



Controversies in oncology: are genomic tests quantifying homologous recombination repair deficiency (HRD) useful for treatment decision making?

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Mechanisms that protect DNA are crucial to preserve the genome integrity from injuries produced by environmental agents or those spontaneously generated during DNA metabolism.¹ DNA single-strand breaks (SSBs) are repaired by mismatch repair (MMR), base excision repair (BER) and nucleotide excision repair (NER).¹ Non-homologous end joining (NHEJ) or homologous recombination repair (HRR) processes double-strand breaks (DSBs).¹ While NHEJ solves DSBs in a potentially inaccurate way, HRR is an error-free pathway that restores the genomic sequence of the broken DNA ends by using the sister chromatid as template for repair.¹ Several proteins are involved in the HRR pathway: some of them act as sensors of DSBs such as γ H2AX, ATM and ATR, leading to the activation of signal mediator proteins (ie, BRCA1, BRCA2 and PALB2). The final event of the HRR pathway is the loading of a small nuclear protein called RAD51 onto single-stranded DNA, where it promotes strand invasion and replication fork stabilisation (figure 1).² Tumours with HRR deficiency (HRD) were described for the first time in cancers that harbour germline mutations of the tumour suppressors *BRCA1* and *BRCA2* (*BRCA1/2*).³ Nonetheless, genetic and epigenetic events can also result in inactivation of other HRR components, leading to HRD in sporadic cancers.³

HRD is harboured by approximately 13% and 15% of ovarian and triple negative breast cancers (TNBC), respectively, and it is attributable to germline *BRCA1/2* (*gBRCA1/2*) mutations.^{4,5} Furthermore, 50% and 40% of ovarian and TNBC, respectively, are characterised by harbouring HRD in the absence of *gBRCA1/2* mutations.^{4,6} Also, 10%–12% of advanced prostate cancer harbour germline or somatic *BRCA2* inactivation and up to 25% contain a DNA repair defect.⁷ As HRR

is required for the repair of DSBs generated during DNA interstrand cross-link (ICL) resolution, HRR-deficient tumours are sensitive to ICL-generating platinum chemotherapy.^{3,8} Moreover, *BRCA1/2*-mutant cells are sensitive to PARP inhibitors (PARPi), a new class of drugs that block SSB repair, favouring accumulation of DSB that HRR-deficient cells cannot repair.⁹ PARPi also trap PARP onto DNA causing replication stress that is toxic in these cells.⁹ Several PARPi have been approved for the treatment of ovarian and breast cancers.^{10–14} EMA approved olaparib and rucaparib as maintenance treatments for platinum-sensitive ovarian cancer with germline/somatic *BRCA1/2*-mutation; and niraparib was labelled as maintenance treatment for patients who are in response to platinum-based chemotherapy.^{10,12,15} Olaparib and talazoparib have been approved for patients with advanced breast cancer and a *gBRCA1/2* mutations who have previously been treated with chemotherapy.^{13–15} The current clinical challenge is the identification of *BRCA1/2* wild type (WT) patients who harbour alterations in the HRR pathway and share molecular features of *BRCA1/2*-mutated tumours (the so-called 'BRCAness' phenotype) and who may also benefit from similar therapeutic approaches.¹⁶

Different approaches are currently being investigated to identify *BRCA1/2* WT tumours that can benefit from DNA-damaging agents and PARPi based on the presence of HRD, that is, (1) scores capturing large genomic aberrations, so-called 'genomic scars', (2) analysis of mutational signatures or (3) point mutations identified in HRR genes using DNA sequencing panels.^{9,17–19} In *BRCA1/2*-mutant cells, chromosomal spreads reveal increased gross chromosomal rearrangements. This led to the development of assays to evaluate the 'genomic scars' caused by the loss of HRR

