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Opinion

A Tale of Two Viruses: Does Heterologous Flavivirus Immunity Enhance Zika Disease?

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The rise of Zika virus (ZIKV) and its unusual clinical manifestations provided ground for speculative debate. The clinical severity of secondary dengue virus (DENV) infections is associated with antibody-dependent enhancement (ADE), and it was recently suggested that previous exposure to DENV may worsen ZIKV clinical outcomes. In this Opinion article we analyze the relationship among different flaviviruses and ADE. We discuss new evidence obtained in non-human primates and human cohorts demonstrating that there is no correlation to ADE when ZIKV infection occurs in the presence of pre-existing DENV immunity. We propose a redefinition of ADE in the context of complex immunological flavivirus interactions to provide a more objective perspective when translating *in vitro* or *in vivo* observations into the clinical setting.

Zika Virus Is a Clinical Outlier Flavivirus

Rising from obscurity 60 years after its discovery, ZIKV caused the first major human epidemic in the Federated States of Micronesia, followed by a major pandemic with its introduction in Brazil sometime in 2013 [1]. Currently, indigenous mosquito-borne ZIKV transmission has been confirmed in 49 countries or territories of the Americas Appendix A. The introduction and spread of ZIKV in the Americas was marked by the appearance of severe adverse outcomes such as fetal loss [2], congenital Zika syndrome (CZS) (see Glossary) [3], Guillain-Barré syndrome (GBS) [4], and rare cases of encephalopathy [5], meningoencephalitis [6], myelitis [7], uveitis [8], and severe thrombocytopenia [9]. Several hypotheses have been put forward to explain the unprecedented observed pathogenicity of ZIKV infection in the Americas, including prior heterologous flavivirus infection, virulence of the virus, host genetics and environmental factors among others.

ADE: From *In Vitro* Evidence to Clinical Relevance

Analogous to antibody-dependent enhancement (ADE), during secondary DENV exposure, the scientific community hypothesized that a ZIKV infection following a previous DENV infection may result in increased ZIKV pathogenesis (for example CZS and GBS) in the Americas. ADE *in vitro* can be considered as a common experimental phenomenon with uncertain clinical relevance, as it has been demonstrated for many viruses (alphaviruses [10], rabies [11], coxsackievirus B3 [12], coronavirus [13], human immunodeficiency virus [14,15], and others) without evidence of worsened disease during secondary infection in mice or in human populations [16]. Such a precise ADE definition is very specific in describing an experimental finding as a fact. In *in vitro* assays, immune sera from patients exposed to a variety of different flaviviruses, including yellow fever and Japanese encephalitis viruses, will also enhance DENV infection [17]. Even the homotypic serotype responsible for a past DENV infection can induce ADE of DENV, if the serum is diluted to subneutralizing concentrations [18]. However, in contrast to ADE described for other viruses, ADE of DENV *in vivo* is commonly associated with a worse clinical outcome [19]. Secondary DENV infections result in dramatic

Trends

Zika virus (ZIKV) caused atypical clinical manifestations in areas with previous exposure to other flaviviruses.

Different dengue-ZIKV cross-reacting antibodies neutralize or enhance ZIKV *in vitro*, but the percentage of dengue immune serum neutralizing ZIKV is very low.

Antibody-dependent enhancement (ADE) of ZIKV by dengue and West Nile immune sera has been shown *in vitro* and induced in immunosuppressed mice by dengue and West Nile immune sera.

No ADE of ZIKV by previous dengue immunity was detected in non-human primates.

No ADE of ZIKV was documented in a human cohort previously exposed to dengue.

ADE needs to be redefined in the context of clinical outcomes.

In vitro and experimental results in small animals need to be carefully weighed when translating results to humans.

Prospective epidemiological and clinical studies are needed to reassure that previous exposure to dengue or other flaviviruses does not increase the pathogenesis of ZIKV.

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clinical impairment along with a cytokine storm characterized by the increase in interleukin-6 (IL-6), IL-8, IL-10, interferon- γ (IFN- γ), IFN- α , and vascular endothelial growth factor (VEGF), combined with tumor necrosis factor- α (TNF- α), indicating a poor prognostic outcome [20] (Figure 1). Because of this, ADE related to flaviviruses should not be seen only as a single biological process of virus-antibody interaction. Defining ADE in the context of pathogenesis, as we usually read the outcome of the biological process, should imply a clinical consequence, including clinical and laboratory evidence of impairment. In this way, ADE would be defined as a common experimental *in vitro* phenomenon but a rare *in vivo* occurrence leading to worsening of the clinical presentation usually associated with hemodynamic changes, increased viremia, proinflammatory cytokine profile, and other detectable laboratory alterations.

