





# Complete Genome Sequence of the Aerobic Facultative Methanotroph *Methylocella tundrae* Strain T4

 Martine A. R. Kox,<sup>a</sup> Muhammad Farhan Ul Haque,<sup>b</sup> Theo A. van Alen,<sup>a</sup> Andrew T. Crombie,<sup>c</sup>  Mike S. M. Jetten,<sup>a</sup> Huub J. M. Op den Camp,<sup>a</sup> Svetlana N. Dedysh,<sup>d</sup> Maartje A. H. J. van Kessel,<sup>a</sup> J. Colin Murrell<sup>b</sup>

<sup>a</sup>Department of Microbiology, IWWR, Radboud University, Nijmegen, The Netherlands

<sup>b</sup>School of Environmental Sciences, University of East Anglia, Norwich, United Kingdom

<sup>c</sup>School of Biological Sciences, University of East Anglia, Norwich, United Kingdom

<sup>d</sup>Winogradsky Institute of Microbiology, Research Center of Biotechnology of the Russian Academy of Sciences, Moscow, Russia

**ABSTRACT** *Methylocella tundrae* T4<sup>T</sup> is a facultative aerobic methanotroph which was isolated from an acidic tundra wetland and possesses only a soluble methane monoxygenase. The complete genome, which includes two megaplasmids, was sequenced using a combination of Illumina and Nanopore technologies. One of the megaplasmids carries a propane monoxygenase gene cluster.

Methane-oxidizing bacteria (MOB) play a major role in the global conversion of methane, since they utilize methane as a source of carbon and energy. MOB are widespread in nature, especially in methane-rich areas (1–3). Most MOB are aerobic Gram-negative bacteria belonging to the *Alphaproteobacteria*, *Gammaproteobacteria*, or *Verrucomicrobia*. MOB are mostly obligate one-carbon utilizers, except for *Methylocella* species, *Methylocapsa aurea*, and some *Methylocystis* species, which also utilize multicarbon compounds (4–8). Unlike most methanotrophs, *Methylocella* species rely entirely on soluble methane monoxygenase (sMMO) for methane oxidation and lack particulate methane monoxygenase (pMMO). The draft genome sequences of two *Methylocella* strains *Methylocella silvestris* BL2<sup>T</sup> (9) and *Methylocella silvestris* TVC, have been published (10). We now report the complete genome sequence of *Methylocella tundrae* T4<sup>T</sup>, isolated from an acidic *Sphagnum* tundra peatland in northern Russia (11).

*M. tundrae* T4<sup>T</sup> was cultivated on M2 agar medium (12) with methane (10% [vol/vol] in the headspace) as the sole carbon and energy source. Multiple colonies were harvested, from which genomic DNA was extracted using the ammonia acetate extraction method (13). Sequencing was performed using a dual sequencing strategy. First, the DNA was sequenced using the MinION access program (Oxford Nanopore Technologies, Oxford, UK). The library was prepared using kit number SQK-LSK108 with the fragmentation step using a g-TUBE (2 × 60 s at 5,000 rpm; Covaris, Inc., Woburn, MA, USA) and subsequently sequenced using a FLO-MIN106 R9.4.1 flow cell. Next, DNA was also sequenced using paired-end (2 × 150-bp) Illumina MiSeq sequencing to obtain high-quality sequences. The library was prepared using the Nextera XT kit (Illumina, San Diego, CA, USA) and sequenced using the MiSeq reagent kit v3 (Illumina).

A total of 6,386,172 raw Illumina paired-end reads (mean length, 149 bp) were obtained, subsequently trimmed using default settings, with a minimum read length of 100 bp and trimming of the first 15 bp, and merged in CLC Genomics Workbench v11 (Qiagen Aarhus A/S, Denmark). Nanopore sequencing yielded 49,146 raw reads (mean length, 6,466 bp; N<sub>50</sub>, 8,738 bp), which were base-called using Albacore v2.1.10, assembled using Canu v1.8 (14), and polished first with Racon v1.3.1 (15) and then with Illumina reads using Pilon v1.23 (16), all with default settings. This resulted in three circular scaffolds. Circularity was further investigated using Repseek v6.6 with default

**Citation** Kox MAR, Farhan Ul Haque M, van Alen TA, Crombie AT, Jetten MSM, Op den Camp HJM, Dedysh SN, van Kessel MAHJ, Murrell JC. 2019. Complete genome sequence of the aerobic facultative methanotroph *Methylocella tundrae* strain T4. *Microbiol Resour Announc* 8:e00286-19. <https://doi.org/10.1128/MRA.00286-19>.

**Editor** J. Cameron Thrash, University of Southern California

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Address correspondence to Maartje A. H. J. van Kessel, [maartje.vankessel@science.ru.nl](mailto:maartje.vankessel@science.ru.nl).

M.A.R.K. and M.F.U.H. contributed equally to this article.

