

MicroRNA in intervertebral disc degeneration

Zheng Li*, Xin Yu*, Jianxiong Shen*, Matthew T.V. Chan† and William Ka Kei Wu†'‡

*Department of Orthopaedic Surgery, Peking Union Medical College Hospital, Peking Union Medical College, Beijing 100007, China, †Department of Anaesthesia and Intensive Care, The Chinese University of Hong Kong, Hong Kong 999077, China and ‡State-Key Laboratory of Digestive Disease and Institute of Digestive Disease, LKS Institute of Health Science, The Chinese University of Hong Kong, Hong Kong 999077, China

Received 23 September 2014; revision accepted 22 November 2014

Abstract

Aetiology of intervertebral disc degeneration (IDD) is complex, with genetic, developmental, biochemical and biomechanical factors contributing to the disease process. It is becoming obvious that epigenetic processes influence evolution of IDD as strongly as the genetic background. Deregulated phenotypes of nucleus pulposus cells, including differentiation, migration, proliferation and apoptosis, are involved in all stages of progression of human IDD. Non-coding RNAs, including microRNAs, have recently been recognized as important regulators of gene expression. Research into roles of microRNAs in IDD has been very active over the past 5 years. Our review summarizes current research enlightenment towards understanding roles of microRNAs in regulating nucleus pulposus cell functions in IDD. These exciting findings support the notion that specific modulation of microRNAs may represent an attractive approach for management of IDD.

Introduction

Intervertebral disc degeneration (IDD) is one of the major causes of low back pain, which compose a global burden with severe health care and socioeconomic costs (1–3). Even though IDD treatment has advanced remarkably over the last decade, many patients still do not achieve sustained remission, constituting a major and unmet clinical need (4). Aetiology of IDD is multifactorial, including genetic predisposition, lifestyles (for

Xin Yu and Zheng Li contributed equally to this work.

example, type of occupation, smoking, alcohol consumption) and aging (1,5–7). Underlying IDD molecular and cellular mechanisms are still largely unknown. Thus, increasing numbers of studies support the observation that nucleus pulposus (NP) cells are important in maintaining integrity of intervertebral discs (IVD) by their roles in producing type II collagen, aggrecan and other components of the extracellularmatrix (ECM) (8-10). There is a growing body of evidence to support that microRNA (miRNA) can regulate many, if not all, aspects of cell activity, from differentiation and proliferation to apoptosis, to mediate their effects on a great range of physiological and pathological processes (11-14). In this review, we summarize the current knowledge on roles of miRNAs in regulation of NP cell functions, including differentiation, proliferation, ECM synthesis, apoptosis, and the possible implications for IDD.

MicroRNA biogenesis and function

Over the last decade it has become better recognized that small ribonucleic acids (RNAs) are important components of gene regulatory networks (15–17). Among these, miRNAs are a class of small molecules and non-coding single-stranded RNAs of 18–22 nucleotides, that act as post-transcriptional gene regulatory elements (18–20). Currently, miRNAs have been found in virtually all species of animals, plants and viruses examined; there are over 2042 mature human miRNA sequences listed in the miRNA registry (21,22). It is estimated that there are approximately 1500 predicted miRNAs in the human genome that have the potential to regulate at least 20–30% of all human genes (23,24).

MiRNAs are transcribed from their respective gene loci as primary miRNAs (pri-miRNA) (25) and their biogenesis consists of a series of maturation steps (26). They can be derived from two sources: (i) Pri-miRNAs can be transcribed from specific miRNA-encoding

Correspondence: Jianxiong Shen, Department of Orthopaedic Surgery, Peking Union Medical College Hospital, Peking Union Medical College, Beijing 100007, China. Tel.: 86-010-69152812; Fax: 86-010-69152812; E-mail: shenjianxiong@medmail.com.cn

regions of the genome (intergenic miRNA) by RNA polymerase II (27,28). These pri-miRNAs are then cleaved in the nucleus by a multiprotein complex, which contains an anchor protein DGCR8 (DiGeorge syndrome critical region gene 8), giving rise to 70-nucleotide pre-miRNAs (29,30). (ii) Pri-miRNAs can be derived from mRNA intronic sequences (mirtrons), whose maturation does not require Drosha/DGCR8 processing. These miR-NAs are debranched and spliced by lariat debranching enzyme (Ldbr) and are co-transcribed with host protein-coding genes, forming pre-miRNA hairpins as above (31,32). Intriguingly, these intronic miRNAs are usually involved in the same biological pathways as host protein-coding genes (33).

