

## A Postmortem Study to Compare Agonist and Antagonist 5-HT<sub>1A</sub> Receptor-binding Sites in Alzheimer's Disease

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### SUMMARY

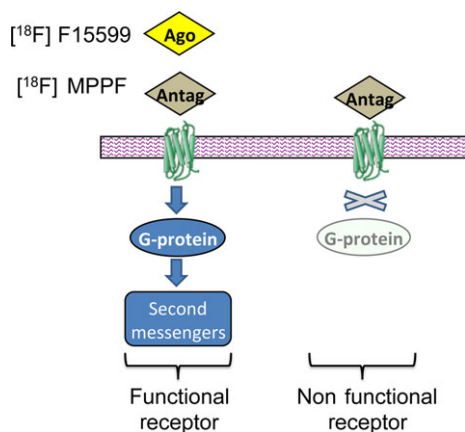
**Aims:** Positron emission tomography (PET) imaging using 5-HT<sub>1A</sub> receptor radioligands shows a decreased expression of this serotonin receptor in the hippocampus of patients with Alzheimer's disease (AD) at advanced stages. However, previous 5-HT<sub>1A</sub> receptor radioligands used in human imaging were antagonists, thought to bind to 5-HT<sub>1A</sub> receptors in different functional states (i.e., both the one which displays high affinity for agonists and is thought to mediate receptor activation, as well as the functional state which has low affinity for agonists). Comparing the PET imaging obtained using an agonist radioligand, which binds selectively to the functional state of the receptors, with the PET imaging obtained using an antagonist radioligand would therefore provide original information on 5-HT<sub>1A</sub> receptor impairment during AD. **Methods:** Quantitative autoradiography using <sup>18</sup>F-F15599 and <sup>18</sup>F-MPPF, a 5-HT<sub>1A</sub> agonist and antagonist, respectively, was measured in hippocampi of 18 patients with AD. **Results:** Functional 5-HT<sub>1A</sub> receptors, labeled by <sup>18</sup>F-F15599, represented ~35% of total receptors, as estimated by <sup>18</sup>F-MPPF labeling. <sup>18</sup>F-F15599 binding decreased in dentate gyrus of patients with AD, as indicated by Braak's stages. In contrast, binding of <sup>18</sup>F-MPPF was statistically unchanged. **Conclusion:** These *in vitro* results support testing the concept of functional PET imaging using agonist radioligands in clinical studies.

### Introduction

Positron emission tomography (PET) is a nuclear medicine imaging modality that can be used to study brain function and neurochemistry of small animals [1], medium-sized animals [2], and human subjects [3]. The development of PET radioligands allows the *in vivo* exploration of an increasing variety of central nervous system targets. These include numerous monoamine receptor subtypes, such as serotonin 5-HT<sub>1A</sub> receptors which are highly expressed in the hippocampus and are known to be important for regulation of memory processes [4]. Because early cognitive deficits in Alzheimer's disease (AD) concern episodic memory associated with neurofibrillary tangles in the hippocampus [5], several PET studies have focused on the status of 5-HT<sub>1A</sub> receptors in the hippocampus of patients with AD. While a majority of these studies, using the radiolabelled antagonist <sup>18</sup>F-MPPF, have shown a decrease of hippocampal 5-HT<sub>1A</sub> receptor density in patients with advanced AD [6,7], divergent results have been

reported in the case of patients at predementia stages of the disease [8,9]. One hypothesis to account for the observations is that compensatory mechanisms of 5-HT<sub>1A</sub> receptor regulation may intervene at early stages of AD. If so, this receptor may constitute a procognitive target in AD [10].

The present study is based on the fact that 5-HT<sub>1A</sub> receptors have been shown to exist in different G-protein-coupling states: (1) a state that has high affinity for agonists (i.e., in an "active state") when the receptor is coupled with G-proteins; and (2) a state that has low affinity for agonists when the receptor is uncoupled from G-proteins [11–14]. The binding of agonists to G-protein-coupled receptors is considered to elicit a functional response and growing evidence suggests that changes in receptor/G-protein coupling may be involved in neurodegenerative disorders [15]. In this context, comparing the receptor binding of an antagonist (which indiscriminately labels the receptor in different states) with that of an agonist (which preferentially labels G-protein-coupled receptors, Figure 1) could therefore provide information concerning the proportion of "functional



