

Update in Cystic Fibrosis 2018

Bonnie W. Ramsey^{1,2,3*}, Gregory P. Downey^{4,5,6,7,8}, and Christopher H. Goss^{3,9,10}

¹Department of Pediatrics, University of Washington School of Medicine, Seattle, Washington; ²Center for Clinical and Translational Research and ¹⁰Cystic Fibrosis Foundation Therapeutics Development Network Coordinating Center, Seattle Children's Research Institute, Seattle, Washington; ³Division of Pediatric Pulmonology, Department of Pediatrics, and ⁹Division of Pulmonary and Critical Care Medicine, Department of Medicine, University of Washington, Seattle, Washington; ⁴Division of Pulmonary, Critical Care, and Sleep Medicine, Department of Medicine, ⁵Department of Pediatrics, and ⁶Department of Biomedical Research, National Jewish Health, Denver, Colorado; and ⁷Division of Pulmonary Sciences and Critical Care Medicine, Department of Medicine, and ⁸Department of Microbiology and Immunology, University of Colorado, Aurora, Colorado

In this review, we have selected several outstanding studies from the *American Journal of Respiratory and Critical Care Medicine* (AJRCCM), the *American Journal of Respiratory Cell and Molecular Biology* (AJRCMB), and the *Annals of the American Thoracic Society* (AnnalsATS) in 2018 to provide readers with an overview of the important contributions to the rapidly evolving literature in the field of cystic fibrosis (CF) published in these three journals. The featured studies focus on advances in five major themes central to the understanding of the pathogenesis and treatment of CF: 1) therapies directed at the underlying defect, 2) early intervention strategies to delay disease progression, 3) respiratory tract infections, 4) inflammation, and 5) epidemiology and outcome measures. Because of word limitations, we regret that not every original contribution could be included.

Advances in Cystic Fibrosis Transmembrane Conductance Regulator Modulator Therapies

Over the past decade, management of CF has been transformed with the approval of

several small molecules, termed CFTR (cystic fibrosis transmembrane conductance regulator) modulators, used alone or in combination. These modulators consist of two classes of drugs: potentiators that augment opening of the CFTR channel function at the apical membrane of epithelial cells and correctors that assist in transport of nascent protein to the cell surface. These oral therapies have been able to restore CFTR function across a range of CFTR mutations. Ivacaftor (potentiator), used in treating patients with at least one copy of *G551D* and other ivacaftor-responsive mutations, was approved by the Food and Drug Administration in January 2012 with extended approval to children aged 12 months and older in 2018. Lumacaftor (corrector)/ivacaftor combination for treatment of patients homozygous for the *F508del* mutation received Food and Drug Administration approval in July 2015 and since August 2018 is approved for use in patients aged 2 years and older. Tezacaftor (corrector)/ivacaftor combination was approved in February 2018 for the homozygous *F508del* populations aged 12 years and older, as well as for some *F508del* heterozygotes with a residual function

mutation on the other allele. Together, these three drug combinations provide therapies for approximately 50% of individuals with CF. All these therapies were approved on the basis of phase 2 and 3 trial efficacy data demonstrating significant improvement in multiple clinical outcomes: percent predicted FEV₁ (ppFEV₁), improved body mass index (BMI), and prolonged time to pulmonary exacerbations (PEs) requiring antibiotics. The clinical changes noted for ivacaftor in the *G551D* population (1) were more robust than seen in the trials of lumacaftor/ivacaftor (2) and tezacaftor/ivacaftor (3, 4). Furthermore, lumacaftor/ivacaftor and tezacaftor/ivacaftor did not demonstrate clinical efficacy in individuals with one *F508del* allele and a second allele with minimal CFTR function. For this reason, a second generation of correctors was added to the tezacaftor/ivacaftor combination to create a triple combination. Two triple-combination therapies (tezacaftor/ivacaftor/445 and tezacaftor/ivacaftor/659) have completed phase 2 trials with good safety profiles and more robust clinical efficacy, both demonstrating at least a 10% increase in ppFEV₁ in both the homozygous *F508del* population and

(Received in original form February 8, 2019; accepted in final form March 25, 2019)

*B.W.R. is Associate Editor of *AJRCCM*. Her participation complies with American Thoracic Society requirements for recusal from review and decisions for authored works.

Supported by the Cystic Fibrosis Foundation and the NIH (P30DK089507, UL1TR002319, U01TR002487, and U01HL114623-05) (B.W.R.); the Cystic Fibrosis Foundation, the NIH (R01HL103965, R01HL113382, R01AI101307, UM1 HL119073, and P30DK089507), and the Food and Drug Administration (R01FD003704) (C.H.G.); and the NIH (HL132950, UG3TR002445, and U01HL131755) and the Department of Defense (W81XWH-16-2-0018 and W81XWH-16-2-0029) (G.P.D.).

Author Contributions: All authors were involved in the drafting of the manuscript for important intellectual content.

Correspondence and requests for reprints should be addressed to Bonnie W. Ramsey, M.D., Center for Clinical and Translational Research, Seattle Children's Research Institute, P.O. Box 5371, M/S CW8-5B, Seattle, WA 98145-5005. E-mail: bonnie.ramsey@seattlechildrens.org.

Am J Respir Crit Care Med Vol 199, Iss 10, pp 1188–1194, May 15, 2019

Copyright © 2019 by the American Thoracic Society

Originally Published in Press as DOI: 10.1164/rccm.201902-0310UP on March 27, 2019

Internet address: www.atsjournals.org

patients with one copy of *F580del* and a second minimal function mutation (5, 6). These two triple combinations are currently in phase 3 trials. If approved, therapies may soon become available to over 90% of the CF population.

The recent availability of these approved modulators for a large portion of individuals with CF has led to clinical questions regarding the optimal patients for initiation of therapy. For this reason, the first clinical practice guideline for the use of CFTR modulators was published in *AnnalsATS* in 2018 (7). These guidelines, endorsed by the American Thoracic Society in November 2017, were based on a systematic review of relevant publications and evaluated using the GRADE (Grading of Recommendations, Assessment, Development and Evaluation) approach. They are not intended to be a standard of care but provide genotype-specific recommendations based on published evidence. For example, the guidelines strongly recommend treatment with ivacaftor/lumacaftor for individuals with two copies of *F580del* older than 12 years of age and an FEV₁ less than 90% predicted. With the addition of new CF therapies, the guidelines will continue to evolve.