MiRNAs inhibit gene expression by selectively binding to complementary 3'untranslated regions (3'UTRs) of target mRNAs through complementary base pairing (34,35). Most identified miRNAs are highly conserved among species (36). Expression of miRNAs has both spatial and temporal specificity, together with tissue and cell type specificity (37). They play crucial roles in diverse pathological conditions, such as in cancer, neurodegeneration and aging (38-40). Recent findings have revealed that miRNAs can be involved in cell proliferation, differentiation and apoptosis, and are thus involved in broader processes, such as animal development, homeostasis and bone metabolism (41-45). Recent progress in biology has shown that miRNAs are dysregulated in different cancer types, including gastric cancer, breast cancer, osteosarcoma, lung cancer and hepatocellular carcinoma (46-50). Thus, miRNAs have considerable potential to become a research focus for prevention and treatment of IDD, specially for targeting IDDrelated cell processes, such as NP cell proliferation and apoptosis.

MiRNAs in intervertebral disc degeneration

MiRNAs and NP cell apoptosis

NP cell death mediated by apoptosis is involved in various deleterious consequences of IDD, such as inflammation, degeneration and ECM degradation (51). NP cell apoptosis may have dual effects on IDD depending on the extent of the cell death (4,52). On the one hand, NP cell apoptosis can offset effects of aberrant NP cell proliferation during IDD (53). On the other hand, apoptosis can be harmful as NP cells produce interstitial collagen fibres, which are critical for maintaining tensile strength of the fibrous cap (54). Thus, apoptosis of NP cells plays an important role in determining their stability.

MiR-155 is one of the well-documented miRNAs involved in regulation of apoptotic pathways and

immunological responses (55). Its aberrant expression has been found to be associated with various diseases, such as lung, gastric, pancreatic and colon cancers (56– 59). Wang *et al.* have shown that 29 miRNAs exhibit significantly differential expression in degenerative NP cells, of which miR-155 was found to be one of the most down-regulated (60). Overexpression of miR-155 can inhibit NP cell apoptosis by repressing FADD and caspase-3 expression. *In situ* hybridization and immunohistochemistry have further revealed that miR-155, when expressed in cytoplasm of human NP cells is inversely correlated with FADD and caspase-3.

MiR-27a is a further well-studied miRNA expressed in diverse tissue types. Its aberrant expression is found to be associated with several diseases, including (once again) colorectal, bladder and gastric cancers (61-63). Moreover, previous studies have shown that miR-27a is involved in cell proliferation, apoptosis and tumourigenesis (62,64). It has been found that it affects apoptotic signalling pathways in initiation and progression of gliomas. Using real-time RT-PCR, Liu et al. (65) discovered that expression of miR-27a is high in degenerative NP cells. Furthermore, its enforced expression inhibits phosphoinositide-3 kinases (PI3K) expression by directly targeting its 3'-UTR, and this inhibition is abolished by mutation of miR-27a binding sites. In short, upregulation of miR-27a initiates apoptosis of NP cells by targeting PI3K.

MiRNAs and NP cell proliferation

Increasing numbers of studies have demonstrated that formation of NP cell clusters and aberrant NP cell proliferation play a crucial role in IDD (8). More evidence has shown that miRNAs play an important role in control of NP cell proliferation by post-transcriptional regulation of a number of genes (66).

Previous investigations revealed that miR-10b is involved in regulation of cell proliferation in various cell types, especially in cancer cells, such as breast, liver, pancreatic and gastric cancers (67-70). It is also deregulated in these cancers and its levels are closely associated with tumour progression and pathological grade (71,72). In addition, our previous results have shown that miR-10b is significantly down-regulated in gastric cancer cell lines and tissues as demonstrated by quantitative real-time PCR. Overexpression of miR-10b in MGC-803 and HGC-27 cells dramatically suppressed cell proliferation, migration and invasion and induced apoptosis (11). Similar to its roles in cancer, miR-10b is up-regulated in degenerative NP tissues and is significantly associated with disc degeneration grade. Moreover, overexpression of miR-10b significantly increased NP cell proliferation. In addition, miR-10b promoted proliferation of NP cells by directly targeting HOXD10. MiR-10b also induces Akt phosphorylation in a RhoC-dependent manner (73).