Dengue Induced ZIKV ADE?

The debate of whether ZIKV ADE by flavivirus immune serum has recently increased because of results showing an increase in ZIKV pathogenesis in a mouse model (Stat2-/-) [21]. Using this model, Bardina *et al.* showed that, by administering DENV and West Nile virus (WNV) immune serum intraperitoneally, in an appropriate concentration before ZIKV infection, this resulted in fever and weight loss with an increased mortality as compared to some of the animals administered serum from flavivirus-naïve individuals [21]. However, results from the same

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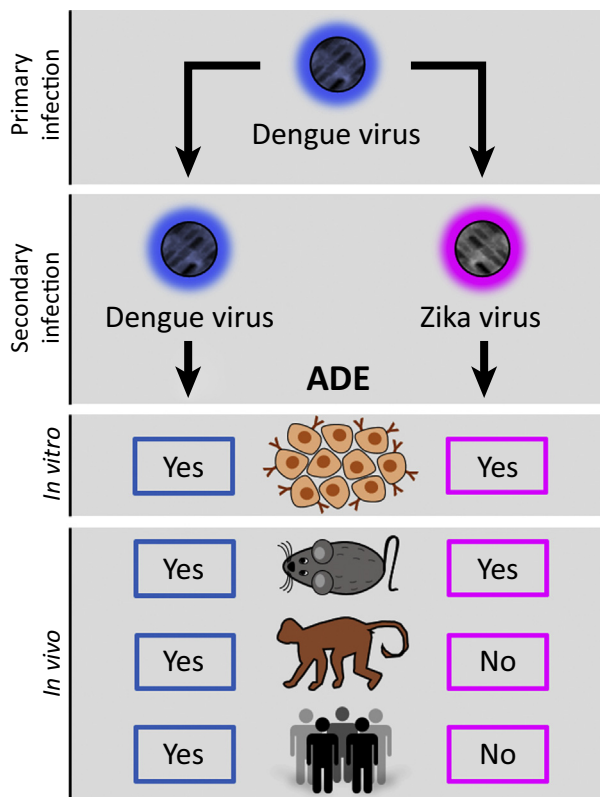
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Trends in Microbiology

Figure 1. Antibody-Dependent Enhancement (ADE) of Dengue and Zika Virus. ADE during a secondary heterologous Dengue virus infection has been documented *in vitro*, in mice, in non-human primates, and in humans playing a key role in the worsening of the clinical presentations. In contrast, ADE of Zika virus by pre-existing immunity to Dengue virus can be induced *in vitro*, and in immunodeficient mice. However, there is no evidence to support ADE occurring in non-human primates or in humans.

work, in a dose-dependent evaluation of mouse survival and the clinical presentation experiment, revealed that control plasma at the highest concentration could also decrease mice survival by about 40%, similar to the effect of DENV immune plasma at the lowest dilutions. In addition to proving that antibodies induced by prior DENV infection, administered under different concentrations, can amplify or neutralize ZIKV disease manifestations *in vivo*, Bardina *et al.* also showed the limited value of ADE *in vivo* in immunosuppressed mice.

Indeed, ADE of ZIKV by previous flavivirus infection is not a novel concept. Back in 1987 Fabgami *et al.* demonstrated that ZIKV replication can be enhanced in P388D1 macrophage cell line by subneutralizing concentrations of antibodies in immune ascitic fluids from six other different flaviviruses, including Wesselsbron, Uganda S, WNV, Dakar bat, yellow fever and Potiskum virus [22]. However, the following facts might anticipate the unlikelihood of DENV-induced ZIKV ADE (as defined above) in humans: (i) there is no epidemiological or clinical evidence of DENV ADE with any other closely related flavivirus or any other viruses; (ii) before its introduction into the Western hemisphere, ZIKV continuously circulated in flavivirus-endemic areas (such as Africa and Southeast Asia), and an increase in ZIKV pathogenesis has not been reported in these locations; (iii) not all heterologous flavivirus immunity is the same, including the sequence in which infection occurs with different DENV serotypes [18,19].