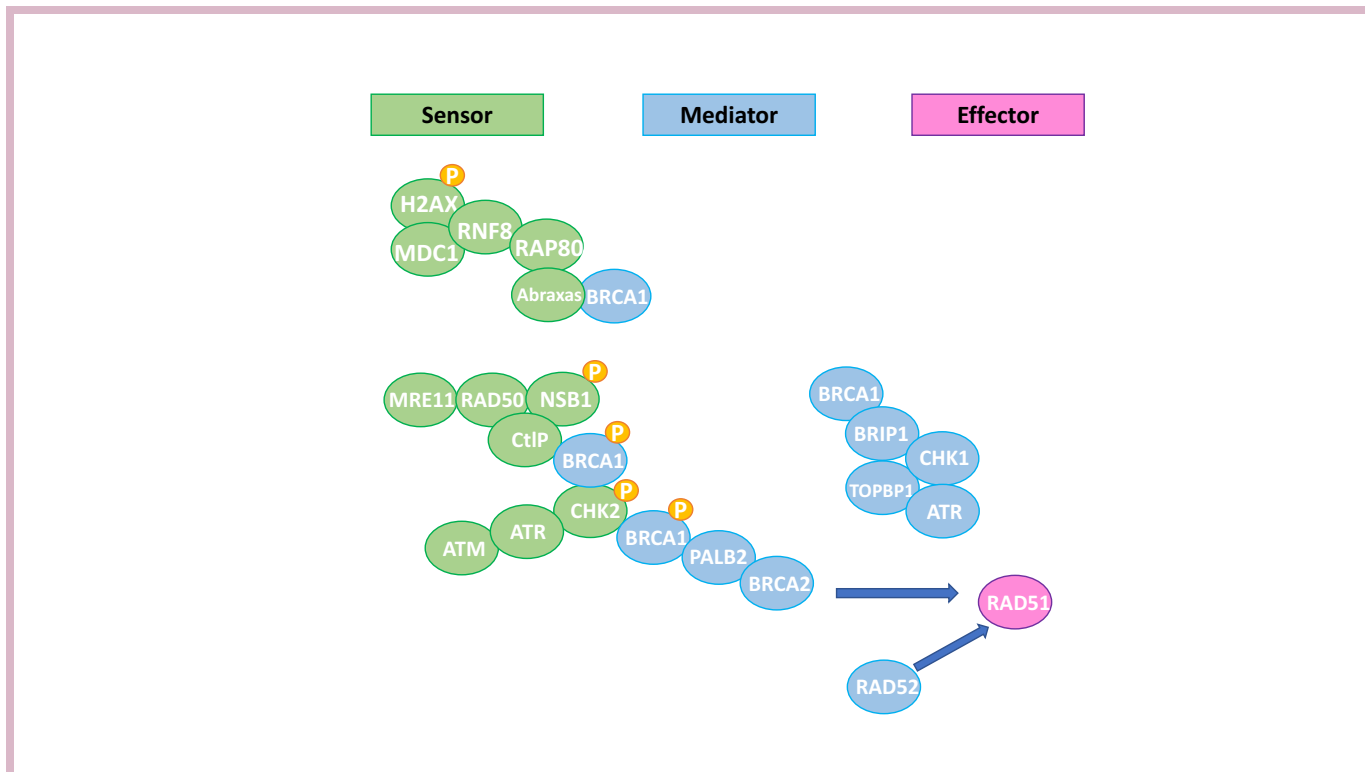


Figure 1 Homologous recombination pathway. Adapted from De Picciotto *et al.*⁵⁰

function using SNP array data.³ Two commercial genomic scar assays have been tested to identify tumours with HRD in clinical trials. The ‘myChoice HRD’ assay by Myriad tests for the presence of loss of heterozygosity (LOH), telomeric allelic imbalance (TAI), and large-scale state transitions (LST) across the genome.¹⁷ The readout of this assay is presented as an ‘HRD score’: a tumour with an HRD score ≥ 42 is labelled as HRD-positive. The ‘FoundationFocus CDx_{BRCA LOH}’ is designed to detect the presence of mutations in the *BRCA1/2* genes and the percentage of the genome affected by LOH in DNA from tumour tissue samples of patients with ovarian cancer.²⁰ According to the FoundationFocus test, tumours are categorised as LOH-high if score is ≥ 16 . On the other hand, mutational signatures are characteristic patterns left on the cancer genome by each mutational process: for example, HRD has been associated with the ‘signature 3’ described by Alexandrov *et al.*^{21–23} ‘Signature 3’ is also able to accurately classify missense *BRCA1/2* mutations with known functional implications and it is associated with silencing of *RAD51C* and *BRCA1* by promoter methylation.²⁴

So, what is the current clinical evidence around the use of genomic scars to quantify HRD and its impact on treatment decision-making? The main open question is whether genomic scars are predictive biomarkers of response to platinum salts or PARPi, beyond *BRCA1/2* mutation.