MiR-21, a well-known miRNA, is most frequently dysregulated in different types of human cancer, such as those of the breast, lung, liver and stomach, and has been shown to be implicated in multiple cell processes, including migration, differentiation, proliferation, apoptosis and invasion (74–77). Liu *et al.* reported that miR-21 was up-regulated in human degenerative NP tissues compared to normal NP (78). Moreover, enforced expression of miR-21 promotes NP cell proliferation. In this regard, overexpression of miR-21 led to increased phosphorylation of Akt by directly targeting PTEN. Furthermore, effects of miR-21 on NP cell proliferation and cyclin D1 induction in human NP cells can be blocked by Ly294002, an AKT inhibitor.

miRNAs and ECM remodelling of NP cells

The disc is a complex structure highly specialized for mechanical functions such as spine connectivity, flexure, rotation and extension (79). Currently available evidence implicates loss of IVD ECM upon IDD, as a major cause of low back pain (80). During degeneration, IVD matrix undergoes structural, mechanical and molecular changes, which result in loss of demarcation between the outer annulus fibrosus and inner NP tissues (81,82). ECM is constantly synthesized and degraded by disc cells, during which rates of the processes are

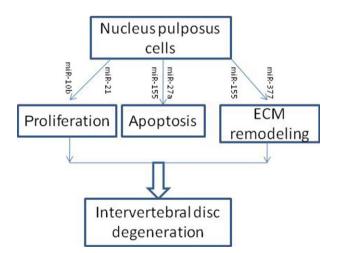


Figure 1. MiRNAs in intervertebral disc degeneration. MiR-10b and miR-21 induce nucleus pulposus (NP) cell proliferation; miR-155 and miR-27a inhibit NP cell apoptosis; miR-155 and miR-377 are involved in NP cell ECM remodelling. ECM, extracellularmatrix.

 Table 1. The expression and function of miRNAs in intervertebral disc degeneration

miRNA	Expression	Function	Target	References
miR-155	Decrease	Apoptosis	FADD, caspase-3	(60)
miR-27a	Increase	Apoptosis	PI3K	(65)
miR-10b	Increase	Proliferation	HOXD10	(73)
miR-21	Increase	Proliferation	PTEN	(78)
miR-155	Decrease	ECM remodelling	Collagen I and III	(85)
miR-377	Increase	ECM remodelling	ADAMTS5	(86)

ECM, extracellularmatrix.

normally in balance (83). However, this balance becomes shifted towards degradation in IDD, with alterations in collagen type and reduction in proteoglycan content, leading to loss of tissue integrity (84). Understanding which factors affect ECM changes is important to fully comprehend the possible implications for IDD.

Chen *et al.* found that ligamentum flavum (LF) thickness and expression of collagens I and III, as well as miR-155, were higher in LF from lumbar spinal stenosis (LSS) patients than from lumbar disc herniation (LDH) patients (85). To this end, expression level of miR-155 positively correlated with LF thickness and levels of types I and III collagen. Overexpression of miR-155 increased mRNA and protein expression of collagens I and III in fibroblasts isolated from LF, while down-regulated expression of miR-155 had opposite effects.

Protein kinase C (PKC) signalling, a major regulator of chondrogenic differentiation, has also been implicated in pathological ECM remodelling. Tsirimonaki *et al.* demonstrated that PKCe activation induced up-regulation of miR-377, which was coupled to reductions in ADAMTS5 and cleaved aggreean (86).

Conclusion

In this review, we summarized roles of miRNAs in function of NP cells and their contributions to IDD (Fig. 1 and Table 1); miRNAs critically affect them. As the aetiology of IDD is multifactorial, these data should improve our insights into pathogenesis of IDD and inform relationships between genetic predisposition and risk factor exposure. Taken together, miRNAs definitively represent an emerging area of research that will provide new insight into IDD pathogenesis with the hope of bringing about novel drug candidates and biomarkers.

Acknowledgements

Funding: This work was supported by the National Natural Science Foundation of China (NSFC) (Grant Numbers: 81401847, 81272053 and 81330044).