**Figure 1** Schematic representation of <sup>18</sup>F-MPPF antagonist binding (Antago) indiscriminately labeling 5-HT<sub>1A</sub> receptors in a G-coupled receptor state (functional and linked to the second messengers) and in uncoupled state (nonfunctional state). The agonist, <sup>18</sup>F-F15599 (Ago), binds specifically the G-coupled receptor state (functional receptors) [19]. Note that PET radioligands, at tracer dose, occupy only a small fraction of the available receptors.

receptors" [16]. To our knowledge, no imaging study has compared in patients with AD the binding of an agonist versus an antagonist 5-HT<sub>1A</sub> PET radiotracer, both of which are amenable to further imaging studies *in vivo*.

The aim of this study was to investigate on human brain tissue whether it is possible to obtain distinct 5-HT<sub>1A</sub>-binding profiles at different stages of AD, depending on the pharmacological profile of the PET radioligand. We hypothesized that an antagonist radioligand, which labels the entire population of 5-HT<sub>1A</sub> receptors, would have a different binding pattern to that of an agonist radioligand which would be anticipated to label only those 5-HT<sub>1A</sub> receptors that are coupled to G-proteins (i.e., in their "active state").

## Methods

### Human Subjects, Tissue Selection, and Neuropathological Staging

Frozen samples were obtained from the brain banks of two University Hospitals (Hospices Civils de Lyon and Assistance Publique - Hôpitaux de Paris). These subjects were selected from specific individuals with cognitive deficits or premortem suspected AD cases (all came from Alzheimer brain collections). Their use was approved by the brain bank's scientific review committee. In total, samples of 18 subjects were dissected from each hemisphere (Table 1).

Anterior hippocampal sections from one hemisphere were fixed in formalin solution and cut into blocks for neuropathological staging. Briefly, after rehydration, 5- $\mu$ m-thick sections were incubated overnight at 4°C with a monoclonal primary antibody directed against hyperphosphorylated tau (Innogenetics Br-03, clone AT8, dilution 1:500; Zymed Lab-SA detection system). Each case was given a Braak's stage based on the intensity of staining of neurofibrillary tangles (NFTs), according to the BrainNet Europe

**Table 1** Demographic data of the brains

Braak stages	0	I–II–III	IV–V–VI
n (total = 16)	4	6	8
Age (years)	61.5 $\pm$ 5.4	74.8 $\pm$ 7.7	81.7 $\pm$ 12.2
Gender (M/F)	4/0	4/2	4/4
PMI (h)	29.8 $\pm$ 3.9	30.5 $\pm$ 12.9	27.8 $\pm$ 5.9

n, number of patients; M/F, male/female; PMI, postmortem interval.

consortium [17], an adaptation of the Braak and Braak classification [18]. For autoradiography experiments (see below), the contralateral hemisphere was cut at the level of anterior hippocampus and immediately stored at  $-80^{\circ}\text{C}$  (without formalin fixation). Consecutive 30- $\mu$ m-thick sections were cut at  $-20^{\circ}\text{C}$  (10/subject), mounted on gelatin-coated slides, and stored at  $-80^{\circ}\text{C}$ .

### PET Radioligand Synthesis and Quality Controls

Because of the short half-life of fluorine-18 (110 min), <sup>18</sup>F-MPPF and <sup>18</sup>F-F15599 were synthesized in an automated radiosynthesizer (NEPTIS Synthesizer, ORA, Philippeville, Belgium) on the same days when autoradiography experiments were carried out (see below), according to previously described radiochemical pathways [19]. Their chemical and radiochemical purities were higher than 98%, as determined by HPLC, and their specific activity at time of autoradiography was systematically calibrated at 37 GBq/ $\mu$ mol (1 Ci/ $\mu$ mol).

### Quantitative Autoradiography with PET Radiotracers

Defrosted slides were incubated for 20 min in Tris phosphate-buffered saline buffer (138 mM NaCl, 2.7 mM KCl, pH adjusted to 7.5) containing 111 kBq/mL of <sup>18</sup>F-MPPF or <sup>18</sup>F-F15599 (i.e., 3 nM). Nonspecific binding was determined in duplicate serial sections co-incubated with 10  $\mu$ M serotonin (Sigma-Aldrich, Saint-Quentin Fallavier, France). For verification of the agonistic binding of <sup>18</sup>F-F15599, the corresponding buffer was supplemented with Gpp(NH)p (10  $\mu$ M), a nonhydrolysable analogue of GTP decoupling G-protein-coupled receptors.

