# Differential Localization of G-proteins $G_i$ and $G_o$ in the Accessory Olfactory Bulb of the Rat

### Haruo Shinohara, Tomiko Asano, and Kanefusa Kato

Department of Biochemistry, Institute for Developmental Research, Aichi Prefectural Colony, Kasugai, Aichi, Japan

To clarify the functional differences among G-proteins, we investigated the localization of G<sub>i</sub> and G<sub>o</sub> in the olfactory bulb of rats by both immunohistochemical and immunochemical techniques, using purified antibodies specific to the  $\alpha$ -subunits of  $G_{i1}$  ( $G_{i1}\alpha$ ),  $G_{i2}$  ( $G_{i2}\alpha$ ), and  $G_{o}$  ( $G_{o}\alpha$ ), respectively. We found that  $G_{i2}\alpha$  is localized exclusively in the accessory olfactory bulb, but it is present at only low levels in the main olfactory bulb. The unique pattern of immunoreactivity specific for  $G_{ij}\alpha$  and  $G_{ij}\alpha$  within the glomeruli of the accessory olfactory bulb and the results of immunoassays indicate that the accessory olfactory bulb is divided into two parts: the anterior region is rich in G<sub>12</sub>, while the posterior region is rich in G. These findings suggest that the accessory olfactory bulb has two different functions. In addition, we found that the concentration of  $G_{ip}\alpha$  in the accessory olfactory bulb increases during puberty and reaches the adult level at 12 weeks after birth, while that in the main olfactory bulb remains constant. By contrast, the concentrations of  $G_{\alpha}\alpha$  in the accessory olfactory bulb and the main olfactory bulb increase with similar kinetics. These findings suggest that G<sub>12</sub> is a key protein in signal transduction in the accessory olfactory bulb, and increases in its level seem to be related to sexual maturation.

There are two olfactory pathways in vertebrates, the main and the accessory pathways. The main olfactory pathway, which originates in the sensory epithelium in the nasal cavity and ends in the main olfactory bulb, is involved in the perception of volatile olfactory stimuli. By contrast, the receptor neurons in the vomeronasal organ, which consists of bilaterally paired tubes lying in the ventral portion of the nasal cavity, project to the accessory olfactory bulb through the vomeronasal nerves (McCotter, 1912), and this pathway seems to be involved in the olfaction of nonvolatiles (Wysocki et al., 1980). Destruction of the accessory olfactory pathway impairs the reproductive functions of experimental animals (Powers and Winans, 1971; Meredith, 1986; Saito and Moltz, 1986), suggesting that the accessory olfactory system plays an important role in the perception of intersex smell.

G-proteins are a family of signal-coupling proteins, and each is a heterotrimer composed of  $\alpha$ -,  $\beta$ -, and  $\gamma$ -subunits. In the

sensory organs of vertebrates, such as the olfactory epithelium and the retina, tissue-specific G-proteins function as active signal transducers. Certain odorants stimulate adenylyl cyclase through G<sub>olf</sub> (Jones and Reed, 1989) and G<sub>s</sub> (Mania-Farnell and Farbman, 1990), and transducin regulates the activity of cGMP phosphodiesterase in response to activation by light of the photoreceptor rhodopsin (Gilman, 1987). There are also many reports suggesting that G<sub>i</sub> and G<sub>o</sub> are coupled with several receptors and regulate adenylyl cyclase activity, Ca<sup>2+</sup> and K<sup>+</sup> channels, and phospholipase C activity (Kikuchi et al., 1986; Hescheler et al., 1987; Neer and Clapham, 1988; VanDongen et al., 1988; Moriarty et al., 1990). However, it is still unclear whether the three subtypes of G<sub>i</sub> (G<sub>i1</sub>, G<sub>i2</sub>, and G<sub>i3</sub>) and G<sub>o</sub> have individua! and specific functions. Go and Gil are mainly localized in the neuropil of the brain. G<sub>12</sub> is widely distributed in various tissues, including the brain, but concentrations of G<sub>1</sub>, in the brain are much lower than those of Gi and Go. To clarify the functional differences between G<sub>i</sub> and G<sub>o</sub>, we investigated the localization of these G-proteins by use of purified antibodies specific to the respective  $\alpha$ -subunits of  $G_{i1}$   $(G_{i1}\alpha)$ ,  $G_{i2}$   $(G_{i2}\alpha)$ , and  $G_{i3}$   $(G_{i4}\alpha)$  in the rat olfactory bulb. We report here that  $G_{i2}\alpha$  is expressed at an extremely high level in the accessory olfactory pathway and that its concentration increases with postnatal development. probably in close association with sexual maturation. We also show that the accessory olfactory bulb is composed of two distinctive regions, one of which is rich in G<sub>12</sub> while the other is rich in Go.

