

HHS Public Access

Author manuscript J Alzheimers Dis. Author manuscript; available in PMC 2019 June 16.

Published in final edited form as: J Alzheimers Dis. 2019 ; 68(3): 1161–1170. doi:10.3233/JAD-180788.

An UNC5C Allele Predicts Cognitive Decline and Hippocampal Atrophy in Clinically Normal Older Adults

Hyun-Sik Yanga,b,c,d, **Jasmeer P. Chhatwal**a,c , **Jishu Xu**d, **Charles C. White**d, **Bernard** Hanseeuw^{a,c,e}, Jennifer S. Rabin^{c,f}, Kathryn V. Papp^{a,b,c}, Rachel F. Buckley^{a,b,c,g,h}, Aaron P. **Schultz**a,c,i , **Michael J. Properzi**a, **Jennifer R. Gatchel**c,f,j,k , **Rebecca E. Amariglio**a,b,c , **Nancy J. Donovan** $^{\text{b,c,f,I}}$ **, Elizabeth C. Mormino** $^{\text{a,c,m}}$ **, Trey Hedden^{c,i}, Gad A. Marshall^{a,b,c}, Dorene M. Rentz**a,b,c , **Keith A. Johnson**a,b,c,i , **Philip L. De Jager**d,n, and **Reisa A. Sperling**a,b,c,* aDepartment of Neurology, Massachusetts General Hospital, Boston, MA, USA

bDepartment of Neurology, Center for Alzheimer Research and Treatment, Brigham and Women's Hospital, Boston, MA, USA

^cHarvard Medical School, Boston, MA, USA

^dCell Circuits Program, Broad Institute of MIT and Harvard, Cambridge, MAs, USA

^eDepartment of Neurology, Cliniques Universitaires Saint-Luc, Institute of Neurosciences, Université Catholique de Louvain, Brussels, Belgium

^fDepartment of Psychiatry, Massachusetts General Hospital, Boston, MA, USA

^gFlorey Institutes of Neuroscience and Mental Health, Melbourne, VIC, Australia

hMelbourne School of Psychological Sciences, University of Melbourne, Melbourne, VIC, Australia

ⁱDepartment of Radiology, Massachusetts General Hospital, Boston, MA, USA

^jDivision of Geriatric Psychiatry, McLean Hospital, Belmont, MA, USA

^kGerontology Research Unit, Massachusetts General Hospital, Boston, MA, USA

^lDepartment of Psychiatry, Brigham and Women's Hospital, Boston, MA, USA

^mDepartment of Neurology and Neurological Sciences, Stanford University, Stanford, CA, USA

ⁿDepartment of Neurology, Center for Translational & Computational Neuroimmunology, Columbia University Medical Center, New York, NY, USA

Abstract

Background: The UNC5C rs3846455^G allele has been linked to poor cognitive resilience against age-related neuropathologies, but this association remains to be replicated, and the allele's effect on hippocampal neurodegeneration needs to be examined.

Authors' disclosures are available online ([https://www.j-alz.com/manuscript-disclosures/18-0788r3\)](https://www.j-alz.com/manuscript-disclosures/18-0788r3).

^{*}Correspondence to: Reisa A. Sperling, MD, Brigham and Women's Hospital, 60 Fenwood Road, Boston, MA 02115, USA. Tel.: +1 617 525 8675; Fax: +1 617 264 5295; reisa@bwh.harvard.edu.

Objective: To further validate the association between rs3846455^G and faster cognitive decline, especially among cognitively normal older adults, and to assess whether $rs3846455^{\text{G}}$ predicts accelerated hippocampal volume loss in older adults.

Methods: We assessed participants in the Harvard Aging Brain Study (HABS), a longitudinal cohort study of older adults who were clinically normal at baseline. To avoid bias from population admixture, analyses were limited to participants of European descent with longitudinal neuroimaging data ($n = 174$). Linear mixed effect models were used to examine the effect of rs3846455^G on longitudinal change of the Preclinical Alzheimer Cognitive Composite (PACC) and MRI-measured bilateral hippocampal volume, adjusting for baseline amyloid-β (Aβ) measured by the cortical Pittsburgh Compound B PET distributed volume ratio. We also tested whether hippocampal atrophy mediates the association between $rs3846455^G$ and greater PACC decline through a mediation analysis.

Results: $rs3846455^G$ was associated with greater PACC decline ($\beta = -0.087$ /year, 95% CI −0.169 to −0.005, $p = 0.039$) after controlling for baseline Aβ. Further, rs3846455^G predicted accelerated hippocampal atrophy after controlling for baseline Aβ ($\beta = -57.3$ mm³/year, 95% CI -102.8 to -11.9 , $p = 0.014$). The association between rs3846455^G and greater PACC decline was partially mediated by accelerated hippocampal atrophy (mediated effect (relative scale) = −0.014, 95% CI –0.032 to –6.0 \times 10⁻⁴, $p = 0.039$).

Conclusion: UNC5C rs3846455^G predicts greater cognitive decline and accelerated hippocampal atrophy in clinically normal older adults.

Keywords

Alzheimer's disease; amyloid plaques; cognitive reserve; genetics; hippocampus; neuroimaging

INTRODUCTION

More than half of the heterogeneity in late-life cognitive decline remains unexplained even after accounting for common neuropathologies such as Alzheimer's disease (AD), vascular brain injury, or Lewy body disease [1]. Understanding the genetic architecture of this variable resilience to neuropathologies could enable personalized prognostication and novel therapeutic target identification.

