[†]Address correspondence to Scott Vrieze (vrieze@umn.edu), University of Minnesota, 75 East River Road, Minneapolis, MN 55455; or Dajiang J. Liu (dajiang.liu@psu.edu), Penn State College of Medicine, HCAR 2020, Hershey, PA.

[*]These authors contributed equally to the work.

[††]See acknowledgments for a list of authors associated with the replication consortia.

Exome chip meta-analysis fine maps causal variants and elucidates the genetic architecture of rare coding variants in smoking and alcohol use.

A full list of authors and affiliations appears at the end of the article.

Abstract

Background: Smoking and alcohol use have been associated with common genetic variants in multiple loci. Rare variants within these loci hold promise in the identification of biological mechanisms in substance use. Exome arrays and genotype imputation can now efficiently

genotype rare nonsynonymous and loss of function variants. Such variants are expected to have deleterious functional consequences, and contribute to disease risk.

Methods: We analyzed ~250,000 rare variants from 16 independent studies genotyped with exome arrays and augmented this dataset with imputed data from the UK Biobank. Associations were tested for five phenotypes: cigarettes per day, pack years, smoking initiation, age of smoking initiation, and alcoholic drinks per week. We conducted stratified heritability analyses, single-variant tests, and gene-based burden tests of nonsynonymous/loss of function coding variants. We performed a novel fine mapping analysis to winnow the number of putative causal variants within associated loci.

Results: Meta-analytic sample sizes ranged from 152,348–433,216, depending on the phenotype. Rare coding variation explained 1.1–2.2% of phenotypic variance, reflecting 11%–18% of the total SNP heritability of these phenotypes. We identified 171 genome-wide associated loci across all phenotypes. Fine mapping identified putative causal variants with double base-pair resolution at 24 of these loci, and between 3 and 10 variants for 65 loci. 20 loci contained rare coding variants in the 95% credible intervals.

Conclusions: Rare coding variation significantly contributes to the heritability of smoking and alcohol use. Fine mapping GWAS loci identifies specific variants contributing to the biological etiology of substance use behavior.

Keywords

Tobacco; Nicotine; Alcohol; GWAS; Heritability; Behavioral Genetics

Introduction

Tobacco and alcohol use together account for more morbidity and mortality in Western society than any other single risk factor or health condition(1). These preventable and modifiable behaviors are heritable(2), but previous human and model organism research, including genome-wide association studies of common variants, have resulted in few associated genetic variants, which most prominently feature genes involved in alcohol/ nicotine metabolism and nicotinic receptors(3–7).

Advances in sequencing, genotyping, and genotype imputation now allow cost effective investigation of rare and low frequency variants. Compared to common variants (minor allele frequency [MAF] > 1%) most commonly used in genome-wide association studies (GWAS), rare variants have greater potential to elucidate biological mechanisms of complex traits, including substance use and addiction(8, 9). In particular, nonsynonymous and loss of function (LoF) coding variants, which result in the loss of normal function of a protein, may have greater phenotypic impact and more direct mechanistic interpretation than other variants that do not have obvious biological consequences(10, 11).

No large-scale genome- or exome-wide study of rare variation has been conducted to date. The vast majority of existing addiction-related rare variant studies have used targeted sequencing of putative addiction-associated loci to discover and test for association in relatively small samples. Existing research has led to intriguing leads, including rare variant

associations in loci that span nicotinic receptor gene clusters(12–21) and alcohol metabolism genes(22–24) for nicotine and alcohol dependence, respectively. This strategy has also produced rare variant associations in novel loci. In one case, gene-level association tests were used to find an association with rare variants in *SERINC2*(24). In another case, a burden test across *PTP4A1*, *PHF3*, and *EYS* showed association with alcohol dependence(25). Unfortunately, these genes are not obviously involved in etiological processes related to addiction, and replications have not been reported to date.

Previous studies have also attempted to leverage information about predicted functional consequences of rare mutations to improve association analyses. One study of nicotine dependence found significant rare single-variant associations in *CHRNB4*, but only when variants were weighted by their predicted effect on the cellular response to nicotine and acetylcholine(26). Such positive findings could benefit from replication, which has not always been straightforward. For example, all rare variant associations in addiction are, to our knowledge, candidate gene analyses with type I error thresholds based only on the number of tests within that region. Historically, such analyses have produced overly optimistic estimates of the number of associated loci(27). Genome-wide analyses with more conservative type I error thresholds have reported null rare variant findings across an array of phenotypes relevant to addiction(28–30). Precisely because genome-wide analyses are conducted on many variants across the genome, they are in principle able to discover novel rare variant associations within new or known loci. One way to improve power in genome-wide analyses is through genetic association meta-analysis, which entails the aggregation of results across many studies to achieve large sample sizes.

Here, we attempted to expand on these previous discoveries by conducting the largest meta-analytic investigation of exonic rare variants to date. We conducted an exome-wide association meta-analysis of nicotine and alcohol use across 16 studies genotyped on the exome array, which genotypes low-frequency nonsynonymous and putative loss of function exonic variants. We combined these data with the UK Biobank, which includes approximately 400,000 individuals of European ancestry with genotype imputation to the Haplotype Reference Consortium(31) imputation reference panel and relevant smoking/drinking phenotypes. Sample sizes for well-imputed variants were thus enlarged and the availability of noncoding variants from UK Biobank enabled comprehensive analysis of genetic architecture(32) and fine mapping(33).