## **Materials and Methods**

Immunohistochemical study. Five male rats (12 weeks old) were deeply anesthetized with diethyl ether and perfused with a fixative solution (Schmechel et al., 1980) composed of 4% paraformaldehyde, 1% glutaraldehyde, 0.2% picric acid, and 2% sucrose in 0.1 m sodium acetate buffer, pH 6.0. This fixative has been shown to be suitable for immunohistochemical studies of the  $\alpha$ -subunits of  $G_0$  (Asano et al., 1987, 1988; Semba et al., 1990), G<sub>i1</sub> (Asano et al., 1990), and G<sub>i2</sub> (Asano et al., 1989a). The dissected olfactory bulbs were kept in the same fixative for 4 hr. After rinsing with and left standing in 50 mm Tris-HCl buffer (pH 7.4) overnight, the tissues were dehydrated through a graded series of alcohols and embedded in paraffin. Serial sections (5  $\mu$ m) of the main and the accessory olfactory bulbs from rats were prepared and immunostained by the indirect peroxidase-labeled antibody method (Nakane, 1975), using purified antibodies against  $G_{i1}\alpha$ ,  $G_{i2}\alpha$ ,  $G_{o}\alpha$ , and  $\beta\gamma$ . For controls, antibodies preabsorbed with the respective purified G-proteins were used, and these antibodies were not associated with any positive staining. The specificity of these antibodies has been described elsewhere (Asano et al., 1987, 1989a, 1990; Morishita et al., 1988), Production of the antibodies can be summarized as follows.  $G_0\alpha$ - and  $\beta\gamma$ -subunits were purified from bovine brain, while  $G_{i1}\alpha$  and  $G_{i2}\alpha$  were purified from bovine lung. The antisera were raised in rabbits by injection of the purified G-proteins, and the antibodies were purified from antisera by the use of appropriate antigen-coupled Sepharose columns. Antibodies against  $G_0\alpha$  reacted only with  $G_0\alpha$  and did not cross-react with  $G_1\alpha$ - or

Received July 5, 1991; revised Oct. 29, 1991; accepted Nov. 11, 1991.

This work was supported in part by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science and Culture of Japan.

Correspondence should be addressed to Haruo Shinohara, M.D., Department of Biochemistry, Institute for Developmental Research, Aichi Prefectural Colony, Kamiya, Kasugai, Aichi 480-03, Japan.

