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#### **Key Points:**

- Bidirectional exchange of HCOOH was observed over a mixed forest, with a canopy‐level source; soil and below‐canopy sources were minor
- The gross canopy HCOOH source is 3% and 38% of isoprene and monoterpene C emissions and is underestimated fifteenfold in a current model
- Correlation analysis points to secondary formation, not direct emission, as the major HCOOH source in this forest ecosystem

#### **[Supporting Information:](http://dx.doi.org/10.1029/2018GL081526)**

- Supporting Information.<br>• [Supporting Information S1](http://dx.doi.org/10.1029/2018GL081526)
- [Figure S1](http://dx.doi.org/10.1029/2018GL081526)
- [•](http://dx.doi.org/10.1029/2018GL081526) [Figure S2](http://dx.doi.org/10.1029/2018GL081526)
- [•](http://dx.doi.org/10.1029/2018GL081526) [Figure S3](http://dx.doi.org/10.1029/2018GL081526)

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## $\bigcap_{i=1}^n$ **Oxidation of Volatile Organic Compounds as the Major Source of Formic Acid in a Mixed Forest Canopy**

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**Abstract** Formic acid (HCOOH) is among the most abundant carboxylic acids in the atmosphere, but its budget is poorly understood. We present eddy flux, vertical gradient, and soil chamber measurements from a mixed forest and apply the data to better constrain HCOOH source/sink pathways. While the cumulative above‐canopy flux was downward, HCOOH exchange was bidirectional, with extended periods of net upward and downward flux. Net above‐canopy fluxes were mostly upward during warmer/drier periods. The implied gross canopy HCOOH source corresponds to 3% and 38% of observed isoprene and monoterpene carbon emissions and is 15× underestimated in a state-of-science atmospheric model (GEOS‐Chem). Gradient and soil chamber measurements identify the canopy layer as the controlling source of HCOOH or its precursors to the forest environment; below‐canopy sources were minor. A correlation analysis using an ensemble of marker volatile organic compounds suggests that secondary formation, not direct emission, is the major source driving ambient HCOOH.

**Plain Language Summary** Formic acid (HCOOH) is one of the most abundant acids in the atmosphere and affects the acidity of precipitation. A number of recent studies have shown that the atmospheric abundance of HCOOH is much higher than predicted, implying some unknown or underrepresented source. Here we present new measurements of HCOOH and related species above, within, and below a mixed forest canopy and use the results to investigate its sources and sinks in this ecosystem. We find that the forest is simultaneously a source and a sink of atmospheric HCOOH, and vertically resolved measurements identify the canopy layer as the major HCOOH source for this environment. Soils have been shown to be a source of HCOOH in some cases, but we show that their influence is unimportant for this ecosystem. The magnitude of the gross HCOOH source from this forest is 15 times higher than predicted in a current atmospheric model. A correlation analysis suggests that the main HCOOH source from this forest is oxidation of other compounds rather than direct emissions from vegetation.

## **1. Introduction**

Organic acids are a significant component of ambient nonmethane organic carbon and contribute to aqueous‐phase acidity and OH reactivity (Galloway et al., 1982; Jacob, 1986). As photochemical products of volatile organic compound (VOC) oxidation, organic acids can also serve as integrated markers of the overall VOC degradation cascade. Formic acid (HCOOH) is among the most abundant carboxylic acids in the atmosphere (Khare et al., 1999; Millet et al., 2015; Paulot et al., 2011); however, its sources are poorly known. Here we present new HCOOH canopy flux, gradient, and soil chamber measurements from a mixed forest in Michigan, United States, and interpret the results in terms of their implications for HCOOH sources and sinks.