These new therapies also provide an unprecedented opportunity to study the changes in pathophysiology and natural history of CF. These drugs are invaluable tools for developing and optimizing biomarkers and clinical outcomes for future therapeutic development. Several studies published in *AJRCCM* in 2018 have furthered understanding of the short-term and long-term consequences of CFTR modulation. A key multicenter observational study of the impact of ivacaftor in individuals responsive to the therapy (GOAL [G551D Observational] study; NCT01521338) was initially published in *AJRCCM* in 2014 (8). The investigators in that study reported clinically significant changes in a range of outcomes, including ppFEV₁, mucociliary clearance, and intestinal pH, after initiation of ivacaftor. The GOAL study linked to Cystic Fibrosis Foundation Patient Registry (CFFPR) data also demonstrated that in the year after initiation of ivacaftor, the odds of *Pseudomonas aeruginosa* (*Pa*) positivity were reduced by 35% compared with the year before drug initiation (odds ratio, 0.65; $P < 0.001$) (9). In 2018, this group (10) used data from the GOAL-linked CFFPR to evaluate

longer-term changes in ppFEV₁, BMI, and rate of PEs requiring hospitalization by comparing patient data ($n = 144$) for the 2 years before ivacaftor initiation with the 2 years after initiation. The key findings were that treated patients demonstrated a 2-year benefit with a reduced decline in FEV₁ and PE rate; an additional encouraging finding was that the 25% of patients who did not demonstrate a short-term improvement in FEV₁ at 1 month had long-term improvements in these parameters. These data are supported by another small study nested within GOAL in which researchers noted improvement in lung clearance index (LCI) with treatment of ivacaftor in children 3–5 years of age (11).

A second study reported the impact of ivacaftor on a key nonpulmonary manifestation: cystic fibrosis–related diabetes (CFRD) and abnormal insulin secretion (12). This is the largest clinical study published on this topic. In it, researchers hypothesized that ivacaftor would improve glucose metabolism as well as insulin secretion. Patients initiating ivacaftor therapy ($n = 12$; 11 children) underwent oral glucose tolerance tests, mixed meal tolerance tests, and arginine-potentiated glucose tests at pretherapy baseline and at 16 weeks after therapy. Seven of the patients had normal and five had abnormal oral glucose tolerance test results at baseline; none had CFRD. The key finding was that multiple measures of insulin secretion improved after therapy, including first-phase responsiveness, even in these young patients with mild glucose abnormalities. This finding provides hope that restoration of CFTR function may impact glucose metabolism and eventually CFRD (13).

To measure the change in CFTR function *in vivo*, Graeber and colleagues (14) evaluated the impact of lumacaftor/ivacaftor on several biomarkers in patients homozygous for *F508del* mutation aged 12 years and older. Fifty-three patients were evaluated before and after 8–16 weeks of therapy. Clinical outcomes of FEV₁ and BMI were compared with biomarkers, sweat chloride, nasal potential difference, and intestinal current measurement. Lumacaftor/ivacaftor resulted in partial rescue of CFTR function of about 10.2% (interquartile range [IQR], 0.0% to 26.1%) in nasal potential difference and 17.7% (IQR, 10.8% to 29.0%) in intestinal current measurement and a 17.8 mmol/L (IQR, –25.9 to –6.1 mmol/L) reduction in sweat chloride. There was no correlation, however,

between the changes in these biomarkers and clinical outcomes. In fact, all patients had some response in these biomarkers, even if no change in FEV₁ was detected. Further studies are needed to validate the role of these biomarkers in development of future modulators (15).

A final study addressed the importance of secondary factors that may impact CFTR expression in the airway and efficacy of CFTR modulators. The authors described the role of the TGF- β (transforming growth factor- β) and microRNA (miR, miRNA)-145 regulatory pathway in reducing CFTR expression in the airways (16). Both TGF- β and miRs are elevated in CF airway secretions, potentially reducing CFTR expression. The authors demonstrated in primary airway epithelial cells that TGF- β , through miRNA-145, inhibits the corrective benefit of lumacaftor. It is possible that the TGF- β and miR pathway contributes to a variable clinical response *in vivo*. More important, antagonism of miRNA-145 reverses this TGF- β inhibition of CFTR function and may be a useful adjunct therapy to enhance CFTR modulation and other therapeutic approaches to restore CFTR function (17).

Clinical Trials Evaluating Early Interventions to Delay Progression of Lung Disease

Over the past two decades, as more therapeutic approaches directed at improving airway clearance and treating airway infection and inflammation have become available, there has been increased interest in starting these therapies as early as possible to prevent the progression of lung disease (18). Three randomized controlled trials in the past year expanded knowledge regarding approved therapies in young patients with CF.

The first study, with a novel antiinflammatory approach in which the investigators used a multivitamin with additional antioxidants, hypothesized that increased systemic antioxidant concentrations would result in an antiinflammatory effect (NCT01859390). Although elevated antioxidant concentrations were achieved, only modest reductions in systemic inflammation after 4 weeks were observed (circulating calprotectin mean difference, –0.13 log₁₀[μ g/ml]; $P = 0.03$) (19). Because

current CFTR modulators such as ivacaftor may not have a robust effect on sputum markers of inflammation, including neutrophil elastase activity (mean change [SD], $-0.1 [0.37] \log_{10}[\mu\text{g/ml}]$) (8), additional antiinflammatory approaches may be needed.

A second study assessed the impact on LCI of preventive hypertonic saline in infants with CF who were less than 4 months of age (NCT01619657) (20). The authors found that early introduction of 6% hypertonic saline significantly improved LCI and weight. This study was small ($N = 42$), and additional endpoints, such as chest magnetic resonance imaging scores and exacerbation rates, did not change. However, the study demonstrated that LCI might be a key indicator of efficacy in infants with CF. As we think of advancing novel therapies to the youngest children with CF, we must have robust endpoints such as LCI in this vulnerable population.