Copyright © 1992 Society for Neuroscience 0270-6474/92/121275-05\$05.00/0

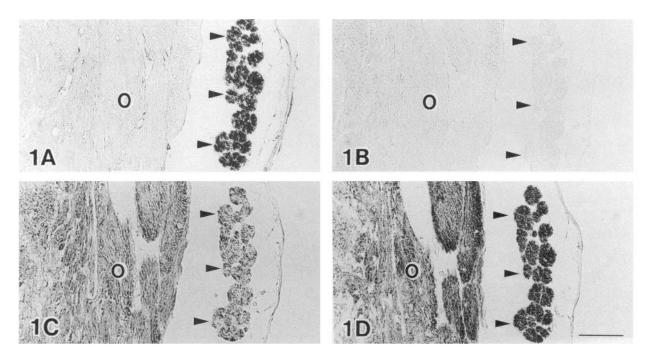


Figure 1. Serial sections of the main olfactory bulb and the vomeronasal nerves, immunostained with antibodies against  $G_{i2}\alpha$  (A),  $G_{i1}\alpha$  (B),  $G_o\alpha$  (C), and  $\beta\gamma$  (D). The vomeronasal nerves (arrowheads) between bilateral olfactory bulbs are intensely immunopositive for  $G_{i2}\alpha$ ,  $G_o\alpha$ , and  $\beta\gamma$  but almost immunonegative for  $G_{i1}\alpha$ . By contrast, the olfactory nerve layer (O) in the main olfactory bulb is immunopositive only for  $G_o\alpha$  and  $\beta\gamma$ . Note that the vomeronasal nerves are homogeneously immunopositive for  $\beta\gamma$ , but they are not homogeneously immunopositive for  $G_{i2}\alpha$  and  $G_o\alpha$ . Scale bar, 50  $\mu$ m.

 $\beta\gamma$ -subunits in an immunoblot assay. Antibodies against  $G_{i_1}\alpha$  (Asano et al., 1990) cross-reacted with  $G_{i_3}\alpha$ , but not with  $G_{i_2}\alpha$  or  $G_o\alpha$ . Because the concentration of  $G_{i_3}$  seems to be very low in the brain (Goldsmith et al., 1988; Kanaho et al., 1989), the antibodies are referred to as anti- $G_{i_1}\alpha$ . Antibodies against  $G_{i_2}\alpha$  reacted only with  $G_{i_2}\alpha$ , but not with  $G_{i_1}\alpha$ ,  $G_{i_3}\alpha$ , or  $G_o\alpha$ . Antibodies against the  $\beta\gamma$ -subunits reacted mainly with the 36 kDa  $\beta$ -subunit. Since  $\beta\gamma$ -subunits are common to all G-proteins, antibodies against  $\beta\gamma$  were also used to confirm the presence of the  $\alpha$ -subunit of  $G_o$  and  $G_i$ .

Enzyme immunoassay procedures. Male rats of various ages were deeply anesthetized with diethyl ether, and the main and the accessory olfactory bulbs were dissected out under a stereomicroscope with ophthalmologist's scissors. The anterior and the posterior parts of the accessory olfactory bulb of five male rats (16 weeks old) were dissected out in the same way. The tissues were kept frozen at −80°C until analysis. The frozen tissues of the accessory olfactory bulb were homogenized at 0°C with a Potter-Elvehjem homogenizer in 100 µl of 20 mm Tris-HCl (pH 8.0), 1 mm EDTA, 5 mm  $\beta$ -mercaptoethanol, and 1% sodium cholate. The main olfactory bulbs were homogenized in 9 vol (v/w) of 20 mm Tris-HCl (pH 8.0), 1 mm EDTA, 5 mm β-mercaptoethanol, and 2% sodium cholate. The homogenate was sonicated for 20 sec and then centrifuged at 4°C at 100,000 × g for 1 hr. The supernatant fractions were used for the immunoassay of  $G_{i2}\alpha$  (Asano et al., 1989a) and  $G_{\alpha}\alpha$  (Asano et al., 1987) after dilution. Less than 10% of total GTP $\gamma$ S-binding activity remained in the insoluble fraction when this procedure was followed (Asano et al., 1987). Proteins were quantitated by the method of Schaffner and Weissmann (1973).

## Results

The vomeronasal nerves running between the right and the left main olfactory bulbs were intensely stained after immunoreactions specific for  $G_{i2}\alpha$ ,  $G_o\alpha$ , and  $\beta\gamma$ , but they were almost immunonegative for  $G_{i1}\alpha$ . In addition, the vomeronasal nerves were homogeneously immunopositive for  $\beta\gamma$ , but they were not homogeneously immunopositive for  $G_{i2}\alpha$  and  $G_o\alpha$ . This observation suggests that not all of the sensory neurons in the vomeronasal organ are  $G_{i2}\alpha$  and  $G_o\alpha$  positive (Fig. 1). By contrast,

