

# Aluminum Modifies the Properties of Alzheimer's Disease PHF $\tau$ Proteins *in vivo* and *in vitro*

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Hyperphosphorylated adult human CNS  $\tau$  (PHF $\tau$  or A68) forms paired helical filaments (PHFs) in neurofibrillary tangles (NFTs), neuropil threads, and dystrophic neurites associated with senile plaques (SPs) during the progression of Alzheimer's disease (AD). While amyloid fibrils in SPs are composed of  $\beta$ -amyloid (A $\beta$ ), NFTs and SPs contain similar associated components such as ubiquitin,  $\alpha_1$ -antichymotrypsin (ACT), apolipoprotein E (ApoE), heparan sulfate proteoglycans (HSPGs), and aluminum salts. Thus, SPs and NFTs may result from specific interactions between PHF $\tau$ , A $\beta$ , and these other components. In fact, intracerebral injections of PHF $\tau$  induce co-deposits of A $\beta$ , ACT, and ubiquitin (Shin et al., 1993). To examine this issue further, we probed interactions between PHF $\tau$ , aluminum salts, and other plaque and tangle components.

We investigated *in vivo* interactions of PHF $\tau$  and aluminum chloride (AlCl<sub>3</sub>) with other plaque and tangle components by injecting PHF $\tau$  with and without AlCl<sub>3</sub> into the rodent brain. PHF $\tau$  co-injected with AlCl<sub>3</sub> formed aggregates that persisted much longer in the rat brain, and induced longer-lived co-deposits of A $\beta$ , ubiquitin, ACT, and ApoE than PHF $\tau$  alone. Injections of PHF $\tau$  with AlCl<sub>3</sub> also induced neurons near the injection site to acquire PHF $\tau$ -like properties as monitored with antibodies (AT8, T3P, PHF1) that recognize defined PHF $\tau$  epitopes containing phosphoserine residues (Ser<sup>202</sup>, Ser<sup>396</sup>, Ser<sup>404</sup>). Injections of AlCl<sub>3</sub> alone as well as injections of normal adult and fetal CNS  $\tau$ , several different synthetic peptides, neurofilament proteins, ACT, HSPGs, or ApoE with and without AlCl<sub>3</sub> failed to induce co-deposits of A $\beta$  or alter the immunoreactivity of  $\tau$  in rodent neurons.

To determine if aluminum salts interact directly and specifically with PHF $\tau$  *in situ*, we pretreated sections of AD hippocampus with 10 mM AlCl<sub>3</sub> and then probed these sections by immunohistochemistry with antibodies to PHF $\tau$  as well as

to a number of other plaque and tangle components. Preincubation of these sections with AlCl<sub>3</sub> diminished PHF $\tau$  immunoreactivity in NFTs and SPs using the PHF $\tau$ -specific antibodies AT8, T3P, and PHF1, while the immunoreactivity of other plaque and tangle proteins (A $\beta$ , ubiquitin, ACT, HSPGs, ApoE) was not abolished. We also examined the effects of AlCl<sub>3</sub> on PHF $\tau$  and normal adult human CNS  $\tau$  *in vitro*. AlCl<sub>3</sub> had no effect on normal adult human CNS  $\tau$ , while increasing concentrations of AlCl<sub>3</sub> (from 0.1 to 1.0 mM) induced PHF $\tau$  to aggregate at the top of the stacking gel, and at high concentrations (0.3 and 1.0 mM) of AlCl<sub>3</sub>, PHF $\tau$  completely failed to enter the gel.

These studies suggest that aluminum binds to PHF $\tau$ , induces these proteins to aggregate, and retards their proteolysis. Further, since intracerebral injections of PHF $\tau$  with and without AlCl<sub>3</sub> in rats appear uniquely capable of inducing co-deposits of a number of proteins found in authentic AD SPs and NFTs (including A $\beta$ , ubiquitin, ACT, and ApoE), we speculate that the contributions of PHF $\tau$  to plaque and tangle formation in AD may be modulated by aluminum.

**[Key words: aluminum, tau proteins, PHFs,  $\beta$ -amyloid, Alzheimer's disease, neurofibrillary tangles]**

The major histopathologic abnormalities that characterize the brains of patients with sporadic and familial Alzheimer's disease (AD) as well as elderly individuals with Down's syndrome include an excess of neurofibrillary lesions, amyloid-rich senile plaques (SPs), and the massive loss of telencephalic neurons (for recent reviews, especially to literature prior to 1990, see Kosik, 1992; Lee and Trojanowski, 1992; Goedert, 1993; Hardy, 1993; Mullan and Crawford, 1993; Selkoe, 1993; Trojanowski et al., 1993b, 1994). The neurofibrillary lesions in AD are composed of paired helical filaments (PHFs) that are biochemically and structurally distinct from the amyloid fibrils in SPs. PHFs accumulate with straight filaments and amorphous material in perikaryal neurofibrillary tangles (NFTs), dystrophic neurites associated with SPs, and in neuropil threads (NTs) throughout the neuropil. PHFs and the straight filaments that dominate ultrastructural images of neurofibrillary lesions are formed from hyperphosphorylated adult CNS  $\tau$  proteins known as PHF $\tau$  or A68 (for reviews, see Goedert et al., 1993; Trojanowski et al., 1993b, 1994). In contrast, the subunit proteins of amyloid fibrils are 39–43 amino acid (aa) long  $\beta$ -amyloid peptides (A $\beta$ ) derived from one or more larger, alternatively spliced amyloid precursor proteins (APPs) encoded by a gene on chromosome 21 (for detailed reviews, see Hardy, 1993; Mullan and Crawford, 1993; Selkoe, 1993). SPs are sites at which abundant accumulations of both PHFs and amyloid fibrils converge (Trojanowski et al.,

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**Table 1. Summary of injections and animals**

Sample injected	Postinjection survivals				
	1 hr	1–2 days	1 week	2/4 weeks	8 weeks
PHF $\tau$	1	4*	3		
PHF $\tau$ + AlCl <sub>3</sub>		4*	4	4/4	2
A $\beta$		2*			
A $\beta$ + AlCl <sub>3</sub>		2*			
ACT	2	3			
ACT + AlCl <sub>3</sub>		2			
APOE		2*			
APOE + AlCl <sub>3</sub>		2*			
APOE3		2*			
APOE3 + AlCl <sub>3</sub>		2*			
APOE4		2*			
APOE4 + AlCl <sub>3</sub>		2*			
HSPG		2*			
HSPG + AlCl <sub>3</sub>		2*			
A $\tau$		2*			
A $\tau$ + AlCl <sub>3</sub>		4*			
F $\tau$		4*			
F $\tau$ + AlCl <sub>3</sub>		2*	2		
Syn $\tau$		5			
NF-H		2			
NF-H + AlCl <sub>3</sub>			2	2/0	
AlCl <sub>3</sub>	2	2	2	2/2	2

This table summarizes the injected samples and the postinjection survival times of the animals studied here. The first column lists the component that was injected, while the subsequent columns indicate the postinjection survival times. The numbers in each column indicate the number of animals studied at each time point. The 2 and 4 week survival times (2/4 weeks) are grouped together in the same column. In at least two of the rats among the groups of animals identified by an asterisk, bilateral injections were performed in which the protein sample alone was injected into one hemisphere, and the same protein sample plus AlCl<sub>3</sub> was injected in the opposite hemisphere. A  $\tau$ , adult CNS  $\tau$ ; F  $\tau$ , fetal CNS  $\tau$ ; Syn  $\tau$ , synthetic  $\tau$  peptides; UB, ubiquitin. Other abbreviations are the same as those used in the text.

1993b, 1994). Despite the presence of APP mutations in rare cases of familial AD, the abundance of NFTs correlates better with the dementia in AD than the density of amyloid plaques (Dickson et al., 1991; Arriagada et al., 1992; Goedert, 1993).