The third randomized controlled study (21) is a multicenter, double-blind, randomized, placebo-controlled, 18-month trial (OPTIMIZE [Optimizing Treatment for Early *Pseudomonas Aeruginosa* Infection in Cystic Fibrosis]; NCT02054156) testing the hypothesis that the addition of thrice-weekly azithromycin (AZ) to tobramycin inhalation solution in children (aged 6 mo to 18 yr) with CF and early *Pa* infection decreases the risk of PEs and prolongs the time to *Pa* recurrence. The study was stopped early by the data safety monitoring board because the prespecified interim boundary for efficacy was reached ($N = 221$; 110 AZ and 111 placebo). The risk for PE requiring antibiotics was reduced by 44% in the AZ group compared with placebo (hazard ratio, 0.56; 95% confidence interval, 0.37–0.83; $P = 0.004$), and there was significant weight gain (1.27 kg) in the AZ group. However, this reduction in PE risk did not track with an effect on *Pa* recurrence or other microbiological endpoints. All three studies demonstrated the importance and challenges of studying the youngest populations and mildest disease states.

Advances in Understanding and Treatment of CF Airway Infections

Chronic infections of airways and sinuses in individuals with CF remain the major cause

of morbidity and mortality in CF (22). Although there is increased interest in the contributions and interactions of the respiratory microbiome early in disease (23), with progression, specific bacterial pathogens predominate, including *Pa*, *Staphylococcus aureus* (*Sa*), and nontuberculous mycobacteria (NTM). In the past year, several articles illuminated the epidemiology of both *Pa* and NTM. In addition, several studies provided new insights into the risk and benefit of three established antimicrobial approaches—antistaphylococcal prophylaxis, chronic AZ administration, and the use of facial masks—for inpatient infection control.

Pa remains the most common bacterial pathogen infecting the CF airway after the first decade (24). Variability in clinical course with chronic *Pa* colonization has been attributed to both the host response and adaptations of the pathogen within the CF airway environment. A key contributor to the pathogenesis of chronic *Pa* pulmonary infections is the pathogen's ability to form structured communities that coat mucosal surfaces (i.e., biofilms), enhancing persistence. An *AJRCCM* review of biofilms (25) provided insights into the clinical impact of and potential therapeutic approaches for *Pa* lung infections. Two articles focused on how genetic evolution of *Pa* isolates may be associated with changes in clinical outcomes. A longitudinal whole-genome deep-sequencing study (26) genotyped *Pa* isolated from 32 patients from first isolate until either death of the patient or eradication of the pathogen. The molecular evolutionary trajectory of *Pa* isolates from patients with mild disease, defined as stable health after 25–35 years, versus severe disease, defined as death in less than 15 years, differed. A higher incidence of loss-of-function mutations and mutations associated with antibiotic resistance was noted in the patients with a severe clinical course compared with those with the milder phenotype, reflecting the dynamic interrelationship of host and pathogen. As noted in the accompanying editorial (27), it is not known whether the *Pa* evolutionary trajectories are a cause or an effect of illness severity. A second study (28) used multilocus sequence typing to define clones of over 1,500 *Pa* isolates from 402 individuals at six large Canadian centers. Clones were defined as six of seven shared alleles, and these clones were then correlated with FEV₁, BMI, PE, mortality,

or transplant. There was a high degree of genetic diversity with very limited sharing of dominant clones, even within centers, and no significant difference in clinical outcomes across the clones. However, within patients, it was found that changes in sequence typing over time were associated with a significant decline in both FEV₁ and BMI. Although both studies suggest that genetic evolution of *Pa* may impact clinical course, it is not known whether these divergent paths are a cause or an effect of illness severity.

A study based on U.S. patients in the CFFPR from 2010 to 2014 (29) reported that sputum positivity for NTM is becoming increasingly prevalent. Of the 16,153 patients in the CFFPR, 3,211 (20%) had at least one positive sputum culture in the 5-year period studied; approximately one-third were infected with *Mycobacterium abscessus* and two-thirds with *Mycobacterium avium* complex. It is noteworthy that during these five years, the annual period prevalence increased from 11% in 2010 to 13.4% in 2014. In addition, a unique subpopulation of patients with CF was identified: Patients over 60 years old diagnosed with CF later in life had a 33% prevalence of *Mycobacterium avium* complex, suggesting that the prevalence may continue to increase as the CF population ages. This report emphasizes the importance of routine screening for these pathogens, including speciation and ongoing research to develop new therapeutic approaches.

The role of antistaphylococcal prophylaxis has been controversial for decades and was the topic of a Cochrane review published in 2017 (30). In the United Kingdom, flucloxacillin is widely used to prevent *Sa* colonization, whereas *Sa* prophylaxis is not recommended in the U.S. Cystic Fibrosis Foundation care guidelines (31) because of possible emergence of *Pa*. Investigators from both the United Kingdom and the United States (32) undertook a longitudinal observational study of children aged 0–4 years using both the UK CF Registry (1,074 children) and the CFFPR in the United States (3,677 children) from 2000 to 2009, testing the hypothesis that *Sa* prophylaxis is associated with decreased *Sa* but no increased risk of *Pa* acquisition. The risk of *Sa* first detection (hazard ratio, 5.79; 95% confidence interval, 4.85–6.90) and *Pa* first detection (hazard

ratio, 1.92; 95% confidence interval, 1.65, 2.24; $P < 0.001$) is greater in the United States than in the United Kingdom. It is unknown whether these differences can be attributed to *Sa* prophylaxis or to environmental differences in the two countries. A second analysis in the same study (32) evaluated U.K. children who were receiving flucloxacillin ($n = 278$) and not prophylaxis ($n = 306$). In this population, prophylaxis did not reduce the detection of *Sa*, but it increased detection of *Pa*. Although the findings are interesting, attributing causality will require a randomized clinical trial, which is currently being conducted in the United Kingdom.

