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# **Radionuclide Imaging of Atherothrombotic Diseases**

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# Abstract

**Purpose of Review:** A variety of approaches and molecular targets have emerged in recent years for radionuclide-based imaging of atherosclerosis and vulnerable plaque using single photon emission computed tomography (SPECT) and positron emission tomography (PET), with numerous methods focused on characterizing the mechanisms underlying plaque progression and rupture. This review highlights the ongoing developments in both the preclinical and clinical environment for radionuclide imaging of atherosclerosis and atherothrombosis.

**Recent Findings:** Numerous physiological processes responsible for the evolution of high-risk atherosclerotic plaque, such as inflammation, thrombosis, angiogenesis, and microcalcification, have been shown to be feasible targets for SPECT and PET imaging. For each physiological process, specific molecular markers have been identified that allow for sensitive non-invasive detection and characterization of atherosclerotic plaque.

**Summary:** The capabilities of SPECT and PET imaging continue to evolve for physiological evaluation of atherosclerosis. This review summarizes the latest developments related to radionuclide imaging of atherothrombotic diseases.

#### Keywords

atherosclerosis; atherothrombosis; molecular imaging; SPECT; PET; radionuclide imaging

# Introduction

Cardiovascular disease remains a leading cause of death worldwide, with many adverse cardiovascular events and deaths being attributable to acute myocardial infarction due to coronary vessel atherosclerosis and/or thrombosis, and atherothrombosis-related ischemic stroke [1]. The underlying atherosclerosis responsible for the majority of adverse cardiovascular events is a complex disease that is characterized by formation and

Human and Animal Rights

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Conflict of Interest

Dr. Stacy has no conflicts of interest to declare for this work.

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progression of occlusive vascular plaques that arise due to a variety of factors, which include, but are not limited to, inflammatory mediators, endothelial injury, infiltration of oxidized low-density lipoproteins, calcium deposition, and plaque neoangiogenesis [2,3]. While the atherosclerotic process can lay dormant for months, years, or decades, erosion or damage to the fibrous cap of an atherosclerotic plaque can have severe health consequences, such as the triggering of occlusive thrombosis and subsequent development of tissue ischemia [4]. Atherothrombosis can be initiated through the exposure of thrombogenic components contained within the plaque following fissure or erosion of the plaque's surface, which ultimately facilitates platelet aggregation on the exposed plaque surface and results in subsequent fibrin and thrombus formation [4,5]. In general, stable plaques typically possess small necrotic cores and thick fibrous caps, while rupture prone vulnerable plaques that are susceptible to triggering atherothrombotic events are usually characterized by larger necrotic cores and thinner fibrous caps [6]. Histologically, rupture- and thrombotic-prone plaques are also commonly characterized by extensive macrophage infiltration, neovascularization, remodeling of the extracellular matrix, apoptosis, and smooth muscle cell proliferation [3,7].

The non-invasive diagnosis of thrombotic diseases and vulnerable plaque has traditionally been limited to indirect detection through the discovery of associated blood flow or tissue perfusion deficits. However, considering the various thrombi-specific epitopes (e.g. fibrin, epitopes on activated platelets, activated coagulation factors) that exist as potential targets for molecular imaging, along with the ongoing developments in modern day imaging technologies that now allow for both high-resolution anatomical and high-sensitivity molecular imaging, a number of non-invasive approaches are emerging that are offering hope for improving the detection of vulnerable plaque and assisting with stratification of atrisk patients. Specifically, intravascular imaging modalities such as optical coherence tomography (OCT) and intravascular ultrasound (IVUS) now provide high-resolution imaging of plaque characteristics in patients undergoing catheterization [8], while more traditional techniques such as x-ray computed tomography (CT) and magnetic resonance (MR) imaging can offer non-invasive morphological insight into plaque calcium burden [9] the presence of plaque hemorrhage [10], respectively.

