuable source for new treatments.^{6,7}

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The metalloenzyme carbonic anhydrases (CAs, EC 4.2.1.1) recently identified in these protozoans are novel promising targets for chemotherapeutic interventions.^{6,8,9} CAs catalyze the reversible hydration of $CO₂$ to bicarbonate and proton, a pivotal reaction for all cells and complex organism[s, w](#page-4-0)hich is also basic in the growth and virulence of pathogenic microorganisms.⁹ CAs from Trypanosoma cruzi (TcCA) and Leishmania donovani chagasi (LdcCA) were cloned and characterized in 2013,^{10−12} resulting in the design of novel antiprotozoal agents that act by a totally new mechanism of action and lack cross-resistan[ce to](#page-4-0) existing drugs. The α -CA TcCA is endowed with a very high catalytic activity for the $CO₂$ hydration reaction and was shown to be inhibited in the nanomolar range by many types of CA inhibitors (CAIs) such as aromatic/heterocyclic sulfonamides, $^{10,13,\hat{14}}$ sulfamates, 10 thiols, 10 anions, $^{15}'$ dithiocarbamates, 15 hydroxamates, 16 and benzoxaboroles. 17 Thiols and hydro[xama](#page-4-0)[tes](#page-5-0) exhibited [in](#page-4-0) vitro [an](#page-4-0)titrypan[oso](#page-5-0)mal activity, inhibi[tin](#page-5-0)g the three p[has](#page-5-0)es of the pathogen's [li](#page-5-0)fe cycle.^{10,16} The β -CA LdcCA also features an effective catalytic activity and was shown to be efficiently inhibited by sulfonamides [a](#page-4-0)[nd](#page-5-0) heterocyclic thiols with nanomolar inhibition constants.^{12,18} Some such thiol derivatives displayed in vitro antileishmania activity in preliminary assays being able to reduce para[sit](#page-4-0)[es](#page-5-0)' growth and causing their death.¹² Identification of new protozoans CAIs with effective antitrypasonomal or antileishmania activities is more than ever [wo](#page-4-0)rth the endeavor due to such targets' remarkable druggability.

Nifurtimox and benznidazole have been the first effective drugs for treating acute-phase human Chagas infection, with the first being no longer available on the market because of undesirable side effects. 19 They feature heteroaromatic nitro moieties that are pivotal for the antiprotozoa mechanism of action. Parasite resistan[ce](#page-5-0) arisen with benznidazole drove the development of alternative therapies. Indeed, combined treatment of benznidazole with drugs with different mechanisms of action such as azoles, nitric oxide, or clomipramine could be a strategy to improve the pharmacotherapy efficacy.⁶

Noteworthy, a new chemotype able to afford α - and β -CAs inhibition was reported by us in 2016, nam[el](#page-4-0)y, N-nitrosulfonamides.²⁰ Interestingly, these latter were shown to inhibit ubiquitous, off-targets isoforms, such as CA II, feebler than lead sulfonamide[s t](#page-5-0)hough holding remarkable submicromolar inhibition of the human (h) tumor-associated CA IX $(\alpha$ -CA) and the β -CA from the pathogen fungus Malassezia Globosa.

Considering the above, we have herein extended the set of Nnitrosulfonamides and screened them on a wider pattern of human and pathogen (from protozoa and fungi) CAs, among which are the targets α -TcCA and β -LdcCA.

Furthermore, we produced silver salts of all such derivatives based on their marked effects against viruses, bacteria, fungi, and protozoa.²⁰ The antimicrobial behaviors of silver, silver ions, and silver-containing compounds have long been investigated with various [ant](#page-5-0)imicrobial mechanisms of action having been proposed to date. $^{21-24}$ The biologically active silver ion (Ag^+) irreversibly damages key enzyme systems in the cell membranes of pathogens. C[onvers](#page-5-0)ely, silver exhibits low toxicity in the human body and minimal risk is expected due to clinical exposure. 21 Recently, silver nanoparticles (Ag-NPs) were demonstrated to produce reactive oxygen species to which Leishman[ia](#page-5-0) parasites are very sensitive. 24 Moreover, the commercially available antibiotic silver sulfadiazine shares a wealth of features with silver N-nitrosulfon[am](#page-5-0)ides. These latter derivatives are thus endowed with multiple potential antiprotozoa entities to be synergistically exploited to overcome resistance issues displayed by single-targeted therapy.