One study examined the risk/benefit of chronic AZ therapy in children with CF. A retrospective cohort study used the CFFPR (33) to ascertain whether long-term AZ administration is associated with an increased risk of emergence of other multidrug-resistant pathogens. Thrice-weekly AZ users were propensity score matched with nonusers to reduce indication and selection bias. Using Kaplan-Meier curves and Cox proportional hazards regression analyses to compare the incidence of new pathogens in users and nonusers, the incidence of methicillin-resistant *Sa*, NTM, and *Burkholderia cepacia* complex was significantly lower in the AZ users. Importantly, there was no increased incidence of other pathogens among AZ users. It is reassuring that in this predominantly pediatric population (mean age, 12 yr), there does not appear to be increased risk of treatment-emergent respiratory pathogens, including NTM infection, contrasting with earlier reports that AZ could potentially increase NTM acquisition (34).

Reducing patient-to-patient spread of CF pathogens is a high priority for clinical management (35), and infection control guidelines strongly recommend wearing face masks in healthcare settings. An Australian study (36) reported the actual efficacy of surgical masks in reducing spread of *Pa* aerosol droplets in 25 adult colonized patients at a distance of 2 m. Although normal talking is rarely associated with viable *Pa* droplets, an uncovered cough has a 75% incidence of *Pa* aerosolization. Covering the mouth with a hand will reduce the incidence to 50%, whereas a mask significantly reduces aerosolization to 8%, demonstrating the potential value of this simple and

inexpensive method to interrupt aerosol spread between patients.

Inflammation and Airway Clearance in CF

Dysfunction of CFTR, a cAMP-regulated anion (Cl^- and HCO_3^-) channel, results in abnormalities in the airway pH, mucus viscoelastic properties, airway surface liquid (ASL) volume, and mucociliary clearance. This abnormal channel predisposes patients to chronic respiratory infections with neutrophil-predominant inflammation involving the lower respiratory tract. A recent study demonstrated that in response to mucopurulent materials, bronchial epithelial cells from patients with CF have normal upregulation of mucin secretion but an absent fluid-secretory response, resulting in more dehydrated mucus and thus further exacerbating mucus adhesion in the airway (37).

There is evidence of dysregulation of both innate and adaptive immunity in the CF lung (38), and pulmonary inflammation is observed very early in life, even in the absence of detectable infection, indicative of dysregulation of the inflammatory response (39). A study published in *AJRCCM* reported that bacterial infection was not necessary for development of inflammatory lung disease in a preclinical model with mucus stasis likely leading to hypersecretion of mucus and bronchiectasis in ferrets genetically deficient in *CFTR*. These ferrets had been treated with lifelong broad-spectrum antibiotics (40), further underscoring the dysregulation of the inflammatory response in the milieu of the CF lung.

Paradoxically, despite the presence of increased numbers of neutrophils in the CF lung, bacteria survive and often thrive in this milieu, supporting a defect in antimicrobial functions of phagocytes in the CF airway (41–43). There are likely multiple factors that contribute to the phagocyte dysfunction in CF, including intrinsic abnormalities in the microbicidal functions of the neutrophils (44–46) and defective epithelial function related to *CFTR* mutations leading to dehydration and increased viscosity of the airway mucus (47). As noted in a study published in *AJRCCM* (46), increased concentrations of proteinases, such as elastase in the ASL, lead to cleavage and inactivation of membrane receptors on the phagocytes and

degradation of antimicrobial factors in the ASL (48–50). Thus, although infection may initiate the inflammatory response, a dysregulated host response is responsible for much of the destruction of lung tissue.

Patients with CF also exhibit an exaggerated inflammatory response driven in part by cytokines such as IL-8 produced by bronchial epithelial cells and amplified by exposure to bacterial pathogens such as *Pa*. Several recent studies have provided important insights into molecular mechanisms responsible for this exaggerated inflammatory response. RGS2 (regulator of G-protein signaling 2), a GTPase-activating protein expressed in airway epithelial cells, is a negative regulator of G-protein signaling (51) and represses IL-8 expression in airway epithelial cells (52). RGS2 is known to enhance responsiveness and promote increased secretion of mucins from human airway epithelial cells (53) and may be directly relevant to the pathogenesis of CF lung disease. Expression of RGS2 is regulated by epigenetic factors, including DNA methylation (54). A recent study by Bouvet and colleagues published in *AJRCCM* (55) provided important insights into abnormalities in the control of RGS2 expression in CF airway epithelial cells. Using methylated DNA immunoprecipitation arrays and methylation-specific PCR, they observed that RGS2 mRNA and protein expression is downregulated in CF airway epithelial cells by a mechanism involving hypermethylation of cytosine residues in the promoters of 13 genes. Importantly, downregulation of RGS2 resulted in enhanced expression of A100A12, a proinflammatory protein known to drive the inflammatory response in the CF lung. Thus, therapeutic approaches to increase RGS2 expression might attenuate proinflammatory responses of CF airway epithelial cells.

Another pathway that regulates the exaggerated inflammatory response in CF lungs involves *PLCB3* (phospholipase C- β 3). Specific genetic variants in *PLCB3* are associated with slow progression of CF pulmonary disease, and silencing of *PLCB3* in cultured human bronchial epithelial cells results in enhanced inflammatory responses triggered by TLRs (Toll-like receptors) (56). A recent study by Rimessi and colleagues (57) built on this knowledge base and identified a genetic variant, *PLCB3*-S3845L,

as a loss-of-function genetic variant. Human bronchial epithelial cells expressing this variant exhibited attenuated inflammatory responses when exposed to *Pa* or respiratory secretions from the airways of patients with CF. This study underscores the importance of *PLCB3* in control of the inflammatory response in epithelial cells and identifies this pathway as a potential therapeutic target to downregulate the injurious inflammatory response in the CF lung. In an accompanying editorial, McElvaney and McElvaney (58) summarized a feedforward system in the CF lung, where neutrophil proteinases such as elastase enhance IL-8 production by bronchial epithelial cells and leukotriene B₄ production by macrophages, thus recruiting more neutrophils. This system is further amplified by bacterial products. The *PLCB3-S3845L* variant described by Rimessi and colleagues (57) functions to disrupt this feedforward loop and thus attenuates the injurious inflammatory response in the CF lung while leaving antibacterial defenses intact.