We conducted single variant and gene-based tests of association with five smoking and drinking phenotypes. We applied a novel fine mapping analysis to prioritize causal variants using statistical and functional information. We also evaluated the contribution of rare exonic variants to the heritability of these phenotypes. Family studies, as well as studies of the aggregate effects of common variants, have found both alcohol use and tobacco use to be heritable behaviors(30, 34–38). Research on the aggregate contribution of rare variants, however, has been scarce, with previous work on related phenotypes in smaller samples failing to detect aggregate effects for smoking and alcohol consumption(28). We used meta-analytic summary statistics to quantify the contribution to heritability of variants in various functional categories and frequency bins.

Methods and Materials

Seventeen studies contributed summary statistics for meta-analysis. These studies, their sample sizes, and available phenotypes are listed in the online supplement (Tables S1 and S2). We augmented our sixteen exome chip cohorts with the UK Biobank, where imputation to the Haplotype Reference Consortium panel was used in lieu of an exome chip array. All individuals were of European ancestry, as determined by genetic principal components.

Phenotypes

Phenotypes were selected to represent multiple stages of smoking. These included initiation, heaviness of use among smokers, and a measure of total lifetime exposure to tobacco. For alcohol use only a measure of amount of alcohol use was systematically available across studies. The selected phenotypes are relevant to prior GWAS of smoking and alcohol use; are commonly available in psychological, medical, and epidemiological data sets; and are known to be correlated with measures of substance dependence(4, 39–41).

1.   Cigarettes per day (CigDay).—The average number of cigarettes smoked in a day among current and former smokers. Studies with binned responses used their existing bins. Studies that recorded an integer value binned responses into one of four categories: 1=1–10, 2=11–20, 3=21–30, 4=31 or more. Anyone reporting 0 cigarettes per day was coded as missing. This phenotype is a component of commonly used measures of nicotine dependence such as the Fagerstrom Test for Nicotine Dependence.

2.   Pack Years (PckYr).—Defined in the same way as cigarettes per day but not necessarily binned, divided by 20 (cigarettes in a pack), and multiplied by number of years smoking. This yielded a measure of total overall exposure to tobacco and is relevant to disease outcomes for which smoking is a risk factor, such as cancer and chronic obstructive pulmonary disease risk.

3.   Age of Initiation of Smoking (AgeSmk).—A measure of early cigarette use. Defined as the age at which a participant first started smoking regularly.

4.   Smoking Initiation (SmkInit).—A binary variable of whether the individual had ever been a regular smoker (1) or not (0), and often defined as having smoked at least 100 cigarettes during one's lifetime.

5.   Drinks per week (DrnkWk).—A measure of drinking frequency/quantity. The average number of drinks per week in current or former drinkers.

Genotypes

Fourteen of the seventeen studies were genotyped with the Illumina HumanExome BeadChip, which contains ~250,000 low-frequency nonsynonymous variants, variants from the GWAS catalog, and a small number of variants selected for other purposes. Two studies were genotyped on the Illumina Human Core Exome, which includes an additional ~250,000 tag SNPs. The remaining study, the UK Biobank, was imputed using Haplotype Reference Consortium panel(31, 42), as well as the reference panel by UK 10K and 1000

Genomes Project. An integrated callset was released by the UK Biobank team[42]. Our UK Biobank genetic association analyses were conducted based on the integrated callset with additional quality control.

Generation of Summary Association Statistics

Seventeen independent studies (see Table S1) with smoking and drinking phenotypes were included in the discovery phase. Individual studies conducted association analysis accounting for age, sex, any study-specific covariates, and ancestry principal components (see Table S2 for genomic controls), and submitted summary statistics for meta-analysis. For studies with related individuals (**see** Table S1), relatedness was accounted for in linear mixed models using empirically estimated kinships from common SNPs[43]. Residuals were inverse-normalized to help ensure well-behaved test statistics for rare variant tests.

Quality control of per-study summary statistics included evaluation and correction of strand flips and allele flips through systematic comparison of alleles and allele frequencies against the reference datasets ExAC v2.0, 1000 Genomes Phase 3, and dbSNP. Variants with call rates<0.9, or Hardy Weinberg $p<1\times10^{-7}$ were also removed. The latter filter was meant to avoid findings that could not be more broadly replicated across the 17 studies.

Meta-analysis

Association testing was done in stages. First, we conducted genome-wide association meta-analysis. Variants with p-values less than the genome-wide significance threshold of 5×10^{-8} were deemed statistically significant. Loci were defined as 1 million basepair windows surrounding a "sentinel" (most significant) variant in the locus. Overlapping or adjacent loci were combined into a single locus. Conditional analysis and fine mapping was then performed within each locus. We attempted to replicate one very rare variant (rs36015615 in *STARD3* associated with CigDay; see results and Table 1) that was available in two other exome chip consortia. These were the CHD Exome+ Consortium (N=17,789) and the Consortium for Genetics of Smoking Behaviour (N=28,583). Both consortia defined their phenotypes, including cigarettes per day similarly, as the usual number of cigarettes smoked in a day corrected for sex, age, principal components (and/or genetic relatedness, as appropriate), and inverse-normalized prior to association analysis.

We also conducted gene-level association tests grouping nonsynonymous, stop gain, stop loss and splice variants within each gene, using rareMETALS version 6.0[44]. Variant annotation was conducted using SEQMINER with RefSeq 1.9[45]. Two complementary gene-level association tests were performed: the sequence kernel association test (SKAT; 46, 47) with a MAF cutoff of 1% and a simple burden test[48] that summed the number of rare alleles within a given gene, again with a maximum MAF=1%. We chose variants with MAF 1% as we were interested in the contribution of variants with a frequency lower than that which has been reliably imputed and tested in past GWAS meta-analyses. We considered a gene association to be significant if the p-value surpassed a Bonferroni correction for the number of genes tested for a given phenotype and test, assuming approximately 20,000 genes in the genome ($.05/20,000 = 2.5\times10^{-6}$).