the olfactory nerve fibers were almost immunonegative for  $G_{i2}\alpha$ and  $G_{il}\alpha$ , but they were intensely immunoreactive for  $G_{o}\alpha$  and  $\beta\gamma$  (Fig. 1). In the accessory olfactory bulb, the pattern of staining of the glomeruli with the various antibodies revealed that the anterior region and the posterior region could be distinguished from one another. The glomeruli in the anterior region were intensely immunoreactive for  $G_{i2}\alpha$  but only faintly so for  $G_{o}\alpha$ , while those in the posterior region were intensely immunopositive for  $G_0\alpha$  but only faintly so for  $G_{i2}\alpha$  (Fig. 2A,C). Although the glomeruli in the anterior and the posterior regions were weakly immunopositive for  $G_{i1}\alpha$ , the immunoreactivity was greater in the posterior region (Fig. 2B). The glomeruli in the anterior and the posterior regions were equally immunoreactive for  $\beta \gamma$  (Fig. 2D). The molecular layer reacted to an equal extent with each of these antibodies (Fig. 2). To extend this finding, we quantitated the concentrations of  $G_0\alpha$  and  $G_{i2}\alpha$  in the anterior region and the posterior region of the accessory olfactory bulb by immunoassay. The results of the immunoassays revealed that the concentration of  $G_{i2}\alpha$  was higher in the anterior region, while the concentration of  $G_0\alpha$  was higher in the posterior region (Fig. 3).

Because the accessory olfactory system seems to participate in sexual behavior, we next examined whether the concentration of  $G_{i2}\alpha$  in the accessory olfactory bulb increases during sexual maturation. We previously reported that the concentration of  $G_{i2}\alpha$  in the rat cerebral cortex was almost constant throughout postnatal development (Asano et al., 1989a). In the main olfactory bulb, the concentration of  $G_{i2}\alpha$  was also constant in rats of all ages examined (Fig. 4). By contrast, the concentration of  $G_{i2}\alpha$  in the accessory olfactory bulb increased markedly at puberty and reached the adult level at 12 weeks after birth. The concentration of  $G_{i2}\alpha$  in the accessory olfactory bulb of rats at 12 weeks after birth (37.4  $\pm$  1.4 pmol/mg protein) was 2.6-fold

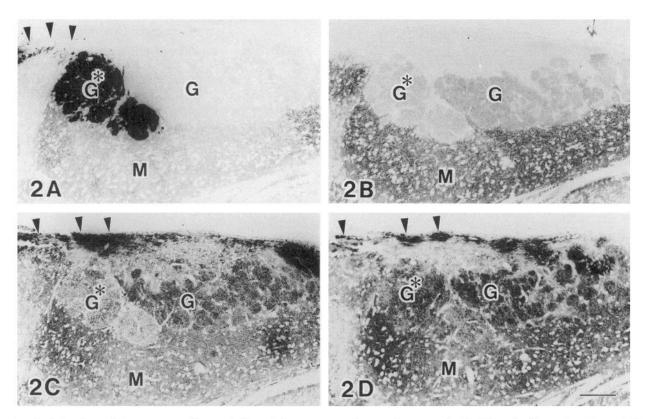


Figure 2. Serial sections of the accessory olfactory bulb and the vomeronasal nerves immunostained with antibodies against  $G_{i2}\alpha$  (A),  $G_{i1}\alpha$  (B),  $G_{o}\alpha$  (C), and  $\beta\gamma$  (D). The vomeronasal nerves are immunopositive for  $G_{i2}\alpha$ ,  $G_{o}\alpha$ , and  $\beta\gamma$  (arrowheads), but they are almost immunonegative for  $G_{i1}\alpha$ . The glomeruli are intensely immunopositive for  $G_{i2}\alpha$  in the anterior region (G\*) but only faintly so in the posterior region (G). By contrast, the glomeruli are intensely immunopositive for  $G_{o}\alpha$  in the posterior region (G) but only weakly so in the anterior region (G\*). The glomeruli are weakly immunopositive for  $G_{i1}\alpha$ , but the immunoreactivity is greater in the posterior region (G) than in the anterior region (G\*). All glomeruli in the accessory olfactory bulb are equally immunopositive for  $\beta\gamma$ . The molecular layers of the anterior and the posterior regions (M) are equally immunopositive for each antibody. Scale bar, 100  $\mu$ m.