Although PHFs and A $\beta$  amyloid fibrils are the most distinct structural elements in NFTs and SPs, respectively, a number of other components selectively accumulate in SPs and NFTs, including ubiquitin (Mori et al., 1987; Perry et al., 1987; Shaw and Chau, 1988),  $\alpha_1$ -antichymotrypsin (ACT) (Abraham et al., 1988; Gollin et al., 1992), heparan sulfate proteoglycans (HSPGs) (Snow et al., 1988; Perlmutter et al., 1991; Perry et al., 1991), and apolipoprotein E (ApoE), a risk factor for sporadic and familial AD (Namba et al., 1991; Wisniewski and Frangione, 1992; Saunders et al., 1993; Schmechel et al., 1993). In addition, aluminum has been detected in NFTs and/or tangle-bearing neurons (Perl and Brody, 1980; Garruto et al., 1984; Picardo et al., 1988; Good et al., 1992; Perl and Good, 1992), as well as in some SPs (Candy et al., 1986, 1992; but see also Stern et al., 1986, and Landsberg et al., 1992, for negative studies). Despite the presence of aluminum and other proteins in NFTs and SPs, the contribution of these components to the pathogenesis of plaques and tangles is not known. However, it has been speculated that components in SPs other than A $\beta$  might contribute to the formation of amyloid plaques in AD (Snow et al., 1988,

1994; Greenberg et al., 1991; Perry, 1993; Perry et al., 1993; Shin et al., 1993; Trojanowski et al., 1993b, 1994).

To develop model systems for studying interactions between different plaque and tangle components, we showed that injections of purified PHF $\tau$  into the rodent brain induced long-lived co-deposits of A $\beta$ , ubiquitin, and ACT at the injection site (Shin et al., 1993). Thus, the convergence of PHF $\tau$  and A $\beta$  in authentic cortical SPs and NFTs (Hyman et al., 1989; Shin et al., 1989; Arai et al., 1990; Spillantini et al., 1990; Perry et al., 1992) may not reflect the stochastic outcome of separate pathological processes. Instead, it is possible that interactions of PHF $\tau$  with other tangle components, including A $\beta$  and other A $\beta$  precursor protein domains (Arai et al., 1990; Gollin et al., 1992; Perry et al., 1993), induce the aggregation of PHFs into perikaryal NFTs. Further, the release of PHF $\tau$  from degenerating neurons in the AD brain may contribute to the formation of SPs by interacting with A $\beta$  secreted by neural cells (Shin et al., 1993). To explore the *in vivo* interactions between PHF $\tau$ , A $\beta$ , aluminum, and other proteins in SPs and NFTs, we injected PHF $\tau$  with aluminum chloride (AlCl<sub>3</sub>) into the rat brain and examined the injection sites for the presence of several proteins found in authentic SPs and NFTs. In complementary studies, we also probed interactions of PHF $\tau$  with AlCl<sub>3</sub> *in situ* and *in vitro* using AD brain samples.

## Materials and Methods

*Intracerebral injections of PHF $\tau$  and other proteins with and without aluminum.* Three different preparations of PHF $\tau$  were purified from the brains of three different AD patients as described earlier (Bramblett et al., 1993; Shin et al., 1993) for intracerebral injection. Briefly, PHF $\tau$  was purified from Sarkosyl-insoluble material and rendered water soluble by extraction with 2 M guanidine isothiocyanate followed by dialysis against water. Normal adult, human CNS  $\tau$  and fetal  $\tau$  from late gestational age normal fetuses (Tohyama et al., 1991; Lee et al., 1993; Yachnis et al., 1993) were prepared as described (Bramblett et al., 1992, 1993). The normal adult and fetal brains as well as the AD brains were evaluated histopathologically and assigned to the control or AD group using consensus criteria for the diagnosis of AD (Khachaturian, 1985) as previously described (Kosik et al., 1988; Arai et al., 1990; Schmidt et al., 1990, 1991; Tohyama et al., 1991; Bramblett et al., 1992; Lee et al., 1993; Yachnis et al., 1993). Human high-molecular-weight neurofilament proteins (NF-H) were purified as described (Balin et al., 1991), and recombinant human ACT was a generous gift of H. Rubin and B. Cooperman, while purified HSPGs and human ApoE (representing a mixture of ApoE2, ApoE3, and ApoE4) were obtained from Becton Dickinson and Chemicon, respectively. Recombinant human ApoE3 and ApoE4 were obtained from Panvera. Peptides based on A $\beta$ <sub>1–42</sub>, as well as on several domains in CNS  $\tau$  were synthesized and characterized as described (Stern et al., 1989; Lee et al., 1991; Lang et al., 1992), and samples of these peptides also were used for injection into rat brains. These synthetic  $\tau$  peptides included sequences extending from aa 389 to 402 (according to the numbering system for the longest CNS  $\tau$  isoform of Goedert et al., 1989) containing Ser<sup>396</sup> in a phosphorylated state (GAEIVYKSPVVSGD), a microtubule (MT)-binding repeat (aa 256–273; VKSKIGSTENLKHQPGGG), and a carboxy-terminal  $\tau$  domain containing phosphorylated Ser<sup>416</sup> (aa 405–421; PRHLSNVSTGSIDMVD). AlCl<sub>3</sub> (20 mM) in 0.5 M Tris pH 6.5 was made as a stock solution and kept at 4°C. The protein samples used for injection were made up with and without added AlCl<sub>3</sub>. For co-injection of proteins with AlCl<sub>3</sub>, the protein samples were mixed at 4°C with the stock solution of AlCl<sub>3</sub> (resulting in a final concentration of 10 mM AlCl<sub>3</sub>/0.25 M Tris pH 6.5) just prior to injection. All of these freshly prepared solutions of protein and aluminum were injected within 2 hr after preparation. Approximately 3  $\mu$ g of PHF $\tau$ , normal adult CNS  $\tau$ , fetal  $\tau$ , HSPGs, synthetic A $\beta$ , ApoE, and NF-H or 9  $\mu$ g of ACT all in 3  $\mu$ l of PBS or in 3  $\mu$ l of 10 mM AlCl<sub>3</sub>/0.25 M Tris buffer pH 6.5 was injected separately at each site. Further, 10  $\mu$ g of the synthetic  $\tau$ <sub>389–402</sub> peptide, 80  $\mu$ g of the synthetic  $\tau$ <sub>256–273</sub> peptide, and 45  $\mu$ g of the synthetic  $\tau$ <sub>405–421</sub> peptide each in 3  $\mu$ l volume of vehicle also were injected at separate sites in different rats.

**Table 2. Summary of antibodies**

Antibody	Specificity	Dilution	Reference
ALZ50	$\tau_{2-10}$	1:10	Wolozin et al., 1988; Goedert et al., 1991
T14	$\tau_{141-178}$	1:500	Kosik et al., 1988; Trojanowski et al., 1989
T46	$\tau_{404-414}$	1:500	Kosik et al., 1988; Trojanowski et al., 1989
Tau1	$\tau$ npSer <sup>202</sup>	1:500d	Binder et al., 1985; Kosik et al., 1989; Goedert et al., 1993
AT8	PHF $\tau$ pSer <sup>202</sup>	1:500	Goedert et al., 1993
T3P*	PHF $\tau$ pSer <sup>396</sup>	1:100	Lee et al., 1991; Bramblett et al., 1993
PHF1	PHF $\tau$ pSer <sup>396&amp;404</sup>	1:2000	Greenberg et al., 1992; Lang et al., 1992; Bramblett et al., 1993
2332*	A $\beta_{1-42}$	1:4000	Shin et al., 1993
2134*	A $\beta_{1-40}$	1:100	Estus et al., 1992
1510	Ubiquitin	1:10000	Chemicon; Shaw et al., 1988
Anti-ACT*	ACT	1:1000	ICN; Abraham et al., 1988
Anti-ACT*	ACT	1:200	DAKO; Abraham et al., 1988
Anti-HSPG*	HSPG	1:100	Narindrasorasak et al., 1991
Anti-Apoe*	APoe	1:4000	Chemicon; Wisniewski et al., 1992
I062	APoe	1:100	Chemicon
RMO24	NF-H	1:10	Lee et al., 1987; Schmidt et al., 1987