In advanced ovarian cancer, the ARIEL2 study demonstrated the efficacy of the PARPi rucaparib as monotherapy in *gBRCA1/2* mutated and/or LOH-high relapsed, platinum-sensitive ovarian cancer, and the ARIEL3 trial

demonstrated the benefit of rucaparib as maintenance therapy in platinum-sensitive recurrent patients who responded to platinum, regardless of the LOH status (table 1).^{12–20} The NOVA trial investigated the role of the PARPi niraparib as maintenance therapy in platinum-sensitive ovarian cancer and showed that patients with *BRCA1/2* mutations or HRD-positive according to myChoice assay benefited from PARPi.¹¹ Nevertheless, niraparib also improved PFS in *BRCA1/2* WT patients with an HRD-negative test, although the magnitude of the benefit was smaller compared to *BRCA1/2*-mutated or HRD-positive patients (table 1).¹¹ From these trials, one may conclude that in the platinum-sensitive population the HRD-genomic scars provide information regarding the magnitude of the clinical benefit, given the high probability of response after platinum sensitivity. Indeed, the magnitude of the benefit is higher among those who are HRD-positive or *BRCA1/2*-mutated. Most importantly, in the ovarian population, further investigations are needed to verify if an HRD test may be useful to select platinum-resistant tumours that may benefit from PARPi or to identify long responders to PARPi and/or platinum salts.

In TNBC, several trials have investigated if HRD-genomic scars predict response to DNA-damaging agents or to the addition of carboplatin to standard chemotherapy beyond *BRCA1/2* mutation (table 2). In the neoadjuvant setting, Telli *et al* retrospectively assessed the predictive value of the ‘myChoice HRD’ assay in three single-arm trials testing platinum-based therapy.²⁵ Patients who were HRD-positive had a higher probability to achieve

Table 1 Efficacy of PARPi according to HRD status in ovarian cancer

| Clinical trial | Drug | Study population | HRD role |
|--------------------------|--------------------------------|---|--|
| ARIEL-2 ¹² | Rucaparib, monotherapy | Relapsed, platinum-sensitive ovarian cancer | Higher efficacy in <i>gBRCA1/2</i> -mutated and/or LOH-high compared with LOH-low tumours. Not powered to show a difference between LOH-high and LOH-low tumours |
| ARIEL-3 ²⁰ | Rucaparib, maintenance therapy | Relapsed, platinum-sensitive ovarian cancer | Efficacy regardless of LOH-status. Magnitude of the benefit dependent on LOH |
| NOVA-trial ¹¹ | Niraparib, maintenance therapy | Relapsed, platinum-sensitive ovarian cancer | Efficacy regardless of HRD-status. Magnitude of the benefit dependent on HRD |

HRD, homologous recombination repair deficiency; LOH, loss of heterozygosity; PARPi, PARP inhibitors.

a complete pathological response or minimal residual disease (RCB 0-I) after platinum chemotherapy, even among *BRCA1/2* WT tumours.²⁵ The GeparSixto trial evaluated the benefit of the addition of carboplatin to anthracycline/taxane-based neoadjuvant chemotherapy in TNBC and analysed the predictive and prognostic value of testing for HRD by the composite biomarker including germline/somatic *BRCA1/2* mutations and the ‘myChoice’ assay.⁶ Among all patients with TNBC, addition of carboplatin resulted in a marked increment in pCR rates in HRD-positive tumours (from 33.9 to 63.5%, $P=0.001$), and in HRD-negative tumours (from 20 to 29.6%, $P=0.399$). However, according to the test of interaction, HRD did not predict for carboplatin benefit in this study. Of note, the control arm in this trial lacked cyclophosphamide, which might have overestimated the carboplatin benefit. Other observations regarding the prognostic value of HRD genomic scars will require a powered study to demonstrate the improvement in disease-free survival (DFS) or overall survival (OS). Litton *et al* have recently showed the efficacy of PARPi talazoparib in the neoadjuvant setting in patients with *BRCA1/2* mutations. In this setting, an HRD test could be useful to identify patients with *BRCA1/2* WT who can also benefit from PARPi.²⁶ Finally, in the adjuvant setting, Sharma *et al*

evaluated the predictive role of the ‘myChoice HRD’ in TNBC to predict outcome of adjuvant anthracycline and cyclophosphamide regimen.²⁷ The study showed a better DFS in patients with high HRD, even beyond *gBRCA1/2* status.