What Non-human Primates (NHPs) Can Tell Us

NHPs are natural hosts (in the sylvatic transmission cycle) supporting the replication of both DENV and ZIKV. For many years NHPs have been used as a surrogate for human infection in order to understand DENV pathogenesis and to test for vaccine immunogenicity and efficacy [23] – and, more recently, for ZIKV replication and pathogenesis [24–28]. In the past, DENV ADE, in terms of viral replication enhancement, has also been proven in NHPs after secondary DENV infection with DENV 2 [29] or by passive administration of optimal dilutions of human DENV-immune serum to the animals [30], or by using specific concentrations of a monoclonal antibody [31]. In addition to being useful for studying DENV pathogenesis, NHPs are a good model for predicting the behavior of different DENV vaccines in humans and for characterizing specific DENV neutralizing antibodies also occurring naturally in human populations [23]. Because of this, it is plausible to anticipate that data on ZIKV pathogenesis in NHPs can also reproduce or predict what will happen in humans. In a recent study, using a limited number of animals, Pantoja *et al.* were unable to show ADE of ZIKV in DENV immune macaques [32]. However, results showed that previous immunity to DENV was able to modulate the innate and cellular immune response to ZIKV with a tendency to lower the average ZIKV viremia days, to limit the increase in liver enzymes, and to induce a significant increase in the plasma perforin as evidence of an increased cytotoxic T cell activity [33]. This cellular immune response in NHPs is supported by recent results from human samples showing that prior DENV infection leads to stronger and faster responses to ZIKV in terms of both CD4 and CD8T cell responses, thus providing evidence of a biological outcome [34].

Human Evidence of ZIKV ADE?

Coincident with the report by Pantoja *et al.*, a study on ADE of a human cohort was published [35]. Terzian *et al.* evaluated a cohort of ZIKV-infected patients and looked for previous DENV exposure and its relationship to viral load, cytokine profile, and clinical symptoms. Despite the suggestions from *in vitro* studies that ADE could occur in DENV-primed ZIKV-infected patients, the authors found no evidence that the presence of DENV antibodies changes the outcome of ZIKV infection in all tested parameters [35]. Collectively, these observations from NHPs and human cohorts strongly suggest that previous exposure to DENV does not have a deleterious

Glossary

Antibody-dependent

enhancement (ADE): a mechanism when non-neutralizing antibodies facilitate virus entry into host cells, leading to increased infectivity in the cells and exacerbation of the clinical disease.

Congenital Zika syndrome (CZS):

a pattern of birth defects found among fetuses and babies infected with ZIKV during pregnancy.

Guillain-Barré syndrome (GBS):

an autoimmune disorder affecting the peripheral nervous system. Initial symptoms are weakness and numbness in the extremities, which eventually develop into paralysis and, if untreated, death.

effect in the clinical outcome of ZIKV infection. Supported by these observations, we can propose that the DENV-induced ZIKV ADE *in vitro* does not exist *in vivo* or that it is so uncommon that it might be not relevant as an epidemiological phenomenon.

Concluding Remarks

In summary, data from NHPs and humans, and from several serological studies [36,37], do not support the suggestion that ZIKV may be enhanced *in vivo* by previous exposure to DENV. On the other hand, the few experimental lines of evidence that have addressed ADE of DENV induced by ZIKV-immune serum, as expected, have shown different degrees of *in vitro* and *in vivo* increase in DENV replication and pathogenesis respectively [38,39].

Recently George *et al.* reported that an infection with DENV, after a short period of exposure to ZIKV, can enhance DENV infection in NHPs [40]. This is a very interesting report as it is expected that ZIKV-induced DENV cross-reacting antibodies, induced early after infection, may either neutralize or have no effect in DENV infection outcome. In any case, this report confirms the need for large studies in NHPs and for epidemiological data from the dengue-naïve human population that has been exposed to ZIKV during the recent epidemic.

Lastly, inferences derived from *in vitro* experiments and from immunologically modified animals will need to be carefully assessed due to the impact they can have on the approaches for ZIKV and DENV vaccines and therapeutics currently under development.