**Received** 22 March 2019

**Accepted** 19 April 2019

**Published** 16 May 2019

**TABLE 1** Genomic attributes of the 3 genetic elements that make up the genome of *Methylocella tundrae* T4<sup>T</sup>

Attribute	Data for:		
	Chromosome	Megaplasmid A	Megaplasmid B
Size (bp)	3,909,804	303,933	208,729
DNA G+C content (%)	61.80	61.84	61.46
No. of DNA scaffolds	1	1	1
Circular	Yes	Yes	Yes
Total no. of genes <sup>a</sup>	4,276	390	275
Protein coding density (%)	86.71	85.56	87.08
No. of RNA genes	59	0	0
No. of rRNA genes	2 (5S,16S, and 23S)	0	0
No. of tRNA genes	53	0	0
No. of pseudogenes	61	13	2
No. of genes with function predicted	616	53	15
No. of genes assigned to COGs <sup>b</sup>	2,957	252	96
Coverage (×)	102	85	114
GenBank accession no.	LR536450	LR536451	LR536452

<sup>a</sup> Without artifacts.<sup>b</sup> COGs, Clusters of Orthologous Groups.

settings (17), which showed overlap (>97% identity for >8,800 bp) at the start and end of every scaffold. Additionally, it was confirmed that there was no overlap between the different scaffolds by calculating dot plots with Gepard v1.40 using default settings (18). Genome completeness was checked with CheckM v1.0.12 (completeness, 98.35%; contamination, 0.73%) (19). Finally, the genome was annotated via the MicroScope platform (20). The largest scaffold consisted of the chromosome (Table 1), and the smaller two scaffolds represented two megaplasms (Table 1, megaplasms A and B), each with their own alphaproteobacterial plasmid replication site (*repABC* operon) (21). This is the first time megaplasms have been observed in *Methylocella* species.

The presence of all genes required for the formation and functioning of sMMO (*mmoXYBZDCRG*) and the absence of pMMO genes was confirmed. Gene operons encoding calcium-dependent methanol dehydrogenase (*mxafJGIRSACKLD*) and lanthanide-dependent methanol dehydrogenase (*soxFJG*) are also present. Like *Methylocella silvestris* BL2<sup>T</sup>, all genes required for carrying out the complete oxidation and assimilation (via the serine cycle) of formaldehyde were identified. Genes related to nitrogen metabolism are located on the chromosome, including *nifHDK* (N<sub>2</sub> fixation), *narGHJ* (membrane-bound respiratory nitrate reduction), and *nosRZDFY* (nitrous oxide reductase). Interestingly, the propane monooxygenase gene cluster (*prmABCD*) is located on megaplasmid A. Megaplasmid B contains mainly genes of unknown function. Further genome analyses and detailed comparative genomics studies of the growing number of *Methylocella* species are required to better understand the phylogeny and evolution of facultative methanotrophy in these unique bacteria.

**Data availability.** This whole-genome sequencing project has been deposited in the ENA within project number [PRJEB31709](#). The raw paired-end Illumina reads were deposited with SRA accession number [ERR3223707](#) and the raw MinION reads with SRA number [ERR3224043](#). The assembled genome is available under GenBank accession numbers [LR536450](#) to [LR536452](#). The version described in this paper is the first version.

## ACKNOWLEDGMENTS

We thank Laura Wenzel for help with the MinION Nanopore sequencing. We thank Geert Cremers and Jeroen Frank for help with bioinformatics. The LABGeM (CEA/Genoscope and CNRS UMR8030), France Génomique, and the French Bioinformatics Institute national infrastructures (funded as part of the Investissement d'Avenir program managed by Agence Nationale pour la Recherche, contracts ANR-10-INBS-09 and ANR-11-INBS-0013) are acknowledged for support within the MicroScope annotation platform.

M.A.R.K. was supported by European Research Council Ecomom 339880 to M.S.M.J.,

who was further supported by the Netherlands Organisation for Scientific Research (SIAM Gravitation grant 024 002 002 and a Spinoza award). M.A.H.J.V.K. was supported by NWO Veni grant 016.veni.192.062. H.J.M.O.D.C. was supported by European Research Council advanced grant VOLCANO 669371. M.F.U.H. was supported by a Leverhulme Trust research project (RPG2016-050) to J.C.M., and A.T.C. was supported by a Leverhulme Trust early career fellowship (ECF2016-626).