This table summarizes the properties of the antibodies used in this study. The name of the antibody, its specificity, the dilution of the antibody used for immunohistochemistry, and the source and/or references on the properties of these antibodies are shown in the columns from left to right, respectively. All of the antibodies are mouse MAbs except for those identified by an asterisk, which are polyclonal antibodies. ALZ50, T14, and T46 are phosphate-independent antibodies to  $\tau$  that recognize fetal and adult human CNS  $\tau$  as well as PHF $\tau$ , but T14 does not recognize rat  $\tau$  proteins. Tau1 recognizes normal CNS  $\tau$  when Ser<sup>202</sup> is not phosphorylated (npSer<sup>202</sup>), but it only binds to PHF $\tau$  after enzymatic dephosphorylation. AT8, T3P, and PHF1 recognize fetal human CNS  $\tau$  and PHF $\tau$ , but not adult human CNS  $\tau$ . These MAbs bind to PHF $\tau$  when the serine residues identified in the table by the superscripts are phosphorylated (as indicated by pSer<sup>202</sup>, pSer<sup>396</sup>, pSer<sup>396&404</sup>). The numbering system for the aa sequences of the epitopes in  $\tau$  recognized by the anti- $\tau$  antibodies listed here (as indicated by the subscript numbers following the  $\tau$  symbol in the second column) is based on the largest human CNS  $\tau$  isoform (Goedert et al., 1989).

In control rats, the same volume of 10 mM AlCl<sub>3</sub>/0.25 M Tris buffer pH 6.5 alone was injected.

The injections of the different proteins into the rodent CNS were performed under anesthesia using procedures similar to those described previously (Trojanowski, 1983; Shin et al., 1993). In some experiments, unilateral injections of the protein samples with or without aluminum were performed at two separate sites in the right hemisphere. In other experiments, bilateral injections of matched protein samples with and without aluminum were performed at two separate sites in the right (protein sample with aluminum) and left (protein sample without aluminum) hemispheres. Following specified postinjection survival intervals, the rats were lethally anesthetized and perfused with PBS prior to removal and immersion fixation of the brains in 70% ethanol with 150 mM NaCl as described (Shin et al., 1993). The number of rats injected with different proteins as well as the postinjection survival times are summarized in Table 1. Finally, sections from a number of the rat brains ( $n = 36$ ) that had received injections of PHF $\tau$  or normal adult human CNS  $\tau$  in a previous study (Shin et al., 1993) were examined further by immunohistochemistry in the experiments described here.

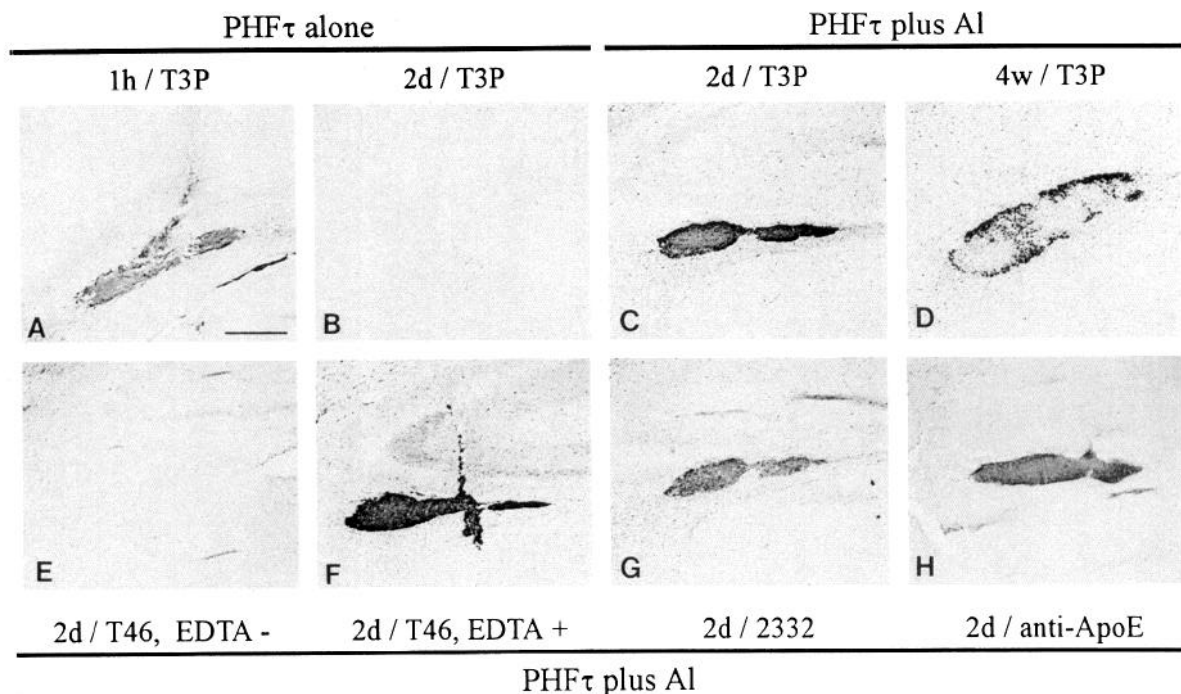
**Light microscopic immunohistochemical studies.** The ethanol-fixed rat brains were embedded in paraffin according to previously described infiltration schedules (Trojanowski et al., 1989), and serial 6- $\mu$ m-thick sections through the injection sites were prepared and examined by immunohistochemistry as outlined in several earlier publications (Schmidt et al., 1991; Tohyama et al., 1991; Bramblett et al., 1992; Shin et al., 1993). The panel of antibodies to normal adult and fetal CNS  $\tau$ , PHF $\tau$ , ubiquitin, ACT, A $\beta$ , HSPGs, ApoE, and NF-H that were used in this study is summarized in Table 2.

Since some PHF $\tau$  epitopes (e.g., the amino-terminal  $\tau$  epitope recognized by the T46 MAb) were not detected at all, or they were weakly immunoreactive in the aggregates that formed following injections of PHF $\tau$  plus AlCl<sub>3</sub> (see Results), we sought to determine if aluminum might bind to these epitopes and "mask" them. To do this, we treated rat brain sections at 37°C for 1 hr with chelating agents including 0.1 M EDTA in 0.1 M Tris pH 7.6, or 0.1 M desferrioxamine mesylate (DFO; Sigma) in 0.1 M Tris pH 7.6 prior to probing the sections with primary antibodies for immunohistochemical analysis. The rat brain sections

were analyzed with and without pretreatment with chelating solutions using antibodies to PHF $\tau$  as well as with antibodies to all of the other proteins examined in this study (see Table 2). In studies performed with the Tau1 MAb, rat brain sections were pretreated with DFO followed by enzymatic dephosphorylation of the sections using *Escherichia coli* alkaline phosphatase (type III-N, Sigma) as described earlier (Lee et al., 1986; Schmidt et al., 1987). Finally, absorption experiments were performed for several of the antibodies as described (Shin et al., 1991).