We performed iterative conditional analysis using a partial correlation based score (PCBS) statistic(49), which can perform proper conditional analysis for meta-analysis that combines datasets measured using different arrays. PCBS takes GWAS meta-analysis summary statistics and LD estimated from the Haplotype Reference Consortium panel as input.

As a key step to evaluate the contribution of variants within a genome-wide significant locus(33), we used our PCBS framework to apply two complementary fine mapping techniques to identify putatively causal genetic variants. The first technique was a Bayesian approach described previously(50) that estimates the posterior probability of association based upon the statistical strength of the association for variants in each locus. We also applied a version of fgwas(51) modified to work within PCBS, which assumes that variants in different functional categories have potentially different prior probability of association. For loci with a single association signal based, effect sizes and variance from single-SNP analyses were used. If a locus contained multiple signals, we used effect sizes and variance from conditional analysis adjusting for all other index variants in this region.

Finally, we attempted to replicate previous rare variant associations referenced in the introduction and listed in Table S4. We attempted replication in our phenotypes for any single variant when that variant was directly genotyped or imputed. We applied a liberal threshold that corrected only for the number of tests conducted for this replication exercise (.05/46=.001).

Genetic Architecture

We performed heritability and genetic correlation analyses using LD score regression(52). The method calculates LD scores from the Haplotype Reference Consortium and the estimation of heritability with these LD scores then follows established methods(53, 54). Heritability was estimated for each trait and partitioned by annotation category and frequency bins. First, we annotated variants on the exome chip based upon gene definitions in RefSeq 1.9, using SEQMINER version 6.0(55). A variant is classified as coding if it belongs to either one of the following categories: nonsynonymous, stop gain, stop loss, and splice. Seven functional categories were considered in the model, including intergenic, intron, common coding (MAF>0.01), rare coding (MAF<0.01), synonymous, and 3'/5' untranslated regions. We fitted the baseline model with seven categories, and estimated phenotypic variance explained by each category.

Results

GWAS analyses behaved well, with genomic control values for the GWAS across exome chip and UK Biobank imputed variants between 1.05 and 1.3. The intercept for LD Score regression ranged between .99 and 1.1, indicating absent or minimal effects of population stratification. (Per-study genomic controls can be found in Table S2.) A total of 171 loci were identified under the genome-wide significance threshold ($p<5\times10^{-8}$), including 3, 11, 17, 93 and 47 loci for AgeSmk, CigDay, PckYr, SmkInit, and DrnkWk. A list of all sentinel variants within each locus is shown in Table S5. QQ plots and Manhattan plots are available in Figures S1 and S2. (Additional exploratory GWAS meta-analysis of individuals with significant African ancestry are provided in the Supplementary Note [including up to 8,974

individuals from three studies]; see also Table S3, Figure S3 and S4.) The genome-wide significant association results included known loci associated with smoking and alcohol use phenotypes. These included associations between smoking phenotypes and variants within the *CHRNA5-CHRNA3-CHRNB4* nicotinic receptor cluster, nicotine metabolism gene *CYP2A6,* and a locus near dopamine receptor *DRD2.* We also replicated previous associations between nonsynonymous variant rs1229984 in *ADH1B* and DrnkWk. Only one very rare variant was associated with any of our five phenotypes. This was rs36015615 (MAF=.0002), a nonsynonymous variant in *STARD3*, associated with CigDay (p=3.2×10^{-8}). This novel variant did not replicate in either of two replication consortium datasets, the CHD Exome+ Consortium (N=17,789, Beta=−.01, *p*=.94) or the Consortium for Genetics of Smoking Behaviour (N=28,583, Beta=.056, *p*=.84). Based upon the estimated genetic effects in the discovery sample ($\beta = 1.2$), the power for replication is >99%. However, if we assume the observed effect sizes in the replication datasets are correct, there is 5% power for replication based upon this estimated effect. The pattern of results may be due to winner's curse, or the discovered variant may be a false positive finding. Additional studies are required to narrow the possible interpretations.

The fine mapping analysis of all 171 GWAS loci pinpointed putatively causal variants with high resolution in some cases. The 95% credible interval for 34% of the loci had <10 SNPs and 24 loci had double basepair resolution, including several instances where the sole putative causal variant was nonsynonymous and of lower frequency, although in only one case with MAF<1%. The resolution increased somewhat when functional information was used to inform the prior, with double base-pair resolution at 32 loci, and 44% of loci having <10 SNPs in the 95% credible interval. Table 1 includes all nonsynonymous or loss of function variants within the genome-wide significant loci that had a posterior probability of association greater than .80 from at least one of the fine mapping methods. Additional results from the fine mapping analysis are available in Tables S6 and S7. Several known functional variants were identified through this method, including: rs16969968(56), a nonsynonymous variant in nicotinic receptor gene *CHRNA5* associated with CigDay (PPA=.92 and .84 from the fine mapping analysis with, and without, functional priors, respectively); rs1229984(57), a nonsynonymous variant in alcohol metabolism gene *ADH1B* associated with DrnkWk (PPA=1.0 and 1.0); and, although with somewhat weaker evidence, rs6265(58), a nonsynonymous variant in brain derived neurotrophic factor *BDNF* associated with SmkInit (MAF=.19; PPA=.83 and .32).