#### Radionuclide Imaging

The radionuclide-based imaging modalities of single photon emission computed tomography (SPECT) and positron emission tomography (PET) have also begun to play a role in the noninvasive evaluation of various molecular processes associated with thrombosis and vulnerable plaque. SPECT and PET are capable of producing three-dimensional (3D) physiological images via detection of gamma rays that are emitted from injected radiopharmaceuticals, with these targeted radiolabeled probes of variable half-lives and energies being utilized for numerous strategies depending on the ultimate goal of the imaging exam. Both SPECT and PET possess superior sensitivity for targeted molecular imaging when compared to other commercially available imaging modalities; however, conventional SPECT and PET scanners also possess relatively poor spatial resolution (SPECT: 5–8 mm<sup>3</sup>; PET: 3–5 mm<sup>3</sup>). Therefore, the majority of modern SPECT and PET scanners now incorporate and are paired with high spatial resolution CT or MR scanners as

part of hybrid systems (i.e. SPECT/CT, PET/CT, PET/MR), which offer the benefits of colocalizing high sensitivity functional imaging and high-resolution anatomical imaging [11].

When comparing the radionuclides available for SPECT and PET imaging, SPECT radionuclides are generally widely available, involve rather simple radiochemistry, and commonly possess relatively long half-lives. Alternatively, PET radionuclides possess characteristics that generally include shorter half-lives and more complex production/ radiochemistry, both qualities that limit the widespread availability of certain PET tracers. With regard to modality comparisons, PET provides numerous advantages over SPECT. Specifically, PET possesses superior sensitivity and spatial resolution when compared to SPECT, and the recent emergence of digital PET scanners is now resulting in further improvements in image resolution down to ultra-high definition at  $1 \times 1 \times 1$  mm<sup>3</sup> voxel sizes [12]. Additionally, PET has established methodology for absolute quantification of radionuclide uptake, while SPECT has traditionally focused on the evaluation of relative radionuclide retention. While PET does possess advantages, SPECT has been, and remains a mainstay in the clinical environment for evaluation of myocardial perfusion due to its lower relative cost and wider availability, and offers potential opportunities for dual isotope targeted molecular imaging that is not always feasible with PET imaging. Additionally, the recent emergence of cadmium zinc telluride solid state detectors for SPECT continues to enhance and expand the translational capabilities of SPECT imaging [13].

The ultimate quantitative assessment of radionuclide imaging in the vasculature remains a topic of debate, but various methods have been evaluated and established for the purposes of SPECT and PET image analysis. Specifically, standardized uptake values (SUVs) and target-to-background ratios (TBRs) have been the traditional methods utilized when quantifying retention of targeted radionuclides in the vasculature. SUVs are calculated based on the radionuclide retention within the region of interest after correcting for the injected radionuclide dose and the patient's body weight. SUVs can then be further analyzed and expressed based on the maximum (SUV<sub>max</sub>) and/or mean (SUV<sub>mean</sub>) count statistics within the vascular region of interest. Alternatively, TBR values, which are commonly used in the analysis of atherosclerotic plaques identified on PET/CT images, are calculated based on acquiring SUVs in both the vessel wall of interest as well as the vessel lumen, so that the vessel wall serves as the "target" and the blood pool serves as the "background" [14].

Regardless of the image analysis techniques utilized, a variety of radionuclides exist and continue to emerge for SPECT- and PET-based molecular imaging of atherothrombosis and vulnerable plaque that are capable of targeting physiological processes such as inflammation, thrombosis, calcium deposition, and angiogenesis [15]. The following sections of this review highlight the recent advancements related to SPECT and PET imaging of atherosclerosis, atherothrombosis, and vulnerable plaque, with particular focus on emerging radionuclides and physiological targets for non-invasive molecular imaging.

#### Radionuclide Imaging of Vascular Inflammation

A physiological process that plays a major role in regulating plaque vulnerability is vascular inflammation. In general, atherosclerosis is promoted by inflammatory responses that occur

following the infiltration, accumulation, and oxidation of low-density lipoproteins (LDL) in the vascular wall. Following LDL oxidation, inflammatory mediators such as T-lymphocytes and macrophages respond to consume LDL and ultimately form foam cells. While this inflammatory response is designed to be protective in nature, macrophage accumulation in the lesion site can ultimately trigger further inflammation and the subsequent creation of a thin fibrous capped atheroma that is susceptible to rupture [10].