A recent genome-wide association study (GWAS) showed a relationship between two independent common alleles, $UNC5C$ rs3846455^G and $ENC1$ rs766629990^A, and worse cognition after accounting for known neuropathologies [2]. UNC5C rs3846455^G is of a particular interest, given other studies that implicated two other independent variants within the gene in cognitive aging: a rare UNC5C mutation (T835M) increases AD dementia risk by increasing hippocampal susceptibility to pathologic insults such as amyloid-β (Aβ) [3], and a common variant, rs28660566T, is associated with a higher burden of cerebral amyloid angiopathy, an Aβ-related cerebrovascular pathology [4]. UNC5C is abundantly expressed in adult human hippocampus, suggesting the gene's potential role in hippocampus [5].

However, the association of $rs3846455^G$ with vulnerability to late-life cognitive decline should be further examined in an independent cohort, and the relationship between

rs3846455^G and longitudinal hippocampal neurodegeneration in older adults needs to be investigated. This knowledge gap should be addressed, especially in clinically normal older adults for whom neuroprotective treatments are most likely to prevent extensive neurodegeneration [6].

Therefore, we sought to do this in the current study, leveraging the structural and molecular neuroimaging data from the Harvard Aging Brain Study (HABS), a longitudinal cohort study that enrolls clinically normal older adults [7]. We aimed to replicate the previously reported association between rs3846455G and greater cognitive decline [2], adjusting for baseline Aβ burden measured in vivo. In addition, we hypothesized that rs3846455G would predict accelerated hippocampal atrophy, which could potentially account for the cognitive decline associated with rs3846455G.

METHODS

Participants

HABS is a longitudinal observational cohort study of aging and preclinical AD that has enrolled over 300 clinically normal community-dwelling older adults. The inclusion criteria included Clinical Dementia Rating Scale global score of 0, Mini-Mental Status Exam (MMSE) score ≥27 (adjusted for age and education), and performance within the normal range on Logical Memory IIa Delayed Recall (adjusted for education; 9 for 16 or more years of education, 5 for 8 to 15 years of education, 3 for 0 to 7 years of education). Annual attrition rate was between 2% to 7%.Further details about HABS areavailable through previous publications [7].

Standard protocol approvals, registrations, and patient consents

Each participant has signed a written informed consent prior to undergoing any study procedure, and the Partners Healthcare Institutional Review Board has approved the protocols and procedures of HABS.

Genotyping

APOE genotyping was done through a targeted Taqman assay for rs7412 and rs429358, and people with at least one *APOE* ϵ 4 haplotype was grouped as *APOE* ϵ 4 carriers.

Genome-wide genotyping was available for 285 HABS participants. The genotyping was performed in two batches, using the same genotyping platform (Axiom™ Biobank Genotyping Array): the first batch ($n = 189$) was genotyped in 2013, while the second batch $(n = 96)$ was completed in 2016. For quality control, we selected single nucleotide polymorphisms (SNPs) with minor allele frequency (MAF) >0.01 in both batches, and subsequently merged the two batches. We performed additional quality control with the following parameters: sex concordance, genotype missing rate (variant) <0.1, genotype missing rate (sample) <0.05, Hardy-Weinberg equilibrium $p > 10^{-50}$, non-extreme heterogyzosity (inbreeding coefficient estimate (F) between −0.1 and 0.1), and identity by descent (IBD) pi-hat < 0.5. All these steps were conducted with PLINK v1.90b3.32. A total of 262 participants and 326,033 SNPs passed quality control.

To assess the population structure, we combined our data with the HapMap 3 samples [8] and used EIGENSTRAT [9] to derive principal components of genotype covariance matrix. We calculated means and standard deviations for each of the first three principal components from the genotype covariance matrix (EV1–3) in HABS participants who reported their race as White. We classified the participants as of European descent only if each of their EV1–3 values were within three standard deviations from these mean values. We confirmed that the EV1–3 values of the HABS participants we classified as of European descent overlapped with the values from the HapMap 3 participants of European descent. UNC5C rs3846455 genotype was available for a total of 259 participants, and 219 participants among them were of European descent. We limited our analyses to participants of European descent to avoid confounding from population admixture. We used rs3846455G carrier status instead of minor allele count in all analyses, as there was only one participant who was a homozygote for the minor allele rs 3846455^{G} (i.e., genotype GG).

Neuropsychological testing

Cognitive performance was measured annually with a modified version of the Preclinical Alzheimer Cognitive Composite (PACC) [10-12], a cognitive composite averaging z-scores of (1) MMSE total score, (2) the Digit Symbol Substitution Test from the Wechsler Adult Intelligence Scale-Revised, (3) the Logical Memory Delayed Recall, and (4) the combined sum of the Free and Cued Selective Reminding Test Free Recall and Total Recall. The zscore for each test was derived using the baseline mean and standard deviation of the entire HABS cohort [10]. Higher scores indicate better cognitive performance. Our study included only participants with baseline and at least one follow-up on the PACC.

Positron emission tomography

To estimate Aβ burden, 11C Pittsburgh Compound B-positron emission tomography (PiB-PET) scans were obtained at the MGH PET facility at baseline. An 8.5–15 mCi bolus of PiB was injected, and the PiB-PET images were acquired through 60 min dynamic acquisitions. PiB-PET data were processed as previously detailed [12]. Baseline PiB distribution volume ratio (DVR) was calculated from a cortical aggregate region including frontal, lateral parietal, lateral temporal, and retrosplenial cortices using a cerebellar gray matter reference [12]. Baseline PiB DVR was used in our study as a measure of Aβ. PiB-PET data were not partial-volume corrected.

