

## Prognostic Value of a Nine-Gene Signature in Glioma Patients Based on mRNA Expression Profiling

Zhao-Shi Bao,<sup>1</sup> Ming-Yang Li,<sup>1</sup> Jia-Yin Wang,<sup>2</sup> Chuan-Bao Zhang,<sup>1</sup> Hong-Jun Wang,<sup>3</sup> Wei Yan,<sup>1</sup> Yan-Wei Liu,<sup>1</sup> Wei Zhang,<sup>1</sup> Ling Chen<sup>4</sup> & Tao Jiang<sup>1,5</sup>

<sup>1</sup> Department of Neurosurgery, Beijing Tiantan Hospital, Capital Medical University, Beijing, China

<sup>2</sup> Cell Therapy Center, Xuanwu Hospital, The Capital Medical University, Beijing, China

<sup>3</sup> Department of Neurosurgery, The Second Affiliated Hospital of Harbin Medical University, Harbin, China

<sup>4</sup> Department of Neurosurgery, Xuanwu Hospital, the Capital Medical University, Beijing, China

<sup>5</sup> Beijing Neurosurgical Institute, Beijing, China

### Keywords

Biomarker; Gliomas; mRNA; Prognosis; Risk score.

### Correspondence

J. Tao, M.D., Ph.D., Beijing Neurosurgical Institute, No.6, Tiantan Xili, Dongcheng District, Beijing 100050, China.  
Tel.: +86-1067021832;  
Fax: +86-1067021832;  
E-mail: taojiang1964@163.com  
and

L. Chen, M.D., Department of Neurosurgery, Xuanwu Hospital, the Capital Medical University, No.45, Changchun Street, Xicheng District, Beijing 100053, China.  
Tel.: +86-1067021832;  
Fax: +86-1067021832;

E-mail: chlyz34@163.com

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### SUMMARY

**Introduction:** Gliomas are the most common primary brain tumors in adults and a significant cause of cancer-related mortality. A 9-gene signature was identified as a novel prognostic model reflecting survival situation obviously in gliomas. **Aims:** To identify an mRNA expression signature to improve outcome prediction for patients with different glioma grades. **Results:** We used whole-genome mRNA expression microarray data of 220 glioma samples of all grades from the Chinese Glioma Genome Atlas (CGGA) database (<http://www.cgga.org.cn>) as a discovery set and data from Rembrandt and GSE16011 for validation sets. Data from every single grade were analyzed by the Kaplan–Meier method with a two-sided log-rank test. Univariate Cox regression and linear risk score formula were applied to derive a gene signature with better prognostic performance. We found that patients who had high risk score according to the signature had poor overall survival compared with patients who had low risk score. Highly expressed genes in the high-risk group were analyzed by gene ontology (GO) and gene set variation analysis (GSVA). As a result, the reason for the divisibility of gliomas was likely due to cell life processes and adhesion. **Conclusion:** This 9-gene-signature prediction model provided a more accurate predictor of prognosis that denoted patients with high risk score have poor outcome. Moreover, these risk models based on defined molecular profiles showed the considerable prospect in personalized cancer management.

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The first three authors contributed equally to this work.

### Introduction

Gliomas are the most common primary central nervous system tumor, which remain one of the most challenging diseases in humans with considerable mortality and posttreatment morbidity [1]. Patients with newly diagnosed glioblastoma multiforme (GBM) have a median survival of approximately 1 year, with generally poor responses to all therapies. As such, scientific and clinical advances are required. Although adjuvant radiotherapy and chemotherapy improves survival, death occurs inevitably from either recurrent or progressive disease [2].

Introduction of molecular biomarkers in the management of patients with cancer may improve their clinical outcomes. Many biomarker candidates have been generated by high-throughput technologies such as microarray gene expression profiling [3], which is a powerful and promising method for evaluating the expression of a large number of genes and evaluating changes in genome-wide expression. The gene expression pattern of the primary tumor has been shown to predict the outcome for several malignancies, including lung cancer, head and neck cancer, and breast cancer. Numerous genes have been discovered to be important in the

management of gliomas, with changes in gene expression having a close relationship with patient prognosis. However, it is unclear whether a signature is available to predict clinical outcomes in patients in every grade in Chinese population. In the present study, we utilized mRNA expression profiling of gliomas to identify a signature that could successfully divide patients into two groups with different overall survival.