Due to the key role that macrophages play in atherosclerotic plaque evolution and vulnerability for rupture, PET imaging of fluorine-18 (<sup>18</sup>F)-fluorodeoxyglucose (FDG), a radiolabeled glucose analog that is retained by metabolically active macrophages, has been one of the most widely used radionuclide-based approaches for assessing atherosclerosis. Clinical research in the last 5 years has demonstrated in a variety of studies that <sup>18</sup>F-FDG PET/CT imaging possesses clinical utility for non-invasively detecting vascular inflammation in the setting of atherosclerosis [16–21]. Recent clinical studies have specifically shown that <sup>18</sup>F-FDG uptake in the ascending aorta possesses prognostic for predicting future cardiovascular events [17] and is associated with high rates of coronary heart disease events [22]. Additionally,<sup>18</sup>F-FDG PET/CT imaging has demonstrated the ability to detect the effectiveness of various statin dosing regimens at reducing plaque inflammation [••16,19]. While <sup>18</sup>F-FDG PET/CT imaging has demonstrated utility for predicting cardiovascular events and tracking therapy in patients, vascular <sup>18</sup>F-FDG uptake has also been shown to be significantly elevated in patients with elevated lipoprotein(a) levels [23] and significantly correlate with circulating levels of myeloperoxidase [18] and total plasma cholesterol [21]. Recent work by Cocker et al. [24] further revealed that <sup>18</sup>F-FDG uptake specifically represents a non-invasive surrogate marker of macrophage and leukocyte infiltration by demonstrating that <sup>18</sup>F-FDG uptake in carotid artery endarterectomy specimens significantly correlated with both CD68 macrophage expression and CD45 leukocyte expression. Along with prior studies that have utilized <sup>18</sup>F-FDG, additional work by Tahara and colleagues [25] has applied a similar macrophage targeted approach by utilizing <sup>18</sup>F-labeled mannose (2-deoxy-2-[<sup>18</sup>F]fluoro-D-mannose; <sup>18</sup>F- FDM), an isomer of glucose, which revealed similar uptake patterns to  ${}^{18}$ F-FDG in the evaluation of atherosclerotic plaques and also showed uptake patterns that were proportional to plaque macrophage infiltration. Recently, various other approaches, such as gallium-68 (<sup>68</sup>Ga)-DOTATATE, a somatostatin receptor subtype-2 targeted PET probe [•26], and zirconium-89 (<sup>89</sup>Zr)-labeled dextran nanoparticles [27], have demonstrated potential for molecular imaging of plaque inflammation and have shown uptake patterns consistent with the extent of macrophage infiltration, thus revealing that various radionuclide techniques may ultimately be feasible and clinically available for targeted imaging of macrophage-mediated plaque inflammation.

Although PET-based assessment of macrophage activity has composed the majority of inflammation targeted imaging investigations, other techniques have emerged in recent years that have focused on various other mediators of inflammation. For example, molecules such as P-selectin and vascular cell adhesion molecule-1 (VCAM-1), which play important roles in the recruitment and adhesion of key inflammatory cells, have served as imaging targets for investigating plaque inflammation in pre-clinical models of atherosclerosis [28–30]. Specifically, Li et al. [28] utilized <sup>68</sup>Ga-Fucoidan for PET/MR imaging of P-selectin

expression in apolipoprotein E-deficient mice and demonstrated increased radionuclide uptake in plaque with thin fibrous caps and a high density of foam cells, while other research teams have utilized both SPECT- [29] and PET-based [30] approaches to target VCAM-1 and have had success using similar mouse and rabbit models of atherosclerosis. In addition to imaging of P-selectin and VCAM-1, less studied techniques have also included the use of SPECT/CT imaging of technetium-99m (<sup>99m</sup>Tc)-labeled peripheral blood mononuclear cells for evaluating the consequences of circulating monocytes on plaque inflammation [23], as well as <sup>68</sup>Gapentixafor PET/CT imaging for assessing CXC-motif chemokine receptor 4 (CXCR4), with the latter approach demonstrating a significant association with various cardiovascular risk factors and showing increased radionuclide uptake in regions of calcified atherosclerotic lesions (Figure 1) [31].