Magnetic resonance imaging

Our study included participants with baseline and at least one follow-up structural brain MRI. There were total 174 participants of European descent with UNC5C rs3846455 genotypes, baseline Aβ, and two or more measurements for both PACC and HV. At baseline (for all $(n = 174)$ subjects included in this study), 18 months from the baseline (for $n = 32$ subjects included in this study), 3 years from the baseline (for $n = 168$ subjects included in this study), and 5 years from the baseline (for $n = 85$ subjects included in this study), structural brain MRIs were obtained on one of two matched Siemens Tim Trio 3T scanners at the Massachusetts General Hospital (MGH) – Athinoula A. Martinos Center for Biomedical Imaging, with a 12-channel phasedarray head coil. Hippocampal volume (HV) was calculated from structural T1-weighted, volumetric magnetization-prepared, rapid

acquisition gradientecho scans (repetition time 6400 ms, echo time 2.8 ms, inversion time 900 ms, flip angle 8°, and resolution $1 \times 1 \times 1.2$ mm) using FreeSurfer v5.1. Summed bilateral HV was used in our analyses, and we included the total intracranial volume (ICV) as a covariate in all HV analyses.

Statistical analysis

All statistical analyses were done with R version 3.3.3, and the threshold for significance was twosided $p \le 0.05$. This study is a hypothesis-driven study that builds on the previous findings related to a SNP, so genome-wide correction for multiple testing was not necessary. We indicated the number of participants included in each analysis in the table legends, if different from $n = 174$ due to missing values.

Baseline demographic characteristics were compared across rs3846455G non-carriers and carriers with two sample *t*-tests (for continuous variables; assuming equal variance) or chisquared tests (for dichotomous variables). Association of rs3846455^G with baseline PACC or HV was tested with linear regression, controlling for covariates that are known predictors of each outcome variable [1, 13]: baseline age, sex, and years of education were adjusted for when PACC was the dependent variable, and baseline age, sex, and ICV were controlled when HV was the dependent variable. In addition, EV1–3 were controlled to further account for a potential confounding due to population stratification. Longitudinal relationships were assessed with the following linear mixed effect models (R "nlme" package), assuming random intercepts and slopes, and using the same set of covariates as the baseline linear models:

> Model 1: PACC or HV ~ rs3846455^G \times time + covariates (+ years of education in PACC mod‐ $els) \times time$ (+ ICV in HV models)

Model 2: HV or PACC \sim rs3846455^G \times time + Baseline $\mathbf{A}\mathbf{\beta} \times \mathbf{time} + \mathbf{covariates}$ (+ years of education in PACC models) \times time (+ ICV in HV models)

All main terms for product terms were also include in the regression analysis. We also performed analyses without covariates to rule out induced correlation from inclusion of covariates.

To evaluate whether the association between $rs3846455^G$ and longitudinal HV change explains the link between rs3846455^G and longitudinal PACC change, we performed a *post*hoc mediation analyses. First, we extracted adjusted random slopes of HV and PACC changes from the following linear mixed effect models:

> $HV \sim (baseline age, sex) \times time + ICV$ (time-varying covariate) covariate) PACC ∼ (baseline age, sex, years of educa‐ tion) \times time

```
Mediator model: HV slope \sim (rs3846455)<sup>G</sup> + baseline
A\beta + EV1-3
Outcome model: PACC slope ∼ HV
slope + rs3846455G+ baseline A\beta + EV1-3
```
Here, $rs3846455^G$ is the binary causal (independent) variable, HV slope is a continuous mediator variable, and PACC slope is the continuous outcome variable. The indirect effect (average causal mediation effect = $ACME$), direct effect (average direct effect = ADE), and total effect, were estimated with the default quasi-Bayesian Monte Carlo method (10,000 simulations) from the R "mediation" package [14, 15].

RESULTS

The characteristics of the study participants ($n = 174$) are summarized in Table 1. Our participants had average 5.7 PACC measurements (range 4 to 7), and average 2.6 HV measurements ($n = 87$ with two measures, $n = 63$ with three measures, and $n = 24$ with four measures). Among these participants, there were 15 rs3846455^G minor allele carriers (14) heterozygotes and one homozygote), and the MAF was 0.05, similar to the European reference population MAF of 0.06 from the 1000 Genomes project [15, 16]. Baseline characteristics were not significantly different between rs3846455G carriers and non-carriers (Table 1).

The rs3846455^G carrier status was not associated with baseline PACC ($p = 0.17$; adjusted for age, sex, years of education, and EV1–3), but the SNP was weakly associated with longitudinal decline in PACC, after controlling for the time interaction terms of baseline Aβ, baseline age, sex, years of education, and EV1–3 (β = −0.087/year, 95% CI −0.169 to −0.005, p = 0.039; Table 2 model 2; Fig. 1B). The standardized effect size (estimated effect ("β-value") divided by standard error) of $rs3846455^G$ carrier status was -2.07, which is about 40% of the standardized effect size of baseline Aβ (−5.42) from the same model. Excluding a participant with rs3846455GG genotype from Table 2 model 2 did not significantly change the result (β = −0.106/year, 95% CI −0.191 to −0.021, $p = 0.015$). Unadjusted association between rs3846455G carrier status and longitudinal PACC also reached statistical significance (β = -0.090/year, 95% CI -0.179 to -0.001, $p = 0.048$), supporting that statistical over-fitting did not drive our result.

The rs3846455^G carrier status was not associated with baseline HV ($p = 0.33$; adjusted for age, sex, intracranial volume, and EV1–3). However, rs3846455G was associated with longitudinal decline in HV ($\beta = -57.1$ mm³/year, 95% confidence interval (CI) –103.0 to −11.2, $p = 0.015$; Table 2 model 3), even after additionally adjusting for baseline A β (β = -57.3 mm³/year, 95% CI -102.8 to -11.9 , $p = 0.014$; Table 2 model 4, Fig. 1A). The standardized effect size of rs3846455G carrier status on longitudinal HV change was −2.49, slightly greater than the standardized effect size of baseline \overrightarrow{AB} (−2.01) from the same model. The association between rs3846455^G and longitudinal HV was consistently observed

in an unadjusted model without covariates (β = −48.1 mm³/year, 95% CI −95.3 to −0.85, p $= 0.046$). Notably, this association was no longer statistically significant when we excluded one participant with rs3846455^{GG} genotype from Table 2 model 4 (β = -42.9 mm³/year, 95% CI −90.1 to 4.3, $p = 0.075$). We note that in a *post-hoc* analysis only assuming random intercepts (i.e., not allowing random slopes) in Table 2 model 4, we observed an association between rs3846455^G and longitudinal HV, even when the rs3846455^{GG} participant was excluded ($\beta = -52.3$ mm³/year, 95% CI –95.0 to –9.6, $p = 0.017$).