After incubation, the slides were dipped in cold buffer and distilled water (4°C) then dried and juxtaposed to a phosphor imaging plate for 60 min (BAS-5000, Fujifilm, Tokyo, Japan). All films were analyzed by a computer-assisted image analysis system (Multigauge, Fujifilm), and regions of interest (CA1, dentate gyrus) were drawn manually, according to a human brain atlas [20] and confirmed by a following eosin-hematoxylin staining. Quantitation in each region of interest was performed by measuring the average optical density in adjacent brain sections.

In parallel, calibration standards were prepared from rat brain tissue homogenates. Briefly, Potter-homogenization of freshly extracted rat brains was performed in controlled temperature. Proteins of the homogenates were quantified by a chemistry analyzer (Architect Ci8200, Abbott Diagnostics, Lake Forest, IL, USA). Their exact wet mass was determined after aliquoting in micro-vials, before  $-80^{\circ}\text{C}$  congelation. On the day of radioligand syntheses, increasing activities of <sup>18</sup>F-radioligand (3.13, 6.25,

12.5, and 25  $\mu$ Ci) were mixed with 50  $\mu$ L of these defrosted homogenates in micro-vials (from 25 to 300 fmol/mg, ligand/protein). These radioactive micro-vials were immediately frozen in 2-methylbutane cooled with dry ice; the frozen core samples were extracted and cut into 30  $\mu$ m coronal sections using a  $-20^{\circ}$ C cryostat. These extemporaneous standards were juxtaposed to the same imaging plates used for the human tissue, and signal to concentration curves were generated. Nonspecific binding was subtracted from the total binding to determine the specific binding, and measurements were converted into fmoles/mg of protein, according to the calibration curve obtained from the standards. In preliminary studies, homogenized human brain tissue was used concurrently to rat brain tissue for calibration standard preparation. As concentration curves with human tissue were strictly superimposable on curves with rat tissue, the latter was then used for the all experiments.

### Statistical Analysis

Statistically significant variations in radioligand binding were measured by Mann–Whitney nonparametric test (GraphPad Prism 6 for Mac OS X, San Diego, CA, USA).

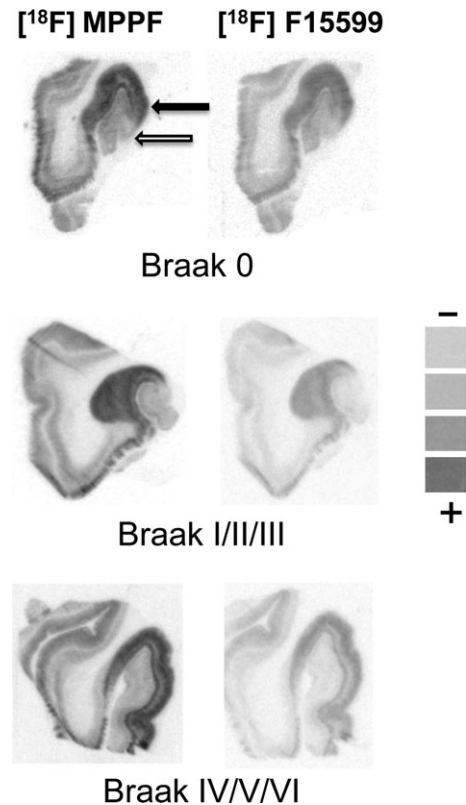
### Results

The neuropathological staging centered on the characteristic distribution of neurofibrillary tangles and NTs and the corresponding six stages were reassembled according to the BrainNet Europe consortium [17]. Stage 0 was characterized by a total absence of neurofibrillary pathology and was used as control group ( $n = 4$ ); stages I, II, and III had changes confined to the transentorhinal and entorhinal regions, and a beginning of AT8 immunopositive NTs in temporo-occipital and temporal cortices, respectively ( $n = 6$ ); stages IV, V, and VI had marked destruction of these regions, extending to isocortical association areas, namely the occipital cortex ( $n = 8$ ).