References

- Masuda K, Oegema TR Jr, An HS (2004) Growth factors and treatment of intervertebral disc degeneration. *Spine (Phila Pa 1976)* 29, 2757–2769.
- 2 Millecamps M, Tajerian M, Naso L, Sage EH, Stone LS (2012) Lumbar intervertebral disc degeneration associated with axial and radiating low back pain in ageing SPARC-null mice. *Pain* **153**, 1167–1179.
- 3 Raj PP (2008) Intervertebral disc: anatomy-physiology-pathophysiology-treatment. *Pain Pract.* 8, 18–44.
- 4 Loreto C, Musumeci G, Castorina A, Martinez G (2011) Degenerative disc disease of herniated intervertebral discs is associated with extracellular matrix remodeling, vimentin-positive cells and cell death. *Ann. Anat.* **193**, 156–162.
- 5 Adams MA, Roughley PJ (2006) What is intervertebral disc degeneration, and what causes it? *Spine (Phila Pa 1976)* **31**, 2151–2161.
- 6 Furukawa T, Ito K, Nuka S, Hashimoto J, Takei H, Takahara M et al. (2009) Absence of biglycan accelerates the degenerative process in mouse intervertebral disc. *Spine (Phila Pa 1976)* **34**, 911– 917.
- 7 Mayer JE, Iatridis JC, Chan D, Qureshi SA, Gottesman O, Hecht AC (2013) Genetic polymorphisms associated with intervertebral disc degeneration. *Spine J.* **13**, 299–317.
- 8 Li Z, Shen J, Wu WK, Yu X, Liang J, Qiu G et al. (2012) Leptin induces cyclin D1 expression and proliferation of human nucleus pulposus cells via JAK/STAT, PI3K/Akt and MEK/ERK pathways. *PLoS ONE* 7, e53176.
- 9 Li Z, Liang J, Wu WK, Yu X, Yu J, Weng X et al. (2014) Leptin activates RhoA/ROCK pathway to induce cytoskeleton remodeling in nucleus pulposus cells. Int. J. Mol. Sci. 15, 1176–1188.
- 10 Li Z, Shen J, Wu WK, Yu X, Liang J, Qiu G et al. (2013) The role of leptin on the organization and expression of cytoskeleton elements in nucleus pulposus cells. J. Orthop. Res. 31, 847–857.
- 11 Li Z, Lei H, Luo M, Wang Y, Dong L, Ma Y *et al.* (2015) DNA methylation downregulated mir-10b acts as a tumor suppressor in gastric cancer. *Gastric Cancer* **18**, 43–54.
- 12 Brenner B, Hoshen MB, Purim O, David MB, Ashkenazi K, Marshak G et al. (2011) MicroRNAs as a potential prognostic factor in gastric cancer. World J. Gastroenterol. 17, 3976–3985.
- 13 Zheng F, Liao YJ, Cai MY, Liu YH, Liu TH, Chen SP *et al.* (2012) The putative tumour suppressor microRNA-124 modulates hepatocellular carcinoma cell aggressiveness by repressing ROCK2 and EZH2. *Gut* **61**, 278–289.
- 14 Zhao H, Guo M, Zhao G, Ma Q, Ma B, Qiu X et al. (2012) miR-183 inhibits the metastasis of osteosarcoma via downregulation of the expression of Ezrin in F5M2 cells. *Int. J. Mol. Med.* **30**, 1013– 1020.
- 15 Malumbres M (2012) miRNAs and cancer: an epigenetics view. Mol. Aspects Med. 34, 863–74.
- 16 Liang W, Gao B, Fu P, Xu S, Qian Y, Fu Q. (2013) The miRNAs in the pathgenesis of osteosarcoma. *Front. Biosci. (Landmark Ed.)* 18, 788–794.
- 17 Farazi TA, Spitzer JI, Morozov P, Tuschl T (2011) miRNAs in human cancer. J. Pathol. 223, 102–115.