higher than that at 2 weeks after birth  $(14.6 \pm 0.8 \text{ pmol/mg})$  protein). The concentration of  $G_o\alpha$  in both the main and the accessory olfactory bulbs increased during postnatal development. However, the rapid increase in concentration of  $G_o\alpha$  in the main olfactory bulb occurred earlier than the increase in concentration of  $G_{i2}\alpha$  in the accessory olfactory bulb; the concentration of  $G_o\alpha$  in the main olfactory bulb of rats at 12 weeks after birth  $(59.4 \pm 1.9 \text{ pmol/mg protein})$  was only 1.2-fold higher than that at 2 weeks after birth  $(47.7 \pm 2.3 \text{ pmol/mg protein})$ . The concentration of  $G_o\alpha$  in the accessory olfactory bulb increased with similar kinetics to the increase in the concentration of  $G_o\alpha$  in the main olfactory bulb, but it reached the adult level slightly later. In addition, there was no significant difference in the concentration of  $G_{i2}\alpha$  between adult males and females (data not shown).

#### Discussion

The use of urine to mark the environment is a well-known behavioral trait of many vertebrates. The vomeronasal organ is involved in the detection of nonvolatile odorants in the urine (Wysocki et al., 1980), and major histocompatibility complex antigens are excreted in rat urine as olfactory recognition cues (Singh et al., 1987) for a wide range of phenotypic characteristics, such as species, strain, and sex of the subject (Beauchamp et al., 1985). In yeast, the mating pheromone response is activated by a G-protein-mediated signaling pathway in which  $\beta \gamma$  is previ-

ously thought to be the active transducer of the signal in *Saccharomyces cerevisiae* (Whiteway et al., 1989). By contrast, it is reported recently that the  $\alpha$ -subunit appears to function as a positive factor that transmits the signal from the mating-factor receptors to a downstream effector(s) in *Schizosaccharomyces pombe* (Obara et al., 1991).

In the mammalian brain,  $G_{i2}\alpha$  appears to be a minor G-protein and it is distributed almost uniformly in the neuropil-rich regions (Asano et al., 1989a, 1990). By contrast, the concentration of  $G_{i2}\alpha$  in the accessory olfactory bulb in the present study is the highest found by us in any tissues or cells examined (Asano et al., 1989a,b). Therefore, the present results suggest that  $G_{i2}\alpha$  is one of the major G-proteins in the accessory olfactory bulb and plays a crucial role in signal transduction, as is the case for transducin (Gilman, 1987) in the retina and for  $G_{olf}$  (Jones and Reed, 1989) in the olfactory epithelium.

During the development of the cerebral cortex in rats, the concentration of  $G_{i2}\alpha$  was constant but that of  $G_o\alpha$  increased markedly and approached the adult level within 3 weeks after birth (Asano et al., 1988). The developmental changes in the concentrations of  $G_{i2}\alpha$  and  $G_o\alpha$  in the main olfactory bulb in the present study were similar to the results obtained in the cerebral cortex. We have also demonstrated a significant increase in the concentration of  $G_o\alpha$ , with little change in that of  $G_{i2}\alpha$ , in pheochromocytoma PC12 cells and in neuroblastoma × glioma hybrid NG108-15 cells that had been stimulated to

Figure 3. Immunoassay of  $G_{12}\alpha$  and  $G_{0}\alpha$  in the two regions of the accessory olfactory bulb. The concentration of  $G_{12}\alpha$  in the anterior region is higher than that in the posterior region. By contrast, the concentration of  $G_{0}\alpha$  in the posterior region is higher than that in the anterior region. Each column shows the mean  $\pm$  SE of results from five samples.

differentiate by treatment with NGF or forskolin (Asano et al., 1989b). From these results, it appears that the concentration of  $G_{\circ}\alpha$  increases during the maturation of nervous tissues while that of  $G_{\circ 2}\alpha$  does not. By contrast, the concentration of  $G_{\circ 2}\alpha$  in the accessory olfactory bulb increased during puberty and reached the adult level at 12 weeks after birth, lagging behind the maturation of nervous tissues. Therefore, our findings support the hypothesis that the gene for  $G_{\circ 2}$  is expressed during sexual maturation in the accessory olfactory bulb and plays an important role in the perception of intersex smells, such as those associated with pheromones. It is also possible that the concentration of  $G_{\circ}\alpha$  in the accessory olfactory bulb increases further as a result of sexual maturation.