The general synthetic strategy proposed by Minksztym²⁴ for the chemoselective mononitration of aminosulfonamides was applied to a set of ten starting compounds being comme[rci](#page-5-0)ally available $(1-3, 7, 8)$ or yielded by methylation $(4-6)$ or deacetylation (17, 19) reactions (Schemes 1 and 2).

Scheme 1. Synthesis of Aromatic N-Nitrosulfonamides: (a) $H₂O$; (b) NH₄OH(aq)

Quenching and workup of the NH_4NO_3/H_2SO_4 based nitration reaction was switched for the most unstable derivatives 16, 18, and 20 from water to $NH₄OH(aq)$ to generate the stable ammonium salts instead of the zwitterion forms of Nnitrosulfonamides (Schemes 1 and 2). $25,26$

The production of the silver salts of the derivatives was achieved by different methods depen[ding](#page-5-0) on the nature of the compound or the form it was produced as in the previous step (Scheme 3). Silver carbonate was used as the base (to remove

Scheme 3. Synthesis of Silver Salts of N-Nitrosulfonamides: (c) Ag_2CO_3 , H₂O; (d) NaOH, AgNO₃, H₂O

the proton) and the source of silver ion in an aqueous phase in case of zwitterion of amino aromatic compounds. NaOH/ $AgNO₃$ was used for the zwitterion of the amino aliphatic compound 15 and for ammonium salts 16, 18, and 20. All compounds silver salts precipitated in aqueous phases and were therefore recrystallized by the same solvent. All the obtained derivatives were properly characterized by means of ¹H NMR,
¹³C NMR, and MS (see Supporting Information). 13 C NMR, and MS (see Supporting Information).
The inhibition profiles of the N-nitrosulfonamide derivatives

were evaluated against six α [-CAs and three](http://pubs.acs.org/doi/suppl/10.1021/acsmedchemlett.8b00430/suppl_file/ml8b00430_si_001.pdf) β -CA isoforms in addition to acetazolamide (AAZ) as standard inhibitor, by a stopped flow $CO₂$ hydrase assay.²

Five human CA isoforms, among which the ubiquitous CA I and II (involved in many phys[iop](#page-5-0)athological processes), the membrane-associated CA IV (involved in ocular aches, stroke and arthritis), and IX and XII (overexpressed in hypoxic tumors) were included in the kinetic study to work out thorough structure−activity relationships (SARs) and selectivity profiles.²⁸ Along with the target TcCA and LdcCA, the activity of two additional $β$ -CA isoforms from pathogenic fungi was studied wit[h th](#page-5-0)e reported inhibitors, namely, MgCA from Malassezia globosa (responsible of the production of dandruff)²⁹ and Can2 from Cryptococcus neoformans (that can cause fungal meningitis and encephalitis).³⁰ The inhibitory efficacy again[st](#page-5-0) nine such isoforms was also assessed with the silver salts of all derivatives to verify whether th[e m](#page-5-0)onovalent metal ion affects the enzymatic activities. The inhibition constants (K_1s) of these latter do not show significant variations out of the error ranges, witnessing no significant action of the $Ag⁺$ ion to each CA activity. Tables 1 (hCAs) and 2 (pathogens CAs) gather the inhibition data of silver salts 21−30 (inhibition data of 9−20 are not shown; comparisons can be made with data previously reported). 20

Table 1. Inhibition Data of CA I, II, IV, IX, and XII wi[th](#page-5-0) N-Nitrosulfonamides 21−30 and the Standard Sulfonamide Inhibitor Acetazolamide (AAZ) by a Stopped Flow CO_2 Hydrase Assay²

The following structure−activity relationships (SAR) can be drawn from the inhibition data reported in Table 1 and 2.