- 18 Fabbri M, Calin GA (2010) Epigenetics and miRNAs in human cancer. Adv. Genet. 70, 87–99.
- 19 Kozlowska E, Krzyzosiak WJ, Koscianska E (2013) Regulation of huntingtin gene expression by miRNA-137, -214, -148a, and their respective isomiRs. *Int. J. Mol. Sci.* 14, 16999–17016.
- 20 Ishikawa K, Ishikawa A, Shoji Y, Imai T (2014) A genotoxic stress-responsive miRNA, miR-574-3p, delays cell growth by suppressing the enhancer of rudimentary homolog gene in vitro. *Int. J. Mol. Sci.* **15**, 2971–2990.
- 21 Fayyad-Kazan H, Bitar N, Najar M, Lewalle P, Fayyad-Kazan M, Badran R *et al.* (2013) Circulating miR-150 and miR-342 in plasma are novel potential biomarkers for acute myeloid leukemia. *J. Transl. Med.* **11**, 31.
- 22 Bianchi N, Zuccato C, Finotti A, Lampronti I, Borgatti M, Gambari R. (2012) Involvement of miRNA in erythroid differentiation. *Epigenomics* 4, 51–65.
- 23 Dassow H, Aigner A (2013) MicroRNAs (miRNAs) in colorectal cancer: from aberrant expression towards therapy. *Curr. Pharm. Des.* 19, 1242–1252.
- 24 Marcinkowska M, Szymanski M, Krzyzosiak WJ, Kozlowski P (2011) Copy number variation of microRNA genes in the human genome. *BMC Genom.* 12, 183.
- 25 Papagiannakopoulos T, Kosik KS (2008) MicroRNAs: regulators of oncogenesis and stemness. BMC Med. 6, 15.
- 26 Sato F, Tsuchiya S, Meltzer SJ, Shimizu K (2011) MicroRNAs and epigenetics. FEBS J. 278, 1598–1609.
- 27 Nelson KM, Weiss GJ (2008) MicroRNAs and cancer: past, present, and potential future. *Mol. Cancer Ther.* 7, 3655–3660.
- 28 Han L, Witmer PD, Casey E, Valle D, Sukumar S (2007) DNA methylation regulates MicroRNA expression. *Cancer Biol. Ther.* 6, 1284–1288.
- 29 Chen Z, Wu J, Yang C, Fan P, Balazs L, Jiao Y *et al.* (2012) Di-George syndrome critical region 8 (DGCR8) protein-mediated microRNA biogenesis is essential for vascular smooth muscle cell development in mice. *J. Biol. Chem.* 287, 19018–19028.
- 30 Fan P, Chen Z, Tian P, Liu W, Jiao Y, Xue Y et al. (2013) miR-NA biogenesis enzyme Drosha is required for vascular smooth muscle cell survival. PLoS ONE 8, e60888.
- 31 Fernandez-Hernando C, Ramirez CM, Goedeke L, Suarez Y (2013) MicroRNAs in metabolic disease. *Arterioscler. Thromb. Vasc. Biol.* 33, 178–185.
- 32 Grant JS, White K, Maclean MR, Baker AH (2013) MicroRNAs in pulmonary arterial remodeling. *Cell. Mol. Life Sci.* 70, 4479–94.
- 33 Rottiers V, Naar AM (2012) MicroRNAs in metabolism and metabolic disorders. *Nat. Rev. Mol. Cell Biol.* 13, 239–250.
- 34 Guancial EA, Bellmunt J, Yeh S, Rosenberg JE, Berman DM (2014) The evolving understanding of microRNA in bladder cancer. Urol. Oncol. 32(41), e31–e40.
- 35 Yoshino H, Seki N, Itesako T, Chiyomaru T, Nakagawa M, Enokida H. (2013) Aberrant expression of microRNAs in bladder cancer. *Nat. Rev. Urol.* 10, 396–404.
- 36 Kobayashi E, Hornicek FJ, Duan Z (2012) MicroRNA Involvement in Osteosarcoma. Sarcoma 2012, 359739.
- 37 Gillen AE, Gosalia N, Leir SH, Harris A (2011) MicroRNA regulation of expression of the cystic fibrosis transmembrane conductance regulator gene. *Biochem. J.* 438, 25–32.
- 38 Bou Kheir T, Futoma-Kazmierczak E, Jacobsen A, Krogh A, Bardram L, Hother C *et al.* (2011) miR-449 inhibits cell proliferation and is down-regulated in gastric cancer. *Mol. Cancer.* 10, 29.
- 39 Montag J, Hitt R, Opitz L, Schulz-Schaeffer WJ, Hunsmann G, Motzkus D. (2009) Upregulation of miRNA hsa-miR-342-3p in experimental and idiopathic prion disease. *Mol. Neurodegener.* 4, 36.