Photochemical production is thought to be the dominant source of atmospheric HCOOH globally, contributing 60–80% of its total known budget (Millet et al., 2015; Stavrakou et al., 2012). In particular, the OH‐ initiated oxidation of isoprene produces HCOOH via several known pathways, with a total yield that has been estimated at 13% (Millet et al., 2015). HCOOH is also formed from formaldehyde oxide (CH<sub>2</sub>OO, a stabilized Criegee intermediate derived from terminal alkene ozonolysis), both directly (Nguyen et al., 2016) and via hydroxymethyl hydroperoxide (HMHP; Allen et al., 2018). Ozonolysis precursors of HCOOH thus encompass a wide suite of alkenes including isoprene, many monoterpene isomers, other terpenoids, and simpler alkenes. Reported HCOOH yields from monoterpene photooxidation and ozonolysis range from

<sup>1</sup>–50% (Larsen et al., 2001; Lee, Goldstein, Kroll, et al., 2006), depending on the species; average yields of 16% (OH) and 8% (ozonolysis) have been assumed in recent large-scale modeling studies (Millet et al., 2015; Paulot et al., 2011). Additional known secondary HCOOH sources include OH‐driven oxidation of terminal alkynes (Hatakeyama et al., 1986) and photo‐tautomerization of acetaldehyde followed by oxidation of the resulting enol (Millet et al., 2015; Peeters et al., 2015; Shaw et al., 2018).

HCOOH also has several primary sources. Direct emissions from soil (Mielnik et al., 2018; Sanhueza & Andreae, 1991) and plants (Kesselmeier et al., 1998) have been observed, with the latter exhibiting light and temperature dependence. Other identified primary sources include agriculture (Ngwabie et al., 2008) and combustion of biomass (Chaliyakunnel et al., 2016; Goode et al., 2000) and fossil fuels (Kawamura et al., 1985; Talbot et al., 1988). Heterogeneous sources have also been proposed, including in‐cloud formaldehyde oxidation (Jacob, 1986; Lelieveld & Crutzen, 1991), as well as aging of organic aerosol via OH (Vlasenko et al., 2008),  $O_3$  (Eliason et al., 2003; Pan et al., 2009), and photolysis (Malecha & Nizkorodov, 2016; Park et al., 2006). Wet and dry deposition are the predominant HCOOH sinks; together with photochemical loss and a minor contribution from dust uptake, these yield an overall atmospheric lifetime of 2–4 days (Chebbi & Carlier, 1996; Paulot et al., 2011; Stavrakou et al., 2012).

Despite the large number of identified primary and secondary HCOOH sources, current chemical transport models typically underestimate observed concentrations by at least twofold (Millet et al., 2015; Paulot et al., 2011; Stavrakou et al., 2012). A number of recent studies have investigated this discrepancy with the aim of characterizing potential missing or underestimated HCOOH sources. For example, Bannan et al. (2014, 2017) measured HCOOH levels over London UK of up to 13 ppb and invoked traffic emissions and biogenic VOC photooxidation as likely sources. In the Colorado Front Range, Mattila et al. (2018) found evidence for photochemical and ground‐level HCOOH sources and speculated that heterogeneous ozonolysis of surface organics might contribute to the latter. Mungall et al. (2018) measured surprisingly high HCOOH concentrations (up to 11 ppb) over the Arctic tundra, which they attributed to unidentified surface- or near-surfacesources; elevated concentrations occurred during cold/overcast as well as warm/sunny periods. Nguyen et al. (2015) observed net downward HCOOH fluxes over a deciduous forest in the Southeastern United States; however, observed deposition velocities were only half of predicted values, and the authors invoked a gross daytime canopy source of ~1 nmol/m $^2$ /s to account for the difference. Eddy flux measurements by Schobesberger et al. (2016) over a boreal pine/spruce forest revealed a net upward HCOOH flux that was both light and temperature dependent. Neither Nguyen et al. (2015) nor Schobesberger et al. (2016) were able to determine whether the observed canopy source was due to direct HCOOH emissions from vegetation or soil, in‐canopy chemical production, or heterogeneous surface processes.

Here we present measurements of HCOOH and related VOCs by high-resolution mass spectrometry from the 2016 PROPHET‐AMOS campaign in a mixed deciduous/coniferous forest at the University of Michigan Biological Station. Observations include (i) canopy‐level flux measurements by eddy covariance; (ii) hourly vertical gradients of VOC mixing ratios through the forest canopy; and (iii) diurnal soil fluxes. Together, these measurements provide a more comprehensive picture for understanding the forest– atmosphere exchange of HCOOH than has been available previously. Based on the flux, gradient, and soil chamber measurements of HCOOH and their correlation patterns with marker VOCs of known origin, we argue that in-canopy secondary production (rather than direct emission from plants or soils) is the main HCOOH source in this forest ecosystem.