According to preliminary data previously reported, 20 N-nitro aromatic sulfonamides exhibited low CA I and II inhibitory effectiveness, with K_I s spanning in a low to medium [mic](#page-5-0)romolar range (2.2−80.6 μM). Heteroaromatic derivatives 29 and 30 turned out as the most potent inhibitors against these ubiquitous hCAs. Whereas CA IV was targeted by all derivatives in a low micromolar range (1.4–39.2 μ M), a wealth of submicromolar K_I values against CA IX and XII (0.23–9.4 μ M) confirmed the favorite efficacy of N-nitrosulfonamides against the tumorassociated isoforms. CA XII was the most affected isozyme among the considered cluster, though the greatest inhibition was measured with the thiadiazole derivative 30 with CA IX (K_I of 0.23 μ M). It should be noted that CA XII features more Thr and

Table 2. Inhibition Data of TcCA, LdcCA, MgCA, and Can2 with N-Nitrosulfonamides and AAZ

		K_{I} $(\mu M)^{a}$			
compd	R	TcCA	LdcCA	MgCA	Can2
21	$2-NH2$	3.2	4.7	0.52	7.4
22	$3-NH2$	0.15	0.49	1.7	0.25
23	$4-NH2$	0.10	0.23	0.76	0.40
24	$2-N(CH_3)$	5.0	4.8	32.2	4.3
25	$3-N(CH_3)$	1.4	0.50	4.5	1.1
26	$4-N(CH_3)$	0.43	0.65	0.30	0.42
27	CH ₂ NH ₂	0.47	0.71	7.1	1.0
28	3-NH ₂ , 4-OH, 5-NO ₂	0.85	1.0	0.57	0.35
29		0.35	0.52	4.1	0.76
30		0.32	0.44	2.7	2.3
AAZ		0.06	0.09	0.076	0.01
^a Mean from three different assays, by a stopped flow technique (errors were in the range of \pm 5–10% of the reported values).					

Ser residues in the active site than other hCAs. Likely extended H-bond networks between N-nitrosulfonamide moieties and such hydrophilic residues could justify the reported low K_I values. Noteworthy, TcCA turned out to be the most affected α -CA among those herein studied (Table 2). Most derivatives inhibited TcCA in a medium nanomolar range $(0.10-0.85 \,\mu\text{M})$, except for compounds bearing 2-NH₂, 2-N(CH₃)₂, or 3- $N(CH_3)_2$ moieties at the phenyl ring (K_Is in the range 1.4− 5.0 μ M). The incorporation of primary amino groups at the *meta* or para position of the phenyl ring confers to 22 and 23, the greatest TcCA inhibitory efficacies as well as the strongest CA inhibition properties of the study. In agreement with the inhibition data previously reported,²⁰ β -CAs were generally more efficiently inhibited by N-nitrosulfonamides than α -CAs. Indeed, most $K₁s$ shown in Table 2 for [Ld](#page-5-0)cCA, MgCA, and Can2 lie into a submicromolar range. While the first isozyme is undoubtedly the most affected one among the three $(K₁s$ in the range 0.23−4.8 μ M), equally efficient inhibitions were measured against the fungal MgCA and Can2. The $4-NH_2$ -phenyl derivative 23 arose again as the most potent one against the target LdcCA (K_I of 0.23 μ M). Unlike against TcCA, 22 inhibited the isozyme comparably with the heterocyclic derivatives 29 and 30 (K _Is of 0.49, 0.52, and 0.44 μ M, respectively).

Striking target/off-target CAs selectivity profiles can be ascribed to many N-nitrosulfonamide derivatives. As a general trend, the designed compounds acted one to more than two orders of magnitude more potently against TcCA and LdcCA than ubiquitous h-isoforms CA I and II; e.g., derivative 23 showed TcCA/CA II and LdcCA/CA II inhibition ratios of 540 and 230, respectively.

The noticeable in vitro inhibition results of N-nitrosulfonamides against TcCA and LdcCA isoforms led us to study the inhibitory activity of some such inhibitors against various Trypanosoma cruzi and Leishmania forms. Compounds 22 and 23 demonstrated the most potent and selective inhibition against the target CAs and were progressed in the study in the silver salt forms.

The percentages of inhibition of the Trypanosoma cruzi epimastigote forms at different concentrations of synthetic compounds are shown in Table 3. The experiments showed that compounds 22 and 23 possess better activity than the reference drug benznidazole (Bnz[\) agains](#page-3-0)t the epimastigotes forms of Trypanosoma cruzi in both Dm28c clone and Y strain. At the

 a Determination of cytotoxicity (CC_{50}) and the selectivity index (SI_{50}) of 22 and 23 was done using RAW 264.7 macrophages. Average values of three independent experiments \pm standard deviations. SI³ = IC₅₀ Raw 267.4 cells/IC₅₀ epimastigote forms of T. cruzi Dm28c and T. cruzi Y.