- 40 Wilfred BR, Wang WX, Nelson PT (2007) Energizing miRNA research: a review of the role of miRNAs in lipid metabolism, with a prediction that miR-103/107 regulates human metabolic pathways. *Mol. Genet. Metab.* **91**, 209–217.
- 41 Cao H, Hu X, Zhang Q, Wang J, Li J, Liu B *et al.* (2014) Upregulation of let-7a inhibits vascular smooth muscle cell proliferation in vitro and in vein graft intimal hyperplasia in rats. *J. Surg. Res.* **192**, 223–33.
- 42 Majid S, Dar AA, Saini S, Shahryari V, Arora S, Zaman MS *et al.* (2012) MicroRNA-1280 inhibits invasion and metastasis by targeting ROCK1 in bladder cancer. *PLoS ONE* 7, e46743.
- 43 Shi Z, Wei Q, Zhang M, She J (2014) MicroRNAs in bladder cancer: expression profiles, biological functions, regulation, and clinical implications. *Crit. Rev. Eukaryot. Gene Expr.* 24, 55–75.
- 44 Bae Y, Yang T, Zeng HC, Campeau PM, Chen Y, Bertin T et al. (2012) miRNA-34c regulates Notch signaling during bone development. *Hum. Mol. Genet.* 21, 2991–3000.
- 45 Xie W, Li Z, Li M, Xu N, Zhang Y (2013) miR-181a and inflammation: miRNA homeostasis response to inflammatory stimuli in vivo. *Biochem. Biophys. Res. Commun.* **430**, 647–652.
- 46 Bandres E, Bitarte N, Arias F, Agorreta J, Fortes P, Agirre X *et al.* (2009) microRNA-451 regulates macrophage migration inhibitory factor production and proliferation of gastrointestinal cancer cells. *Clin. Cancer Res.* **15**, 2281–2290.
- 47 Pichiorri F, Palmieri D, De Luca L, Consiglio J, You J, Rocci A et al. (2013) In vivo NCL targeting affects breast cancer aggressiveness through miRNA regulation. J. Exp. Med. 210, 951–968.
- 48 Duan Z, Choy E, Harmon D, Liu X, Susa M, Mankin H et al. (2011) MicroRNA-199a-3p is downregulated in human osteosarcoma and regulates cell proliferation and migration. *Mol. Cancer Ther.* **10**, 1337–1345.
- 49 Furuta M, Kozaki KI, Tanaka S, Arii S, Imoto I, Inazawa J. (2010) miR-124 and miR-203 are epigenetically silenced tumor-suppressive microRNAs in hepatocellular carcinoma. *Carcinogenesis* 31, 766–776.
- 50 Kumar MS, Armenteros-Monterroso E, East P, Chakravorty P, Matthews N, Winslow MM *et al.* (2014) HMGA2 functions as a competing endogenous RNA to promote lung cancer progression. *Nature* 505, 212–217.
- 51 Wei A, Brisby H, Chung SA, Diwan AD (2008) Bone morphogenetic protein-7 protects human intervertebral disc cells in vitro from apoptosis. *Spine J.* 8, 466–474.
- 52 Jiang L, Zhang X, Zheng X, Ru A, Ni X, Wu Y *et al.* (2013) Apoptosis, senescence, and autophagy in rat nucleus pulposus cells: implications for diabetic intervertebral disc degeneration. *J. Orthop. Res.* **31**, 692–702.
- 53 Ha KY, Kim BG, Kim KW, Oh IS, Seo JY (2011) Apoptosis in the sequestrated nucleus pulposus compared to the remaining nucleus pulposus in the same patient. *Spine (Phila Pa 1976)* 36: 683–689.
- 54 Murata Y, Nannmark U, Rydevik B, Takahashi K, Olmarker K (2008) The role of tumor necrosis factor-alpha in apoptosis of dorsal root ganglion cells induced by herniated nucleus pulposus in rats. *Spine (Phila Pa 1976)* **33**: 155–162.
- 55 Wei Y, Nazari-Jahantigh M, Neth P, Weber C, Schober A (2013) MicroRNA-126, -145, and -155: a therapeutic triad in atherosclerosis? *Arterioscler. Thromb. Vasc. Biol.* **33**, 449–454.
- 56 Wang Y, Li J, Tong L, Zhang J, Zhai A, Xu K *et al.* (2013) The prognostic value of miR-21 and miR-155 in non-small-cell lung cancer: a meta-analysis. *Jpn. J. Clin. Oncol.* 43, 813–820.
- 57 Li CL, Nie H, Wang M, Su LP, Li JF, Yu YY et al. (2012) microRNA-155 is downregulated in gastric cancer cells and involved in cell metastasis. Oncol. Rep. 27, 1960–1966.