## Methods

### Datasets

Whole-genome mRNA expression microarray data were deposited from the Chinese Glioma Genome Atlas (CGGA) database [4] as a training set, and the following two datasets were obtained for validation: Repository for Molecular Brain Neoplasia Data (REMBRANDT, <http://caintegrator.nci.nih.gov/rembrandt>) and GSE16011 data (<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE16011>).

### Statistical Analysis

In those 220 glioma samples of all grades from the CGGA database, there were 97 grade II tumors, 34 grade III tumors, and 89 GBMs. Overall survival time (OS) was defined as the interval from the date of diagnosis to death or the last follow-up. The prognostic difference of patients with high or low expression of a certain gene (higher or lower than the median value) was calculated by the Kaplan–Meier method with the two-sided log-rank test by two packages (survival and KMsurv) of R. The number of significant genes ( $P < 0.05$ ) was 1095, 3397, and 1906 in grade II, III, and IV, respectively. We then chose the overlap of the three groups of genes above. As a result, nine genes remained, which were used for signature development.

To investigate the effectiveness of these nine genes as a mRNA-based gene signature for clinical outcome prediction, a mathematical formula for survival prediction was constructed. More specifically, we assigned each patient a risk score according to a linear combination of the expression level of the mRNAs, weighted by the regression coefficients derived from the univariate Cox regression analyses [5]. From our nine-gene signature, the risk score for each patient was calculated as follows:

$$\text{Risk score} = \text{expr}_{\text{gene1}} \times \beta_{\text{gene1}} + \text{expr}_{\text{gene2}} \times \beta_{\text{gene2}} + \dots + \text{expr}_{\text{genen}} \times \beta_{\text{genen}}$$

Patients having higher risk scores are expected to have shorter overall survival. We divided patients in the training dataset into high-risk and low-risk groups using the median mRNA signature risk score as the cutoff point. With regard to the validation sets, we used the same  $\beta$ . Considering genes with multiple probes, we calculated their average expression value and then subsequently excluded samples without prognostic information. The Kaplan–Meier method was used to estimate overall survival, and the differences in survival between high-risk and the low-risk patients were analyzed using the two-sided log-rank test.

### Gene Ontology (GO) Analysis of Associated Genes in Every Grade

Significant analysis of microarray (SAM) was performed in every grade of gliomas identifying the differently expressed genes, and then, GO analysis of the top 3000 genes, highly expressed in the high-risk group, was performed using DAVID [6] for function annotation (Table 1).

### Gene Set Variation Analysis (GSVA) with Gene Lists Expression

For another functional annotation, we also conducted GSVA by GSVA package [7] of R. Gene lists were from the following GO terms: 0000084, 0000236, 0043065, 0005925, 0045773, 0050771, and 0042789.

## Results

### Identification of 9-Gene Signature and its Association with Survival and Expression from the Training Set

In the 220 glioma samples of all grades, we used a two-sided log-rank test to analyze each grade mRNA expression microarray data in the training set and identified nine genes that were the overlap of gene lists in each grade that were significantly associated with OS ( $P < 0.05$ , Table 2). We then applied the nine genes to develop a signature using the risk score method. The risk score was calculated for each of the 220 patients in the training set, and patients in every grade were then successfully divided into a high-risk group and a low-risk group based on the cutoff value (median risk score). We observed that patients in the high-risk group had a shorter median OS than those in the low-risk group (Figure 1A–C). Subsequently, we also found that nine genes were significantly differently expressed from II to IV grade (Figure 2).