## **2. Methods**

Canopy and vertical gradient measurements were performed on the PROPHET tower during the July 2016 AMOS field campaign at University of Michigan Biological Station, Michigan, United States (Millet et al., 2018). The surrounding forest is dominated by aspen, birch, and red oak (upper canopy) and by white pine, red maple, beech, and red oak (lower canopy; Bryan et al., 2015; Palik & Pregitzer, 1992). Soils at the site are excessively drained spodosols (Seok et al., 2015). Measurements of HCOOH and related VOCs during PROPHET‐AMOS were made with a Proton Transfer Reaction‐Quadrupole Interface Time‐Of‐Flight Mass Spectrometer (PTR‐QiTOF; Ionicon Analytic Gmbh). The hourly sampling procedure included direct ecosystem‐scale VOC flux measurements by eddy covariance, plus vertical profiling to quantify in‐canopy VOC gradients. A detailed description of instrumental performance, calibration, inlet configuration,



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**Figure 1.** Time series of HCOOH and related measurements during PROPHET-AMOS. Plotted is the observed (a) photosynthetic photon flux density (PPFD) and friction velocity (*u*\*), (b) temperature and dew point, (c) rainfall and soil moisture, (d) HCOOH mixing ratios, (e) net canopy HCOOH flux, and (f) net canopy HCOOH exchange velocity. Also shown in (d)–(f) are the simulated HCOOH mixing ratios, fluxes, and exchange velocities as predicted by GEOS‐Chem (in red). For (e) and (f), the gray and orange lines show the full observational and model data sets, respectively, while the black and red show the same datasets after filtering for turbulent conditions ( $u^*$  > 0.3 m/s).

sampling procedure, flux calculations, and quality control is provided by Millet et al. (2018) and summarized in supporting information S1.

During this study we also measured diurnal fluxes of HCOOH and other VOCs from the forest floor using four dynamic soil chambers. Details on these measurements and on other supporting observations used here are provided in supporting information S1 (Pape et al., 2009).

Finally, we compare the observed HCOOH concentrations and fluxes with predictions from a high-resolution ( $0.25^\circ \times 0.3125^\circ$ ) GEOS-Chem model simulation nested over North America, with model details provided in supporting information S1.

## **3. Results and Discussion**

### **3.1. Observations Reveal Bidirectional HCOOH Exchange**

Figure 1 shows the above-canopy HCOOH mixing ratios, fluxes, and exchange velocities measured throughout the campaign. Mixing ratios ranged from 0.1–5.3 ppb, with an overall mean of 1.3 ppb and a daytime (10:00–17:00 EDT) mean of 1.6 ppb. When aggregated over the entire campaign, the net above-canopy HCOOH flux was downward (into the canopy), averaging  $-3.9 \times 10^8$  molec/cm<sup>2</sup>/s (mean exchange velocity: −0.07 cm/s). However, the time series in Figure 1 reveals extended periods of net upward as well as net downward HCOOH exchange, indicating a bidirectional flux with counteracting gross source and sink effects.

Figure 2 shows the corresponding mean diel profiles of the HCOOH mixing ratios, fluxes, and exchange velocities. On average, the HCOOH mixing ratios peak during afternoon (at ~1.7 ppb) before dropping after sunset to <1 ppb. The diel mean fluxes (which as shown in Figure 1 combine distinct periods with net downward and upward exchange) exhibit net upward exchange during daytime, again peaking during afternoon, with net fluxes into the canopy at night. In‐canopy storage changes (HCOOH burden fluctuations below the sampling inlet; Figure 2b) account for ~15% of the net 24‐hr integrated above‐canopy HCOOH flux and do not significantly change the diel cycle. The weak measured soil fluxes (Figure 2b) argue against an important role for soil HCOOH emissions in this ecosystem.