A study by Jones-Nelson and colleagues described a novel mechanism whereby activation of TLR5 by *Pa* flagellin results in enhanced neutrophil recruitment into the lung and release of neutrophil elastase leading to epithelial barrier dysfunction and lung injury and increased pathogenicity of *Klebsiella pneumoniae* during coinfection (59). This response could be attenuated by sivelestat, a neutrophil elastase inhibitor, or by antibodies to TLR5. The accompanying editorial by Bratcher and Malcolm (60) discussed the mechanisms underlying polymicrobial infection, focusing on both bacteria-derived toxins such as flagellin and host-derived factors such as proteinases (elastase and other matrix metalloproteinases) in microbial pathogenicity, and suggested therapeutic approaches such as promoting neutrophil phagocytosis to mitigate lung injury.

Advances in CF Epidemiology and Clinical Outcomes

During 2018, significant advances occurred in understanding of both clinical epidemiology and clinical outcome assessment in CF. The advances fall into two general areas: 1) clinical features associated with survival and decline in lung function

and 2) gaining a better understanding of disparities of care in CF.

Several studies in 2018 brought new insights into the use of biomarkers and clinical outcomes. One of the most essential outcomes in a life-shortening disease such as CF is survival. In an interesting study published in *AnnalsATS*, Saavedra and colleagues (61) used clustering techniques with genomic data from whole blood to identify four different clusters of genes that were associated with very different clinical outcomes. At 5 years, all subjects in the very low-risk cluster were alive and well, whereas 90% of subjects in the high-risk cluster had experienced a major event, defined as mechanical ventilation or ICU admission, referral for lung transplant, undergoing lung transplant, or death ($P = 0.0001$). Another international collaboration evaluated the role of cardiopulmonary exercise testing (CPET) in predicting survival. Ten CF centers in Australia, Europe, and North America provided data to support the use of CPET in a retrospective cohort of 433 patients with CF, the largest study of its kind (62). Using both multivariate Cox proportional hazards modeling and cluster analysis, they identified novel predictors of clinical outcomes based on key CPET endpoints (i.e., $\dot{V}O_2$ peak in percent predicted, peak work rate, and ventilatory equivalent for oxygen and carbon dioxide). Given the challenges that survival prediction has encountered in CF (63), adding more specificity to clinical prediction with a functional test such as CPET will likely improve risk stratification (64). Besides improving understanding of the overall CF population, additional work from the Canadian CF Registry has assessed the survival of persons diagnosed with CF as adults (65). Median 10-year transplant-free survival of this patient population (median age of diagnosis, 34.3 yr) was 87.7%.

Although assessing survival is important, one of the key intermediate endpoints remains lung function. In fact, a recent study published in *AnnalsATS* successfully correlated improved lung function with improved survival (66). Predictors of lung function recovery and, importantly, lung function decline can provide insights into both pathophysiology of disease and clinical care. Recent analyses of data from the NHLBI's "Grand Opportunity" Exome Sequencing Project (LungGO) have revealed that specific single-nucleotide variants in genes involving

cilia were found to be associated with both lung disease progression and lung function preservation, adding to the ever-expanding understanding of modifiers of CF lung disease (67). Although genetic factors impact lung function, day-to-day care substantially impacts lung function. One can see the impact of care practices by comparing countries with disparate healthcare systems, such as the United States and Canada. In a longitudinal analysis of lung function and BMI in both the United States and Canada, nutritional status and lung function improved in both Canada and the United States from 1990 to 2013, with the improvements most prominent in the BMI trajectories in the United States, especially in patients born after 1990 (68). This suggests that national efforts to improve nutritional status and lung function in the early 1990s are likely now being seen. Assessments within U.S. healthcare systems can also yield important associations that can inform clinical decision making. An observational study of treatment of exacerbation in the United States suggested that the proportion of the treatment occurring on an inpatient basis was more important than duration of treatment in achieving successful recovery (69).

Other important predictors of patient survival and well-being are disparities based on race, ethnicity, and socioeconomic status (70, 71). A study published in *AJRCCM* reported an analysis of CFFPR data revealing that Hispanic patients with CF have a markedly increased risk of death compared with non-Hispanic patients with CF (hazard ratio, 1.27; 95% confidence interval, 1.05–1.53), even after adjusting for key confounders (69). In two additional studies, one of the key features associated with gaps of care when transferring from pediatric to adult centers was lack of health insurance (72, 73). Gaps in care were much more likely in those U.S. patients with CF who were younger at transfer, lacked health insurance, and had relocated around the time of transition (72). It is key that future studies delve further into how such disparities in outcome can be reduced, be they based on sex, race, ethnicity, or socioeconomic status.

Conclusions

The studies summarized in this review have provided new insights into the molecular pathophysiology of CF and helped improve

understanding of the impact of novel therapies on patients with this genetic disorder. In the next decade, as highly effective CFTR modulators become more widely available to individuals with CF, the

community must continue to closely observe and record the physiological and clinical changes that evolve. There will be many important questions to answer about onset and progression of respiratory

infection and inflammation and progression of functional and structural lung disease. ■

Author disclosures are available with the text of this article at www.atsjournals.org.