#### **Radionuclide Imaging of Thrombosis**

Along with targeted imaging of plaque inflammation, recent research has also focused on non-invasive imaging of various molecular markers associated with thrombosis. The end product of the coagulation cascade, fibrin, has served as one of the most widely investigated imaging markers of thrombus formation, with fibrin being non-invasively detected in preclinical models of thrombosis using radiolabeled probes targeting various fibrin binding peptides (FBPs). For example, Ciesienski and colleagues [32] conjugated three different FBPs (FBP1, FBP2, FBP3) as 1,4.7,10-tetraazacyclododecane-1,4.7,10, tetraacetic acid (DOTA)- monoamides for labeling with copper-64 (<sup>64</sup>Cu) and demonstrated with hybrid PET/MR imaging that uptake of FBP1 and FBP2 probes was elevated approximately 4-fold above control levels when evaluating a rat model of carotid artery thrombosis. Additional work by Ay et al. [••33] utilized a <sup>64</sup>Cu-labeled FB7 probe for the *in vivo* evaluation of a rat model of embolic stroke that underwent thrombolysis with recombinant tissue-type plasminogen activator (rtPA), which revealed the ability of <sup>64</sup>Cu-FB7 PET/CT imaging to non-invasively detect response to treatment. Similarly, Blasi and colleagues [34] utilized a <sup>64</sup>Cu-labeled FBP8 probe for PET/CT imaging and revealed the ability of <sup>64</sup>Cu-FBP8 to detect blood clots of varying ages (i.e. 1, 3, or 7 days after induction of thrombosis) in a rat model, with imaging findings confirmed by both histology and autoradiography. Although <sup>64</sup>Cu-labeled FBPs have proven to be effective for PET imaging of fibrin, recent work by Oliveira et al. [35] has also demonstrated that FBP targeted probes such as <sup>68</sup>Ga-labeled FBP14 and indium-111 (<sup>111</sup>In)-FBP15 possess high affinity for fibrin and good specificity for thrombus detection, thereby representing potential radionuclides that could be utilized for PET- or SPECT-based imaging of thrombus formation.

While fibrin targeted imaging has represented the majority of thrombosis focused investigations, factor XIII (FXIII), an enzyme involved in the coagulation cascade and crosslinking of fibrin, has also served as a molecular probe for thrombosis research. Specifically, Chen and colleagues [36] demonstrated that an <sup>111</sup>In-labeled peptide targeting FXIII was capable of detecting elevated activation of the coagulation cascade in a mouse model of silent brain ischemia by using SPECT/CT imaging, and showed further utility of <sup>111</sup>In-FXIII SPECT/CT imaging through the non-invasive tracking of therapy-induced reductions in FXIII activity. Additionally, recent work by Zhuang et al. [37] has utilized a

<sup>99m</sup>Tc-labeled FXIII targeted probe for SPECT/CT imaging and shown sensitivity for detecting the formation of cross-linked fibrin in the microcirculation of the mouse heart.

In addition to fibrin and FXIII imaging, SPECT and PET research has also been conducted in the pre-clinical setting to non-invasively evaluate activated platelets as potential biomarkers of atherothrombosis. In an attempt to image activated platelets, previous research in mouse models of carotid artery thrombosis has focused on radionuclides that selectively targeted the ligand induced binding sites (LIBS) epitope of the receptor site for glycoprotein IIb/IIIa that resides on the surface of platelets [38,39]. Specific work by Heidt and colleagues [38] have demonstrated that <sup>111</sup>In-labeled LIBS specifically binds to activated platelets and can be utilized for SPECT/CT imaging of carotid artery thrombosis, while Ardipradja and colleagues [39] have more recently shown the feasibility of utilizing an <sup>18</sup>Flabeled LIBS probe, <sup>18</sup>F-scFv<sub>anti-LIBS</sub>, for PET/CT imaging of carotid artery thrombosis in a mouse model.