Measured net exchange velocities (Figure 2c) follow a similar diel profile as the fluxes. The observed daytime (10:00–17:00 EDT) mean exchange velocity for the campaign of  $0.42 \pm 0.48$  ( $\mu \pm \sigma$ ) cm/s is lower than that reported by Schobesberger et al. (2016) for a boreal forest

 $(0.7 \pm 1.7 \text{ cm/s})$  and larger than the net downward exchange observed for a mixed temperate forest in the Southeastern United States ( $-1.0 \pm 0.4$  cm/s; Nguyen et al., 2015) and for a tropical forest in Amazonia (−0.2 cm/s; Kuhn et al., 2002).

Also plotted in Figures 1 and 2 are the HCOOH mixing ratios, fluxes (net, gross upward, and gross downward), and exchange velocities predicted by GEOS‐Chem. The observed HCOOH mixing ratios are underpredicted in the model by a mean factor of 3–5, consistent with prior findings (Millet et al., 2015; Paulot et al., 2011; Yuan et al., 2015). Furthermore, while the observations reveal upward as well as downward net HCOOH fluxes over the canopy, the simulated net fluxes are consistently downward (Figures 1 and 2). The net deposition measured at night (Figure 2b) is, on average, well-captured by the model; however, the model is clearly missing the large gross canopy HCOOH source apparent in the daytime observations.

We see in Figure 2b that the gross canopy HCOOH source estimated from the model output is significantly less than the mean net flux observed during daytime, despite the fact that the latter reflects the sum of gross upward (emission  $+$  in-canopy photochemical production) and downward (deposition  $+$  in-canopy





Figure 2. Mean diel profiles of HCOOH (a) mixing ratios, (b) fluxes, and (c) exchange velocities as measured during PROPHET‐AMOS and simulated by GEOS‐Chem. The solid black lines and shaded gray regions show the observed mean and associated 95% confidence interval, with the corresponding model values shown in red. Shaded regions for the model fluxes and exchange velocities span upper and lower limits for in‐canopy production and loss as described in supporting information S1. Along with the measured above‐canopy flux, (b) shows the observed contributions from canopy storage (orange) and soils (brown; the shaded region here spans the range from the three chambers with exposed soil after correcting for wall effects based on the blank chamber). Also shown in (b) are the model‐derived gross upward (green) and downward (blue) fluxes.

photochemical loss) exchange. We can estimate the gross HCOOH source flux that is implied by the observations by correcting for deposition (incanopy chemical loss is negligible for HCOOH due to its ~25‐day photochemical lifetime; Atkinson et al., 2006). To accomplish this, we apply the HCOOH deposition velocities computed by the model to the observed HCOOH number densities and add the resulting loss term to the measured above-canopy net flux. In this way, we derive from the observations a 24-hr mean gross HCOOH canopy source of  $(5.1 \pm 0.3) \times 10^{10}$ molec/cm<sup>2</sup>/s (mean  $\pm$  standard error), with corresponding daytime (10:00–17:00 EDT) and nighttime (22:00–05:00 EDT) values of  $(10.9 \pm 0.6) \times 10^{10}$  and  $(1.1 \pm 0.2) \times 10^{10}$  molec/cm<sup>2</sup>/s, respectively. This implied gross canopy HCOOH source corresponds to 2.9% and 38% (on a carbon basis) of the observed 24‐hr mean isoprene and monoterpene fluxes, respectively. It is also  $15\times$  larger than the 24-hr gross HCOOH source (emissions + photochemical production) predicted by GEOS-Chem  $(20 \times$  and  $5 \times$  larger for the daytime-only and nighttimeonly values).

The measured HCOOH mixing ratios and fluxes are the highest (most positive) under warm and low‐relative humidity (RH) conditions. For example, we see in Figure 1 that upward net fluxes tend to occur under large dew point depressions. Figure 3 quantifies this relationship, plotting the measured fluxes as a function of temperature, light, and humidity: On average, the daytime net fluxes become more positive with increasing light and temperature and more negative with increasing RH (or dew point). The daytime temperature dependence for the modeled net HCOOH fluxes is the reverse of that seen in the observations, with fluxes becoming more negative rather than more positive under warmer conditions—a tendency driven by increasing model deposition fluxes at warmer temperatures. Figure 3 also shows that the light dependence for the modeled net HCOOH flux is weaker than observed.