The present results also indicate that the accessory olfactory bulb consists of two regions; the anterior region is rich in  $G_{i2}\alpha$  and the posterior region is rich in  $G_{o}\alpha$ . It has been reported that carbohydrate antigens that are detectable with monoclonal antibodies are localized heterogeneously in the accessory olfactory bulb (Schwarting and Crandall, 1991) and in the vomeronasal nerve (Mori, 1987), as was observed in the case of G-proteins in the present study. It seems likely, therefore, that the vomeronasal-accessory pathway consists of at least two systems, one in which  $G_{i2}$  is involved and one in which  $G_{o}$  is involved.

#### References

Asano T, Semba R, Ogasawara N, Kato K (1987) Highly sensitive immunoassay for the α subunit of the GTP-binding protein G₀ and its regional distribution in bovine brain. J Neurochem 48:1617–1623. Asano T, Kamiya N, Semba R, Kato K (1988) Ontogeny of the GTP-binding protein G₀ in rat brain and heart. J Neurochem 51:1711–1716.

Asano T, Morishita R, Semba R, Itoh H, Kaziro Y, Kato K (1989a) Identification of lung major GTP-binding protein as G<sub>12</sub> and its distribution in various rat tissues determined by immunoassay. Biochemistry 28:4749–4754.

Asano T, Morishita R, Sano M, Kato K (1989b) The GTP-binding

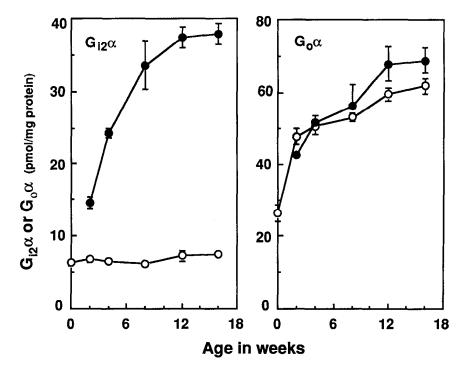


Figure 4. The changes in concentrations with development of  $G_{12}\alpha$  and  $G_{o}\alpha$  in the accessory (solid circles) and the main olfactory bulb (open circles). The concentration of  $G_{12}\alpha$  in the accessory olfactory bulb increases at puberty and reaches the adult level 12 weeks after birth, while that in the main olfactory bulb remains relatively constant. Concentrations of  $G_{o}\alpha$  in the accessory and the main olfactory bulb increased in parallel during postnatal development. Each point represents the mean  $\pm$  SE of results from five animals.