**Immunoelectron microscopic studies of PHF $\tau$  injection sites.** To monitor the ability of the guanidine isothiocyanate-treated, water-soluble PHF $\tau$  proteins to reassemble into filaments in the rat brain, immunoelectron microscopic studies were performed as reported elsewhere (Springall et al., 1984; Balin et al., 1991; Hill et al., 1991; V. M.-Y. Lee et al., 1991; J. H. Lee et al., 1993; Shin et al., 1993). These studies were carried out on sections through PHF $\tau$  accumulations produced in three additional rats that received injections of PHF $\tau$  alone. These rats survived 1 d postinjection, and they were perfused with buffer as described above. The brains were removed immediately thereafter and small pieces of tissue from the injection sites were excised and immersion fixed in 1% glutaraldehyde/2% paraformaldehyde in phosphate buffer for 4 hr, embedded in paraffin, and then sectioned as described above. Sequential sections were probed with the antibodies to PHF $\tau$  and normal CNS  $\tau$  summarized in Table 2 using the same immunohistochemical methods outlined earlier except that the peroxidase-antiperoxidase complex was replaced with an immunogold reporter system with and without silver intensification of the gold (5 nm) particles (Springall et al., 1984; Shin et al., 1993). Portions of the rat brain adjacent to the injection sites were excised, embedded in araldite, sectioned, and examined using a Hitachi 600 electron microscope.

**Assessment of interactions between aluminum salts and AD pathology in situ.** In complementary studies of the ability of aluminum salts to interact with proteins found in SPs and NFTs of the AD brain *in situ*, we prepared sections of hippocampus from four different AD brains and treated these sections with 10 mM AlCl<sub>3</sub> in 0.25 M Tris pH 6.5 at 37°C overnight prior to immunohistochemical analysis using the same antibodies described in Table 2. The diagnostic evaluation of these AD brains was performed as outlined above.



**Figure 1.** Photomicrographs of rat brains injected with PHF $\tau$  alone (*A* and *B*) or with PHF $\tau$  plus aluminum (*C–H*) at postinjection survival times of 1 hr (*A*), 2 d (*B*, *C*, and *E–H*), and 4 weeks (*D*). Some of the sections were not pretreated with chelators (*A*, *B*, and *E*), while others were pretreated with EDTA (*F*) or DFO (*C*, *D*, *G*, and *H*) for 1 hr prior to immunostaining with antibodies specific for PHF $\tau$  (T3P in *A–D*), normal adult, and fetal CNS  $\tau$  as well as PHF $\tau$  (T46 in *E* and *F*), A $\beta$  (2332 in *G*), or ApoE (*H*). The sections were counterstained with hematoxylin. All of the photomicrographs are at the same magnification. Scale bar (in *A*), 300  $\mu$ m.

**In vitro assessment of interactions between PHF $\tau$  and aluminum.** Samples from tangle-rich regions of the brains of three AD patients as well as brain samples from three neurologically and neuropathologically normal subjects were homogenized in reassembly buffer (0.1 M MES, 0.5 mM MgSO<sub>4</sub>, 1 mM EGTA, 2 mM dithiothreitol pH 6.8) containing 0.75 M NaCl, and a cocktail of protease inhibitors (0.5 M phenylmethylsulfonyl fluoride and TPCK, TLCK, leupeptin, pepstatin, and soybean trypsin inhibitor, each at 1 gm/ml) and centrifuged at 90,000  $\times$  g for 30 min. The resulting supernatants were boiled for 10 min, followed by centrifugation at 90,000  $\times$  g for 60 min, and then once again at 350,000  $\times$  g for 60 min. The final 350,000  $\times$  g supernatants from the AD brains (which contained an aqueous-soluble fraction of PHF $\tau$  as well as aqueous-soluble normal adult CNS  $\tau$ ; see Results for further details) and from the normal brains (which only contained aqueous-soluble normal adult CNS  $\tau$ ) were incubated with different concentrations of AlCl<sub>3</sub> (ranging from 0 to 1 mM) in 0.25 M Tris pH 6.5 at 37°C for 1 hr, and centrifuged at 90,000  $\times$  g for 30 min. The resultant supernatants and pellets were analyzed by 8.5% SDS-PAGE followed by immunoblot analysis with MABs specific for PHF $\tau$  (i.e., PHF1, T3P, AT8) and normal adult CNS  $\tau$  (i.e., Tau1) as described (Lee et al., 1991; Bramblett et al., 1992, 1993; Shin et al., 1993). The supernatants from the AD brains also were applied to nickel grids, negatively stained, and monitored by electron microscopy using a Hitachi 600 electron microscope for the presence of particulate material, straight filaments, or PHFs as outlined in previous reports (Balin et al., 1991; Lee et al., 1991).

## Results

### Co-injection of aluminum salts with PHF $\tau$ enhances the resistance of PHF $\tau$ to *in vivo* proteolysis

To investigate the effects of aluminum on the *in vivo* biological activities of PHF $\tau$ , we used a rodent model in which intracerebral injections of PHF $\tau$  have been shown to produce clustered aggregates of PHF $\tau$  that resist rapid proteolysis relative to normal adult CNS  $\tau$  and fetal human (Shin et al., 1993). Accordingly, several different preparations of PHF $\tau$  from different AD brains ( $n = 6$ ) were screened for their relative resistance to *in*

*vivo* proteolysis by injecting aliquots of each of the preparations into the rat CNS. We confirmed that all of these PHF $\tau$  isolates were more resistant to *in vivo* proteolysis than normal adult CNS  $\tau$ , which is eliminated almost entirely within 1 hr following intracerebral injection. However, the different PHF $\tau$  preparations resisted proteolysis to varying degrees; that is, different isolates remained in the rat brain injection site for periods of time ranging from <1 week to >4 weeks. The reasons for this variation are uncertain, but similar variations have been noted earlier (Shin et al., unpublished observations), and we took advantage of this variation to assess rapidly the effects of AlCl<sub>3</sub> on the *in vivo* proteolysis of PHF $\tau$ . Specifically, in the studies summarized below, we used one of the more rapidly proteolyzed PHF $\tau$  preparations that was eliminated routinely from the injection site within 1 week.

Injections of aliquots from this preparation of PHF $\tau$  without aluminum produced large aggregates of immunoreactive PHF $\tau$  at 1 hr postinjection (Fig. 1*A*), but most of these aggregates were removed from the injection site by 1 d postinjection, and only trace amounts of PHF $\tau$  were detected at 2 d to 1 week postinjection (Fig. 1*B*). As described earlier (Shin et al., 1993), co-deposits of A $\beta$  appeared in these aggregates at 1 hr postinjection, and this A $\beta$  immunoreactivity disappeared over the ensuing week (data not shown). When the same amount of the same PHF $\tau$  preparation (i.e., 3  $\mu$ g/3  $\mu$ l PBS) was co-injected with 10 mM AlCl<sub>3</sub>, aggregates of immunoreactive PHF $\tau$  also formed rapidly (within 1 hr of injection) in the rat brain. However, these PHF $\tau$  aggregates persisted for up to at 8 weeks postinjection (Fig. 1*C,D*), after which time they were eliminated from the injection site. When sections containing the injected PHF $\tau$  plus aluminum were pretreated with metal chelators (i.e., EDTA or DFO), the pretreatment enhanced the immunodetection of the

PHF $\tau$  aggregates with antibodies that recognize both normal  $\tau$  and PHF $\tau$  (e.g., compare Fig. 1E,F), or only PHF $\tau$  (see Fig. 1C,D), and this effect was most evident at the postinjection survival times of <1 week.

The effect of co-injecting aluminum with PHF $\tau$  and the effect of pretreating sections with chelators were replicated at each postinjection survival time (i.e., two or four different rats per time point) mentioned above (data not shown). These observations also were confirmed by injecting two different PHF $\tau$  samples with and without aluminum in opposite sides of the same rat brains ( $n = 2$ ) and examining these brains at 1 week postinjection survival times (data not shown). Thus, co-injection of aluminum with PHF $\tau$  dramatically enhanced the resistance of PHF $\tau$  to *in vivo* proteolysis, and this resulted in the prolonged persistence of PHF $\tau$  aggregates in the rat brain relative to the injections of aliquots of the same preparation of PHF $\tau$  without aluminum. Further, the "unmasking" of PHF $\tau$  epitopes in sections through the injection sites containing PHF $\tau$  plus aluminum by pretreatment with chelators suggests that aluminum remained closely associated with PHF $\tau$  in the injection site.