Table 4. Analysis of Cytotoxicity and Trypanocidal Effect of Compounds^a

Vero cells toxicity $CC_{50} (\mu M)$ intracellular amastigotes trypomastigotes trypomastigotes compd		$IC_{50}(\mu M)$			selectivity index (SI)	
						intracellular amastigotes
	benznidazole	15.6 ± 1.9	1.7 ± 0.3	>500	>32	>294.1
0.8 ± 0.3 21.1 ± 2.8 26.4 5.2 ± 1.1 22 4.1						
8.3 ± 1.5 24.1 ± 3.6 2.9 3.9 ± 1.1 6.2 23						

a
Average values of three independent experiments \pm standard deviations. SI = IC₅₀ Vero cells/IC₅₀ trypomastigote and intracellular amastigote forms of T. cruzi, Dm28c-Luc clone.

concentration of 5.03 \pm 0.95 μ M compound 22 inhibited by 50% (IC₅₀) the proliferation of *T. cruzi* Dm28c. For *T. cruzi* Y, IC₅₀ values were reached at the concentrations of 12.00 ± 1.06 and $2.51 \pm 0.40 \mu M$ for 22 and 23, respectively. Anyhow, the two derivatives possess higher toxicity than Bnz for Raw 267.4 macrophages cells (Table 3). As a result, only 22 shows a better SI (5.87 \pm 1.14) than benznidazole (4.77 \pm 0.91) for *T. cruzi* Dm28c, whereas uniquely 23 display a higher SI for the Y strain of the parasite 11.58 \pm 2.72 with respect to the standard (8.09 \pm 0.40). Compounds 22 and 23 were also screened against both T. cruzi forms relevant to human infection. Table 4 summarizes the trypanocidal activity against the nonreplicative (trypomastigotes) and replicative (amastigotes) stages of T. cruzi, Dm28c-Luc clone. Both inhibitors 22 and 23 showed a potent activity against trypomastigotes, reaching IC_{50} values of 4- to 19.5-fold better than Bnz, respectively. For intracellular amastigotes, 22 $(IC_{50} = 5.2 \pm 1.1)$ and 23 $(IC_{50} = 8.3 \pm 1.5)$ showed lower efficacy than Bnz (IC₅₀ = 1.7 \pm 0.3). The higher toxicity than Bnz against Vero cells found for compounds 22 and 23 resulted in reduced selectivity index with respect to the reference drug (Table 4).

Minimum inhibitory concentrations (MIC) of 22 and 23 against Leishamnia infantum and L. amazonensis are shown in Table 5, whereas IC_{50} , CC_{50} , and SI values are represented in Figure 1. While compound 22 shows MIC values of 25 against both Leishmania spp, 23 exhibits a 2-fold greater activity against [L.amazo](#page-4-0)nensis than L. infantum (Table 5). These values are higher than the reference drug amphotericin B (AMP).

Table 5. Minimum Inhibitory Concentration (MIC) Assays of L. amazonensis and L. infantum \mathbf{u}

		MIC (µM)		
compd	promastigotes forms L. amazonensis	promastigotes forms L. infantum		
amphotericin B				
22	25	25		
23	12.5	25		

"Average values of three independent experiments \pm standard deviations.

Derivatives 22 and 23, respectively, display IC_{50} of 16.61 and 8.43 μ M against the promastigote forms of *L.amazonensis* and 16.64 and 17.67 μ M for *L. infantum*. The standard AMP is more effective with IC₅₀ values of 1.65 and 1.77 μ M against the two Leishmania sp. Nonetheless, the concentration of AMP, which reduced 50% of RAW 267.4 macrophage cells viability $(CC₅₀)$, is very low $(1 \mu M)$, whereas inhibitors 22 and 23 possess lower toxicity (CC₅₀ values of 29.28 and 34.89 μ M, respectively; Figure 1B). As a result of the latter, whereas the SI of the reference compound AMP was more than 2-fold higher than 22 [against](#page-4-0) [b](#page-4-0)oth Leishmanias, 23 showed a better selectivity profile than AMP against L. amazonensis (Figure 1C).