Finally, we performed a post-hoc mediation analysis to assess whether the association between $rs3846455^G$ and hippocampal volume loss (Table 2 model 4) mediates the more rapid PACC decline observed with the rs3846455G allele (Table 2 model 2). In this mediation model with $rs3846455^G$ as the independent variable, HV slope as the mediator variable, and PACC slope as the dependent variable, the effect of rs3846455G on PACC decline was partially mediated by faster progression of hippocampal atrophy in rs3846455^G carriers (mediated effect on PACC slope = -0.014 , 95% CI -0.032 to -6.0×10^{-4} , $p = 0.039$; Fig. 2). The direct effect of rs3846455^G on PACC decline not mediated through HV slope was not statistically significant ($p = 0.29$; Fig. 2).

DISCUSSION

In this study, leveraging neuroimaging data from HABS, we replicated the previous finding from clinical-pathologic cohorts (the Religious Orders Study (ROS) and the Rush Memory and Aging Project (MAP)) that an UNC5C haplotype captured by $rs3846455^G$ is associated with worse cognitive trajectories in older adults, even after controlling for measurable neuropathology burden [2]. As HABS and ROS-MAP are independent, nonoverlapping observational studies of cognitive aging, our study provides an independent validation of a role of the UNC5C rs3846455^G haplotype in interpersonal variability in cognitive decline, and demonstrates the utility of neuroimaging biomarkers in the ante-mortem studies of genetically determined susceptibility to age-related cognitive decline.

UNC5C is a gene with roles in axon guidance during development [17, 18] and apoptosis regulation [19]. UNC5C is a functional dependence receptor that can induce apoptosis in the absence of its ligand (netrin) [19] and has an enriched expression in adult human and rodent hippocampus [3, 5]. Overexpression of UNC5C increased susceptibility of cultured rat hippocampal neurons to various neurotoxic insults [3], suggesting the role of UNC5C in modulating hippocampal susceptibility to neurodegeneration. Moreover, the previously observed association of rs3846455G with cognitive decline was specific to the episodic memory domain that depends on hippocampus [2], further implicating UNC5C in hippocampal susceptibility to neurodegeneration.

In this context, our hypothesis-driven characterization of a candidate polymorphism shows that the $UNC5C$ rs3846455^G haplotype is associated with accelerated hippocampal atrophy in older adults who were clinically normal at baseline. This association partially mediated the association of rs3846455^G with accelerate cognitive decline, in a *post-hoc* mediation modeling. This finding is line with previous experimental and clinical-pathologic studies of $UNC5C$ [2, 3], and expands our understanding of the rs3846455^G haplotype. However, our

study has a modest sample size, and the association between rs3846455^G carrier status and accelerated hippocampal atrophy was no longer statistically significant after excluding the single participant that is homozygous for the risk allele (rs3846455^{GG} genotype) ($p = 0.075$). Therefore, the previously unreported association between rs3846455^G and accelerated hippocampal atrophy requires further validation by replication studies with larger sample size.

UNC5C rs3846455^G haplotype was not associated with baseline A β , hippocampal volume, or cognition in our study. This is consistent with the previous clinical-pathologic study with close to 1,000 participants that did not find a significant relationship of $rs3846455^{\text{G}}$ with any of the measured post-mortem neuropathologies or baseline cognition [2], as well as a large GWAS meta-analysis on a cross-sectional mid-adulthood hippocampal volume in more than 26,000 participants [20]. Thus, the clinical implication of the haplotype captured by rs3846455G is likely specific to the older adults at risk of cognitive decline.

The molecular mechanism by which the UNC5C haplotype captured by $rs3846455^G$ increases the susceptibility to cognitive decline in older adults remains unclear. As cellular overexpression of wild-type UNC5C increases apoptosis in cultured hippocampal neurons [3], it is plausible to hypothesize that functional variants increasing UNC5C expression in hippocampus could lead to accelerated hippocampal neurodegeneration. UNC5C $rs3846455^G$ is an intronic SNP within an enhancer region in human hippocampus, per the ChromHMM core 15-chromatin state prediction [21] based on the Roadmap Epigenomics Project [22]. This functional prediction suggests that the SNP might alter UNC5C gene expression in the hippocampus, but rs3846455 was not significantly associated with hippocampal UNC5C mRNA expression in the Genotype-Tissue Expression database ($n =$ 111) [22]. Given the SNP's relatively low MAF (0.06), larger number of hippocampal samples from older adults will be required to examine the expression quantitative trait locus association.