In metastatic TNBC, Isakoff *et al* conducted a phase II trial aimed to investigate the predictive role of genomic scars to platinum salts. Higher HRD scores were reported in responding patients, independent of *BRCA1/2* mutational status.²⁸ However, the predictive role of this HRD test was not confirmed in the TNT trial, a randomised phase III trial comparing the efficacy of first-line carboplatin versus docetaxel in patients with advanced TNBC.²⁹ According to the preplanned biomarker analysis, carboplatin resulted in higher overall response rates (ORR) among patients harbouring a *gBRCA1/2* mutation, but not in subjects with other profiles associated with HRR dysfunction such as high HRD-score, *BRCA1* methylation, or *BRCA1* mRNA-low, mostly evaluated in the primary tumours.²⁹ These results could be partially explained by the fact that genomic scars tested in the primary tumour may have lower prediction power for response in the advanced setting because metastatic tumours may have restored the HRR function and become resistant to platinum. As in the GeparSixto trial, HRD-positive tumours

Table 2 Efficacy of platinum or DNA-damaging chemotherapy according to HRD status in breast cancer

| Clinical trial | Drug | Study population | HRD role |
|---|----------------------------------|-------------------------------------|--|
| PrECOG 0105 Cisplatin-1 trial Cisplatin-2 trial ⁵¹ | Platinum salts | Neoadjuvant TNBC | Patients who were HRD-positive had higher complete pathological response |
| Gepar-Sixto trial ⁶ | Carboplatin | Neoadjuvant TNBC | Patients who were HRD-positive had a better prognosis compared with HRD-negative. No robust conclusions regarding the predictive role of HRD for addition of carboplatin |
| SWOG S9313 trial ²⁷ | Doxorubicin and cyclophosphamide | Adjuvant TNBC | Patients who were HRD-positive had a better DFS, even beyond <i>gBRCA1/2</i> status |
| TBCRC009 trial ²⁸ | Platinum salts | Advanced, first or second line TNBC | Higher HRD scores were reported in responding patients, independent of <i>BRCA1/2</i> mutational status. |
| TNT trial ²⁹ | Carboplatin | Advanced, first line TNBC | ORR did not correlate with HRD-score of the primary tumours. |

HRD, homologous recombination repair deficiency; ORR, overall response rate; TNBC, triple negative breast cancers.

were more likely to respond to any chemotherapy regimens compared with the HRD-negative ones. Several open questions may raise from the previous statements: first, that no data are available comparing the HRD status in early and advanced breast cancer, and second, that further studies are required to dissect the role of recovering the HRR function in predicting resistance to PARPi and platinum salts.³⁰ Furthermore, despite the OlympiAD and EMBRACA trials demonstrated the efficacy of PARPi in *BRCA1/2*-mutated metastatic breast cancer,^{13 14} still a relevant proportion of patients did not respond. Further research is needed to investigate if an HRD test would help to refine the subgroup more likely to benefit.

In prostate cancer, the interest in developing DNA damaging and PARPi-based therapeutic strategies arises from the enrichment for DNA damage response gene mutations among cases with advanced disease. The TOPARP-A trial evaluated the antitumor activity of olaparib in advanced prostate cancer, identifying a strong association between the presence of certain DNA repair defects and response to olaparib.³¹ Preliminary data from the TRITON2 study, evaluating rucaparib, confirmed the high response rate to PARPi of *BRCA1/2*-deficient prostate cancers.³² Nevertheless, the predictive value of other defects in DNA repair genes such as *ATM*, *FANCA* or *CHEK2* remains yet to be validated in randomised studies. None of the HRD score tests has been validated yet in prostate cancer clinical trials. Of note, in two recent studies, the prevalence of LOH-high signatures, based on the FoundationOne assay, among *BRCA2*-mutated prostate cancer was lower than for the *BRCA1/2*-mutated ovarian cancer setting.^{33 34}

A current limitation of the genomic scar assays is the impossibility to capture tumour evolution processes, such as a restoration of the HRR function in response to therapy-selective pressure. As an alternative, it could be useful to incorporate functional biomarkers based on dynamic assays that assess the activity of a repair pathway. A crucial step of HRR is mediated by the RAD51 protein.¹ In vivo and in vitro studies supported the highly sensitive and specific predictive power of lack of nuclear RAD51 foci to PARPi response.^{30 35–37} One limitation is that the RAD51 assay may fail to identify *ATM*-mutated tumours that can benefit from PARPi.^{31 38–40} There are several other tests currently used in research to identify tumours with a similar biological behaviour as the *BRCA1/2*-mutated ones. Those tests take into account not only the copy number changes but also methylation or gene expression profiles.^{41–43} Even if preclinical and retrospective data suggest that these tests are predictive of high dose alkylating chemotherapy and PARPi response, the lack of prospective validation and concerns regarding their large-scale feasibility seem to be major issues for their clinical application.^{43–49}