The related clinical information such as The Cancer Genome Atlas (TCGA) and CGGA subtype, which were annotated as previously reported [4], was listed as well as the isocitrate dehydrogenase (IDH) mutation status, histology, gender, age, Karnofsky

**Table 1** Ten gene ontology (GO) terms of associated genes in every grade

Name	Count	P-Value	Grade
GO:0060284~regulation of cell development	39	0.010906	II
GO:0010721~negative regulation of cell development	13	0.015282171	II
GO:0022403~cell cycle phase	150	1.21E-32	III
GO:0007049~cell cycle	222	1.15E-30	III
hsa04510~focal adhesion	58	4.78E-07	III
GO:0042981~regulation of apoptosis	146	6.89E-05	III
GO:0006915~apoptosis	99	0.020368	III
GO:0006350~transcription	333	1.10E-08	IV
GO:0007049~cell cycle	269	5.49E-65	IV
GO:0000278~mitotic cell cycle	171	2.47E-61	IV

**Table 2** Nine genes associated significantly with overall survival time (OS)

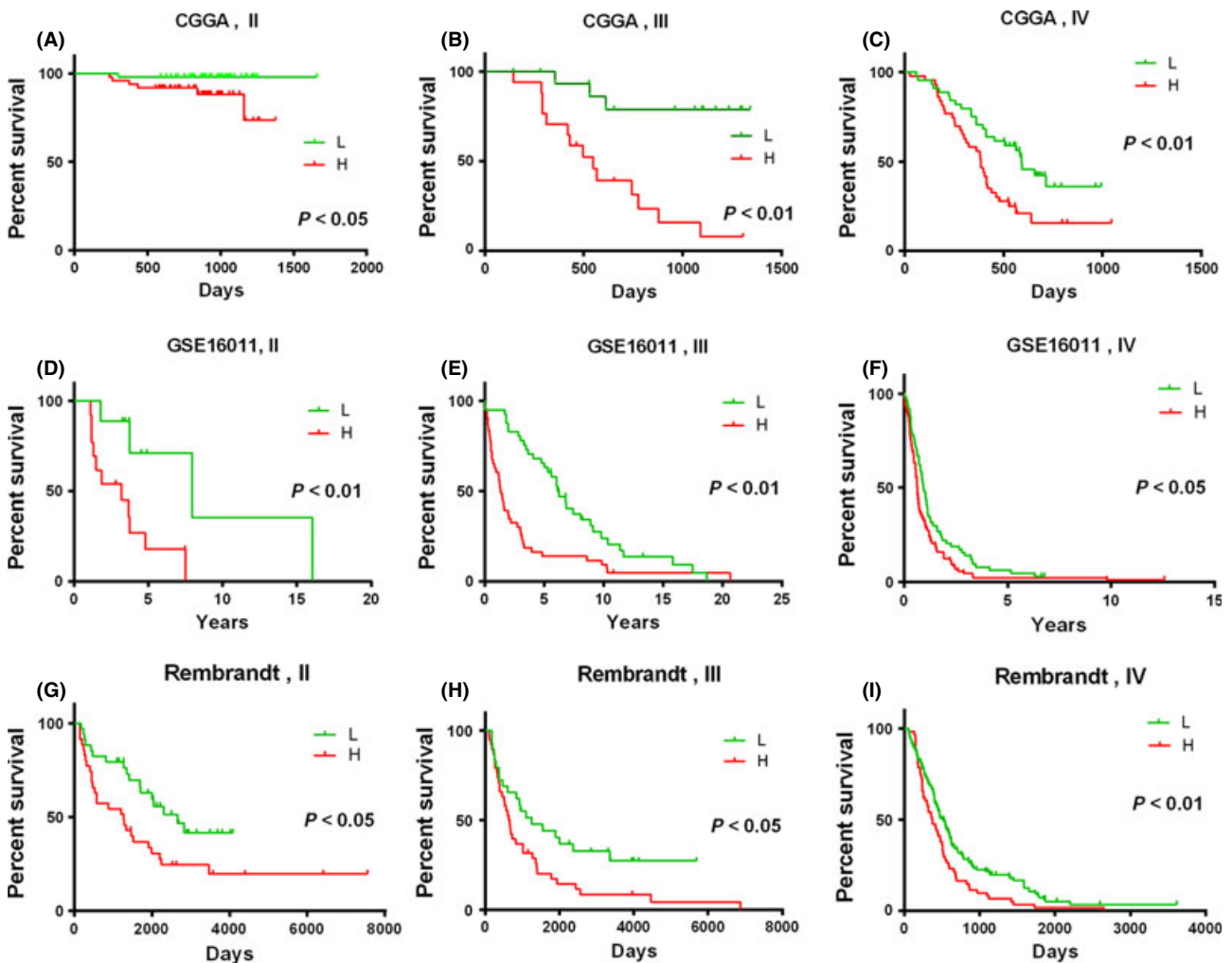
Symbol	Hazard ratio	Parametric <i>P</i> -value
BIRC5	1.653	<0.001
TEAD2	2.382	<0.001
TUBA1B	2.942	<0.001
MT1E	1.365	0.004
RAB1A	0.584	0.001
SFXN4	0.22	<0.001
TPX2	2.588	<0.001
HDAC4	0.548	<0.001
FAM125B	2.447	<0.001