## REFERENCES

- Farhan Ul Haque M, Crombie AT, Ensminger SA, Baciu C, Murrell JC. 2018. Facultative methanotrophs are abundant at terrestrial natural gas seeps. *Microbiome* 6:118. <https://doi.org/10.1186/s40168-018-0500-x>.
- Ghashghavi M, Jetten MSM, Luke C. 2017. Survey of methanotrophic diversity in various ecosystems by degenerate methane monooxygenase gene primers. *AMB Express* 7:162. <https://doi.org/10.1186/s13568-017-0466-2>.
- Knief C. 2015. Diversity and habitat preferences of cultivated and uncultivated aerobic methanotrophic bacteria evaluated based on *pmoA* as molecular marker. *Front Microbiol* 6:1346. <https://doi.org/10.3389/fmicb.2015.01346>.
- Dedysh SN, Knief C, Dunfield PF. 2005. *Methylocella* species are facultatively methanotrophic. *J Bacteriol* 187:4665–4670. <https://doi.org/10.1128/JB.187.13.4665-4670.2005>.
- Crombie AT, Murrell JC. 2014. Trace-gas metabolic versatility of the facultative methanotroph *Methylocella silvestris*. *Nature* 510:148–151. <https://doi.org/10.1038/nature13192>.
- Dunfield PF, Dedysh SN. 2014. *Methylocella*: a gourmand among methanotrophs. *Trends Microbiol* 22:368–369. <https://doi.org/10.1016/j.tim.2014.05.004>.
- Vorobev A, Jagadevan S, Jain S, Anantharaman K, Dick GJ, Vuilleumier S, Semrau JD. 2014. Genomic and transcriptomic analyses of the facultative methanotroph *Methylocystis* sp. strain SB2 grown on methane or ethanol. *Appl Environ Microbiol* 80:3044–3052. <https://doi.org/10.1128/AEM.00218-14>.
- Dedysh SN, Dunfield PF. 2018. Facultative methane oxidizers, p 1–20. *In* McGenety T (ed), *Taxonomy, genomics and ecophysiology of hydrocarbon-degrading microbes. Handbook of hydrocarbon and lipid microbiology*. Springer, Cham, Switzerland.
- Chen Y, Crombie A, Rahman MT, Dedysh SN, Liesack W, Stott MB, Alam M, Theisen AR, Murrell JC, Dunfield PF. 2010. Complete genome sequence of the aerobic facultative methanotroph *Methylocella silvestris* BL2. *J Bacteriol* 192:3840–3841. <https://doi.org/10.1128/JB.00506-10>.
- Wang J, Geng K, Farhan Ul Haque M, Crombie A, Street L, Wookey P, Ma K, Murrell JC, Pratscher J. 2018. Draft genome sequence of *Methylocella silvestris* TVC, a facultative methanotroph isolated from permafrost. *Genome Announc* 6:e00040-18. <https://doi.org/10.1128/genomeA.00040-18>.
- Dedysh SN, Berestovskaya YY, Vasylieva LV, Belova SE, Khmel'nina VN, Suzina NE, Trotsenko YA, Liesack W, Zavarzin GA. 2004. *Methylocella tundrae* sp. nov., a novel methanotrophic bacterium from acidic tundra peatlands. *Int J Syst Evol Microbiol* 54:151–156. <https://doi.org/10.1099/ijs.0.02805-0>.
- Dedysh SN, Panikov NS, Liesack W, Grosskopf R, Zhou J, Tiedje JM. 1998. Isolation of acidophilic methane-oxidizing bacteria from northern peat wetlands. *Science* 282:281–284. <https://doi.org/10.1126/science.282.5387.281>.
- Kowalchuk GA, De Bruijn F, Head IM, Van der Zijpp AJ, van Elsas JV. 2008. *Molecular microbial ecology manual*, 2nd ed. Springer, Dordrecht, The Netherlands.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive *k*-mer weighting and repeat separation. *Genome Res* 27:722–736. <https://doi.org/10.1101/gr.215087.116>.
- Vaser R, Sović I, Nagarajan N, Sikić M. 2017. Fast and accurate *de novo* genome assembly from long uncorrected reads. *Genome Res* 27:737–746. <https://doi.org/10.1101/gr.214270.116>.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
- Achaz G, Boyer F, Rocha EP, Viari A, Coissac E. 2007. Repseek, a tool to retrieve approximate repeats from large DNA sequences. *Bioinformatics* 23:119–121. <https://doi.org/10.1093/bioinformatics/btl519>.
- Krumsiek J, Arnold R, Rattei T. 2007. Gepard: a rapid and sensitive tool for creating dotplots on genome scale. *Bioinformatics* 23:1026–1028. <https://doi.org/10.1093/bioinformatics/btm039>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
- Medigue C, Calteau A, Cruveiller S, Gachet M, Gautreau G, Josso A, Lajus A, Langlois J, Pereira H, Planel R, Roche D, Rollin J, Rouy Z, Vallenet D. 2017. MicroScope: an integrated resource for community expertise of gene functions and comparative analysis of microbial genomic and metabolic data. *Brief Bioinform*, in press. <https://doi.org/10.1093/bib/bbx113>.
- Pinto UM, Pappas KM, Winans SC. 2012. The ABCs of plasmid replication and segregation. *Nat Rev Microbiol* 10:755–765. <https://doi.org/10.1038/nrmicro2882>.