Our study has several important limitations. First and foremost, we had a very limited sample size for a genetic association study ($n = 177$), and the study sample included only fifteen $rs3846455^G$ carriers. Therefore, our results should be interpreted with caution, and the newly found association between $rs3846455^G$ and accelerated hippocampal atrophy as well as the subsequent mediation analysis requires further validation. Also, due to our limited sample size, we could not assess interactive association of $\mathbf{A}\beta$ and rs3846455^G, a potential genetic risk factor of hippocampal neurodegeneration, despite multiple previous reports that Aβ and neurodegeneration have synergistic predictive value for cognitive decline among clinically normal older adults [23-25]. Nonetheless, our study is an independent replication of the previously reported link between rs3846455G and cognitive decline [2], and the link between rs3846455G and hippocampal neurodegeneration was found through a hypothesis-driven candidate polymorphism analysis based on previous reports [2, 3]. Thus, our results are likely capturing a true effect rather than spurious associations. Second, our analyses were restricted to participants from European ancestry, and the results cannot be easily extrapolated to people of non-European ancestry. Third, HABS participants are highly educated and in good physical and cognitive health at the time of enrollment. As such, they are not entirely representative of the general population of older adults. Still, MAF of

 $rs3846455^G$ in our participants was not significantly different from the reference panel [15, 16], indicating a genetically representative sampling. Fourth, there were only 14% of APOE e^4 carriers among the rs3846455^G carriers, while 31% of the rs3846455^C homozygotes were APOE $e4$ carriers. Although this imbalance did not reach statistical significance, it might have led to a slight underestimation of the effect of the rs3846455G allele on hippocampal atrophy and cognitive decline. Finally, the mediation analysis should be interpreted cautiously, given that this was a simplified post-hoc model assuming a linear relationship among rs3846455G carrier status, HV change, and PACC change.

In conclusion, leveraging longitudinal data with approximately five-year average follow-up, we validated the previously reported association of the UNC5C rs3846455^G haplotype and faster cognitive decline in older adults. Moreover, our results suggest that accelerated hippocampal atrophy might partially mediate the association between 3846455G and faster cognitive decline in clinically normal older adults. This study also demonstrates the utility of modestly sized, but deeply phenotyped cohorts in investigating the genetics of complex endophenotypes such as cognitive decline not explained by neuropathology [26]. Nonetheless, neuroimaging studies with larger numbers of participants including individuals of non-European descent are required to further validate our results and understand the role of the UNC5C rs3846455^G haplotype in more diverse populations. Further investigations of the functional mechanism of the UNC5C rs3846455^G haplotype may open additional therapeutic avenues for neuroprotection in older adults at high risk of cognitive decline.

ACKNOWLEDGMENTS

We thank the participants of the Harvard Aging Brain Study (HABS). In this study, we also used the data from the Genotype-Tissue Expression (GTEx) Project, which was supported by the Common Fund of the Office of the Director of the National Institutes of Health, and by NCI, NHGRI, NHLBI, NIDA, NIMH, and NINDS. The data used for the analyses described in this manuscript were obtained from: the GTEx Portal on 02/23/2018.

Our study was funded by NIH grants P01 AG036694, P50 AG005134, and K24 AG035007, and Alzheimer's Association Clinical Fellowship (AACF-17-505359). A generous gift from the Marr family funded the genomewide genotyping in HABS. This research was carried out in part at the Athinoula A. Martinos Center for Biomedical Imaging at the Massachusetts General Hospital, using resources provided by the Center for Functional Neuroimaging Technologies, P41EB015896, a P41 Biotechnology Resource Grant supported by the National Institute of Biomedical Imaging and Bioengineering (NIBIB), National Institutes of Health. This work also involved the use of instrumentation supported by the NIH Shared Instrumentation Grant Program and/or High-End Instrumentation Grant Program; specifically, grant numbers S10RR021110, S10RR023401, and S10RR023043.

REFERENCES

- [1]. Yu L, Boyle PA, Segawa E, Leurgans S, Schneider JA, Wilson RS, Bennett DA (2015) Residual decline in cognition after adjustment for common neuropathologic conditions. Neuropsychology 29, 335–343. [PubMed: 25495832]
- [2]. White CC, Yang HS, Yu L, Chibnik LB, Dawe RJ, Yang J, Klein HU, Felsky D, Ramos-Miguel A, Arfanakis K, Honer WG, Sperling RA, Schneider JA, Bennett DA, De Jager PL (2017) Identification of genes associated with dissociation of cognitive performance and neuropathological burden: Multistep analysis of genetic, epigenetic, and transcriptional data. PLoSMed 14, e1002287.
- [3]. Wetzel-Smith MK, Hunkapiller J, Bhangale TR, Srinivasan K, Maloney JA, Atwal JK, Sa SM, Yaylaoglu MB, Foreman O, Ortmann W, Rathore N, Hansen DV, Tessier-Lavigne M, Alzheimer's Disease Genetics C, Mayeux R, Pericak-Vance M, Haines J, Farrer LA, Schellenberg GD, Goate A, Behrens TW, Cruchaga C, Watts RJ, Graham RR (2014) A rare

mutation in UNC5C predisposes to late-onset Alzheimer's disease and increases neuronal cell death. NatMed 20,1452–1457.