References

- Ramsey BW, Davies J, McElvaney NG, Tullis E, Bell SC, Dřevinek P, et al.; VX08-770-102 Study Group. A CFTR potentiator in patients with cystic fibrosis and the G551D mutation. *N Engl J Med* 2011;365:1663–1672.
- Wainwright CE, Elborn JS, Ramsey BW, Marigowda G, Huang X, Cipolli M, et al.; TRAFFIC Study Group; TRANSPORT Study Group. Lumacaftor-ivacaftor in patients with cystic fibrosis homozygous for Phe508del CFTR. *N Engl J Med* 2015;373:220–231.
- Donaldson SH, Pilewski JM, Griese M, Cooke J, Viswanathan L, Tullis E, et al.; VX11-661-101 Study Group. Tezacaftor/ivacaftor in subjects with cystic fibrosis and F508del/F508del-CFTR or F508del/G551D-CFTR. *Am J Respir Crit Care Med* 2018;197:214–224.
- Rowe SM, Daines C, Ringshausen FC, Kerem E, Wilson J, Tullis E, et al. Tezacaftor-ivacaftor in residual-function heterozygotes with cystic fibrosis. *N Engl J Med* 2017;377:2024–2035.
- Keating D, Marigowda G, Burr L, Daines C, Mall MA, McKone EF, et al.; VX16-445-001 Study Group. VX-445-tezacaftor-ivacaftor in patients with cystic fibrosis and one or two Phe508del alleles. *N Engl J Med* 2018;379:1612–1620.
- Davies JC, Moskowitz SM, Brown C, Horsley A, Mall MA, McKone EF, et al.; VX16-659-101 Study Group. VX-659-tezacaftor-ivacaftor in patients with cystic fibrosis and one or two Phe508del alleles. *N Engl J Med* 2018;379:1599–1611.
- Ren CL, Morgan RL, Oermann C, Resnick HE, Brady C, Campbell A, et al. Cystic fibrosis foundation pulmonary guidelines: use of cystic fibrosis transmembrane conductance regulator modulator therapy in patients with cystic fibrosis. *Ann Am Thorac Soc* 2018;15:271–280.
- Rowe SM, Heltshe SL, Gonska T, Donaldson SH, Borowitz D, Gelfond D, et al.; GOAL Investigators of the Cystic Fibrosis Foundation Therapeutics Development Network. Clinical mechanism of the cystic fibrosis transmembrane conductance regulator potentiator ivacaftor in G551D-mediated cystic fibrosis. *Am J Respir Crit Care Med* 2014;190:175–184.
- Heltshe SL, Mayer-Hamblett N, Burns JL, Khan U, Baines A, Ramsey BW, et al.; GOAL (the G551D Observation-AL) Investigators of the Cystic Fibrosis Foundation Therapeutics Development Network. *Pseudomonas aeruginosa* in cystic fibrosis patients with G551D-CFTR treated with ivacaftor. *Clin Infect Dis* 2015;60:703–712.
- Heltshe SL, Rowe SM, Skalland M, Baines A, Jain M; GOAL Investigators of the Cystic Fibrosis Foundation Therapeutics Development Network. Ivacaftor-treated patients with cystic fibrosis derive long-term benefit despite no short-term clinical improvement. *Am J Respir Crit Care Med* 2018;197:1483–1486.
- Ratjen F, Klingel M, Black P, Powers MR, Grasemann H, Solomon M, et al. Changes in lung clearance index in preschool-aged patients with cystic fibrosis treated with ivacaftor (GOAL): a clinical trial. *Am J Respir Crit Care Med* 2018;198:526–528.
- Kelly A, De Leon DD, Sheikh S, Camburn D, Kubrak C, Pelecks AJ, et al. Islet hormone and incretin secretion in cystic fibrosis after four months of ivacaftor therapy. *Am J Respir Crit Care Med* 2019;199:342–351.
- Norris AW. Is cystic fibrosis-related diabetes reversible? New data on CFTR potentiation and insulin secretion. *Am J Respir Crit Care Med* 2019;199:261–263.
- Graeber SY, Dopfer C, Naehrlich L, Gyulumyan L, Scheuermann H, Hirtz S, et al. Effects of lumacaftor-ivacaftor therapy on cystic fibrosis transmembrane conductance regulator function in Phe508del homozygous patients with cystic fibrosis. *Am J Respir Crit Care Med* 2018;197:1433–1442.
- Bell SC, Wood ME. Biomarkers: their role in CFTR modulator therapies from early development to the clinic. *Am J Respir Crit Care Med* 2018;197:1375–1376.
- Lutfal Kabir F, Ambalavanan N, Liu G, Li P, Solomon GM, Lal CV, et al. MicroRNA-145 antagonism reverses TGF- β inhibition of F508del CFTR correction in airway epithelia. *Am J Respir Crit Care Med* 2018;197:632–643.
- Kramer EL, Clancy JP. MicroRNA-145, cystic fibrosis transmembrane conductance regulator, and transforming growth factor- β : an (un)tangled regulatory web. *Am J Respir Crit Care Med* 2018;197:551–552.
- Ranganathan SC, Hall GL, Sly PD, Stick SM, Douglas TA; Australian Respiratory Early Surveillance Team for Cystic Fibrosis (AREST-CF). Early lung disease in infants and preschool children with cystic fibrosis: what have we learned and what should we do about it? *Am J Respir Crit Care Med* 2017;195:1567–1575.
- Sagel SD, Khan U, Jain R, Graff G, Daines CL, Dunitz JM, et al. Effects of an antioxidant-enriched multivitamin in cystic fibrosis: a randomized, controlled, multicenter clinical trial. *Am J Respir Crit Care Med* 2018;198:639–647.
- Stahl M, Wielpütz MO, Ricklefs I, Dopfer C, Barth S, Schlegel A, et al. Preventive Inhalation of Hypertonic Saline in Infants with Cystic Fibrosis (PRESIS): a randomized, double-blind, controlled study. *Am J Respir Crit Care Med* [online ahead of print] 9 Nov 2018; DOI: 10.1164/rccm.201807-1203OC.
- Mayer-Hamblett N, Retsch-Bogart G, Kloster M, Accurso F, Rosenfeld M, Albers G, et al.; OPTIMIZE Study Group. Azithromycin for early *Pseudomonas* infection in cystic fibrosis: the OPTIMIZE randomized trial. *Am J Respir Crit Care Med* 2018;198:1177–1187.
- Mogayzel PJ Jr, Flume PA. Update in cystic fibrosis 2009. *Am J Respir Crit Care Med* 2010;181:539–544.
- Huang YJ, LiPuma JJ. The microbiome in cystic fibrosis. *Clin Chest Med* 2016;37:59–67.
- Cystic Fibrosis Foundation. Cystic Fibrosis Foundation Patient Registry 2016 annual data report. Bethesda, MD: Cystic Fibrosis Foundation; 2017.
- Maurice NM, Bedi B, Sadikot RT. *Pseudomonas aeruginosa* biofilms: host response and clinical implications in lung infections. *Am J Respir Cell Mol Biol* 2018;58:428–439.
- Klockgether J, Cramer N, Fischer S, Wiehlmann L, Tümmler B. Long-term microevolution of *Pseudomonas aeruginosa* differs between mildly and severely affected cystic fibrosis lungs. *Am J Respir Cell Mol Biol* 2018;59:246–256.
- Winstanley C, Brockhurst MA. Can we manipulate the evolutionary biology of pathogens for clinical benefit? *Am J Respir Cell Mol Biol* 2018;59:143–144.
- Middleton MA, Layeghifard M, Klingel M, Stanojevic S, Yau YCW, Zlosnik JEA, et al. Epidemiology of clonal *Pseudomonas aeruginosa* infection in a Canadian cystic fibrosis population. *Ann Am Thorac Soc* 2018;15:827–836.
- Adjemian J, Olivier KN, Prevots DR. Epidemiology of pulmonary nontuberculous mycobacterial sputum positivity in patients with cystic fibrosis in the United States, 2010–2014. *Ann Am Thorac Soc* 2018;15:817–826.
- Langton Hower SC, Smyth AR. Antibiotic strategies for eradicating *Pseudomonas aeruginosa* in people with cystic fibrosis. *Cochrane Database Syst Rev* 2017;4:CD004197.
- Flume PA, O'Sullivan BP, Robinson KA, Goss CH, Mogayzel PJ Jr, Willey-Courand DB, et al.; Cystic Fibrosis Foundation, Pulmonary Therapies Committee. Cystic fibrosis pulmonary guidelines: chronic medications for maintenance of lung health. *Am J Respir Crit Care Med* 2007;176:957–969.
- Hurley MN, Fogarty A, McKeever TM, Goss CH, Rosenfeld M, Smyth AR. Early respiratory bacterial detection and antistaphylococcal antibiotic prophylaxis in young children with cystic fibrosis. *Ann Am Thorac Soc* 2018;15:42–48.