Novel variants in novel genes were also prioritized at high resolution. To take the most statistically compelling examples in Table 1, we found rs28929474, a low frequency nonsynonymous variant in *SERPINA1*, associated with DrnkWk (MAF=.02; PPA=1.0 and .95). When homozygous, the alternate T (allele frequency = .02; frequency of TT genotype under Hardy Weinberg = 4 in 10,000) allele is a leading cause of alpha-1 antitrypsin deficiency. Here, we find the same risk allele, the T allele, is associated with an approximately .05 standard deviation decrease in drinks per week. We also discovered rs35891966, a variant in *NAV2*, associated with SmkInit (MAF=.07; PPA = 1.0 and .98) at single base-pair resolution. *NAV2* is involved in neuronal development and previously shown to be differentially expressed between smokers and non-smokers, but not previously implicated in GWAS(59).

Results of gene-based tests are provided in Table 2. A novel gene, rho guanine nucleotide exchange factor 37 (*ARHGEF37*), was associated with Age of Initiation of Smoking (p=1.9×10⁻⁶). *ARHGEF37* has not been widely studied and its function in not well known. Another novel gene without an immediate biological interpretation, was HEAT Repeat Containing 5A (*HEATR5A*), associated with Smoking Initiation (p=1.4×10⁻⁸). We also discovered a significant gene-based association between known alcohol metabolism gene *ADH1C* and Drinks per Week (p=1.4×10⁻²⁷ and p=1.9×10⁻⁴⁰ from the burden and SKAT tests, respectively). Finally, even with relaxed p-value thresholds, we failed to replicate genes identified in previous rare variant association studies referenced in the introduction (Table S4), with the exception of *ADH1C* and *CHRNA5*, two loci long known to be associated with alcohol use and smoking, respectively.

The estimated total SNP heritability for AgeSmk, CigDay, PckYr, SmkInit, and DrnkWk was 6%, 9%, 10%, 14% and 16%. Significant phenotypic variance was explained by rare nonsynonymous variants for all traits, ranging from 1.0%−2.2% (Table 3). As a fraction of the SNP heritability, rare nonsynonymous variants accounted for 11%−18%. Results for all seven functional categories are listed in Table S8; appreciable heritability was accounted for by common and rare coding variants, and intergenic variants. Variants in the untranslated regions and intronic regions contributed less. Almost all pairs of phenotypes were genetically correlated (Table 4, **Panel A**), and the direction of the genetic correlations were in the expected direction. For instance, CigDay was positively correlated with DrnkWk (0.2 ± 0.09), consistent with the observation that increased alcohol consumption is correlated with increased tobacco consumption. Age of initiation has a negative correlation with all other traits, which is consistent with the observation that an earlier age of smoking initiation is correlated with increased tobacco and alcohol consumption in adulthood. The patterns and magnitudes of correlation are highly similar when considering only rare nonsynonymous variants (Table 4, **Panel B**).

Discussion

With a maximum sample size ranging from 152,348 to 433,216, the present study is the largest study to date of low-frequency nonsynonymous and loss of function variants in smoking and alcohol use. Our meta-analytic study design combined studies genotyped on the exome array with imputed genotypes in the UK Biobank, allowed us to comprehensively evaluate the contribution of rare and low frequency variants to the etiology of tobacco and alcohol use. All told, we identified 171 genome-wide significant loci for the five phenotypes.

We showed that the rare variants (MAF 1%) together explain 1.0% - 2.2% of the phenotypic variance for the five traits, amounting to 11–18% of the total SNP heritability. A number of putatively causal low frequency nonsynonymous variants in novel genes were identified through two complementary fine mapping techniques. These include a variant known to affect alpha-1 antitrypsin deficiency in *SERPINA1*. The effect of the risk allele resulted in a decrease in drinks per week. One interpretation is that this variant leads to impaired liver function through alpha-1 antitrypsin deficiency which, in turn, reduces alcohol consumption. Interestingly, neither this particular variant or the locus surrounding it was associated with smoking phenotypes, even though alpha-1 antitrypsin deficiency also affects lung function

over time. Other mechanisms by which *SERPINA1* exerts its effect on alcohol consumption are certainly possible. Another novel nonsynonymous variant was in neuron navigator 2 (*NAV2*), associated with smoking initiation. *NAV2* has not previously been associated with substance use or addiction. Given its suspected involvement in neuronal growth and migration, a putatively causal nonsynonymous variant is a strong candidate for functional follow up experiments. Other genes implicated in the fine mapping analysis have less direct interpretations (e.g., *HEATR5A*) and such results will benefit from replication and/or follow-up experiments. In general, fine mapping studies narrowed the credible set of likely causal variants to single or double base pair resolution for 24 loci (Table S6). Some loci were not amenable to fine mapping, with credible intervals containing thousands of SNPs in some cases. Given the cost in money and time of conducting functional experiments at the cellular or organismal level, fine mapping likely causal variants can be extremely useful in predicting functional consequences and prioritizing variants for further work.

Gene based tests identified a small number of associated genes, including an expected association with *ADH1C* and drinks per week. The other two associated genes, *ARHGEF37* and *HEATR5A*, do not lend themselves to ready biological interpretations.