We can summarise that genomic scars associated with HRR defects in ovarian cancer identify patients who obtain maximum benefit from PARPi maintenance after platinum response. In early breast cancer, HRD-genomic

scars have shown a high correlation to DNA-damaging chemotherapy response, and a definitive word to predict the benefit of adding carboplatin is warranted. In prostate cancer, recent data suggest that HRR gene-mutated tumours, including *gBRCA2*, are sensitive to PARPi. Nonetheless, it is of concern that HRD is observed less-frequently than in ovarian cancers.

As future perspectives, research is needed to confirm if clinical implementation of HRD tests might be useful to identify patients with platinum-resistant ovarian cancer who may benefit from PARPi. Likewise, some *BRCA1/2* WT, HRD-positive breast cancers could respond to PARPi. Also, the prognostic role of HRD should be further investigated with *ad hoc* trials in order to recognise patients with early breast cancer candidates for a targeted strategy. Prospective comparison between HRD-genomic scars and functional dynamic tests such as the RAD51 assay is encouraged.

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REFERENCES

- Roy R, Chun J, Powell SN. *BRCA1* and *BRCA2*: important differences with common interests. *Nat Rev Cancer* 2012;12:372.
- Murai J, Huang S-yinN, Das BB, *et al*. Trapping of PARP1 and PARP2 by clinical PARP inhibitors. *Cancer Res* 2012;72:5588–99.
- Hoppe MM, Sundar R, Tan DSP, *et al*. Biomarkers for homologous recombination deficiency in cancer. *J Natl Cancer Inst* 2018;110:704–13.
- Bell D, Berchuck A, Birrer M, *et al*. Integrated genomic analyses of ovarian carcinoma. *Nature* 2011;474:609–15.
- Akashi-Tanaka S, Watanabe C, Takamaru T, *et al*. BRCAness predicts resistance to taxane-containing regimens in triple negative breast cancer during neoadjuvant chemotherapy. *Clin Breast Cancer* 2015;15:80–5.
- Loibl S, Weber KE, Timms KM, *et al*. Survival analysis of carboplatin added to an anthracycline/taxane-based neoadjuvant chemotherapy and HRD score as predictor of response-final results from GeparSixto. *Ann Oncol* 2018;29:2341–7.
- Robinson D, Van Allen EM, Wu Y-M, *et al*. Integrative clinical genomics of advanced prostate cancer. *Cell* 2015;161:1215–28.
- Heeke AL, Baker T, Lynce F, *et al*. Prevalence of homologous recombination deficiency among all tumor types. *JCO* 2017;35(15_suppl):1502.
- O'Connor MJ. Targeting the DNA damage response in cancer. *Mol Cell* 2015;60:547–60.