- [4]. Yang HS, White CC, Chibnik LB, Klein HU, Schneider JA, Bennett DA, De Jager PL (2017) UNC5C variants are associated with cerebral amyloid angiopathy. Neurol Genet 3, e176. [PubMed: 28761931]
- [5]. Hawrylycz MJ, Lein ES, Guillozet-Bongaarts AL, Shen EH, Ng L, Miller JA, van de Lagemaat LN, Smith KA, Ebbert A, Riley ZL, Abajian C, Beckmann CF, Bernard A, Bertagnolli D, Boe AF, Cartagena PM, Chakravarty MM, Chapin M, Chong J, Dalley RA, David Daly B, Dang C, Datta S, Dee N, Dolbeare TA, Faber V, Feng D, Fowler DR, Goldy J, Gregor BW, Haradon Z, Haynor DR, Hohmann JG, Horvath S, Howard RE, Jeromin A, Jochim JM, Kinnunen M, Lau C, Lazarz ET, Lee C, Lemon TA, Li L, Li Y, Morris JA, Overly CC, Parker PD, Parry SE, Reding M, Royall JJ, Schulkin J, Sequeira PA, Slaughterbeck CR, Smith SC, Sodt AJ, Sunkin SM, Swanson BE, Vawter MP, Williams D, Wohnoutka P, Zielke HR, Geschwind DH, Hof PR, Smith SM, Koch C, Grant SGN, Jones AR (2012) An anatomically comprehensive atlas of the adult human brain transcriptome. Nature 489, 391–399. [PubMed: 22996553]
- [6]. Sperling R, Mormino E, Johnson K (2014) The evolution of preclinical Alzheimer's disease: Implications for prevention trials. Neuron 84, 608–622. [PubMed: 25442939]
- [7]. Dagley A, LaPoint M, Huijbers W, Hedden T, McLaren DG, Chatwal JP, Papp KV, Amariglio RE, Blacker D, Rentz DM, Johnson KA, Sperling RA, Schultz AP (2017) Harvard Aging Brain Study: Dataset and accessibility. Neuroimage 144, 255–258. [PubMed: 25843019]
- [8]. The International HapMap 3 Consortium (2010) Integrating common and rare genetic variation in diverse human populations. Nature 467, 52–58. [PubMed: 20811451]
- [9]. Price AL, Patterson NJ, Plenge RM, Weinblatt ME, Shadick NA, Reich D (2006) Principal components analysis corrects for stratification in genome-wide association studies. Nat Genet 38, 904–909. [PubMed: 16862161]
- [10]. Mormino EC, Papp KV, Rentz DM, Donohue MC, Amariglio R, Quiroz YT, Chhatwal J, Marshall GA, Donovan N, Jackson J, Gatchel JR, Hanseeuw BJ, Schultz AP, Aisen PS, Johnson KA, Sperling RA (2017) Early and late change on the preclinical Alzheimer's cognitive composite in clinically normal older individuals with elevated amyloid beta. Alzheimers Dement 13, 1004–1012. [PubMed: 28253478]
- [11]. Donohue MC, Sperling RA, Salmon DP, Rentz DM, Raman R, Thomas RG, Weiner M, Aisen PS, Australian Imaging B, Lifestyle Flagship Study of Ageing, Alzheimer's Disease Neuroimaging Initiative, Alzheimer's Disease Cooperative Study (2014) The preclinical Alzheimer cognitive composite: Measuring amyloid-related decline. JAMA Neurol 71, 961–970. [PubMed: 24886908]
- [12]. Buckley RF, Schultz AP, Hedden T, Papp KV, Hanseeuw BJ, Marshall G, Sepulcre J, Smith EE, Rentz DM, Johnson KA, Sperling RA, Chhatwal JP (2017) Functional network integrity presages cognitive decline in preclinical Alzheimer disease. Neurology 89, 29–37. [PubMed: 28592457]
- [13]. Jack CR Jr., Barkhof F, Bernstein MA, Cantillon M, Cole PE, Decarli C, Dubois B, Duchesne S, Fox NC, Frisoni GB, Hampel H, Hill DL, Johnson K, Mangin JF, Scheltens P, Schwarz AJ, Sperling R, Suhy J, Thompson PM, Weiner M, Foster NL (2011) Steps to standardization and validation of hippocampal volumetry as a biomarker in clinical trials and diagnostic criterion for Alzheimer's disease. Alzheimers Dement 7, 474–485 e474. [PubMed: 21784356]
- [14]. Tingley D, Yamamoto T, Hirose K, Keele L, Imai K (2014) mediation: R package for causal mediation analysis. J Stat Softw 59, 10.18637/jss.v059.i05.
- [15]. Ward LD, Kellis M (2012) HaploReg: A resource for exploring chromatin states, conservation, and regulatory motif alterations within sets of genetically linked variants. Nucleic Acids Res 40, D930–934. [PubMed: 22064851]
- [16]. The 1000 Genomes Project Consortium (2012) An integrated map of genetic variation from 1,092 human genomes. Nature 491, 56–65. [PubMed: 23128226]
- [17]. Leonardo ED, Hinck L, Masu M, Keino-Masu K, Ackerman SL, Tessier-Lavigne M (1997) Vertebrate homologues of C. elegans UNC-5 are candidate netrin receptors. Nature 386, 833– 838. [PubMed: 9126742]