We showed that rare coding variants available on the exome chip or imputable by the Haplotype Reference Consortium, with frequency <1%, explain significant proportions of phenotypic variance, and a substantial proportion of the total SNP heritability. The exome chip was designed to genotype coding variants uncovered in ~12,000 sequenced exomes. By design, it comprehensively ascertained high confidence rare nonsynonymous, splice, and stop variants within those sequences and only sparsely genotypes other classes of variation, including common variants. The Haplotype Reference Consortium panel imputed data also have limited accuracy when the underlying genetic variants are rare. Therefore, our current investigation did not fully explore the genetic architecture of very rare variants (i.e. with MAF<0.1%). With the development of larger imputation reference panels, and the availability of large scale deep whole genome sequences (such as the Trans-Omics for Precision Medicine Study [TOPMed]), we expect to be able to conduct an even more comprehensive analysis of the genetic architecture for variants with ever lower frequencies. Ultimately, the discovery of low frequency with small effects will require even larger sample sizes. For example, for rare variant with MAF of .1% and effects of .2, .15, and 0.1 standard deviations on the phenotype, to identify associations at $a = 5 \times 10^{-8}$ with 80% of power, sample sizes of 500,000 890,000 and 1,990,000 are required. While such numbers seemed astronomical just a few years ago, they will indeed be attainable in the next few years with the availability of large biobank datasets and ever improving imputation. Another limitation of the present study is the limited samples sizes from non-European ancestries, where only exploratory analyses were possible. Substantial improvements can be made to the resolution of fine mapping analysis by leveraging disparate LD information across samples with diverse ancestry(33). Future research will do well to include individuals of diverse ancestry.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Authors

David M. Brazel[*],
Institute for Behavioral Genetics, University of Colorado Boulder Department of Molecular, Cellular, and Developmental Biology, University of Colorado Boulder

Yu Jiang[*],
Department of Public Health Sciences, Penn State College of Medicine, Hershey, PA

Jordan M. Hughey[*],
Department of Public Health Sciences, Penn State College of Medicine, Hershey, PA

Valérie Turcot,
Montreal Heart Institute, Montreal, Quebec, H1T 1C8, Canada Department of Medicine, Faculty of Medicine, Université de Montréal, Montreal, Quebec, H3T 1J4, Canada

Xiaowei Zhan,
Department of Clinical Science, Center for Genetics of Host Defense, University of Texas Southwestern

Jian Gong,
Public Health Sciences Division, Fred Hutchinson Cancer Research Center

Chiara Batini,
Department of Health Sciences, University of Leicester

J. Dylan Weissenkampen,
Department of Public Health Sciences, Penn State College of Medicine, Hershey, PA

MengZhen Liu,
Department of Psychology, University of Minnesota

CHD Exome+ Consortium[††], Consortium for Genetics of Smoking Behaviour[††],
Daniel R. Barnes,
Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK

Sarah Bertelsen,
Department of Neuroscience, Icahn School of Medicine at Mount Sinai

Yi-Ling Chou,
Washington University

A. Mesut Erzurumluoglu,
Department of Health Sciences, University of Leicester

Jessica D. Faul,
Survey Research Center, Institute for Social Research, University of Michigan

Jeff Haessler,
Public Health Sciences Division, Fred Hutchinson Cancer Research Center

Anke R. Hammerschlag,
Department of Complex Trait Genetics, Center for Neurogenomics and Cognitive Research, Amsterdam Neuroscience, VU University Amsterdam

Chris Hsu,
University of Southern California

Manav Kapoor,
Department of Neuroscience, Icahn School of Medicine at Mount Sinai

Dongbing Lai,
Department of Medical and Molecular Genetics, Indiana University School of Medicine

Nhung Le,
Department of Medical Microbiology, Immunology and Cell Biology, Southern Illinois University School of Medicine

Christiaan A de Leeuw,
Department of Complex Trait Genetics, Center for Neurogenomics and Cognitive Research, Amsterdam Neuroscience, VU University Amsterdam

Anu Loukola,
Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Helsinki, Finland; Department of Public Health, University of Helsinki, Helsinki, Finland

Massimo Mangino,
Department of Twin Research and Genetic Epidemiology, Kings College London, London SE1 7EH, UK; NIHR Biomedical Research Centre at Guy's and St Thomas' Foundation Trust, London SE1 9RT, UK

Carl A. Melbourne,
Department of Health Sciences, University of Leicester

Giorgio Pistis,
Istituto di Ricerca Genetica e Biomedica, Consiglio Nazionale delle Ricerche (CNR), Monserrato, Cagliari, Italy.

Beenish Qaiser,
Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Helsinki, Finland; Department of Public Health, University of Helsinki, Helsinki, Finland

Rebecca Rohde,
University of North Carolina, Chapel Hill

Yaming Shao,
University of North Carolina, Chapel Hill

Heather Stringham,
Department of Biostatistics, University of Michigan

Leah Wetherill,
Department of Medical and Molecular Genetics, Indiana University School of Medicine

Wei Zhao,
Department of Epidemiology, School of Public Health, University of Michigan

Arpana Agrawal,
Department of Psychiatry, Washington University School of Medicine

Laura Bierut,
Department of Psychiatry, Washington University School of Medicine

Chu Chen,
Public Health Sciences Division, Fred Hutchinson Cancer Research Center
Department of Epidemiology and Department of Otolaryngology; Head and Neck Surgery, University of Washington, Seattle, WA

Charles B. Eaton,
Department of Family Medicine, Brown University, Providence, RI

Alison Goate,
Department of Neuroscience, Icahn School of Medicine at Mount Sinai

Christopher Haiman,
Department of Preventative Medicine, Keck School of Medicine, University of Southern California

Andrew Heath,
Department of Psychiatry, Washington University

William G. Iacono,
Department of Psychology, University of Minnesota

Nicholas G. Martin,
Queensland Institute for Medical Research

Tinca J. Polderman,
Department of Complex Trait Genetics, Center for Neurogenomics and Cognitive Research, Amsterdam Neuroscience, VU University Amsterdam

Alex Reiner,
Public Health Sciences Division, Fred Hutchinson Cancer Research Center, Department of Epidemiology, University of Washington, Seattle, WA

John Rice,
Departments of Psychiatry and Mathematics, Washington University St. Louis

David Schlessinger,
National Institute on Aging, National Institutes of Health

H Steven Scholte,
Department of Psychology, University of Amsterdam & Amsterdam Brain and Cognition, University of Amsterdam