For quantitative autoradiography, the addition of Gpp(NH)p or of an excess of serotonin in the buffer led to a 70% and 90% decreases of the  $^{18}$ F-F15599 binding in the dentate gyrus and the CA1 area tissues, respectively (experiments in duplicate for the 18 subjects). In the same conditions, no significant modification of  $^{18}$ F-MPPF binding was measured after addition of Gpp(NH)p. At each Braak's stage, the total amount of 5-HT<sub>1A</sub> binding sites ( $^{18}$ F-MPPF) was significantly 3-fold higher in the CA1 area than in the dentate gyrus (Figure 2). During the first Braak's stage (0), the proportion of 5-HT<sub>1A</sub> agonist binding sites measured by the ratio of  $^{18}$ F-MPPF/ $^{18}$ F-F15599 was 40% and 35% in the CA1 area and in the dentate gyrus, respectively. In the dentate gyrus,  $^{18}$ F-F15599 labeling fell significantly by 40% and 50% during the I/II/III and IV/V/VI Braak's stages, respectively ( $P < 0.01$ ), but remained unchanged in the CA1 area during all Braak's stages (Figure 3).

### Discussion and Conclusion

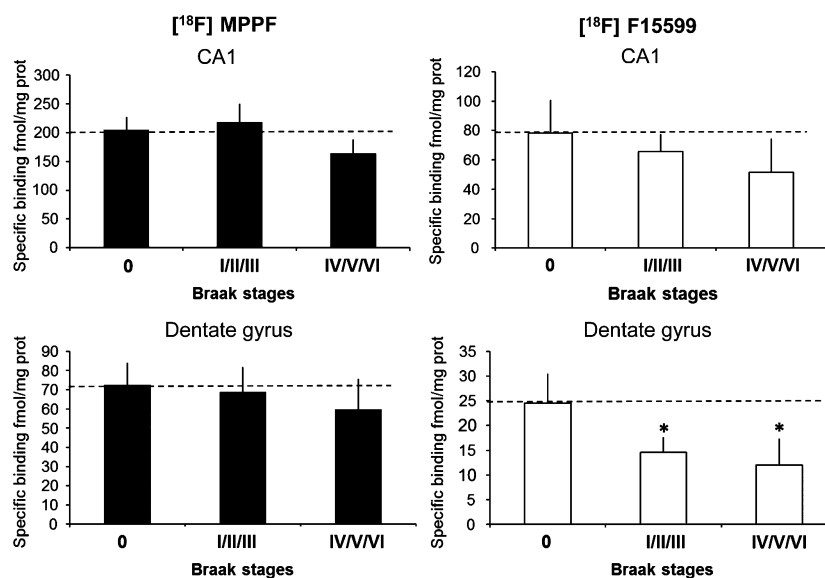
This study compared for the first time in patients with AD the binding of two PET radioligands directed toward the same serotonin receptor, but differing in their agonist or antagonist



**Figure 2** The regional distribution of  $^{18}$ F-MPPF and  $^{18}$ F-F15599 binding sites in hippocampi of Alzheimer's disease patients at different Braak's stages. Black and white arrows show the prominent signal in CA1 and dentate gyrus, respectively. Pseudocolor scale obtained from extemporaneous standards is shown from low (–) to high radioactive level (+).

pharmacological properties. We hypothesized that an antagonist radioligand ( $^{18}$ F-MPPF), which labels the entire population of 5-HT<sub>1A</sub> receptors, would display a different binding pattern to that of an agonist radioligand ( $^{18}$ F-F15599) which was anticipated to label only those 5-HT<sub>1A</sub> receptors which are G-coupled and linked to the second messengers (i.e., in their “active” or “functional” state).

Although the present study utilized brain tissue samples from a modest number of patients, the validity of its results is supported by its robust methodology: Braak's staging adapted by the BrainNet Europe consortium was chosen because of the better correlation of hyperphosphorylated tau deposits than  $\beta$ -amyloid with the cognitive status [17,18];  $^{18}$ F-F15599 and  $^{18}$ F-MPPF bindings were compared in serial sections limiting interindividual variability;  $^{18}$ F-F15599 and  $^{18}$ F-MPPF are both highly selective for 5-HT<sub>1A</sub> receptors and display similar nanomolar affinity for this target (approximately 3 nM in each case) [21,22], allowing a direct comparison of their binding (if identical experimental conditions are used) [19]; their incubation concentrations were optimally chosen for autoradiography so that they were three times their K<sub>d</sub> values in human brain [23]; the fact that  $^{18}$ F-F15599 binds preferentially to G-protein-coupled “functional” 5-HT<sub>1A</sub> receptors [19] was confirmed by the receptor/G-protein decoupling action of Gpp(NH)p;