We see in Figures 3 and S1 that, for the most part, the net canopy HCOOH fluxes are upward for RH  $\leq 60\%$  and downward for RH  $\geq 70\%$ . It is possible that this behavior partly reflects the adsorption of HCOOH to wet surfaces with subsequent release from thin film water upon drying—as observed for the weak acid HONO at the same site (He et al., 2006). Karl et al. (2004) previously estimated the HCOOH uptake velocity to dew‐ wetted surfaces in a tropical forest at 0.06–0.08 cm/s, versus the nighttime deposition velocities seen here approaching ~1 cm/s (Figure 2). However, the extent of aqueous‐phase uptake will depend on several poorly known factors, including leaf surface area, the pH of the liquid, presence of deposited mineral dust, and any HCOOH/formate metabolism by vegetation or surface‐inhabiting microbes.

## **3.2. In‐Forest HCOOH Gradients Point to a Canopy‐Driven Secondary Source**

The bidirectional exchange (and implied in‐forest source) of HCOOH seen in the above-canopy fluxes is also evident in the measured vertical gradients. Figure 4 shows a diel curtain plot of HCOOH in‐canopy vertical gradients, averaged over the entire PROPHET‐AMOS campaign. The nighttime HCOOH mixing ratios decrease with depth into the canopy,

consistent with the net downward exchange measured at that time (Figure 2c). During daytime, an HCOOH maximum appears in the midcanopy (~20 m), implying a direct and/or indirect source from foliage. The gradient measurements do not reveal a surface-level HCOOH enhancement at any time of day, which



**Figure 3.** Dependence of canopy HCOOH fluxes on environmental drivers. The top row shows measured (black) and modeled (red) HCOOH fluxes plotted as a function of temperature, light (photosynthetic photon flux density [PPFD]), and relative humidity (RH). For the temperature and RH plots, daytime (PPFD > ~1,500 µmol/m<sup>2</sup>/s; black and red) and other (gray and orange) values are indicated separately. The large symbols and fit lines show the mean fluxes binned by quantile of the independent variable. The thin green and blue lines show the corresponding dependencies for the modeled gross upward and downward HCOOH fluxes, respectively. The bottom row shows curtain plots of the measured HCOOH fluxes as a function of temperature, light, RH, and dew point.

supports the soil chamber data in showing that the forest floor is not an appreciable HCOOH source in this environment.

Along with HCOOH, we also measured hourly vertical gradients for all the VOCs across the mass spectrum detected by PTR‐QiTOF (636 ions). These in‐canopy gradients for other VOCs of known origin can



Figure 4. In-canopy vertical gradients of HCOOH and correlations with tracers of known origin. (a) Curtain plot showing the mean diel cycle of HCOOH vertical gradients for the PROPHET‐AMOS campaign. (b) Mean hourly correlations between the HCOOH vertical gradients and those for selected other VOCs: *m/z* 33.034 = methanol; *m/z* 59.050 = acetone; *m/z* 61.029 = acetic acid/glycolaldehyde; *m/z* 65.021 = CH5O3 +; *m/z* 69.070 = isoprene; *m/z* 71.050 = MVK + MACR; *m/z* 73.027 = methylglyoxal;  $m/z$  75.045 = hydroxyacetone;  $m/z$  89.022 = C<sub>3</sub>H<sub>5</sub>O<sub>3</sub><sup>+</sup>;  $m/z$  113.055 = C<sub>6</sub>H<sub>9</sub>O<sub>2</sub><sup>+</sup>;  $m/z$  $137.132$  = monoterpenes;  $m/z$  205.196 = sesquiterpenes.