#### Radionuclide Imaging of Plaque Angiogenesis

Angiogenesis, which is initiated and promoted when plaques develop thick necrotic and hypoxic cores, remains another important physiological process that can contribute to both plaque growth and vulnerability to rupture [40]. Therefore, molecular targets, such as the  $\alpha v\beta 3$  integrin and vascular endothelial growth factor (VEGF), have emerged in recent years for SPECT- and PET-based molecular imaging of angiogenesis in high-risk plaques [41-44]. For example, Yoo and colleagues [•41] developed the αvβ3 integrin targeted probe, <sup>99m</sup>Tc-IDA-D-[c(RGDfk)]<sub>2</sub>, for the application of SPECT/CT imaging in a mouse model of atherosclerosis and demonstrated significant elevations in radionuclide uptake within atherosclerotic aortas, with confirmatory histological analysis that revealed probe uptake colocalized to plaques possessing large necrotic cores and thin fibrous caps. Additional preclinical work by Jiang and colleagues [42] utilized a PET probe, <sup>64</sup>Cu-NOTA-3-4A, in a mouse model of atherosclerosis to image  $\alpha v\beta 3$  integrin expression and showed significant elevations in radionuclide uptake within atherosclerotic lesions of the carotid arteries, as well as specificity of the tracer for targeting plaque (Figure 2). Along with research in preclinical models of atherosclerosis, Golestanie et al. [43] have also evaluated the potential clinical translatability of a  ${}^{18}$ F-RGD probe for assessing  $\alpha v\beta 3$  integrin expression by incubating human carotid endarterectomy specimens in the radionuclide and then applying microPET imaging, which demonstrated significant correlations between radionuclide uptake and integrin staining score, as well as radionuclide uptake and CD31 staining score. While various radionuclides have proven to be effective for imaging of  $\alpha v\beta 3$  integrin expression, more recent work by Tekabe et al. [44] has applied a <sup>99m</sup>Tc-labeled probe for SPECT/CT targeted imaging of VEGF receptor-1 (VEGFR-1) and -2 (VEGFR-2) in apolipoprotein E deficient mice and shown significant increases specifically in VEGFR-1 signaling within atherosclerotic aortas and brachiocephalic vessels, thus revealing a novel target for imaging of intraplaque angiogenesis that may assist in guiding future research in this field.

#### Radionuclide Imaging of Plaque Microcalcification

Generally regarded as a natural healing response to extensive inflammation within plaque, calcification of atherosclerotic lesions has been traditionally characterized as latestage macrocalcification, where plaque has reached a phase of stabilization, or early-stage microcalcification, where plaque remains in an unstable and vulnerable state due to disruption of the fibrous cap and/or ongoing inflammation in the region of the plaque surrounding the calcium deposits [10]. The radionuclide primarily utilized for investigating active microcalcification in atherosclerotic plaques has been <sup>18</sup>F-sodium fluoride (NaF) due to its high affinity and specificity for calcified deposits [45••]. The mechanism of uptake for  $^{18}$ F-NaF is based on NaF replacing the hydroxyl groups of hydroxyapatite, which has been shown to be present in areas of active microcalcification [46]. Early research with <sup>18</sup>F-NaF PET/CT imaging in the setting of carotid artery atherosclerosis demonstrated that <sup>18</sup>F-NaF uptake co-localized with regions of arterial calcification (Figure 3), was more prevalent in patients at high-risk for atherothrombotic events, and significantly correlated with the presence of multiple cardiovascular risk factors, such as diabetes, history of smoking, and prior history of cardiovascular events [47•]. Additional work by other research teams has similarly revealed that patients with high coronary artery <sup>18</sup>F-NaF uptake also had a higher prevalence of arterial calcification and history of myocardial infarction or unstable angina [48], as well as higher rates of prior cardiovascular events and higher Framingham risk scores [49]. While *in vivo* clinical imaging studies have shown a close link between <sup>18</sup>F-NaF uptake and cardiovascular history/risk, more recent research has also focused on histological validation of <sup>18</sup>F-NaF uptake to better understand the mechanisms associated with <sup>18</sup>F-NaF retention in high-risk plaques. Specifically, recent work by Joshi et al. [50] has demonstrated that <sup>18</sup>F-NaF uptake in carotid artery endarterectomy specimens is associated with histological evidence of active calcification, macrophage infiltration, apoptosis, and necrosis, while additional research by Irkle et al. [45••] has shown that <sup>18</sup>F-NaF can differentiate between regions of macro- and microcalcification in atherosclerotic plaque. Furthermore, Cocker and colleagues [51] have recently revealed in excised human carotid artery plaques that <sup>18</sup>F-NaF uptake significantly correlates with Goldner's trichrome staining, a method for histologically detecting the presence of hydroxyapatite, thus demonstrating that <sup>18</sup>F-NaF is related to the process of active microcalcification in plaque and may serve as a useful noninvasive biomarker for characterizing high-risk plaque that is vulnerable to rupture.