**Figure 3** 5-HT<sub>1A</sub> receptor-binding site densities in the CA1 area and the dentate gyrus of Alzheimer's disease patients at different Braak's stages determined from film autoradiography with <sup>18</sup>F-MPPF and <sup>18</sup>F-F15599. \**P* < 0.01.

and, finally, autoradiography data were obtained in a quantitative manner (calibration curves).

The binding of the antagonist, <sup>18</sup>F-MPPF, which provides a measure of total 5-HT<sub>1A</sub> receptor density, revealed a similar distribution and quantitative labeling at Braak's stage 0 as in previous postmortem brain studies in non-AD subjects [24]. This indicates that, in terms of serotonin neurotransmission, the Braak's stage 0 can be considered as a control group. It has to be mentioned that in terms of patient demography, there was a tendency for an effect of Braak staging on age of the subjects, that is, higher Braak staging for higher age of patients. This nonhomogeneity of patient ages (particularly for the Braak's stage 0 vs. other Braak's stages) is inherent in studies using brain banks (indeed, availability of brain tissue from very old patients without neurodegenerative diseases is rare). Nevertheless, even if the age effect has an influence on 5-HT<sub>1A</sub> receptor density [25,26], it should not be a source of bias in the present study because the two radioligands (<sup>18</sup>F-MPPF and <sup>18</sup>F-F15599) were directly compared on brain tissue from the same patients. Differences in the densities of receptors labeled by the two radioligands can therefore be attributed to their distinct agonist/antagonist properties rather than to the ages of the patients with AD.

Interestingly, the ratio between the number of receptors labeled by the antagonist (total receptor density) and the number of receptors labeled by the agonist (receptors coupled to G-proteins only) provides an indication of the level of 5-HT<sub>1A</sub> receptor G-protein-coupling in different areas of human hippocampus, in accordance with results obtained in a feline model [15]. Whereas total 5-HT<sub>1A</sub> receptor sites measured by the antagonist, <sup>18</sup>F-MPPF, were unchanged at all AD stages, there was a significant decrease in the density of binding sites labeled by the 5-HT<sub>1A</sub> agonist, <sup>18</sup>F-F15599, at the early Braak's stages I/II/III and at the advanced stages IV/V/VI. Our results revealed therefore an early decrease of functional 5-HT<sub>1A</sub> hippocampus receptors, undetectable at the same stages by the prototypical radiolabelled MPPF. Other post-mortem studies have described a decrease of G-protein-mediated GTP hydrolysis with the progression of AD [27,28], but the *in vitro*

approaches of those studies (i.e., [<sup>35</sup>S]GTPγS or [<sup>3</sup>H]8-OH-DPAT autoradiography) were unsuitable for transfer to *in vivo* studies. In contrast, our approach using PET radioligands is directly transferable to imaging studies *in vivo*. The present results therefore support a new concept consisting in comparing the binding profiles of a PET agonist and an antagonist in patients to determine, *in vivo*, the extent of G-protein coupling of 5-HT<sub>1A</sub> receptors and to follow its modifications in functionality during the neurodegenerative process.

The biological underpinnings of an early loss of 5-HT<sub>1A</sub> receptor functionality in AD brain could be linked with the up-regulation of 5-HT<sub>1A</sub> receptors associated with preserved cognitive function during dementias [29]. This suggests that comparison of agonist and antagonist PET measures could shed light on why 5-HT<sub>1A</sub> agonist therapies have been unsuccessful thus far in the treatment of AD [10]. Finally, future *in vitro* studies should not only focus on replication of the present findings in larger patient populations but also on exploring other brain regions, that is, cortical regions, and on carrying out full quantitation of binding parameters (i.e., K<sub>d</sub> and B<sub>max</sub> quantitation using Scatchard analysis), before adaptation of this paradigm to *in vivo* imaging studies in patients with AD.

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## Conflict of Interest

Dr. Newman-Tancredi is an employee and stockholder of Neurolix but has no financial disclosures associated with this project. The other authors report no conflict of interest and have nothing to disclose.

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