Previous studies showed that topographically separate PHF $\tau$  epitopes are eliminated from the PHF $\tau$  aggregates according to a distinct temporal sequence (Shin et al., 1993). For example, the carboxy-terminal  $\tau$  epitope containing phosphorylated Ser<sup>396</sup> (recognized by the PHF1 and T3P antibodies) persisted the longest (i.e., for many days to several weeks) in the rat brain, while amino-terminal  $\tau$  epitopes (recognized by the ALZ50 MAb) were eliminated shortly (i.e., within a few hours) after an injection of PHF $\tau$  alone (Shin et al., 1993). In view of these observations, we characterized the temporal profile for the elimination of PHF $\tau$  epitopes in the injection sites containing PHF $\tau$  plus aluminum. These studies showed that aggregates of PHF $\tau$  co-injected with aluminum retained immunoreactivity for the epitopes recognized by the PHF1, T3P, and AT8 (which recognizes the epitope containing phosphorylated Ser<sup>202</sup>) antibodies longer than any of the other epitopes studied here (i.e., >4 weeks postinjection; see Fig. 1D), while the amino-terminal epitope recognized by the ALZ50 MAb was eliminated far more rapidly (i.e., within <2 weeks postinjection; data not shown). However, all of the epitopes recognized by the antibodies to normal CNS  $\tau$  and PHF $\tau$  were eliminated much more slowly from the injection sites containing PHF $\tau$  plus aluminum compared to PHF $\tau$  alone (data not shown). Thus, the temporal profile for the elimination of topographically separate epitopes in PHF $\tau$  is similar for injections of PHF $\tau$  with and without aluminum, but the rate at which these epitopes is eliminated appears to be far more attenuated when aluminum is co-injected with PHF $\tau$ .

Previous studies also showed that PHF $\tau$  induced co-deposits of A $\beta$ , ubiquitin, and ACT at the injection sites (Shin et al., 1993). Further studies here confirmed these observations. In addition, we noted that injections of PHF $\tau$  alone also induced co-deposits of ApoE (data not shown). Similar co-deposits of A $\beta$  (Fig. 1G), ubiquitin, ACT, and ApoE (Fig. 1H) were seen following injections of PHF $\tau$  with aluminum, and these co-deposits remained at the injection site for prolonged periods of time (i.e., for 2 weeks postinjection) relative to injections of the same PHF $\tau$  preparation without aluminum. Specifically, co-deposits of A $\beta$ , ACT, ubiquitin, and ApoE were induced by injections of the same preparation of PHF $\tau$  without aluminum, but these co-deposits remained at the injection site for <1 d. Thus, injections of PHF $\tau$  plus aluminum lead to longer-lived

co-deposits of A $\beta$ , ubiquitin, ACT, and ApoE in the rodent brain than injections of the same preparation of PHF $\tau$  without aluminum.

#### *Specificity of in vivo co-deposition of A $\beta$ with PHF $\tau$*

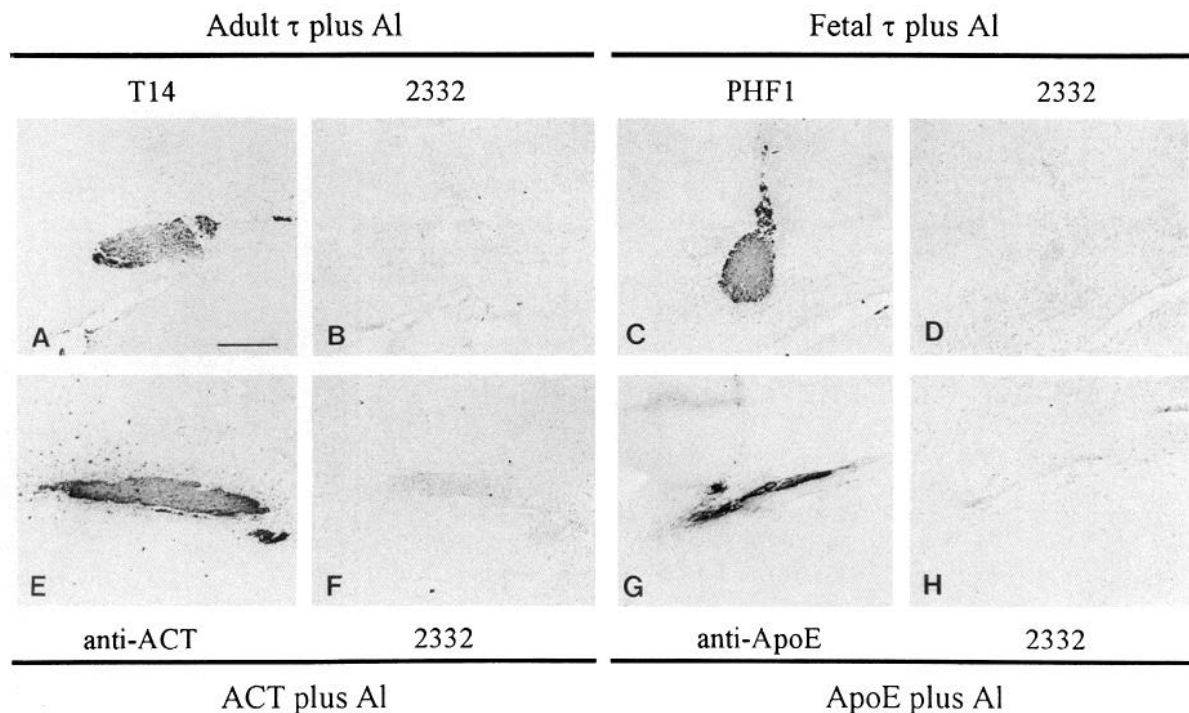
In contrast to PHF $\tau$ , adult and fetal human CNS  $\tau$  proteins injected into rat brains were eliminated far more rapidly (i.e., within 1 hr) from the injection site without forming aggregates (Shin et al., 1993). Further, injections of these  $\tau$  proteins as well as injections of various synthetic peptides failed to induce co-deposits of A $\beta$ , ubiquitin, ACT, and ApoE (data not shown). To determine if the proteolysis of adult and fetal human CNS  $\tau$  would be influenced by co-injecting these forms of  $\tau$  with aluminum, we performed additional injections of adult and fetal human CNS  $\tau$  with and without aluminum as described above. Both adult (Fig. 2A) and fetal (Fig. 2C) CNS  $\tau$  proteins co-injected with aluminum formed aggregates that were somewhat longer lived (e.g., as long as 2 d postinjection for adult  $\tau$  and 1 week for fetal  $\tau$ ) relative to injections of the same proteins alone in the contralateral hemisphere of the same rats (data not shown).