be used as spatial-temporal fingerprints for understanding the factors driving observed HCOOH fluxes. Figure S2 shows diel mean vertical gradients for selected marker VOCs: isoprene, monoterpenes, methanol, acetone, acetic acid/glycolaldehyde, *m/z* 65.021 (CH<sub>5</sub>O<sub>3</sub><sup>+</sup>), methyl vinyl ketone + methacrolein (MVK + MACR), and  $m/z$  113.055 ( $C_6H_9O_2^+$ ). For isoprene, we see peak concentrations from midday through afternoon, with a relatively sharp in‐canopy maximum around 20 m (and low concentrations at night), consistent with light‐ and temperature‐driven emissions from the aspen‐dominated overstory. Monoterpenes exhibit a very different pattern, with concentration peaks during morning and evening in the lower canopy, reflecting the influences of temperature-dependent (and partially light-independent; Millet et al., 2018; Ortega et al., 2007) emissions, a direct source from the white pine and red maple understory, and atmospheric mixing.

Methanol and acetone are likewise directly emitted by vegetation (Guenther et al., 2012). While both species can also be produced from the oxidation of biogenic precursors, this is thought to be a small source compared to the direct biogenic flux (Fischer et al., 2012; Millet et al., 2008). We should thus expect in-canopy variability of methanol and acetone during PROPHET‐AMOS to be driven by direct emissions and depositional uptake. Figure S2 shows that very similar gradient patterns are seen for both species. At night, concentrations decrease with depth in the canopy due to deposition, while during daytime, an afternoon peak emerges that appears to originate in both the upper and lower canopies—perhaps because the capacity to emit methanol and acetone is relatively uniform across woody plants (Guenther et al., 2012).

Sources for other species in Figure S2 are believed to be dominated by secondary production, that is, oxidation of precursor VOCs. Acetic acid/glycolaldehyde (produced from monoterpene and isoprene oxidation; Dibble, 2004; Lee, Goldstein, Keywood, et al., 2006; Lee, Goldstein, Kroll, et al., 2006; Paulot et al., 2011), *m/z* 65.021 (either HMHP or a fragment of a larger oxygenated VOC [OVOC]), *m/z* 75.045 (hydroxyacetone), and *m/z* 113.055 (a likely monoterpene oxidation product; Lee, Goldstein, Keywood, et al., 2006; Lee, Goldstein, Kroll, et al., 2006) all exhibit gradient patterns that are very similar to those of HCOOH (Figure 4), with net nighttime uptake and a diffuse midafternoon peak that is centered around midcanopy. The diel cycle for  $MVK + MACR$  (first-generation products of isoprene oxidation) is somewhat distinct, with a peak around 10:00 EST that suggests a larger role for entrainment in this case.

We performed a correlation analysis to quantify the level of agreement between the hourly averaged vertical gradients of HCOOH and those of other species. Results are shown in Figure 4 for the marker VOCs discussed above, plus sesquiterpenes,  $m/z$  73.027 ( $C_3H_5O_2^+$ ; likely methyl glyoxal),  $m/z$  75.045 ( $C_3H_7O_2^+$ ; hydroxyacetone/propanoic acid), and  $m/z$  89.022 (C<sub>3</sub>H<sub>5</sub>O<sub>3</sub><sup>+</sup>; possibly pyruvic acid). Figure 4 shows that the monoterpene and sesquiterpene gradients are weakly or negatively correlated with that of HCOOH at all times, indicating dissimilar in‐canopy source/sink distributions. The isoprene gradients exhibit only a weak correlation with HCOOH during daytime (mean  $R = 0.38$  for 10:00–17:00 EDT) that disappears at night. Acetone and methanol both have similar vertical structure to that of HCOOH at night (mean  $R = 0.94$  and 0.99, respectively, for 22:00–05:00 EDT), likely reflecting a common net sink to depositional uptake, but the daytime vertical gradients for these directly emitted VOCs do not match those of HCOOH (mean  $R < 0.3$  for both species; Figure 4).