10. Ledermann JA, Harter P, Gourley C, *et al.* Overall survival in patients with platinum-sensitive recurrent serous ovarian cancer receiving olaparib maintenance monotherapy: an updated analysis from a randomised, placebo-controlled, double-blind, phase 2 trial. *Lancet Oncol* 2016;17:1579–89.
11. Mirza MR, Monk BJ, Herrstedt J, *et al.* Niraparib maintenance therapy in platinum-sensitive, recurrent ovarian cancer. *N Engl J Med* 2016;375:2154–64.
12. Swisher EM, Lin KK, Oza AM, *et al.* Rucaparib in relapsed, platinum-sensitive high-grade ovarian carcinoma (ARIEL2 Part 1): an international, multicentre, open-label, phase 2 trial. *Lancet Oncol* 2017;18:75–87.
13. Robson M, Im S-A, Senkus E, *et al.* Olaparib for Metastatic Breast Cancer in Patients with a Germline *BRCA* Mutation. *N Engl J Med* 2017;377:523–33.
14. Litton JK, Rugo HS, Ettl J, *et al.* Talazoparib in Patients with Advanced Breast Cancer and a Germline *BRCA* Mutation. *N Engl J Med* 2018;379:753–63.
15. European Medicines Agency. Available: <https://www.ema.europa.eu/medicines/human/>
16. Lord CJ, Ashworth A. BRCAness revisited. *Nat Rev Cancer* 2016;16:110–20.
17. Watkins JA, Irshad S, Grigoriadis A, *et al.* Genomic scars as biomarkers of homologous recombination deficiency and drug response in breast and ovarian cancers. *Breast Cancer Res* 2014;16.
18. Hodgson DR, Dougherty B, Lai Z, *et al.* 435 candidate biomarkers of PARP inhibitor sensitivity in ovarian cancer beyond the *BRCA* genes. *European Journal of Cancer* 2015;51.
19. Abkevich V, Timms KM, Hennessy BT, *et al.* Patterns of genomic loss of heterozygosity predict homologous recombination repair defects in epithelial ovarian cancer. *Br J Cancer* 2012;107:1776–82.
20. Coleman RL, Oza AM, Lorusso D, *et al.* Rucaparib maintenance treatment for recurrent ovarian carcinoma after response to platinum therapy (ARIEL3): a randomised, double-blind, placebo-controlled, phase 3 trial. *The Lancet* 2017;390:1949–61.
21. Helleday T, Eshtad S, Nik-Zainal S. Mechanisms underlying mutational signatures in human cancers. *Nat Rev Genet* 2014;15:585–98.
22. Alexandrov LB, Nik-Zainal S, Wedge DC, *et al.* Signatures of mutational processes in human cancer. *Nature* 2013;500:415–21.
23. Peng G, Chun-Jen Lin C, Mo W, *et al.* Genome-wide transcriptome profiling of homologous recombination DNA repair. *Nat Commun* 2014;5.
24. Polak P, Kim J, Braunstein LZ, *et al.* A mutational signature reveals alterations underlying deficient homologous recombination repair in breast cancer. *Nat Genet* 2017;49:1476–86.
25. Telli ML, Timms KM, Reid J, *et al.* Homologous recombination deficiency (HRD) score predicts response to Platinum-Containing neoadjuvant chemotherapy in patients with triple-negative breast cancer. *Clin Cancer Res* 2016;22:3764–73.
26. Litton JK, Scoggins M, Hess KR, *et al.* Neoadjuvant talazoparib (TALA) for operable breast cancer patients with a *BRCA* mutation (*BRCA+*). *JCO* 2018;36(15_suppl):508.
27. Sharma P, Barlow WE, Godwin AK, *et al.* Impact of homologous recombination deficiency biomarkers on outcomes in patients with triple-negative breast cancer treated with adjuvant doxorubicin and cyclophosphamide (SWOG S9313). *Ann Oncol* 2018;29:654–60.
28. Isakoff SJ, Mayer EL, He L, *et al.* TBCRC009: a multicenter phase II clinical trial of platinum monotherapy with biomarker assessment in metastatic triple-negative breast cancer. *JCO* 2015;33:1902–9.
29. Tutt A, Tovey H, Cheang MCU, *et al.* Carboplatin in *BRCA1/2*-mutated and triple-negative breast cancer *BRCA*ness subgroups: the TnT trial. *Nat Med* 2018;24:628–37.
30. Cruz C, Castroviejo-Bermejo M, Gutiérrez-Enríquez S, *et al.* *RAD51* foci as a functional biomarker of homologous recombination repair and PARP inhibitor resistance in germline *BRCA*-mutated breast cancer. *Ann Oncol* 2018;29:1203–10.
31. Mateo J, Carreira S, Sandhu S, *et al.* DNA-repair defects and olaparib in metastatic prostate cancer. *N Engl J Med Overseas Ed* 2015;373:1697–708.