- proteins,  $G_o$  and  $G_{12}$ , of neural cloned cells and their changes during differentiation. J Neurochem 53:1195–1198.
- Asano T, Shinohara H, Morishita R, Kato K (1990) Immunochemical and immunohistochemical localization of the G protein  $G_{i1}$  in rat central nervous tissues. J Biochem (Tokyo) 108:988–994.
- Beauchamp GK, Yamazaki K, Boyse EA (1985) The chemosensory recognition of genetic individuality. Sci Am 253:66-72.
- Gilman AG (1987) G proteins: tranducers of receptor-generated signals. Annu Rev Biochem 56:615-649.
- Goldsmith P, Backlund PS Jr, Rossiter K, Carter A, Milligan G, Unson CG, Spiegel AM (1988) Purification of heterotrimeric GTP-binding proteins from brain: identification of a novel form of G<sub>o</sub>. Biochemistry 27:7085–7090.
- Hescheler J, Rosenthal W, Trautwein W, Schultz G (1987) The GTP-binding protein, G<sub>o</sub>, regulates neuronal calcium channels. Nature 325: 445–447.
- Jones DT, Reed RR (1989) G<sub>oir</sub>: an olfactory neuron specific-G protein involved in odorant signal transduction. Science 244:790–795.
- Kanaho Y, Katada T, Hoyle K, Crooke ST, Stadel JM (1989) Immunochemical comparison of pertussis toxin substrates in brain and peripheral tissues. Cell Signal 1:553-560.
- Kikuchi A, Kozawa O, Kaibuchi K, Katada T, Ui M, Takai Y (1986) Direct evidence of involvement of a guanine nucleotide-binding protein in chemotactic peptide-stimulated formation of inositol biphosphate and triphosphate in differentiated human leukemic (HL-60) cells. J Biol Chem 261:11558-11562.
- Mania-Farnell B, Farbman AI (1990) Immunohistochemical localization of guanine nucleotide-binding proteins in rat olfactory epithelium during development. Dev Brain Res 51:103-112.
- McCotter RE (1912) The connection of the vomeronasal nerves with the accessory olfactory bulb in the opossum and other mammals. Anat Rec 6:299–318.
- Meredith M (1986) Vomeronasal organ removal before sexual experience impairs male hamster mating behavior. Physiol Behav 36:737–744.
- Mori K (1987) Monoclonal antibodies (2C5 and 4C9) against lactoseries carbohydrates identify subsets of olfactory and vomeronasal receptor cells and their axons in the rabbit. Brain Res 408:215–221.
- Moriarty TM, Padrell E, Carty DJ, Omri G, Landau EM, Iyengar R (1990) G<sub>o</sub> protein as signal transducer in the pertussis toxin-sensitive phosphatidylinositol pathway. Nature 343:79–82.

- Morishita R, Kato K, Asano T (1988) Major pertussis-toxin-sensitive GTP-binding protein of bovine lung. Purification, characterization and production of specific antibodies. Eur J Biochem 174:87–94.
- Nakane P (1975) Recent progress in peroxidase-labeled antibody method. Ann NY Acad Sci 254:203–211.
- Neer EJ, Clapham DE (1988) Roles of G protein subunits in transmembrane signalling. Nature 333:129-134.
- Obara T, Nakafuku M, Yamamoto M, Kaziro Y (1991) Isolation and characterization of a gene encoding a G-protein α subunit from *Schizosaccharomyces pombe*: involvement in mating and sporulating pathways. Proc Natl Acad Sci USA 88:5877–5881.
- Powers JB, Winans SS (1971) Vomeronasal organ: critical role in mediating sexual behavior of the male hamster. Science 187:961-963
- Saito TR, Moltz H (1986) Sexual behavior in the female rat following removal of the vomeronasal organ. Physiol Behav 38:81-87.
- Schaffner W, Weissmann C (1973) A rapid, sensitive, and specific method for the determination of protein in dilute solution. Anal Biochem 56:502-514.
- Schmechel DE, Brightman MW, Marangos PJ (1980) Neurons switch from non-neuronal enolase to neuron-specific enolase during differentiation. Brain Res 190:195-214.
- Schwarting GA, Crandall JE (1991) Subsets of olfactory and vomeronasal sensory epithelial cells and axons revealed by monoclonal antibodies to carbohydrate antigens. Brain Res 547:239–248.
- Semba R, Asano T, Kato K (1990) Physiological expression of neural marker proteins in the heart of young rats. Dev Brain Res 54:217–220
- Singh PB, Brown RE, Roser B (1987) MHC antigens in urine as olfactory recognition cues. Nature 327:161-164.
- VanDongen AMJ, Codina J, Olate J, Mattera R, Joho R, Birnbaumer L, Brown AM (1988) Newly identified brain potassium channels gated by the guanine nucleotide binding protein G<sub>o</sub>. Science 242: 1433-1437.
- Whiteway M, Hougan L, Dignard D, Thomas DY, Bell L, Saari GC, Grant FJ, O'hara P, MacKay VL (1989) The STE4 and STE18 genes of yeast encode potential  $\beta$  and  $\gamma$  subunits of the mating factor receptor-coupled G protein. Cell 56:467-477.
- Wysocki CJ, Wellington JL, Beauchamp GK (1980) Access of urinary nonvolatiles to the mammalian vomeronasal organ. Science 207:781–783.