Overall, of the 636 detected ions, 25 have mean vertical gradients that have  $R > 0.5$  with that of HCOOH during every hour of the day, thus implying similar in-canopy source-sink distributions. Of these, 14 can be assigned a molecular formula according to the procedure outlined in Millet et al. (2018), excluding isotopes:  $C_4H_5O_3^+$ ,  $C_2H_5O_2^+$  (acetic acid/glycolaldehyde),  $C_5H_7O_2^+$ ,  $C_5H_7O_3^+$ ,  $C_2H_3O^+$  (common OVOC fragment),  $C_3H_5O_3^+$ ,  $C_6H_9O_2^+$  (likely terpene oxidation product),  $O_2N^+$  (fragment of peroxyacetyl nitrate and other organic nitrates),  $C_3H_7O_2^+$  (hydroxyacetone/propanoic acid),  $C_5H_5O_2^+$ ,  $C_4H_7O_4^+$ ,  $CH_5O_3^+$  (HMHP or OVOC fragment),  $C_6H_{11}O_3^+$ , and  $C_3H_5O_2^+$  (methylglyoxal).

A similar correlation analysis was performed with respect to the HCOOH mean diel cycles at each sampling height, with example results shown in Figure S3. Here, 313 detected ions exhibited *R* > 0.5 with the HCOOH diel cycle for all heights (75 had minimum *R* > 0.9), showing that the diel cycles at any given height are a less sensitive diagnostic for resolving VOC source/sink patterns than are the vertical gradients. Finally, since the observed HCOOH flux time series (Figure 1) reveals distinct periods with net upward and net downward exchange, we repeated the above analyses for data subsets reflecting each of those scenarios: 3–<sup>5</sup> July + 22–23 July (upward HCOOH flux tendency) and 11–12 July (downward HCOOH flux tendency). Among identified ions, only *m/z* 61.029 (acetic acid/glycolaldehyde), *m/z* 65.021 (HMHP or OVOC fragment), and *m/z* 75.045 (hydroxyacetone/propanoic acid) exhibited high correlation with HCOOH in all cases (i.e.,  $R > 0.5$  for vertical gradients at all hours in the diel mean and  $R > 0.9$  for mean diel cycles at all heights). As before, the VOCs with the most similar vertical and diel patterns to HCOOH all correspond to oxidized compounds, most with two or more oxygen atoms.

Together, the above analyses suggest a rapid in‐canopy photochemical (rather than direct emission) HCOOH source: The HCOOH gradients do not match well with those for tracers of direct biogenic emissions (isoprene, monoterpenes, sesquiterpenes, methanol, and acetone) and instead strongly resemble those of oxidized VOCs whose main atmospheric source is thought to be secondary chemical production. Potential mechanisms for such a secondary HCOOH source are discussed in supporting information S2 (Crounse et al., 2012; Finlayson‐Pitts et al., 2003; Jud et al., 2016; Li et al., 2018; Neeb et al., 1997; Paulot et al., 2009; Welz et al., 2014). A separate possibility is that the canopy source of HCOOH arises from direct emissions that are dissimilar from those of any available primary emission tracers (thus causing the poor correlations in Figure 4b). Overall, however, while a mainly primary HCOOH source cannot be definitively ruled out based on the data at hand, a mainly secondary HCOOH source seems the simplest and most likely explanation.

## **4. Conclusions**

We presented new measurements of HCOOH mixing ratios, net above-canopy fluxes, vertical in-canopy gradients, and soil fluxes by high‐resolution PTR‐QiTOF over a mixed forest in Michigan, United States, during the PROPHET-AMOS study. Detected net fluxes were bidirectional, mostly upward for RH < 60% and mostly downward for  $RH > 70\%$ . A state-of-science chemical transport model (GEOS-Chem CTM) underestimates the implied gross canopy HCOOH source by on average fifteenfold and misdiagnoses the temperature and light dependence of the net exchange (implying a missing *<sup>T</sup>*‐ and *<sup>h</sup>ν*‐sensitive source). Soils are a negligible HCOOH source in this environment. A correlation analysis of the vertical and diel patterns of HCOOH variability based on other VOC tracers points to secondary chemical production, rather than primary emission from plants or soil, as the most likely HCOOH source for this forest ecosystem.

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