The aggregates of adult human CNS  $\tau$  plus aluminum retained only the T14 epitope (Fig. 2A) and not the epitopes recognized by the ALZ50 or T46 MAbs at 2 d postinjection. In contrast, the aggregates of human fetal  $\tau$  plus aluminum retained the epitopes recognized by all these  $\tau$ -specific antibodies in addition to the PHF $\tau$  and fetal  $\tau$ -specific epitopes recognized by PHF1 (Fig. 2C), T3P, and AT8 at 2 d postinjection. Among the epitopes recognized by the aforementioned antibodies, those detected by ALZ50, T14, and T46 were removed from the injection sites containing fetal  $\tau$  plus aluminum by 1 week postinjection, while the epitopes detected by PHF1, T3P, and AT8 were retained in these aggregates at this postinjection time point (data not shown). Thus, aluminum conveys a greater resistance to *in vivo* proteolysis for the epitopes recognized by PHF1, T3P, and AT8 in both PHF $\tau$  and in fetal human CNS  $\tau$ . However, neither the aggregates of adult CNS  $\tau$  plus aluminum (Fig. 2B) nor of fetal  $\tau$  plus aluminum (Fig. 2D) exhibited the ability to induce co-deposits of A $\beta$ . We also examined the effects of aluminum on several other proteins associated with plaques and tangles including ACT, HSPGs, and ApoE. All of the proteins injected without aluminum were rapidly cleared from the injection sites, while injections of ACT, HSPGs, or ApoE with aluminum delayed proteolysis and resulted in aggregates that were immunoreactive for either ACT (Fig. 2E), HSPGs (data not shown), or ApoE (Fig. 2G), respectively. However, none of these proteins injected with or without aluminum were able to induce co-deposits of A $\beta$  (Fig. 2F,H). Further, no deposition of A $\beta$  was seen in the rat brain following injections of aluminum alone (data not shown). Thus, only injections of PHF $\tau$  with and without aluminum were able to induce co-deposits of A $\beta$ . These findings, together with data demonstrating that other proteins (e.g., NF-H) and  $\tau$  peptides fail to induce co-deposits of A $\beta$  (Table 3), suggest that the ability to induce co-deposits of A $\beta$  in rat brain is a highly specific property that may be unique to PHF $\tau$ .

#### *Several components of plaques and tangles induce co-deposits of ApoE*

Although injections of AlCl<sub>3</sub> alone did not induce co-deposits of ApoE, injections of PHF $\tau$  with and without AlCl<sub>3</sub> and injections of adult or fetal CNS  $\tau$  as well as ACT, HSPGs, and NF-H with AlCl<sub>3</sub> did induce co-deposits of ApoE. Further, injections





**Figure 2.** Photomicrographs of rat brains injected with purified human adult CNS  $\tau$  (A and B), fetal CNS  $\tau$  (C and D), recombinant ACT (E and F), and purified human ApoE (G and H) at a 2 d postinjection survival time. Each of the injections also included  $\text{AlCl}_3$ . All of the sections were pretreated with DFO at 37°C for 1 hr followed by immunostaining with antibodies to adult and fetal CNS  $\tau$  as well as PHF $\tau$  (T14 in A), PHF $\tau$  (PHF1 in C), ACT (E), ApoE (G), and A $\beta$  (2332 in B, D, F, and H). The sections were counterstained with hematoxylin. All of the photomicrographs are at the same magnification. Scale bar (in A), 300  $\mu\text{m}$ .

**Table 3. Summary of injection data**

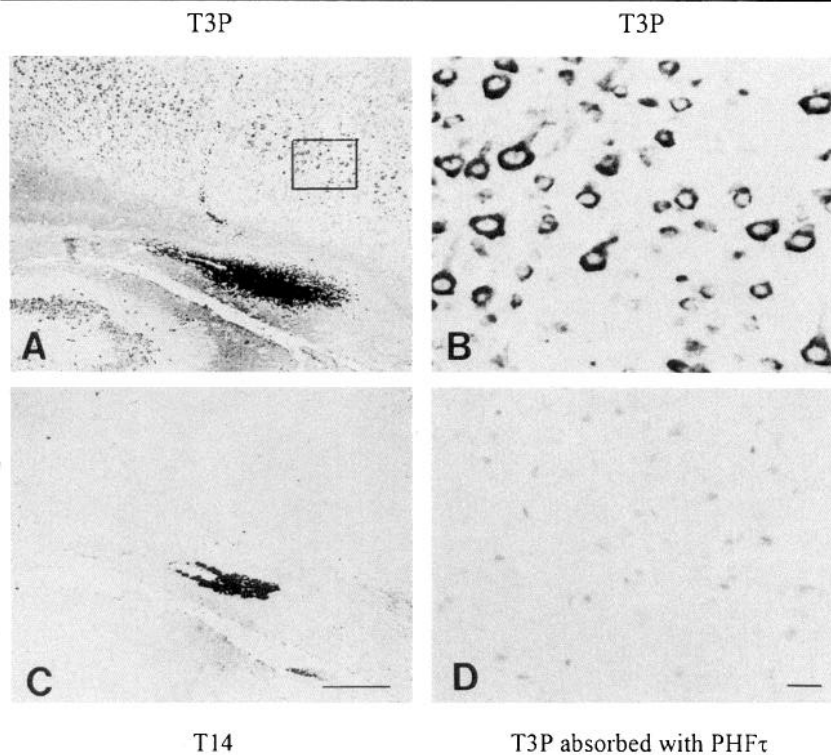
Sample injected	Protein co-deposits				
	A $\beta$	ApoE	HSPG	ACT	UB
PHF $\tau$	+	+	-	+	+
PHF $\tau$ + $\text{AlCl}_3$	+	+	-	+	+
A $\beta$	+	+	-	-	-
A $\beta$ + $\text{AlCl}_3$	+	+	-	-	-
ACT	-	-	-	-	-
ACT + $\text{AlCl}_3$	-	+	-	+	-
APOE	-	-	-	-	-
APOE + $\text{AlCl}_3$	-	+	-	-	-
APOE3	-	-	-	-	-
APOE3 + $\text{AlCl}_3$	-	+	-	-	-
APOE4	-	-	-	-	-
APOE4 + $\text{AlCl}_3$	-	+	-	-	-
HSPG	-	-	-	-	-
HSPG + $\text{AlCl}_3$	-	+	-	-	-
A $\tau$	-	-	-	-	-
A $\tau$ + $\text{AlCl}_3$	-	+	-	-	-
F $\tau$	-	-	-	-	-
F $\tau$ + $\text{AlCl}_3$	-	+	-	-	-
Syn $\tau$	-	-	-	-	-
NF-H	-	-	-	-	-
NF-H + $\text{AlCl}_3$	-	+	-	-	-
$\text{AlCl}_3$	-	-	-	-	-

This table summarizes the major results of the injection data described here. The first column lists the component that was injected while the subsequent columns indicate the rodent proteins that were (+) or were not (-) co-deposited with the injected material at the injection sites. The abbreviations are the same as those used in the text and in Table 1.

of A $\beta$ , with and without  $\text{AlCl}_3$ , produced large A $\beta$  aggregates. These injections also induced co-deposits of ApoE, but no co-deposits of the other plaque or tangle components studied here (data not shown). Thus, the induction of co-deposits of ApoE is not specific to PHF $\tau$  since several different plaque and tangle components were capable of inducing co-deposits of ApoE (see Table 2).

#### Acquisition of PHF $\tau$ -like immunoreactivity by rodent neurons adjacent to aggregates of PHF $\tau$ plus aluminum

Although injections of PHF $\tau$  with or without  $\text{AlCl}_3$  did not induce overt neuropathological changes in the adjacent rat brain (i.e., changes over and above those seen after injections of vehicle alone or following injections of the other material described here), we did observe alterations in immunoreactivity in neurons lying within <1000  $\mu\text{m}$  of the injection sites containing PHF $\tau$  plus aluminum. However, these changes were only observed in sections treated with chelators prior to immunohistochemical analysis. For example, preincubation of rat brain sections with DFO for 6 or more hours revealed that only those neurons near the injection sites containing PHF $\tau$  plus aluminum acquired *de novo* immunoreactivity for PHF1, AT8, and T3P (Fig. 3A,B). In addition, this same population of neurons also acquired Tau1 immunoreactivity only after the rat brain sections were pretreated with DFO and then subjected to enzymatic dephosphorylation with alkaline phosphatase (data not shown). However, using the same experimental procedures, these neurons were not immunostained with other phosphate-independent anti- $\tau$  antibodies such as ALZ50, T14 (Fig. 3C), and T46. Further, glial cells (which also contain low amounts of  $\tau$  proteins; Shin et al., 1991) did not acquire PHF $\tau$  immunoreactivity. Thus,