performance status (KPS), which were obtained from CGGA database, some parameters also had a corresponding trend from low risk score to high risk score. Patients with high risk scores

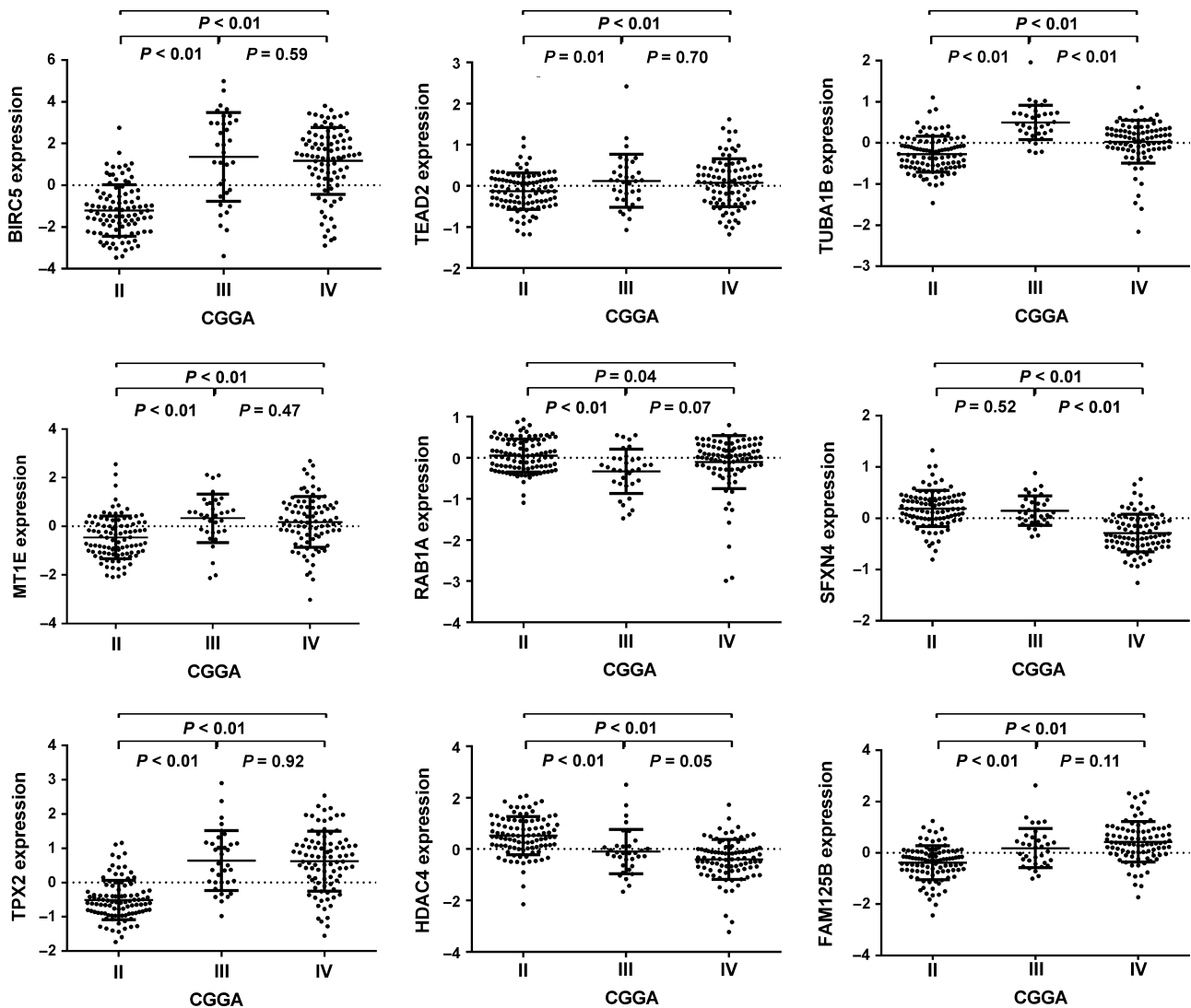
obviously appeared to display G3 and wild-type IDH1 accumulation, and patients with low risk scores tended to show G1, G2, and IDH1 mutation accumulation. Patients with low risk scores had longer OS than those with high risk scores (Figure 3).