Jennifer A. Smith,
Department of Epidemiology, School of Public Health, University of Michigan

Jean-Claude Tardif,
Montreal Heart Institute, Montreal, Quebec, H1T 1C8, Canada Department of
Medicine, Faculty of Medicine, Université de Montréal, Montreal, Quebec, H3T 1J4,
Canada

Hilary A. Tindle,
Department of Medicine, Vanderbilt University, Nashville, TN

Andries R van der Leij,
Department of Psychology, University of Amsterdam & Amsterdam Brain and
Cognition, University of Amsterdam

Michael Boehnke,
Department of Biostatistics, School of Public Health, University of MIchigan

Jenny Chang-Claude,
Division of Cancer Epidemiology, German Cancer Research Center

Francesco Cucca,
Istituto di Ricerca Genetica e Biomedica, Consiglio Nazionale delle Ricerche (CNR),
Monserrato, Cagliari, Italy.

Sean P. David,
Department of Medicine, Stanford University, Stanford, CA

Tatiana Foroud,
Department of Medical and Molecular Genetics, Indiana University School of
Medicine

Joanna M.M. Howson,
Department of Public Health and Primary Care, University of Cambridge

Sharon L.R. Kardia,
Department of Epidemiology, School of Public Health, University of Michigan

Charles Kooperberg,
Public Health Sciences Division, Fred Hutchinson Cancer Research Center

Markku Laakso,
University of Eastern Finland, Finland

Guillaume Lettre,
Montreal Heart Institute, Montreal, Quebec, H1T 1C8, Canada Department of
Medicine, Faculty of Medicine, Université de Montréal, Montreal, Quebec, H3T 1J4,
Canada

Pamela Madden,
Department of Psychiatry, Washington University

Matt McGue,
Department of Psychology, University of Minnesota

Kari North,
Department of Epidemiology, University of North Carolina, Chapel Hill

Danielle Posthuma,
Department of Complex Trait Genetics, Center for Neurogenomics and Cognitive Research, Amsterdam Neuroscience, VU University Amsterdam, Department of Clinical Genetics, VU University Medical Centre Amsterdam, Amsterdam Neuroscience

Timothy Spector,
Department of Genetic Epidemiology, Kings College, London

Daniel Stram,
Department of Preventative Medicine, Keck School of Medicine, University of Southern California

Martin D. Tobin,
Department of Health Sciences, University of Leicester

David R. Weir,
Survey Research Center, Institute for Social Research, University of Michigan

Jaakko Kaprio,
Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Helsinki, Finland; Department of Public Health, University of Helsinki, Helsinki, Finland

Gonçalo R. Abecasis,
Regeneron Pharmaceuticals; Department of Biostatistics, School of Public Health, University of Michigan

Dajiang J. Liu[†], and
Institute of Personalized Medicine, Penn State College of Medicine

Scott Vrieze[†]
Department of Psychology, University of Minnesota

Affiliations

Institute for Behavioral Genetics, University of Colorado Boulder Department of Molecular, Cellular, and Developmental Biology, University of Colorado Boulder

Department of Public Health Sciences, Penn State College of Medicine, Hershey, PA

Department of Public Health Sciences, Penn State College of Medicine, Hershey, PA

Montreal Heart Institute, Montreal, Quebec, H1T 1C8, Canada Department of Medicine, Faculty of Medicine, Université de Montréal, Montreal, Quebec, H3T 1J4, Canada

Department of Clinical Science, Center for Genetics of Host Defense, University of Texas Southwestern

Public Health Sciences Division, Fred Hutchinson Cancer Research Center

Department of Health Sciences, University of Leicester

Department of Public Health Sciences, Penn State College of Medicine, Hershey, PA

Department of Psychology, University of Minnesota

Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK

Department of Neuroscience, Icahn School of Medicine at Mount Sinai

Washington University

Department of Health Sciences, University of Leicester

Survey Research Center, Institute for Social Research, University of Michigan

Public Health Sciences Division, Fred Hutchinson Cancer Research Center

Department of Complex Trait Genetics, Center for Neurogenomics and Cognitive Research, Amsterdam Neuroscience, VU University Amsterdam

University of Southern California

Department of Neuroscience, Icahn School of Medicine at Mount Sinai

Department of Medical and Molecular Genetics, Indiana University School of Medicine

Department of Medical Microbiology, Immunology and Cell Biology, Southern Illinois University School of Medicine

Department of Complex Trait Genetics, Center for Neurogenomics and Cognitive Research, Amsterdam Neuroscience, VU University Amsterdam

Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Helsinki, Finland; Department of Public Health, University of Helsinki, Helsinki, Finland

Department of Twin Research and Genetic Epidemiology, Kings College London, London SE1 7EH, UK; NIHR Biomedical Research Centre at Guy's and St Thomas' Foundation Trust, London SE1 9RT, UK

Department of Health Sciences, University of Leicester

Istituto di Ricerca Genetica e Biomedica, Consiglio Nazionale delle Ricerche (CNR), Monserrato, Cagliari, Italy.

Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Helsinki, Finland; Department of Public Health, University of Helsinki, Helsinki, Finland

University of North Carolina, Chapel Hill

University of North Carolina, Chapel Hill

Department of Biostatistics, University of Michigan

Department of Medical and Molecular Genetics, Indiana University School of Medicine

Department of Epidemiology, School of Public Health, University of Michigan

Department of Psychiatry, Washington University School of Medicine

Department of Psychiatry, Washington University School of Medicine

Public Health Sciences Division, Fred Hutchinson Cancer Research Center Department of Epidemiology and Department of Otolaryngology; Head and Neck Surgery, University of Washington, Seattle, WA

Department of Family Medicine, Brown University, Providence, RI

Department of Neuroscience, Icahn School of Medicine at Mount Sinai

Department of Preventative Medicine, Keck School of Medicine, University of Southern California

Department of Psychiatry, Washington University

Department of Psychology, University of Minnesota

Queensland Institute for Medical Research

Department of Complex Trait Genetics, Center for Neurogenomics and Cognitive Research, Amsterdam Neuroscience, VU University Amsterdam

Public Health Sciences Division, Fred Hutchinson Cancer Research Center, Department of Epidemiology, University of Washington, Seattle, WA

Departments of Psychiatry and Mathematics, Washington University St. Louis

National Institute on Aging, National Institutes of Health

Department of Psychology, University of Amsterdam & Amsterdam Brain and Cognition, University of Amsterdam

Department of Epidemiology, School of Public Health, University of Michigan

Montreal Heart Institute, Montreal, Quebec, H1T 1C8, Canada Department of Medicine, Faculty of Medicine, Université de Montréal, Montreal, Quebec, H3T 1J4, Canada

Department of Medicine, Vanderbilt University, Nashville, TN

Department of Psychology, University of Amsterdam & Amsterdam Brain and Cognition, University of Amsterdam

Department of Biostatistics, School of Public Health, University of MIchigan

Division of Cancer Epidemiology, German Cancer Research Center

Istituto di Ricerca Genetica e Biomedica, Consiglio Nazionale delle Ricerche (CNR), Monserrato, Cagliari, Italy.

Department of Medicine, Stanford University, Stanford, CA

Department of Medical and Molecular Genetics, Indiana University School of Medicine

Department of Public Health and Primary Care, University of Cambridge

Department of Epidemiology, School of Public Health, University of Michigan

Public Health Sciences Division, Fred Hutchinson Cancer Research Center

University of Eastern Finland, Finland

Montreal Heart Institute, Montreal, Quebec, H1T 1C8, Canada Department of Medicine, Faculty of Medicine, Université de Montréal, Montreal, Quebec, H3T 1J4, Canada

Department of Psychiatry, Washington University

Department of Psychology, University of Minnesota

Department of Epidemiology, University of North Carolina, Chapel Hill

Department of Complex Trait Genetics, Center for Neurogenomics and Cognitive Research, Amsterdam Neuroscience, VU University Amsterdam, Department of Clinical Genetics, VU University Medical Centre Amsterdam, Amsterdam Neuroscience

Department of Genetic Epidemiology, Kings College, London

Department of Preventative Medicine, Keck School of Medicine, University of Southern California

Department of Health Sciences, University of Leicester

Survey Research Center, Institute for Social Research, University of Michigan

Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Helsinki, Finland; Department of Public Health, University of Helsinki, Helsinki, Finland

Regeneron Pharmaceuticals; Department of Biostatistics, School of Public Health, University of Michigan

Institute of Personalized Medicine, Penn State College of Medicine

Department of Psychology, University of Minnesota

Acknowledgements:

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Table 1.

All nonsynonymous/loss of function variants with posterior probability of association > .80 from one of the two fine mapping methods.

SNP	REF/ALT	N	ALT AF	GWAS p-value	Beta	SE	Direction	Annotation	Posterior Probability of Association		Number SNPs (Low Frequency Coding SNPs) in 95% Credible Interval	
									W/out Functional Prior	W/ Functional Prior (fgwas)	W/out Functional Prior	W/ Functional Prior (fgwas)
Cigarettes per Day (CigDay)												
rs36015615[a]	G/A	69,951	.0002	3.2×10^{-8}	1.2	.210	==+=+=+=X=X+X+++	Nonsynonymous [STARD3]	.82	.62	8,997 (6211)	11302 (6232)
rs16969968	G/A	153,918	.34	2.5×10^{-139}	.096	.0038	+-+-+-+++-++-++++	Nonsynonymous [CHRNA5]	.84	.92	2(0)	2 (0)
Drinks per Week (DrnkWk)												
rs1260326	T/C	357,854	.61	4.6×10^{-40}	0.032	.0024	++++++++++-+++++	Nonsynonymous [GCKR]	1.0	1.0	1 (0)	1 (0)
rs1229984	T/C	334,588	.98	2.3×10^{-173}	0.25	.0088	=+-XXXX+XXXX=+++	Nonsynonymous [ADH1B]	1.0	1.0	1 (1)	1 (1)
rs28929474	C/T	357,854	.02	2.2×10^{-11}	-.057	.0085	-++-----++----+	Nonsynonymous [SERPINA1]	.95	1.0	1 (1)	1 (1)
rs1800566	G/A	357,854	.18	2.00×10^{-8}	0.017	.0031	+++++++++++-++-+	Nonsynonymous [NQO1]	.32	.97	103 (0)	1 (0)
Smoking Initiation (SmkInit)												
rs2232423	A/G	433,216	.11	1.40×10^{-8}	-0.019	.0034	-+-+---+------	Nonsynonymous [ZSCAN12]	.84	.64	502 (0)	2 (0)
rs35891966	G/A	433,216	.07	1.30×10^{-8}	-0.024	.0042	---+----++-----	Nonsynonymous [NAV2]	.98	1.0	1 (0)	1 (0)
rs14052174	G/T	433,216	.02	1.2×10^{-7}	.043	.0080	++++++++++-+++++	Nonsynonymous [FAM163A]	.81	1.0	2432(66)	1 (0)
rs6265	C/T	433,216	.19	1.9×10^{-10}	-.017	.0030	++-++-++++-+----	Nonsynonymous [BDNF]	.32	.83	25(0)	2 (0)
rs61754158	C/T	433,216	.01	1.44×10^{-6}	-.055	.0114	---+----=+=-++-	Nonsynonymous [HEATR5A]	.39	.87	9742(195)	9742 (195)
rs34967813	A/G	433,216	.31	8.1×10^{-7}	-.011	.0023	-----+----+-----	Nonsynonymous [RYR2]	.14	.98	7413(56)	1 (0)

[a] rs36015615 did not replicate in two additional datasets. See results section.