- 58 Greither T, Grochola LF, Udelnow A, Lautenschlager C, Wurl P, Taubert H. (2010) Elevated expression of microRNAs 155, 203, 210 and 222 in pancreatic tumors is associated with poorer survival. *Int. J. Cancer* **126**, 73–80.
- 59 Pu J, Bai D, Yang X, Lu X, Xu L, Lu J. (2012) Adrenaline promotes cell proliferation and increases chemoresistance in colon cancer HT29 cells through induction of miR-155. *Biochem. Biophys. Res. Commun.* **428**, 210–215.
- 60 Wang HQ, Yu XD, Liu ZH, Cheng X, Samartzis D, Jia LT *et al.* (2011) Deregulated miR-155 promotes Fas-mediated apoptosis in human intervertebral disc degeneration by targeting FADD and caspase-3. J. Pathol. 225, 232–242.
- 61 Zhao X, Yang L, Hu J (2011) Down-regulation of miR-27a might inhibit proliferation and drug resistance of gastric cancer cells. J. Exp. Clin. Cancer Res. 30, 55.
- 62 Drayton RM, Dudziec E, Peter S, Bertz S, Hartmann A, Bryant HE *et al.* (2014) Reduced expression of miRNA-27a modulates cisplatin resistance in bladder cancer by targeting the cystine/glutamate exchanger SLC7A11. *Clin. Cancer Res.* **20**, 1990–2000.
- 63 Hezova R, Kovarikova A, Bienertova-Vasku J, Sachlova M, Redova M, Vasku A *et al.* (2012) Evaluation of SNPs in miR-196-a2, miR-27a and miR-146a as risk factors of colorectal cancer. *World J. Gastroenterol.* 18, 2827–2831.
- 64 Liu T, Tang H, Lang Y, Liu M, Li X (2009) MicroRNA-27a functions as an oncogene in gastric adenocarcinoma by targeting prohibitin. *Cancer Lett.* 273, 233–242.
- 65 Liu G, Cao P, Chen H, Yuan W, Wang J, Tang X. (2013) MiR-27a regulates apoptosis in nucleus pulposus cells by targeting PI3K. *PLoS ONE* **8**, e75251.
- 66 Wang W, Zhao LJ, Tan YX, Ren H, Qi ZT (2012) MiR-138 induces cell cycle arrest by targeting cyclin D3 in hepatocellular carcinoma. *Carcinogenesis* **33**, 1113–1120.
- 67 Ma L (2010) Role of miR-10b in breast cancer metastasis. *Breast Cancer Res.* **12**, 210.
- 68 Wang YY, Ye ZY, Zhao ZS, Li L, Wang YX, Tao HQ et al. (2013) Clinicopathologic significance of miR-10b expression in gastric carcinoma. *Hum. Pathol.* 44, 1278–85.
- 69 Li QJ, Zhou L, Yang F, Wang GX, Zheng H, Wang DS et al. (2012) MicroRNA-10b promotes migration and invasion through CADM1 in human hepatocellular carcinoma cells. *Tumour Biol.* 33, 1455– 1465.
- 70 Frampton AE, Krell J, Zhang Y, Stebbing J, Castellano L, Jiao LR. (2012) The role of miR-10b in metastatic pancreatic ductal adenocarcinoma. *Surgery* 152, 936–8.
- 71 Ma L, Teruya-Feldstein J, Weinberg RA (2007) Tumour invasion and metastasis initiated by microRNA-10b in breast cancer. *Nature* 449, 682–688.
- 72 Nakata K, Ohuchida K, Mizumoto K, Kayashima T, Ikenaga N, Sakai H. (2011) MicroRNA-10b is overexpressed in pancreatic cancer, promotes its invasiveness, and correlates with a poor prognosis. *Surgery* **150**, 916–922.
- 73 Yu X, Li Z, Shen J, Wu WK, Liang J, Weng X et al. (2013) MicroRNA-10b promotes nucleus pulposus cell proliferation through RhoC-Akt pathway by targeting HOXD10 in intervetebral disc degeneration. PLoS ONE 8, e83080.
- 74 Cao Z, Yoon JH, Nam SW, Lee JY, Park WS (2012) PDCD4 expression inversely correlated with miR-21 levels in gastric cancers. J. Cancer Res. Clin. Oncol. 138, 611–619.
- 75 Iyevleva AG, Kuligina E, Mitiushkina NV, Togo AV, Miki Y, Imyanitov EN. (2012) High level of miR-21, miR-10b, and miR-31 expression in bilateral vs. unilateral breast carcinomas. *Breast Cancer Res. Treat.* **131**, 1049–1059.