33. Cogen JD, Onchiri F, Emerson J, Gibson RL, Hoffman LR, Nichols DP, *et al.* Chronic azithromycin use in cystic fibrosis and risk of treatment-emergent respiratory pathogens. *Ann Am Thorac Soc* 2018;15:702–709.
34. Renna M, Schaffner C, Brown K, Shang S, Tamayo MH, Hegyi K, *et al.* Azithromycin blocks autophagy and may predispose cystic fibrosis patients to mycobacterial infection. *J Clin Invest* 2011;121:3554–3563.
35. Saiman L, Siegel J. Infection control in cystic fibrosis. *Clin Microbiol Rev* 2004;17:57–71.
36. Stockwell RE, Wood ME, He C, Sherrard LJ, Ballard EL, Kidd TJ, *et al.*; CF Cough Aerosol Group. Face masks reduce the release of *Pseudomonas aeruginosa* cough aerosols when worn for clinically relevant periods. *Am J Respir Crit Care Med* 2018;198:1339–1342.
37. Abdullah LH, Coakley R, Webster MJ, Zhu Y, Tarran R, Radicioni G, *et al.* Mucin production and hydration responses to mucopurulent materials in normal versus cystic fibrosis airway epithelia. *Am J Respir Crit Care Med* 2018;197:481–491.
38. Bruscia EM, Bonfield TL. Innate and adaptive immunity in cystic fibrosis. *Clin Chest Med* 2016;37:17–29.
39. Khan TZ, Wagener JS, Bost T, Martinez J, Accurso FJ, Riches DW. Early pulmonary inflammation in infants with cystic fibrosis. *Am J Respir Crit Care Med* 1995;151:1075–1082.
40. Rosen BH, Evans TIA, Moll SR, Gray JS, Liang B, Sun X, *et al.* Infection is not required for mucoinflammatory lung disease in CFTR-knockout ferrets. *Am J Respir Crit Care Med* 2018;197:1308–1318.
41. Konstan MW, Hilliard KA, Norvell TM, Berger M. Bronchoalveolar lavage findings in cystic fibrosis patients with stable, clinically mild lung disease suggest ongoing infection and inflammation. *Am J Respir Crit Care Med* 1994;150:448–454.
42. Balough K, McCubbin M, Weinberger M, Smits W, Ahrens R, Fick R. The relationship between infection and inflammation in the early stages of lung disease from cystic fibrosis. *Pediatr Pulmonol* 1995;20:63–70.
43. Rosenfeld M, Gibson RL, McNamara S, Emerson J, Burns JL, Castile R, *et al.* Early pulmonary infection, inflammation, and clinical outcomes in infants with cystic fibrosis. *Pediatr Pulmonol* 2001;32:356–366.
44. Witko-Sarsat V, Allen RC, Paulais M, Nguyen AT, Bessou G, Lenoir G, *et al.* Disturbed myeloperoxidase-dependent activity of neutrophils in cystic fibrosis homozygotes and heterozygotes, and its correction by amiloride. *J Immunol* 1996;157:2728–2735.
45. Hayes E, Pohl K, McElvaney NG, Reeves EP. The cystic fibrosis neutrophil: a specialized yet potentially defective cell. *Arch Immunol Ther Exp (Warsz)* 2011;59:97–112.
46. Pohl K, Hayes E, Keenan J, Henry M, Meleady P, Molloy K, *et al.* A neutrophil intrinsic impairment affecting Rab27a and degranulation in cystic fibrosis is corrected by CFTR potentiator therapy. *Blood* 2014;124:999–1009.
47. Stoltz DA, Meyerholz DK, Welsh MJ. Origins of cystic fibrosis lung disease. *N Engl J Med* 2015;372:1574–1575.
48. Alexis NE, Muhlebach MS, Peden DB, Noah TL. Attenuation of host defense function of lung phagocytes in young cystic fibrosis patients. *J Cyst Fibros* 2006;5:17–25.
49. Vandivier RW, Fadok VA, Hoffmann PR, Bratton DL, Penvari C, Brown KK, *et al.* Elastase-mediated phosphatidylserine receptor cleavage impairs apoptotic cell clearance in cystic fibrosis and bronchiectasis. *J Clin Invest* 2002;109:661–670.
50. Margaroli C, Garratt LW, Horati H, Dittrich AS, Rosenow T, Montgomery ST, *et al.*; AREST-CF; IMPEDE-CF. Elastase exocytosis by airway neutrophils is associated with early lung damage in children with cystic fibrosis. *Am J Respir Crit Care Med* 2019;199:873–881.
51. Kehrl JH, Srikumar D, Harrison K, Wilson GL, Shi CS. Additional 5' exons in the RGS3 locus generate multiple mRNA transcripts, one of which accounts for the origin of human PDZ-RGS3. *Genomics* 2002;79:860–868.
52. Holden NS, George T, Rider CF, Chandrasekhar A, Shah S, Kaur M, *et al.* Induction of regulator of G-protein signaling 2 expression by long-acting β_2 -adrenoceptor agonists and glucocorticoids in human airway epithelial cells. *J Pharmacol Exp Ther* 2014;348:12–24.
53. Liu C, Li Q, Zhou X, Kolosov VP, Perelman JM. Regulator of G-protein signaling 2 inhibits acid-induced mucin5AC hypersecretion in human airway epithelial cells. *Respir Physiol Neurobiol* 2013;185:265–271.
54. Cacan E. Epigenetic regulation of RGS2 (regulator of G-protein signaling 2) in chemoresistant ovarian cancer cells. *J Chemother* 2017;29:173–178.