### Confirmation of the Prognostic Value of the Gene Signature in Validation Sets

For validation, we downloaded the whole-genome mRNA expression profiling of glioma patients from Rembrandt and GSE16011. With the remaining 319 and 256 glioma patients in the two datasets, we then used the same risk score formula obtained from the training set to get a risk score for each individual in each respective situation. In each grade, patients were divided into the high-risk and low-risk groups in line with the risk score, which was higher or lower than the cutoff. The prognostic value of the signature was validated in both of the datasets (Figure 1D–I).



**Figure 1** These Kaplan–Meier estimates of overall survival in patients with each grade glioma were constructed by the signature. *P*-values are indicated for the high-risk and low-risk groups stratified according to the signature risk score in the Chinese Glioma Genome Atlas (CGGA) data (A, B, and C), the GSE16011 data (D, E, and F), and the Rembrandt data (G, H, and I). H, high-risk group; L, low-risk group.



**Figure 2** The expression difference of 9-gene signature in Chinese Glioma Genome Atlas (CGGA) dataset. A single spot was the gene expression value of an individual patient. Lines in the middle were the mean expression value.

### Functional Annotation of the Signature

Using GO analysis, we found that the associated genes, which were obtained from those highly expressed in the high-risk group, were mainly associated with evolution of cell life and adhesion. We also performed GSEA that showed that patients with a higher risk score tended to have a lower expression of anticell development-, cell apoptosis- and adhesion-, transcription-associated genes, and a higher expression of regulation of cell development-, mitotic cell cycle-associated genes in each grade, respectively (Figure 4). These data may explain the different prognoses of the two groups divided by the signature.