We proposed herein an innovative approach to afford agents for the treatment of Chagas di[sease and](#page-4-0) leishmaniasis based on a new CA inhibitory chemotype and on the antimicrobial properties of silver. A set of N-nitrosulfonamides and silver salts thereof were screened against a panel of nine human and pathogens isoforms, among which the targets are TcCA and LdcCA. Most such derivatives showed selective (nanomolar) inhibition of the parasite CAs over human ubiquitous ones. The best inhibitors 22 and 23 showed potent inhibition activity against various developmental forms and spp of Trypanosoma cruzi and Leishmania. The two compounds showed to be more effective than the reference drug benznidazole in inhibiting epimastigote proliferation of both T. cruzi stocks belonging to TcI (Dm28c clone) and TcII (Y strain) lineage. Anyhow, their higher toxicity than Bnz against macrophage cells led to SI comparable to the reference drug. Moreover, both tested compounds displayed 4- to 19.5-fold greater efficacy against T. cruzi forms relevant to human infection, but rather low SI values were calculated owing to low micromolar inhibition of Vero cells in comparison to Bnz.

Both L. amazonensis and L. infantum were inhibited by 22 and 23 in a low micromolar range, but less efficiently than the reference drug amphotericin B. Nevertheless, comparable SI values with the standard were calculated based on a rather less toxic effect of 22 and 23 against RAW 264.7 macrophages. The SI of 23 against L.infantum is even better than amphotericin B.

The present results make N-nitrosulfonamides innovative chemotypes to yield the selective inhibition of the target pathogens CAs over human isoforms. The reported in vitro

Figure 1. (A) Concentration that reduced the proliferation of promastigotes by 50% (IC₅₀ μ M). (B) Cytotoxic concentration that reduced 50% of RAW 267.4 cells $(CC₅₀ µM)$. (C) Selectivity index: RAW 267.4 cells $(CC₅₀)/IC₅₀$ against L. amazonensis and L. infantum.

assays against various strains of T. cruzi and L. donovani are of remarkable interest in the field of NTDs and represent a new interesting starting point for the rational CAI optimization for the treatment of Chagas disease and leishmaniasis.

■ ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsmedchemlett.8b00430.

[Synthetic procedures,](http://pubs.acs.org) characteriz[ation of compounds,](http://pubs.acs.org/doi/abs/10.1021/acsmedchemlett.8b00430) in [vitro](http://pubs.acs.org/doi/abs/10.1021/acsmedchemlett.8b00430) kinetic procedure, and antiparasitic and cytotoxicity assays (PDF)

■ AUTHO[R INF](http://pubs.acs.org/doi/suppl/10.1021/acsmedchemlett.8b00430/suppl_file/ml8b00430_si_001.pdf)ORMATION

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Notes

The authors declare no competing financial interest.

■ ABBREVIATIONS

CA, carbonic anhydrase; CAI, carbonic anhydrase inhibitor; K_I , inhibition constant; Bzn, benznidazole; AMP, amphotericin B

■ REFERENCES

(1) World Health Organization. Available from: http://www.who.int/ chagas/en/ and http://www.who.int/leishmaniasis/en/.

(2) Mackey, T. K.; Liang, B. A.; Cuomo, R.; Hafen, R.; Brouwer, K. C.; Lee, D. E. Emerging and reemerging neglecte[d](http://www.who.int/chagas/en/) [tropical](http://www.who.int/chagas/en/) [diseases:](http://www.who.int/chagas/en/) [a](http://www.who.int/chagas/en/) [review](http://www.who.int/chagas/en/) [of](http://www.who.int/chagas/en/) [ke](http://www.who.int/chagas/en/)y ch[aracteristics,](http://www.who.int/leishmaniasis/en/) [risk](http://www.who.int/leishmaniasis/en/) [factors,](http://www.who.int/leishmaniasis/en/) [and](http://www.who.int/leishmaniasis/en/) [the](http://www.who.int/leishmaniasis/en/) [policy](http://www.who.int/leishmaniasis/en/) [a](http://www.who.int/leishmaniasis/en/)nd innovation environment. Clin. Microbiol. Rev. 2014, 27, 949−979.

(3) Barrett, M. P.; Croft, S. L. Management of trypanosomiasis and leishmaniasis. Br. Med. Bull. 2012, 104, 175−196.