# **Emerging Targets for Radionuclide Imaging of Atherothrombosis**

While a variety of radionuclide-based approaches have been applied in the setting of atherosclerosis, novel methods for targeted imaging of vulnerable plaque are continuing to emerge. One recent technique that has recently emerged is molecular imaging of oxidized LDL (oxLDL), which is a hallmark signal representing proinflammatory conditions within plaque. During the process of LDL oxidation, various byproducts are generated that have proinflammatory chemical modifications of lipid and protein moieties that are known as oxidation-specific epitopes (OSEs), which could signal dangerously high levels of inflammation in plaque [52]. Therefore, Senders et al. [53] investigated the utility of a novel PET probe, <sup>89</sup>Zr-LA25, that targeted malondialdehyde-acetaldehyde epitopes and found that <sup>89</sup>Zr-LA25 specifically co-localized to advanced and ruptured atherosclerotic plaques that

had accompanying thrombi. Recent work by Sasaki and colleagues [54] has further demonstrated the potential of molecular imaging of oxLDL in atherosclerosis by developing a PET targeted probe, <sup>64</sup>Cu-3H3-scFv, targeting the B2-glycoprotein/oxLDL complex, with the probe showing co-localization to lipid deposits in atherosclerotic lesions in a rabbit model of hyperlipidemia.

Along with molecular imaging of oxLDL, imaging of hypoxia has also emerged as a potential target for assessing plaque stability. Recent investigations focused on molecular imaging of plaque hypoxia have revealed that uptake of a previously developed radionuclide, <sup>18</sup>F-fluoromisonidazole (FMISO), increases in macrophage rich hypoxic regions of plaque in a rabbit model of atherosclerosis [•55]. Additionally, a novel <sup>18</sup>F-labeled probe, <sup>18</sup>F-2-(4-((2-nitro- 1H-imidazol-1-yl)methyl)-1H-1,2,3-triazol-1-yl)propan-1-ol (HX4), has recently demonstrated potential for the non-invasive detection of plaque hypoxia in patients with carotid artery stenosis, where a combination of PET, CT, and MR images showed that <sup>18</sup>F-HX4 uptake was specific to regions of atherosclerotic plaque, correlated with <sup>18</sup>F-FDG uptake (macrophage infiltration) in plaque, and correlated with MR-derived measures of carotid artery wall dimensions [56].

#### Conclusions

Novel approaches continue to emerge for radionuclide imaging of atherosclerosis and atherothrombosis, with a variety of radiolabeled probes already available in both the preclinical and clinical environment for SPECT and PET imaging of vulnerable plaque (Table 1). Although numerous physiologic targets have been identified and non-invasively evaluated as potential biomarkers associated with high-risk plaque, the debate continues as to which molecular target provides the most relevant and significant prognostic value for general and clinical populations. Additionally, validation studies that confirm the specificity of radionuclides for their molecular target-of-interest is still warranted before many of these probes can become part of routine clinical use. Regardless of the technique utilized, the high sensitivity imaging characteristics of SPECT and PET should continue to provide unique opportunities for radionuclide-based investigations and offer novel tools for non-invasive detection of, and physiological insight into, atherosclerosis.

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study was one of the first to demonstrate the feasibility of hypoxia targeted imaging in atherosclerotic plaque.

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#### Figure 1.

<sup>68</sup>Ga-pentixafor PET/CT imaging of abdominal aorta inflammation. Transaxial views of (A) CT and (B) fused PET/CT images reveal localization of <sup>68</sup>Ga-pentixafor uptake to regions of partial vascular calcification (denoted by white arrows). Sagittal views of (C) CT and (D) fused PET/CT images demonstrate <sup>68</sup>Ga-pentixafor uptake in partially calcified and non-calcified atherosclerotic lesions (dashed arrows). (This research was originally published in JNM. Weiberg, Thackeray, Daum, et al. Clinical Molecular Imaging of Chemokine Receptor CXCR4 Expression in Atherosclerotic Plaque Using <sup>68</sup>Ga-Pentixafor PET: Correlation with Cardiovascular Risk Factors and Calcified Plaque Burden. J Nucl Med. 2018;59:266–272. ©SNMMI.)