### Gene Function Interpretation

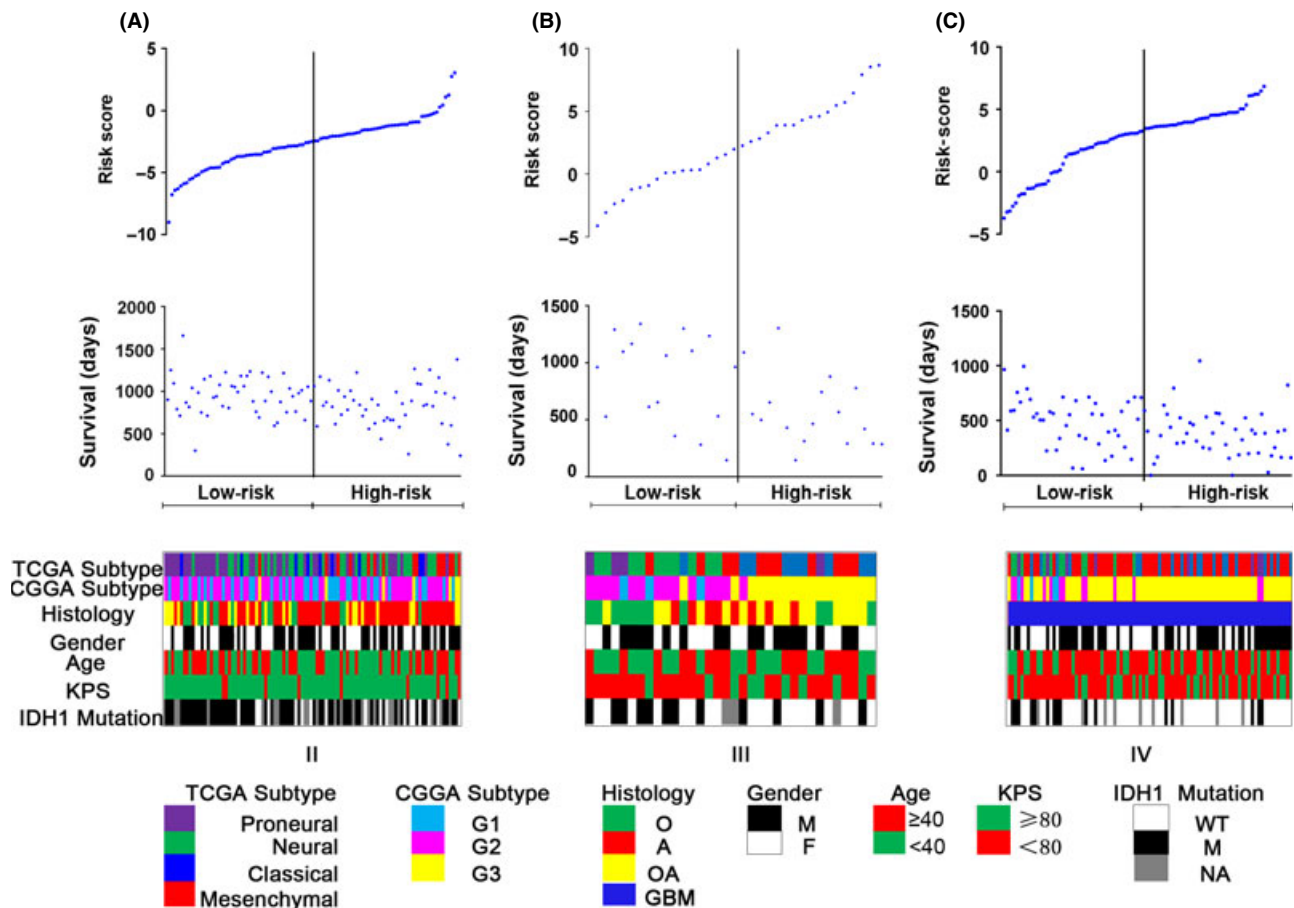
The nine genes used in our signature were RAB1A, BIRC5, TEAD2, TUBA1B, MT1E, SFXN4, TPX2, HDAC4, and FAM125B.

We found that the majority of these genes have similar functions including cell development, apoptosis, proliferation, and migration.

**RAB1A** is a member of the Rab family of small GTPases with a well-characterized function in the regulation of vesicle trafficking from the endoplasmic reticulum to the Golgi apparatus and within Golgi compartments [8].

**BIRC5** is preferentially expressed in human cancer cells and has multiple functions, including the inhibition of cell apoptosis [9], control of the cell cycle [10,11], promotion of tumor angiogenesis [12,13] resistance to chemotherapy or radiotherapy [14], acceleration of metastasis and recurrence [15], and regulation of cancer cell autophagy [16], all of which favor cancer cell survival and tumor maintenance.

**TEAD2** encodes a protein that regulates a wide range of developmental processes, including skeletal and cardiac muscle development, skeletal muscle regeneration, neural crest development,



**Figure 3** Analysis of the signature risk score is illustrated for patients from grade II to IV (A, B, and C), including (Top) signature risk score distribution, (Middle) patient survival duration, and (Bottom) clinical and molecular information.

and notochord development [17–21]. The major roles of Tead2 appear to be the promotion of cell proliferation and suppression of cell death.

**TUBA1B** was more highly expressed in hepatocellular carcinoma (HCC) tumor tissues than in adjacent nontumor tissues, which was a significant predictor for poor overall survival of HCC patients [22].

**MT1E (Metallothionein 1E)** has been found to be highly expressed in motile cell lines and can enhance the migration and invasion of human glioma cells by inducing MMP-9 inactivation via the upregulation of NF- $\kappa$ B p50 [23].

**SFXN4** (comprising SFXN4a and SFXN4b) is widely expressed in almost all tissues examined, which suggests that functional redundancy is likely [24].

**TPX2** is a mitotic regulator involved in the formation of the mitotic spindle and in oncogene-induced mitotic stress. This protein is frequently overexpressed in human cancer, and its deregulation may participate in chromosome numeric aberrations as well as other forms of genomic instability in cancer cells [25].