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Resources

<http://www.who.int/entity/csr/resources/publications/zika/classification/en/>

Supplemental Information

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References

1. Aliota, M.T. *et al.* (2017) Zika in the Americas, year 2: what have we learned? What gaps remain? A report from the Global Virus Network. *Antiviral Res.* 144, 223–246
2. Campos, G.C. *et al.* (2016) Zika virus infection, a new public health challenge. *Braz. J. Infect. Dis.* 20, 227–228
3. Brasil, P. *et al.* (2016) Zika virus infection in pregnant women in Rio de Janeiro. *N. Engl. J. Med.* 375, 2321–2334
4. Cao-Lormeau, V.M. *et al.* (2016) Guillain-Barré syndrome outbreak associated with Zika virus infection in French Polynesia: a case-control study. *Lancet* Published online February 29, 2016. [http://dx.doi.org/10.1016/S0140-6736\(16\)00562-6](http://dx.doi.org/10.1016/S0140-6736(16)00562-6)
5. Roze, B. *et al.* (2016) Zika virus detection in cerebrospinal fluid from two patients with encephalopathy, Martinique, February 2016. *Euro. Surveill.* Published online April 21, 2016. <http://dx.doi.org/10.2807/1560-7917.ES.2016.21.16.30205>
6. Carteaux, G. *et al.* (2016) Zika virus associated with meningoencephalitis. *N. Engl. J. Med.* 374, 1595–1596
7. Mecharles, S. *et al.* (2016) Acute myelitis due to Zika virus infection. *Lancet* 387, 1481
8. Furtado, J.M. *et al.* (2016) Uveitis associated with Zika virus infection. *N. Engl. J. Med.* 375, 394–396
9. Sharp, T.M. *et al.* (2016) Zika virus infection associated with severe thrombocytopenia. *Clin. Infect. Dis.* 63, 1198–1201
10. Peiris, J.S. and Porterfield, J.S. (1982) Antibody-dependent plaque enhancement: its antigenic specificity in relation to Togaviridae. *J. Gen. Virol.* 58, 291–296
11. King, A.A. *et al.* (1984) Antibody-mediated enhancement of rabies virus infection in a mouse macrophage cell line (P388D1). *J. Gen. Virol.* 65, 1091–1093
12. Jarasch-Althof, N. *et al.* (2010) Antibody-dependent enhancement of coxsackievirus B3 infection of primary CD19⁺ B lymphocytes. *Viral Immunol.* 23, 369–376
13. Wang, S.F. *et al.* (2014) Antibody-dependent SARS coronavirus infection is mediated by antibodies against spike proteins. *Biochem. Biophys. Res. Commun.* 451, 208–214
14. Fust, G. (1997) Enhancing antibodies in HIV infection. *Parasitology* 115 (Suppl), S127–S140

Outstanding Questions

What is the role of the time interval between a primary DENV and a subsequent ZIKV infection in the antibody and T cell immune response?

How will this scenario compare to having ZIKV infection following two or more DENV or other flavivirus infections?

How will this timing between infections impact the clinical outcome, if at all?

With the recent ZIKV epidemic there is a substantial population that is ZIKV-positive/DENV-negative. Are these people at risk of having worse clinical presentations or are they partially protected against DENV?

What will be the effect of a ZIKV component in the effectiveness of DENV vaccines currently in clinical trials or the one that has been licensed in some countries?

What is the susceptibility conferred on the population by the DENV and ZIKV vaccines to subsequent heterogenous natural infections with those viruses?