- [18]. Ackerman SL, Kozak LP, Przyborski SA, Rund LA, Boyer BB, Knowles BB (1997) The mouse rostral cerebellar malformation gene encodes an UNC-5-like protein. Nature 386, 838–842. [PubMed: 9126743]
- [19]. Bernet A, Mazelin L, Coissieux MM, Gadot N, Ackerman SL, Scoazec JY, Mehlen P (2007) Inactivation of the UNC5C Netrin-1 receptor is associated with tumor progression in colorectal malignancies. Gastroenterology 133, 1840–1848. [PubMed: 17967459]
- [20]. Hibar DP, Adams HHH, Jahanshad N, Chauhan G, Stein JL, Hofer E, Renteria ME, Bis JC, Arias-Vasquez A, Ikram MK, Desrivieres S, Vernooij MW, Abramovic L, Alhusaini S, Amin N, Andersson M, Arfanakis K, Aribisala BS, Armstrong NJ, Athanasiu L, Axelsson T, Beecham AH, Beiser A, Bernard M, Blanton SH, Bohlken MM, Boks MP, Bralten J, Brickman AM, Carmichael O, Chakravarty MM, Chen Q, Ching CRK, Chouraki V, Cuellar-Partida G, Crivello F, Den Braber A, Doan NT, Ehrlich S, Giddaluru S, Goldman AL, Gottesman RF, Grimm O, Griswold ME, Guadalupe T, Gutman BA, Hass J, Haukvik UK, Hoehn D, Holmes AJ, Hoogman M, Janowitz D, Jia T, Jorgensen KN, Karbalai N, Kasperaviciute D, Kim S, Klein M, Kraemer B, Lee PH, Liewald DCM, Lopez LM, Luciano M, Macare C, Marquand AF, Matarin M, Mather KA, Mattheisen M, McKay DR, Milaneschi Y, Munoz Maniega S, Nho K, Nugent AC, Nyquist P, Loohuis LMO, Oosterlaan J, Papmeyer M, Pirpamer L, Putz B, Ramasamy A, Richards JS, Risacher SL, Roiz-Santianez R, Rommelse N, Ropele S, Rose EJ, Royle NA, Rundek T, Samann PG, Saremi A, Satizabal CL, Schmaal L, Schork AJ, Shen L, Shin J, Shumskaya E, Smith AV, Sprooten E, Strike LT, Teumer A, Tordesillas-Gutierrez D, Toro R, Trabzuni D, Trompet S, Vaidya D, Van der Grond J, Van der Lee SJ, Van der Meer D, Van Donkelaar MMJ, Van Eijk KR, Van Erp TGM, Van Rooij D, Walton E, Westlye LT, Whelan CD, Windham BG, Winkler AM, Wittfeld K, Woldehawariat G, Wolf C, Wolfers T, Yanek LR, Yang J, Zijdenbos A, Zwiers MP, Agartz I, Almasy L, Ames D, Amouyel P, Andreassen OA, Arepalli S, Assareh AA, Barral S, Bastin ME, Becker DM, Becker JT, Bennett DA, Blangero J, van Bokhoven H, Boomsma DI, Brodaty H, Brouwer RM, Brunner HG, Buckner RL, Buitelaar JK, Bulayeva KB, Cahn W, Calhoun VD, Cannon DM, Cavalleri GL, Cheng CY, Cichon S, Cookson MR, Corvin A, Crespo-Facorro B, Curran JE, Czisch M, Dale Am, Davies GE, De Craen AJM, De Geus EJC, De Jager PL, De Zubicaray GI, Deary IJ, Debette S, DeCarli C, Delanty N, Depondt C, DeStefano A, Dillman, Djurovic S, Donohoe G, Drevets WC, Duggirala R, Dyer TD, Enzinger C, Erk S, Espeseth T, Fedko IO, Fernandez G, Ferrucci L, Fisher SE, Fleischman DA, Ford I, Fornage M, Foroud TM, Fox PT, Francks C, Fukunaga M, Gibbs JR, Glahn DC, Gollub RL, Goring HHH, Green RC, Gruber O, Gudnason V, Guelfi S, Haberg AK, Hansell NK, Hardy J, Hartman CA, Hashimoto R, Hegenscheid K, Heinz A, Le Hellard S, Hernandez DG, Heslenfeld DJ, Ho BC, Hoekstra PJ, Hoffmann W, Hofman A, Holsboer F, Homuth G, Hosten N, Hottenga JJ, Huentelman M, Hulshoff Pol HE, Ikeda M, Jack CR Jr., Jenkinson M, Johnson R, Jonsson EG, Jukema JW, Kahn RS, Kanai R, Kloszewska I, Knopman DS, Kochunov P, Kwok JB, Lawrie SM, Lemaitre H, Liu X, Longo DL, Lopez OL, Lovestone S, Martinez O, Martinot JL, Mattay VS, McDonald C, McIntosh AM, McMahon FJ, McMahon KL, Mecocci P, Melle I, Meyer-Lindenberg A, Mohnke S, Montgomery GW, Morris DW, Mosley TH, Muhleisen TW, Muller-Myhsok B, Nalls MA, Nauck M, Nichols TE, Niessen WJ, Nothen MM, Nyberg L, Ohi K, Olvera RL, Ophoff RA, Pandolfo M, Paus T, Pausova Z, Penninx B, Pike GB, Potkin SG, Psaty BM, Reppermund S, Rietschel M, Roffman JL, Romanczuk-Seiferth N, Rotter JI, Ryten M, Sacco RL, Sachdev PS, Saykin AJ, Schmidt R, Schmidt H, Schofield PR, Sigursson S, Simmons A, Singleton A, Sisodiya SM, Smith C, Smoller JW, Soininen H, Steen VM, Stott DJ, Sussmann JE, Thalamuthu A, Toga AW, Traynor BJ, Troncoso J, Tsolaki M, Tzourio C, Uitterlinden AG, Hernandez MCV, Van der Brug M, van der Lugt A, van der Wee NJA, Van Haren NEM, van 't Ent D, Van Tol MJ, Vardarajan BN, Vellas B, Veltman DJ, Volzke H, Walter H, Wardlaw JM, Wassink TH, Weale ME, Weinberger DR, Weiner MW, Wen W, Westman E, White T, Wong TY, Wright CB, Zielke RH, Zonderman AB, Martin NG, Van Duijn CM, Wright MJ, Longstreth WT, Schumann G, Grabe HJ, Franke B, Launer LJ, Medland SE, Seshadri S, Thompson PM, Ikram MA (2017) Novel genetic loci associated with hippocampal volume. Nat Commun 8, 13624. [PubMed: 28098162]
- [21]. Ernst J, Kellis M (2012) ChromHMM: Automating chromatin-state discovery and characterization. Nat Methods 9, 215–216. [PubMed: 22373907]
- [22]. Consortium GTEx (2017) Genetic effects on gene expression across human tissues. Nature 550, 204–213. [PubMed: 29022597]