#### Figure 2.

<sup>64</sup>Cu-NOTA-3–4A PET/CT imaging for detection of integrin expression in a mouse model of carotid artery atherosclerosis. (A) PET, CT, and fused PET/CT imaging 1 hour after injection of <sup>64</sup>Cu-NOTA-3–4A revealed radionuclide uptake in the site of the carotid artery atherosclerotic lesion. (B) Coronal PET images of carotid artery plaques at 1, 2, 4, and 24 hours after injection of <sup>64</sup>Cu-NOTA-3–4A without or with c(RGDyK), a blocking agent, which demonstrated suppression of <sup>64</sup>Cu-NOTA-3–4A uptake. (C) Quantitative analysis of radionuclide retention in organs at various time points following injection of <sup>64</sup>Cu-NOTA-3– 4A, expressed as %ID/g. (D) Quantitative analysis of plaque–to–normal tissue ratios at various time points. %ID/g = percent injected dose per gram; B = blood; H = heart; K = kidney; Li = liver; M = muscle; N = normal vessel wall; P = plaque. (This research was originally published in JNM. Jiang, Tu, Kimura, et al. <sup>64</sup>Cu-Labeled Divalent Cystine Knot

Peptide for Imaging Carotid Atherosclerotic Plaques. J Nucl Med. 2015;56:939–944. ©SNMMI.)



#### Figure 3.

*In vivo* <sup>18</sup>F-NaF PET/CT imaging of the common carotid arteries. Transaxial views of (A) CT, (B) <sup>18</sup>F-NaF PET, and (C) fused PET/CT images demonstrate bilateral uptake of <sup>18</sup>F-NaF in common carotid artery plaques that is localized to sites of vascular calcification. (This research was originally published in JNM. Derlin, Wisotzki, Richter, et al. In Vivo Imaging of Mineral Deposition in Carotid Plaque Using <sup>18</sup>F-Sodium Fluoride PET/CT: Correlation with Atherogenic Risk Factors. J Nucl Med. 2011;52:362–368. ©SNMMI.)

#### Table 1

Radionuclides for Non-Invasive Imaging of Atherothrombosis and Plaque Vulnerability

Physiologic Target	Radionuclide	Imaging Modality	Reference
Inflammation	<sup>68</sup> Ga-DOTATATE	PET/CT	[26]
	68Ga-fucoidan	PET/MR	[28]
	68Ga-pentixafor	PET/CT	[31]
	<sup>99m</sup> Tc-mononuclear cells	SPECT/CT	[23]
	99mTc-scFv-VCAM1	SPECT/CT	[29]
	<sup>18</sup> F-FB-anti-VCAM-1 nanobody	PET/CT	[30]
	<sup>89</sup> Zr-dextran nanoparticle	PET/MR	[27]
	<sup>18</sup> F-FDG	PET/CT	[16-22,24]
	<sup>18</sup> F-FDM	PET	[25]
Thrombosis	<sup>64</sup> Cu-FBP1, -FBP2, -FBP3	PET/MR	[32]
	<sup>64</sup> Cu-FBP7	PET/CT	[33]
	<sup>64</sup> Cu-FBP8	PET/CT	[34]
	<sup>68</sup> Ga-FBP14	PET/CT	[35]
	<sup>111</sup> ln-FBP15	SPECT/CT	[35]
	99mTc-FBP16	SPECT/CT	[35]
	<sup>111</sup> ln-FXIII	SPECT/CT	[36]
	<sup>99m</sup> Tc-FXIII	SPECT/CT	[37]
	<sup>111</sup> ln-LIBS	SPECT/CT	[38]
	<sup>18</sup> F-SCFV <sub>anti-LIBS</sub>	PET/CT	[39]
Plaque Angiogenesis	<sup>99m</sup> Tc-IDA-D-[c(RGDfK)] <sub>2</sub>	SPECT/CT	[41]
	<sup>64</sup> Cu-NOTA-3–4A	PET/CT	[42]
	<sup>18</sup> F-RGD-K5	PET/CT	[43]
	<sup>99m</sup> Tc-scVR1, <sup>99m</sup> Tc-scVR2	SPECT/CT	[44]
Plaque Microcalcification	<sup>18</sup> F-NaF	PET/CT	[45,47–51]
LDL Oxidation	<sup>64</sup> Cu-3H3-scFv	PET/CT	[54]
	<sup>89</sup> Zr-LA25	PET/MR	[53]
Plaque Hypoxia	<sup>18</sup> F-FMISO	PET/CT	[55]
	<sup>18</sup> F-HX4	PET/CT/MR	[56]