Note: REF=reference allele on GRCh37, ALT=alternate allele, N=sample size across all studies that genotyped the variant, ALT AF=allele frequency of the alternate allele estimated in the meta-analysis. A variant is considered "rare" if MAF < .01, and low frequency if MAF < .05. In the DIRECTION column each symbol represents the contribution of one of the studies. A "+" indicates the ALT allele had a positive effect in that study; "−" indicates a negative effect. A "=" indicates the variant was monomorphic and "X" indicates it was absent in that study. The order of studies for CigDay and DrnkWk was ARIC, UKB, COGA, FINNTWIN, FUSION, GECCO, HRS, ID1000, MEC, METSIM, MHI, MCTFR, NAGOZALC, NESCOG, SardiNIA, TwinsUK, and WHI. For SmkInit the order is the same except COGA and MCTFR were not available. See the supplemental materials for study acronym explanations.

Table 2.

Significant gene based test results, assuming a Bonferroni threshold of $.05/20{,}000 = 2.5 \times 10^{-6}$.

Phenotype	Gene	N	Number Variants	Beta	SE	p−value	Method
Age of Initiation of Smoking	*ARHGEF37*	147,010	17	.08	.017	1.9×10^{-6}	Burden
Smoking Initiation	*HEATR5A*	427,262	41	−.02	.009	1.4×10^{-8}	SKAT
Drinks per Week	*ADH1C*	353,265	4	−.15	.014	1.8e−27	Burden
Drinks per Week	*ADH1C*	353,265	4	−.15	.014	1.9e−40	SKAT

Note: no significant genes were identified for the other two phenotypes.

Table 3:

Estimation of Heritability Explained by Variants on Exome Array. We estimate the heritability based upon a baseline model with seven different functional categories. The reported heritability $\hat{h}^2$ is based upon the cumulative value from the functional categories with significant heritabilities. We also report the standard deviation ($se\left(\hat{h}^2\right)$) and p–values, estimated using jackknife.

Annotation	Phenotype	Heritability Estimates		
		$\hat{h}^2$	(se ($\hat{h}^2$))	P–Value
All Variants	Age of Initiation of smoking	.06	.0049	7.7×10^{-35}
	Cigarettes per Day	.09	.0019	$< 2.2\times10^{-303}$
	Pack Years	.10	.0022	$< 2.2\times10^{-303}$
	Smoking Initiation	.14	.0007	$< 2.2\times10^{-303}$
	Drinks per Week	.16	.0089	7.3×10^{-73}
Rare Coding Variants (MAF<.01)	Age of Initiation of smoking	.011	.0015	2.8×10^{-2}
	Cigarettes per Day	.010	.0006	1.7×10^{-2}
	Pack Years	.018	.0007	8.5×10^{-6}
	Smoking Initiation	.022	.0002	3.9×10^{-16}
	Drinks per Week	.020	.0013	1.8×10^{-7}

Table 4:
Estimation of Genetic Correlation Between Smoking and Drinking Traits.

We estimate genetic correlations between five smoking and drinking traits. Genetic correlation estimates $\left(\hat{r}_g\right)$, their standard deviation $(se\left(\hat{r}_g\right))$ and p–values are reported.

Trait 1	Trait 2	Genetic Correlation		
		$\hat{r}_g$	$se\left(\hat{r}_g\right)$	P–value
A. Aggregated Genetic Correlation Induced by All Variants on the Exome Array				
Drinks per Week	Smoking Initiation	.43	.06	1.7×10^{-11}
Drinks per Week	Age of Initiation of Smoking	.01	.13	9.3×10^{-1}
Drinks per Week	Pack Years	.22	.10	2.6×10^{-2}
Drinks per Week	Cigarettes per Day	.20	.09	3.1×10^{-2}
Smoking Initiation	Age of Initiation of Smoking	−.64	.11	1.1×10^{-8}
Smoking Initiation	Pack Years	.45	.08	4.9×10^{-8}
Smoking Initiation	Cigarettes per Day	.10	.07	1.5×10^{-1}
Age of Initiation of Smoking	Pack Years	−.63	.17	2.1×10^{-4}
Age of Initiation of Smoking	Cigarettes per Day	−.26	.16	9.9×10^{-2}
Pack Years	Cigarettes per Day	.77	.13	2.2×10^{-9}
B. Genetic Correlation Induced by Rare (MAF < 1%) Nonsynonymous Variants				
Drinks per Week	Smoking Initiation	.49	.08	1.2×10^{-10}
Drinks per Week	Age of Initiation of Smoking	−.04	.30	8.9×10^{-1}
Drinks per Week	Pack Years	.08	.02	2.7×10^{-4}
Drinks per Week	Cigarettes per Day	.09	.02	5.2×10^{-5}
Smoking Initiation	Age of Initiation of Smoking	−1.10	.21	1.3×10^{-7}
Smoking Initiation	Pack Years	.63	.08	1.5×10^{-14}
Smoking Initiation	Cigarettes per Day	.23	.08	3.3×10^{-3}
Age of Initiation of Smoking	Pack Years	−1.10	.33	1.5×10^{-3}
Age of Initiation of Smoking	Cigarettes per Day	−.69	.32	3.2×10^{-2}
Pack Years	Cigarettes per Day	.87	.14	1.4×10^{-9}