(4) Guedes, P. M.; Silva, G. K.; Gutierrez, F. R.; Silva, J. S. Current status of Chagas disease chemotherapy. Expert Rev. Anti-Infect. Ther. 2011, 9, 609−620.

(5) World Health Organization. Sustaining the drive to overcome the global impact of neglected tropical diseases. Second WHO report on neglected tropical diseases; 2013. Available from: http://www.who.int/ neglected_diseases/9789241564540/en/.

(6) Vermelho, A. B.; Capaci, G. R.; Rodrigues, I. A.; Cardoso, V. S.; Mazotto, A. M.; Supuran, C. T. Carbon[ic anhydrases from](http://www.who.int/neglected_diseases/9789241564540/en/) [Trypanosoma](http://www.who.int/neglected_diseases/9789241564540/en/) [and](http://www.who.int/neglected_diseases/9789241564540/en/) [Leishmania](http://www.who.int/neglected_diseases/9789241564540/en/) [as](http://www.who.int/neglected_diseases/9789241564540/en/) [anti-pro](http://www.who.int/neglected_diseases/9789241564540/en/)tozoan drug targets. Bioorg. Med. Chem. 2017, 25, 1543−1555.

(7) Ortiz, C.; Moraca, F.; Medeiros, A.; Botta, M.; Hamilton, N.; Comini, M. A. Binding Mode and Selectivity of Steroids towards Glucose-6-phosphate Dehydrogenase from the Pathogen Trypanosoma cruzi. Molecules 2016, 21, 368.

(8) Supuran, C. T. Inhibition of carbonic anhydrase from Trypanosoma cruzi for the management of Chagas disease: an underexplored therapeutic opportunity. Future Med. Chem. 2016, 8, 311−324.

(9) Capasso, C.; Supuran, C. T. Bacterial, fungal and protozoan carbonic anhydrases as drug targets. Expert Opin. Ther. Targets 2015, 19, 1689−1704.

(10) Pan, P.; Vermelho, A. B.; Capaci, G. R.; Scozzafava, A.; Tolvanen, M. E. E.; Parkkila, S.; Capasso, C.; Supuran, C. T. Cloning, characterization, and sulfonamide and thiol inhibition studies of an acarbonic anhydrase from Trypanosoma cruzi, the causative agent of Chagas disease. J. Med. Chem. 2013, 56, 1761−1771.

(11) de Menezes, D. R.; Calvet, C. M.; Rodrigues, G. C.; de Souza Pereira, M. C.; Almeida, I. R.; de Aguiar, A. P.; Supuran, C. T.; Vermelho, A. B. Hydroxamic acid derivatives: a promising scaffold for rational compound optimization in Chagas disease. J. Enzyme Inhib. Med. Chem. 2016, 31, 964−973.

(12) Syrjanen, L.; Vermelho, A. B.; Rodrigues, I. A.; Corte-Real, S.; Salonen, T.; Pan, P.; Vullo, D.; Parkkila, S.; Capasso, C.; Supuran, C. T. Cloning, characterization, and inhibition studies of a b-carbonic anhydrase from Leishmania donovani chagasi, the protozoan parasite responsible for leishmaniasis. J. Med. Chem. 2013, 56, 7372−7381.

(13) Guzel-Akdemir, O.; Akdemir, A.; Pan, P.; Vermelho, A. B.; Parkkila, S.; Scozzafava, A.; Capasso, C.; Supuran, C. T. A class of sulfonamides with strong inhibitory action against the a-carbonic

anhydrase from Trypanosoma cruzi. J. Med. Chem. 2013, 56, 5773– 5781.

(14) Alafeefy, A. M.; Ceruso, M.; Al-Jaber, N. A.; Parkkila, S.; Vermelho, A. B.; Supuran, C. T. A new class of quinazolinesulfonamides acting as efficient inhibitors against the a-carbonic anhydrase from Trypanosoma cruzi. J. Enzyme Inhib. Med. Chem. 2015 , 30, 581 −585.

(15) Pan, P.; Vermelho, A. B.; Scozzafava, A.; Parkkila, S.; Capasso, C.; Supuran, C. T. Anion inhibition studies of the a-carbonic anhydrase from the protozoan pathogen Trypanosoma cruzi, the causative agent of Chagas disease. Bioorg. Med. Chem. 2013, 21, 4472–4476.