PHF $\tau$  plus Al

**Figure 3.** Photomicrographs of rat brains injected with PHF $\tau$  plus aluminum at 2 weeks postinjection. The sections were pretreated with DFO at 37°C overnight and probed with T3P (*A*, *B*, and *D*) or T14 (*C*). The T3P antiserum used in *D* was absorbed with PHF $\tau$  (20  $\mu$ g/ml), which eliminated staining of neurofibrillary lesions in sections of AD brain. The sections were counterstained with hematoxylin. Note that many normal-appearing rodent neurons exhibit PHF $\tau$  (T3P) immunoreactivity near the injection site following pretreatment of the sections with DFO (*A* and *B*), and that this immunoreactivity is abolished after absorption of T3P with PHF $\tau$  (*D*). In contrast, T14 stains the aggregates formed at the injection site of PHF $\tau$  plus aluminum, but not any of the rodent neurons. The photomicrographs in *A* and *C* are at the same magnification, as are the photomicrographs in *B* and *D*. The boxed area in *A* is shown at higher power in *B*. Scale bars: *C*, 300  $\mu$ m for *A* and *C*; *D*, 20  $\mu$ m for *B* and *D*.

injections of PHF $\tau$  with AlCl<sub>3</sub> induced changes in the phosphorylation state of normal adult rodent CNS  $\tau$  in neurons near the PHF $\tau$ /aluminum injection site such that several PHF $\tau$ -specific antibodies (i.e., PHF1, T3P, AT8) immunostained these neurons and this PHF $\tau$  staining was absorbed out by incubating antibodies with PHF $\tau$  (Fig. 3*D*). Notably, the detection of PHF $\tau$  immunoreactivity in the neurons described here required prolonged exposure to DFO. For example, similar results were not obtained following preincubations with DFO for brief periods of time (e.g., for 1 hr), or with the chelator EDTA. Finally, the rodent neurons that acquired *de novo* PHF $\tau$  immunoreactivity were confined to the region adjacent to the larger aggregates of PHF $\tau$  plus aluminum (i.e., within 1000  $\mu$ m of the injection site).

Using the same DFO pretreatment, we examined sections from the rat brains ( $n = 62$ ) that had received intracerebral injections of the other samples described above (e.g., CNS  $\tau$ , ACT, ubiquitin, A $\beta$ , ApoE, HSPGs, NF-H). In some of the brains ( $n = 3$ , 1 hr and 2 d postinjection survival times) from the series of rats described in an earlier study (Shin et al., 1993) that received injections of PHF $\tau$  alone, pretreatment with DFO revealed a small population of neurons that were immunostained by AT8, T3P, and PHF1. Further, these neurons also acquired Tau1 immunoreactivity following enzymatic dephosphorylation with alkaline phosphatase after initial treatment of the sections with DFO. These neurons were also confined to the injection site and were most abundant in the injection sites that contained large aggregates of PHF $\tau$ . In contrast, no PHF $\tau$  immunoreactivity was induced in rat neurons surrounding injection sites containing normal adult or fetal human CNS  $\tau$ , ACT, HSPGs, ApoE, or NF-H with and without AlCl<sub>3</sub>, and injections of AlCl<sub>3</sub> alone did not produce similar changes in protein immunoreactivity (data not shown).

Taken together, the observations from all of the rats studied

here are most compatible with one of the following possibilities: (1) PHF $\tau$  injected with aluminum is capable of inducing the hyperphosphorylation of Ser<sup>202</sup>, Ser<sup>396</sup>, and Ser<sup>404</sup> in rodent CNS  $\tau$  proteins by activating kinases and/or blocking phosphatases; (2) PHF $\tau$  (or proteolytic fragments thereof) is internalized with the co-injected aluminum by rat neurons and that the PHF $\tau$  epitopes recognized by T3P, AT8, and PHF1 are protected from degradation within neurons by aluminum.

#### Immunoelectron microscopic studies of PHF $\tau$ injection sites

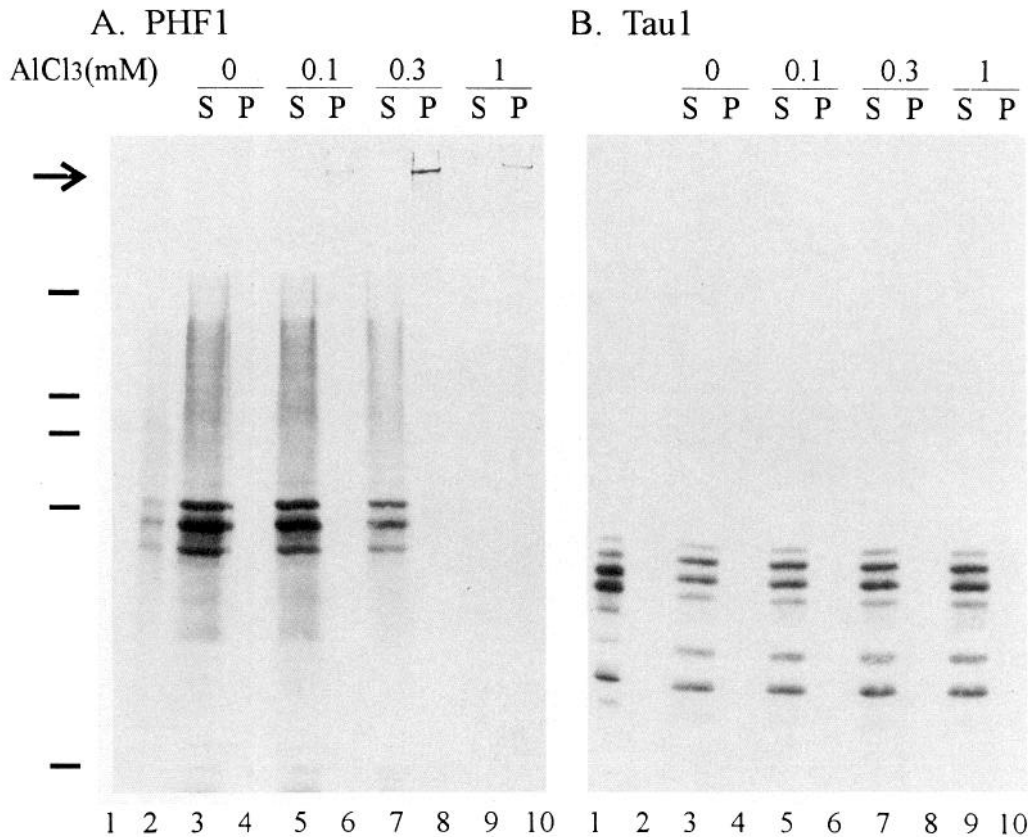
Previous studies showed that PHF $\tau$  proteins do not reassemble into PHFs or straight filaments *in vitro* following treatment with guanidine isothiocyanate, which is used to render these proteins water soluble for MT-binding studies (Bramblett et al., 1993) or for injection into the rat brain (Shin et al., 1993). Although rare T3P-positive straight filaments (~10–25 nm in diameter) were observed in one of two rats that survived 1 d following injections of PHF $\tau$  alone (Shin et al., 1993), examination of three additional rats injected with PHF $\tau$  without AlCl<sub>3</sub> did not reveal PHF $\tau$ -immunoreactive filaments (data not shown). Additional *in vitro* electron microscopic studies of the guanidine isothiocyanate-treated PHF $\tau$  preparations prior to injection into the rat brain also failed to reveal reassembled PHFs. Hence, the ability of PHF $\tau$  proteins to reassemble into filaments in the rat brain following guanidine isothiocyanate treatment appears to be very limited under the experimental conditions described here. Finally, recent *in vitro* studies indicate that aluminum does not induce  $\tau$  to form PHFs (Scott et al., 1993).