- 76 Haigl B, Vanas V, Setinek U, Hegedus B, Gsur A, Sutterlüty-Fall H. (2014) Expression of microRNA-21 in non-small cell lung cancer tissue increases with disease progression and is likely caused by growth conditional changes during malignant transformation. *Int. J. Oncol.* 44, 1325–1334.
- 77 Huang YH, Lin YH, Chi HC, Liao CH, Liao CJ, Wu SM et al. (2013) Thyroid hormone regulation of miR-21 enhances migration and invasion of hepatoma. *Cancer Res.* **73**, 2505–2517.
- 78 Liu H, Huang X, Liu X, Xiao S, Zhang Y, Xiang T et al. (2014) miR-21 promotes human nucleus pulposus cell proliferation through PTEN/AKT signaling. Int. J. Mol. Sci. 15, 4007–4018.
- 79 Tian Y, Yuan W, Fujita N, Wang J, Wang H, Shapiro IM *et al.* (2013) Inflammatory cytokines associated with degenerative disc disease control aggrecanase-1 (ADAMTS-4) expression in nucleus pulposus cells through MAPK and NF-kappaB. *Am. J. Pathol.* **182**, 2310–2321.
- 80 Roughley PJ (2004) Biology of intervertebral disc aging and degeneration: involvement of the extracellular matrix. *Spine (Phila Pa 1976)* **29**: 2691–2699.
- 81 Huang M, Wang HQ, Zhang Q, Yan XD, Hao M, Luo ZJ. (2012) Alterations of ADAMTSs and TIMP-3 in human nucleus pulposus

cells subjected to compressive load: implications in the pathogenesis of human intervertebral disc degeneration. *J. Orthop. Res.* **30**, 267–273.

- 82 Hayes AJ, Benjamin M, Ralphs JR (2001) Extracellular matrix in development of the intervertebral disc. *Matrix Biol.* 20, 107–121.
- 83 Clouet J, Pot-Vaucel M, Grimandi G, Masson M, Lesoeur J, Fellah BH *et al.* (2011) Characterization of the age-dependent intervertebral disc changes in rabbit by correlation between MRI, histology and gene expression. *BMC Musculoskelet. Disord.* **12**, 147.
- 84 Chen WH, Lo WC, Lee JJ, Su CH, Lin CT, Liu HY et al. (2006) Tissue-engineered intervertebral disc and chondrogenesis using human nucleus pulposus regulated through TGF-beta1 in plateletrich plasma. J. Cell. Physiol. 209, 744–754.
- 85 Chen J, Liu Z, Zhong G, Qian L, Li Z, Qiao Z *et al.* (2014) Hypertrophy of ligamentum flavum in lumbar spine stenosis is associated with increased miR-155 level. *Dis. Markers* **2014**, 786543.
- 86 Tsirimonaki E, Fedonidis C, Pneumaticos SG, Tragas AA, Michalopoulos I, Mangoura D. (2013) PKCepsilon signalling activates ERK1/2, and regulates aggrecan, ADAMTS5, and miR377 gene expression in human nucleus pulposus cells. *PLoS ONE* 8, e82045.