55. Bouvet GF, Voisin G, Cyr Y, Bascunana V, Massé C, Berthiaume Y. DNA methylation regulates RGS2-induced S100A12 expression in airway epithelial cells. *Am J Respir Cell Mol Biol* 2018;59:601–613.
56. Bezzerri V, d'Adamo P, Rimessi A, Lanzara C, Crovella S, Nicolis E, *et al.* Phospholipase C- β 3 is a key modulator of IL-8 expression in cystic fibrosis bronchial epithelial cells. *J Immunol* 2011;186:4946–4958.
57. Rimessi A, Bezzerri V, Salvatori F, Tamanini A, Nigro F, Dececchi MC, *et al.* PLCB3 loss of function reduces *Pseudomonas aeruginosa*-dependent IL-8 release in cystic fibrosis. *Am J Respir Cell Mol Biol* 2018;59:428–436.
58. McElvaney OJ, McElvaney NG. Targeting IL-8 in cystic fibrosis: enough but not too much. *Am J Respir Cell Mol Biol* 2018;59:401–402.
59. Jones-Nelson O, Hilliard JJ, DiGiandomenico A, Warrener P, Alfaro A, Cheng L, *et al.* The neutrophilic response to *Pseudomonas* damages the airway barrier, promoting infection by *Klebsiella pneumoniae*. *Am J Respir Cell Mol Biol* 2018;59:745–756.
60. Bratcher PE, Malcolm KC. Neutrophils and bacterial coinfection: aiding and abetting. *Am J Respir Cell Mol Biol* 2018;59:668–669.
61. Saavedra MT, Quon BS, Faino A, Caceres SM, Poch KR, Sanders LA, *et al.* Whole blood gene expression profiling predicts severe morbidity and mortality in cystic fibrosis: a 5-year follow-up study. *Ann Am Thorac Soc* 2018;15:589–598.
62. Hebestreit H, Hulzebos EH, Schneiderman JE, Karila C, Boas SR, Kriemler S, *et al.* Prognostic Value of CPET in CF Study Group; Cardiopulmonary exercise testing provides additional prognostic information in cystic fibrosis. *Am J Respir Crit Care Med* [online ahead of print] 15 Oct 2018; DOI: 10.1164/rccm.201806-1110OC.
63. Mayer-Hamblett N, Rosenfeld M, Emerson J, Goss CH, Aitken ML. Developing cystic fibrosis lung transplant referral criteria using predictors of 2-year mortality. *Am J Respir Crit Care Med* 2002;166:1550–1555.
64. Ramos KJ. Cardiopulmonary exercise testing: another tool in the prognostication tool kit for cystic fibrosis. *Am J Respir Crit Care Med* [online ahead of print] 13 Nov 2018; DOI: 10.1164/rccm.201810-2053ED.
65. Desai S, Wong H, Sykes J, Stephenson AL, Singer J, Quon BS. Clinical characteristics and predictors of reduced survival for adult-diagnosed cystic fibrosis: analysis of the Canadian CF Registry. *Ann Am Thorac Soc* 2018;15:1177–1185.
66. Konstan MW, VanDevanter DR, Sawicki GS, Pasta DJ, Foreman AJ, Neiman EA, *et al.* Association of high-dose ibuprofen use, lung function decline, and long-term survival in children with cystic fibrosis. *Ann Am Thorac Soc* 2018;15:485–493.
67. Blue E, Louie TL, Chong JX, Hebring SJ, Barnes KC, Rafaels NM, *et al.*; U.S. National Heart, Lung, and Blood Institute “Grand Opportunity” Exome Sequencing Project (LungGO). Variation in cilia protein genes and progression of lung disease in cystic fibrosis. *Ann Am Thorac Soc* 2018;15:440–448.
68. Goss CH, Sykes J, Stanojevic S, Marshall B, Petren K, Ostrenga J, *et al.* Comparison of nutrition and lung function outcomes in patients with cystic fibrosis living in Canada and the United States. *Am J Respir Crit Care Med* 2018;197:768–775.
69. Rho J, Ahn C, Gao A, Sawicki GS, Keller A, Jain R. Disparities in mortality of hispanic patients with cystic fibrosis in the United States: a national and regional cohort study. *Am J Respir Crit Care Med* 2018;198:1055–1063.
70. Schechter MS, Shelton BJ, Margolis PA, Fitzsimmons SC. The association of socioeconomic status with outcomes in cystic fibrosis patients in the United States. *Am J Respir Crit Care Med* 2001;163:1331–1337.
71. Quon BS, Psoter K, Mayer-Hamblett N, Aitken ML, Li Cl, Goss CH. Disparities in access to lung transplantation for patients with cystic fibrosis by socioeconomic status. *Am J Respir Crit Care Med* 2012;186:1008–1013.
72. Sawicki GS, Ostrenga J, Petren K, Fink AK, D'Agostino E, Strassle C, *et al.* Risk factors for gaps in care during transfer from pediatric to adult cystic fibrosis programs in the United States. *Ann Am Thorac Soc* 2018;15:234–240.
73. Schechter MS, VanDevanter DR, Pasta DJ, Short SA, Morgan WJ, Konstan MW; Scientific Advisory Group and the Investigators and Coordinators of the Epidemiologic Study of Cystic Fibrosis. Treatment setting and outcomes of cystic fibrosis pulmonary exacerbations. *Ann Am Thorac Soc* 2018;15:225–233.