15. Morens, D.M. (1994) Antibody-dependent enhancement of infection and the pathogenesis of viral disease. *Clin. Infect. Dis.* 19, 500–512
16. Miner, J.J. and Diamond, M.S. (2017) Dengue antibodies, then Zika: a fatal sequence in mice. *Immunity* 46, 771–773
17. Halstead, S.B. *et al.* (1980) Enhancement of dengue virus infection in monocytes by flavivirus antisera. *Am. J. Trop. Med. Hyg.* 29, 638–642
18. de Alwis, R. *et al.* (2014) Dengue viruses are enhanced by distinct populations of serotype cross-reactive antibodies in human immune sera. *PLoS Pathog.* 10, e1004386
19. Guzman, M.G. *et al.* (2013) Secondary infection as a risk factor for dengue hemorrhagic fever/dengue shock syndrome: an historical perspective and role of antibody-dependent enhancement of infection. *Arch. Virol.* 158, 1445–1459
20. Srikiatkachorn, A. *et al.* (2017) Immune-mediated cytokine storm and its role in severe dengue. *Semin. Immunopathol.* 39, 563–574
21. Bardina, S.V. *et al.* (2017) Enhancement of Zika virus pathogenesis by preexisting antinflavivirus immunity. *Science* 356, 175–180
22. Fagbami, A.H. *et al.* (1987) Cross-infection enhancement among African flaviviruses by immune mouse ascitic fluids. *Cytobios* 49, 49–55
23. Sariol, C.A. and White, L.J. (2014) Utility, limitations, and future of non-human primates for dengue research and vaccine development. *Front. Immunol.* 5, 452
24. Dudley, D.M. *et al.* (2016) A rhesus macaque model of Asian-lineage Zika virus infection. *Nat. Commun.* 7, 12204
25. Haddow, A.D. *et al.* (2017) High infection rates for adult macaques after intravaginal or intrarectal inoculation with Zika virus. *Emerg. Infect. Dis.* 23, 1274–1281
26. Hirsch, A.J. *et al.* (2017) Zika virus infection of rhesus macaques leads to viral persistence in multiple tissues. *PLoS Pathog.* 13, e1006219
27. Nguyen, S.M. *et al.* (2017) Highly efficient maternal–fetal Zika virus transmission in pregnant rhesus macaques. *PLoS Pathog.* 13, e1006378
28. Osuna, C.E. *et al.* (2016) Zika viral dynamics and shedding in rhesus and cynomolgus macaques. *Nat. Med.* 22, 1448–1455
29. Halstead, S.B. *et al.* (1973) Studies on the pathogenesis of dengue infection in monkeys. II. Clinical laboratory responses to heterologous infection. *J. Infect. Dis.* 128, 15–22
30. Halstead, S.B. (1979) *In vivo* enhancement of dengue virus infection in rhesus monkeys by passively transferred antibody. *J. Infect. Dis.* 140, 527–533
31. Goncalvez, A.P. *et al.* (2007) Monoclonal antibody-mediated enhancement of dengue virus infection *in vitro* and *in vivo* and strategies for prevention. *Proc. Natl. Acad. Sci. U. S. A.* 104, 9422–9427
32. Petraleigh Pantoja, E.X.P.-G. *et al.* (2016) *Secondary Zika Virus Infection Do Not Support Evidences of Antibody-Dependent Enhancement in vivo in Dengue Pre-exposed Rhesus Macaques*, pp. 1–16, University of Puerto Rico Medical Sciences Campus Caribbean Research Center
33. Pantoja, P. *et al.* (2017) Zika virus pathogenesis in rhesus macaques is unaffected by pre-existing immunity to dengue virus. *Nat. Commun.* 8, 15674
34. Grifoni, A. *et al.* (2017) Prior Dengue virus exposure shapes T cell immunity to Zika virus in humans. *J. Virol.* Published online October 4, 2017. <http://dx.doi.org/10.1128/jvi.01469-17>
35. Terzian, A.C.B. *et al.* (2017) Viral load and cytokine response profile does not support antibody-dependent enhancement in dengue-primed Zika-infected patients. *Clin. Infect. Dis.* 65, 1260–1265
36. Collins, M.H. *et al.* (2017) Lack of durable cross-neutralizing antibodies against Zika virus from Dengue virus infection. *Emerg. Infect. Dis.* 23, 773–781
37. Swanstrom, J.A. *et al.* (2016) Dengue virus envelope dimer epitope monoclonal antibodies isolated from dengue patients are protective against Zika virus. *mBio* Published online July 19, 2016. <http://dx.doi.org/10.1128/mBio.01123-16>
38. Kawiecki, A.B. and Christofferson, R.C. (2016) Zika virus-induced antibody response enhances Dengue virus serotype 2 replication *in vitro*. *J. Infect. Dis.* 214, 1357–1360
39. Stettler, K. *et al.* (2016) Specificity, cross-reactivity and function of antibodies elicited by Zika virus infection. *Science* 353, 823–826
40. George, J. *et al.* (2017) Prior exposure to Zika virus significantly enhances peak Dengue-2 viremia in Rhesus macaques. *Sci. Rep.* 7, 10498