32. Abida W, Bryce AH, Vogelzang NJ, *et al.* 793PDPreliminary results from TRITON2: a phase II study of rucaparib in patients (PTS) with metastatic castration-resistant prostate cancer (mCRPC) associated with homologous recombination repair (HRR) gene alterations. *Ann Oncol* 2018;29(suppl_8).
33. Chowdhury S, McDermott R, Piulats JM, *et al.* 795PDGenomic profiling of circulating tumour DNA (ctDNA) and tumour tissue for the evaluation of rucaparib in metastatic castration-resistant prostate cancer (mCRPC). *Ann Oncol* 2018;29(suppl_8).
34. Agarwal N, Sokol ES, Lara P, *et al.* 510Pan-cancer assessment of *BRCA1/2* genomic alterations (GAS) by comprehensive genomic profiling (CGP) of tissue and circulating tumor DNA (ctDNA). *Ann Oncol* 2018;29(suppl_8).
35. Castroviejo-Bermejo M, Cruz C, Llop-Guevara A, *et al.* A *Rad51* assay feasible in routine tumor samples calls PARP inhibitor response beyond *BRCA* mutation. *EMBO Mol Med* 2018;10:e9172.
36. Naipal KAT, Verkaik NS, Ameziane N. *Functional ex vivo assay to select homologous recombination – deficient breast tumors for PARP inhibitor treatment functional ex vivo assay to select homologous recombination – de FI cient breast tumors for PARP*, 2014: 4816–26.
37. Graeser M, Mccarthy A, Lord CJ. *A marker of homologous recombination predicts pathologic complete response to neoadjuvant chemotherapy in primary breast cancer*, 2011.
38. Goodall J, Mateo J, Yuan W, *et al.* Circulating cell-free DNA to guide prostate cancer treatment with PARP inhibition. *Cancer Discov* 2017;7:1006–17.
39. Bakr A, Oing C, Köcher S, *et al.* Involvement of ATM in homologous recombination after end resection and *Rad51* nucleofilament formation. *Nucleic Acids Res* 2015;43:3154–66.
40. McCabe N, Turner NC, Lord CJ, *et al.* Deficiency in the repair of DNA damage by homologous recombination and sensitivity to poly(ADP-ribose) polymerase inhibition. *Cancer Res* 2006;66:8109–15.
41. Lips EH, Laddach N, Savola SP, *et al.* Quantitative copy number analysis by multiplex ligation-dependent probe amplification (MLPA) of *BRCA1*-associated breast cancer regions identifies *BRCA*ness. *Breast Cancer Res* 2011;13.
42. Schouten PC, Grigoriadis A, Kuilman T, *et al.* Robust *BRCA1*-like classification of copy number profiles of samples repeated across different datasets and platforms. *Mol Oncol* 2015;9:1274–86.
43. Konstantinopoulos PA, Spentzos D, Karlan BY, *et al.* Gene Expression Profile of *BRCA* ness that correlates with responsiveness to chemotherapy and with outcome in patients with epithelial ovarian cancer. *JCO* 2010;28:3555–61.
44. Schouten PC, Marmé F, Aulmann S, *et al.* Breast cancers with a *BRCA1*-like DNA copy number profile recur less often than expected after high-dose alkylating chemotherapy. *Clin Cancer Res* 2015;21:763–70.
45. Vollebergh MA, Lips EH, Nederlof PM, *et al.* An aCGH classifier derived from *BRCA1*-mutated breast cancer and benefit of high-dose platinum-based chemotherapy in *HER2*-negative breast cancer patients. *Ann Oncol* 2011;22:1561–70.
46. Lehmann BD, Bauer JA, Chen X, *et al.* Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. *J Clin Invest* 2011;121:2750–67.
47. Masuda H, Baggerly KA, Wang Y, *et al.* Differential response to neoadjuvant chemotherapy among 7 triple-negative breast cancer molecular subtypes. *Clin Cancer Res* 2013;19:5533–40.
48. Severson TM, Peeters J, Majewski I, *et al.* *BRCA1*-like signature in triple negative breast cancer: molecular and clinical characterization reveals subgroups with therapeutic potential. *Mol Oncol* 2015;9:1528–38.
49. Severson TM, Wolf DM, Yau C, *et al.* The *BRCA1*ness signature is associated significantly with response to PARP inhibitor treatment versus control in the I-SPY 2 randomized neoadjuvant setting. *Breast Cancer Res* 2017;19.
50. De Picciotto N, Cacheux W, Roth A, *et al.* Ovarian cancer: Status of homologous recombination pathway as a predictor of drug response. *Crit Rev Oncol Hematol* 2016;101:50–9.
51. Telli ML, Jensen KC, Vinayak S, *et al.* Phase II Study of Gemcitabine, Carboplatin, and Iniparib As Neoadjuvant Therapy for Triple-Negative and *BRCA1/2* Mutation-Associated Breast Cancer With Assessment of a Tumor-Based Measure of Genomic Instability: PrECOG 0105. *JCO* 2015;33:1895–901.