(16) Rodrigues, G. C.; Feijo, D. F.; Bozza, M. T.; Pan, P.; Vullo, D.; Parkkila, S.; Supuran, C. T.; Capasso, C.; Aguiar, A. P.; Vermelho, A. B. Design, synthesis,and evaluation of hydroxamic acid derivatives as promising agents for the management of Chagas disease. J. Med. Chem. 2014 , 57, 298 −308.

(17) Nocentini, A.; Cadoni, R.; Dumy, P.; Supuran, C. T.; Winum, J. Y. Carbonic anhydrases from Trypanosoma cruzi and Leishmania donovani chagasi are inhibited by benzoxaboroles. J. Enzyme Inhib. Med. Chem. 2018, 33, 286-289.

(18) Ceruso, M.; Carta, F.; Osman, S. M.; Alothman, Z.; Monti, S. M.; Supuran, C. T. Inhibition studies of bacterial, fungal and protozoan bclass carbonic anhydrases with Schiff bases incorporating sulfonamide moieties. Bioorg. Med. Chem. 2015 , 23, 4181 −4187.

(19) Crespillo-Andujar, C.; Chamorro-Tojeiro, S.; Norman, F.; ́ Monge-Maillo, B.; López-Velez, R.; Pérez-Molina, J. A. Toxicity of nifurtimox as second-line treatment after benznidazole intolerance in patients with chronic Chagas disease: When available options fail. Clin. Microbiol. Infect. 2018 , 24, 1344.

(20) Nocentini, A.; Vullo, D.; Bartolucci, G.; Supuran, C. T. N-Nitrosulfonamides: A new chemotype for carbonic anhydrase inhibition. Bioorg. Med. Chem. 2016 , 24, 3612 −3617.

(21) Lansdown, A. B. Silver in health care: antimicrobial effects and safety in use. Curr. Probl. Dermatol. 2006, 33, 17–34.

(22) Lok, C. N.; Ho, C. M.; Chen, R.; He, Q. Y.; Yu, W. Y.; Sun, H.; Tam, P. K.; Chiu, J. F.; Che, C. M. Silver nanoparticles: partial oxidation and antibacterial activities. JBIC, J. Biol. Inorg. Chem. 2007, 12, 527– 534.

(23) Rai, M.; Yadav, A.; Gade, A. Silver nanoparticles as a new generation of antimicrobials. Biotechnol. Adv. 2009, 27, 76–83.

(24) Allahverdiyev, A. M.; Abamor, E. S.; Bagirova, M.; Ustundag, C. B.; Kaya, C.; Kaya, F.; Rafailovich, M. Antileishmanial effect of silver nanoparticles and their enhanced antiparasitic activity under ultraviolet light. Int. J. Nanomed. 2011, 6, 2705–2714.

(25) Minksztym, K. Synthesis of Aromatic Aminosulfonic Acid Nitroamides Synthesis. Synthesis 2007 , 12, 1819.

(26) Mathews, B. R. Benzene Sulfonnitramide, Toluene-4-sulfonnitramide, 2-nitroluene-4-sulfonnitrani ide and Some of their Salts. J. Phys. Chem. 1920, 24, 108.

(27) Khalifah, R. G. The carbon dioxide hydration activity of carbonic anhydrase. J. Biol. Chem. 1971 , 246, 2561 −2573.

(28) Supuran, C. T. Carbonic anhydrases: novel therapeutic applications for inhibitors and activators. Nat. Rev. Drug Discovery 2008 , 7, 168.

(29) Hewitson, K. S.; Vullo, D.; Scozzafava, A.; Mastrolorenzo, A.; Supuran, C. T. Molecular cloning, characterization, and inhibition studies of a beta-carbonic anhydrase from Malassezia globosa, a potential antidandruff target. J. Med. Chem. 2012, 55, 3513–3520.

(30) Schlicker, C.; Hall, R. A.; Vullo, D.; Middelhaufe, S.; Gertz, M.; Supuran, C. T.; Muehlschlegel, F. A.; Steegborn, C. Structure and Inhibition of the CO 2-Sensing Carbonic Anhydrase Can2 from the Pathogenic Fungus Cryptococcus neoformans. J. Mol. Biol. 2009, 385, 1207 −1220.