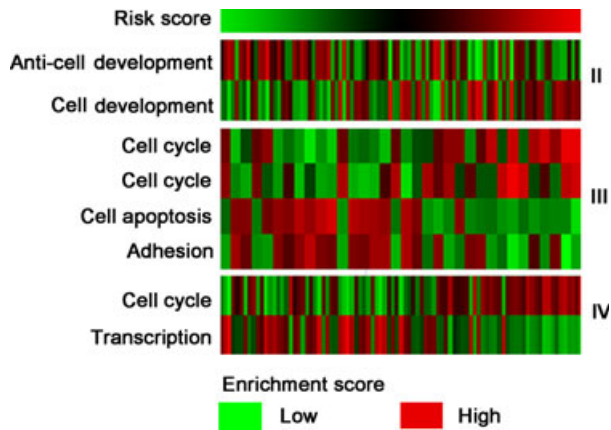
**HDAC4** (histone deacetylase 4) belongs to a class IIa of histone deacetylases, which are important regulators of gene expression, and controls pleiotropic cellular functions [26]. HDAC4 was found to impinge on multiple and apparently contradictory cellular fates, including differentiation, apoptosis, survival, cell growth, and

proliferation. HDAC4 is massively expressed in the proliferative compartment of the colon, and is downregulated during intestinal differentiation [27].

**FAM125B** is a component of the ESCRT-I complex, a heterotetramer, which mediates the sorting of ubiquitinated cargo protein from the plasma membrane to the endosomal vesicle. However, it is unknown whether there is any correlation between its function and human tumors.

## Discussion

As a common fatal central nervous system tumor, gliomas are diagnosed by histopathological criteria. The robust prognostic factors for the majority of these tumors are limited to tumor grade and gene variation. In recent years, the molecular classification of gliomas has developed rapidly. Several groups have reported classification systems based on mRNA expression [28–30], microRNA expression [3], or methylation [31,32], and the majority of the classification systems are focused on mRNA expression. We hypothesized that a set of genes or gene profile would be predictive of survival after surgical resection in patients with glioma. The main objective of the present study was to evaluate the gene expression profiles, identify genes associated with outcome, and build a gene expression signature based on the risk score method



**Figure 4** Gene set variation analysis samples from Chinese Glioma Genome Atlas (CGGA). The risk score (upper) was calculated with the formula described above and ranked from left to right. Gene set enrichment scores (lower) of cell cycle, development, apoptosis, adhere, and transcription were analyzed by gene set variation analysis (GSVA) package of R. Patients with high risk score tended to have a lower expression of anticell development (II), cell apoptosis and adhesion (III), transcription (IV), and higher expression of cell development (II), cell cycle (III), the first cell cycle denotes “phase of cell cycle”, and the second one means “mitotic prometaphase” (IV).

to assess high- and low-risk patients. Hence, we investigated the mRNA expression profile of CGGA data to identify a prognostic signature and found that patients with a high risk score had a poor survival time compared with patients who had a low risk score.

By GO and GSVA analysis, we inferred that the divisibility of glioma patients is likely related to cell life processes and adhesion. However, only MT1E that can enhance the migration and invasion of human gliomas cell had been published in gliomas. So, the other genes need us to further explore.

Accurate staging before treatment is important and facilitates the selection of appropriate treatment strategies. Despite improve-

ments, however, the current clinical staging modalities have not proved very accurate [33–35]. A better understanding of the biological behavior of a tumor will help to determine appropriate therapies, with the potential to improve the outcomes of patients with gliomas.

The analysis of the gene expression profiles associated with different outcomes may be useful for the careful selection of therapies and could also aid in tailoring treatment to the individual patient [36,37].

Additionally, this approach may help to reduce the complexity and dimensionality of genomic data to provide biological insights that may translate into improved management of gliomas.

These observations lay the foundation for future development of a mechanistically based molecular risk estimation model in high-grade gliomas. The signature might lend itself better to translation into clinical practice for two reasons. First, while the expression of an individual gene can vary over short periods of time in cancers, a gene signature is more static and more amenable to predictive screens. Second, the highly expressed genes of the signature in human cancers were associated with promotion of cell proliferation and enhancing cell migration, invasion, and frequency of genetic mutation. Further, in complex glioblastoma tumors, it is considered that multiple, rather than single, genes drive the disease process.

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## Conflict of Interest

The authors declare no conflict of interest.

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