- [23]. Knopman DS, Jack CR Jr, Wiste HJ, Weigand SD, Vemuri P, Lowe V, Kantarci K, Gunter JL, Senjem ML, Ivnik RJ, Roberts RO, Boeve BF, Petersen RC (2012) Short-term clinical outcomes for stages of NIA-AA preclinical Alzheimer disease. Neurology 78, 1576–1582. [PubMed: 22551733]
- [24]. Mormino EC, Betensky RA, Hedden T, Schultz AP, Amariglio RE, Rentz DM, Johnson KA, Sperling RA (2014) Synergistic effect of beta-amyloid and neurodegeneration on cognitive decline in clinically normal individuals. JAMA Neurol 71, 1379–1385. [PubMed: 25222039]
- [25]. Vos SJ, Xiong C, Visser PJ, Jasielec MS, Hassenstab J, Grant EA, Cairns NJ, Morris JC, Holtzman DM, Fagan AM (2013) Preclinical Alzheimer's disease and its outcome: A longitudinal cohort study. Lancet Neurol 12, 957–965. [PubMed: 24012374]
- [26]. De Jager PL, Yang HS, Bennett DA (2018) Deconstructing and targeting the genomic architecture of human neurodegeneration. Nat Neurosci 21, 1310–1317. [PubMed: 30258235]

Fig. 1.

rs3846455G carrier status and longitudinal changes in bilateral hippocampal volume and Preclinical Alzheimer Cognitive Composite. A) PACC change over time was predicted from a linear mixed effect model, controlling for interaction of time (years from baseline) with baseline Aβ, baseline age, sex, years of education, and EV1–3. Predicted trajectories of PACC were plotted by *UNC5C* rs3846455^G minor allele carrier status. B) HV change over time was predicted from a linear mixed effect model, controlling for total intracranial volume and interaction of time (years from baseline) with baseline Aβ, baseline age, sex, and EV1-3. Predicted trajectories of HV were plotted by UNC5C rs3846455^G minor allele carrier status. Aβ, baseline amyloid-β measured by Pittsburgh compound B (PiB) positron emission tomography distributed volume ratio (DVR) from composite cortical regions; EV1–3, first three principal components from the genotype covariance matrix; HV, hippocampal volume; PACC, preclinical Alzheimer cognitive composite.

Fig. 2.

Mediation of the association of rs3846455^G with cognitive decline by accelerated hippocampal atrophy. Causal mediation analysis was performed using quasi-Bayesian Monte Carlo method (with 10,000 simulations), having $rs3846455^G$ as an independent (causal) binary variable, HV slope (adjusted for baseline age, sex, and ICV) as a continuous mediator variable, and PACC slope (adjusted for baseline age, sex, and years of education) as the continuous outcome variable. We also adjusted for baseline $\mathbf{A}\mathbf{\beta}$ and $\mathbf{E}\mathbf{V}1-\mathbf{3}$ in the mediator and outcome models that we used for the simulation. Mediated (indirect) effect (ACME) and direct effect (ADE) were calculated. Solid arrow indicates a significant mediated effect (p) \leq 0.05), and dotted arrow indicates a direct effect that did not reach statistical significance (p) $= 0.29$). Aβ, amyloid-β measured by Pittsburgh compound B (PiB) positron emission tomography distributed volume ratio (DVR) from composite cortical regions; ACME, average causal mediation effect (the effect of the independent variable on the outcome that is mediated through the mediator); ADE, average direct effect (the effect of the independent variable on the outcome that is independent from the mediator); EV1–3, first three principal components from the genotype covariance matrix; HV, hippocampal volume; ICV, intracranial volume; PACC, Preclinical Alzheimer Cognitive Composite.

Table 1

Demographic characteristics of the study participants Demographic characteristics of the study participants

p-value for difference (either by two-sample t-test or chi-squared test) between two genotype groups are noted. ļ ă lyk. á ļ Ļ 5 L

 σ n = 167. Aβ, baseline amyloid-β measured by Pittsburgh compound B (PiB) positron emission tomography distributed volume ratio (DVR) from composite cortical regions; f/u, follow-up; HV, bilateral hippocampal volume; PACC, preclinical Alzheimer Cognitive Composite; SD, standard deviation. Author Manuscript

Author Manuscript

Table 2

UNC5C rs3846455^G and longitudinal change in the Preclinical Alzheimer Cognitive Composite (PACC) and hippocampal volume (HV) G and longitudinal change in the Preclinical Alzheimer Cognitive Composite (PACC) and hippocampal volume (HV) UNC5C rs3846455

(years from baseline) with baseline age, sex, years of education, and first three principal components from genotype covariance matrix (EV1–3). In models 3 and 4, estimated effects (slopes) on longitudinal (years from baseline) with baseline age, sex, years of education, and first three principal components from genotype covariance matrix (EV1-3). In models 3 and 4, estimated effects (slopes) on longitudinal ("β-value") by standard error. Aβ, baseline amyloid-β measured by Pittsburgh compound B positron emission tomography distributed volume ratio from composite cortical regions; CI, confidence interval; In models 1 and 2, estimated effects (slopes) on longitudinal change of PACC were reported for each predictor from linear mixed effect models. In these two models, we adjusted for the interaction of time In models 1 and 2, estimated effects (slopes) on longitudinal change of PACC were reported for each predictor from linear mixed effect models. In these two models, we adjusted for the interaction of time ("β-value") by standard error. Aβ, baseline amyloid-β measured by Pittsburgh compound B positron emission tomography distributed volume ratio from composite cortical regions; CI, confidence interval; hippocampal volume measurement, as well as the interaction of time (years from baseline) with baseline age, sex, and EV1-3. Standardized effect sizes were calculated by dividing each estimated effect hippocampal volume measurement, as well as the interaction of time (years from baseline) with baseline age, sex, and EV1–3. Standardized effect sizes were calculated by dividing each estimated effect change of bilateral hippocampal volume were reported for each predictor from linear mixed effect models. In these two models, we adjusted for the total intracranial volume from the MRI of each change of bilateral hippocampal volume were reported for each predictor from linear mixed effect models. In these two models, we adjusted for the total intracranial volume from the MRI of each HV, bilateral hippocampal volume; PACC, Preclinical Alzheimer Cognitive Composite. HV, bilateral hippocampal volume; PACC, Preclinical Alzheimer Cognitive Composite.