#### Exogenous aluminum salts specifically interact with PHF $\tau$ in AD neurofibrillary lesions

To determine if aluminum salts interact with proteins in AD lesions *in situ*, we incubated 6- $\mu$ m-thick paraffin sections of the







**Figure 5.** Immunoblot analysis of the effects of aluminum on PHF $\tau$  versus adult human CNS  $\tau$ . Aliquots of the same 350,000  $\times$  g supernatant prepared from an AD brain were incubated with 0 mM (lanes 3 and 4), 0.1 mM (lanes 5 and 6), 0.3 mM (lanes 7 and 8), 1.0 mM (lanes 9 and 10) AlCl<sub>3</sub> at 37°C for 1 hr followed by centrifugation at 90,000  $\times$  g for 30 min. For each of these samples, the resulting supernatants (lanes 3, 5, 7, and 9) and pellets (lanes 4, 6, 8, and 10) were analyzed using 8.5% SDS-PAGE and immunoblots. After transfer to nitrocellulose paper, the gel replicas were probed with PHF1 (A) and Tau1 (B). As controls, lanes 1 and 2 were loaded with a sample of normal adult CNS  $\tau$  purified from a control brain (lane 1 in A and B) as well as a sample of PHF $\tau$  purified from another AD brain (lane 2 in A and B). The arrow at the top of the Western blot in A indicates the position of the top of the stacking gel. The position of M, markers is indicated at the left of A. From top to bottom, these markers correspond to 205, 116, 97, 66, and 45 kDa, respectively.

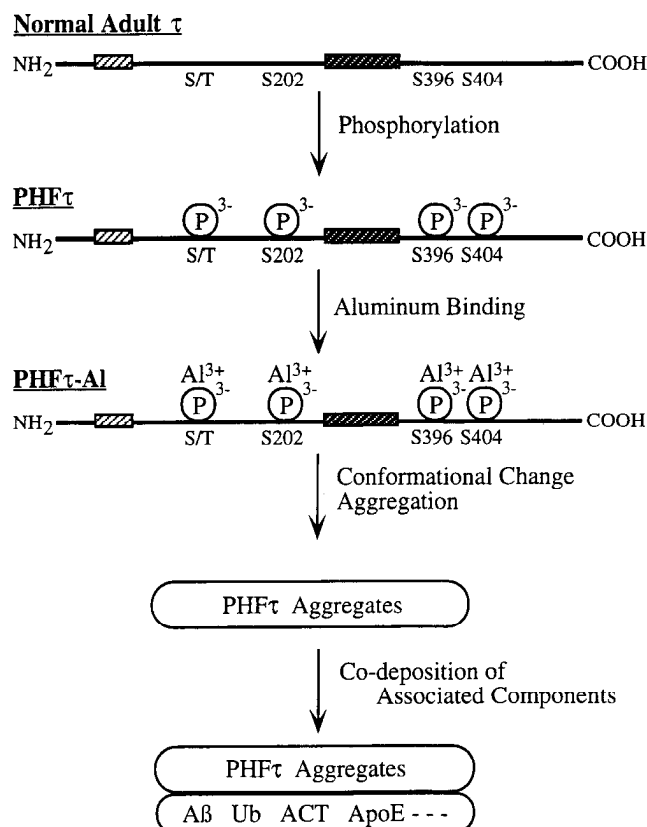
phoretic mobility of the PHF $\tau$  proteins in the AD brain supernatants were dramatically altered (Fig. 5A), while no changes were noted in the normal adult CNS  $\tau$  proteins in the AD (Fig. 5B) and control brain samples (data not shown). For example, increasing concentrations of AlCl<sub>3</sub> reduced the abundance of PHF1-positive PHF $\tau$  bands and induced the accumulation of PHF1-positive material at the top of the stacking gel. This SDS-insoluble material presumably represents aluminum-induced aggregates of PHF $\tau$  that did not enter the stacking gel (Fig. 5A), and similar effects of other aluminum salts on PHF $\tau$  were described recently (Scott et al., 1993). Indeed, at the higher concentrations of aluminum (i.e., at 1 mM AlCl<sub>3</sub>), almost no immunoreactive PHF $\tau$  was present in the Western blots (see lanes 9 and 10 in Fig. 5A). In contrast, the normal adult CNS  $\tau$  proteins that were present with PHF $\tau$  in the same AD brain supernatant fraction were unaffected by the same incubations with increasing concentrations of AlCl<sub>3</sub> (Fig. 5B). Similarly, no changes were observed in the normal adult CNS  $\tau$  proteins that were present in the 350,000  $\times$  g supernatant obtained from the control brains (data not shown). Thus, although aluminum may interact with a number of different polypeptides including normal proteins under some conditions (Scott et al., 1993), we show here that aluminum preferentially interacts with and modifies specific

physical and chemical properties of PHF $\tau$  relative to adult human CNS  $\tau$  under the conditions described here.

## Discussion

The data described here significantly extend previous findings on the induction of co-deposits of several different plaque and tangle components (i.e., A $\beta$ , ubiquitin, ACT) in the rodent brain by intracerebral injections of PHF $\tau$  (Shin et al., 1993). The unique ability of PHF $\tau$  to induce co-deposits of A $\beta$ , ubiquitin, and ACT in the rodent brain was reinforced here by showing that injections of several other plaque and tangle components (i.e., ACT, ApoE, and HSPGs) with and without aluminum failed to induce similar co-deposits. More importantly, we demonstrated that aluminum salts modulate the properties of injected PHF $\tau$  proteins in the rat brain by retarding the proteolysis of these proteins at the injection site, thereby extending the longevity of the co-deposits of A $\beta$ , ACT, and ubiquitin. Furthermore, the complementary *in situ* and *in vitro* studies have partially characterized the more salient aspects of the interactions of AlCl<sub>3</sub> with PHF $\tau$  that might account for the *in vivo* observations reported here. Specifically, the *in situ* and *in vitro* data suggest that the effects of AlCl<sub>3</sub> on the physical and chemical properties of PHF $\tau$  injected into the rat brain may be due to

### HYPOTHETICAL SEQUENCE OF EVENTS LEADING TO PHF $\tau$ AGGREGATION WITH OTHER COMPONENTS IN AD NFTs AND SPs



**Figure 6.** How interactions between aluminum and PHF $\tau$  could represent steps in a hypothetical sequence of events that lead to the aggregation of PHFs in NFTs and SPs. Normal CNS  $\tau$  is shown in the top portion of the schematic oriented with the amino terminus (NH<sub>2</sub>) on the left and the carboxy terminus (COOH) on the right. The amino-terminal (light cross-hatching) and MT-binding (dark cross-hatching) inserts are shown for this 441-aa-long form of adult CNS  $\tau$ . Also shown are unidentified Ser/Thr (S/T) motifs and representative Ser residues (i.e., Ser<sup>202</sup>, Ser<sup>396</sup>, and Ser<sup>404</sup>) that are known to be phosphorylated (circled P with a 3- superscript) in PHF $\tau$ , which is shown below normal CNS  $\tau$ . The interaction of aluminum (Al<sup>3+</sup>) with PHF $\tau$  (PHF $\tau$ -Al) also is depicted in the schematic. The figure shows the conversion of CNS  $\tau$  into PHF $\tau$  by phosphorylation and the interaction of aluminum with PHF $\tau$ , which leads to the increased resistance of PHF $\tau$  to proteolysis, its aggregation, and association with other proteins found in SPs and NFTs such as A $\beta$ , ubiquitin (Ub), ACT, and ApoE (lower portion of the schematic).

the ability of aluminum to bind selectively to domains in PHF $\tau$  that contain phosphorylated Ser or other residues, and this in turn might lead to the aggregation, diminished solubility, and delayed proteolysis of PHF $\tau$  *in vivo* (see schematic summary of these events in Fig. 6). The results described here are all the more remarkable, considering the fact that AlCl<sub>3</sub> may not remain soluble at the pH of the vehicle (pH 6.5) used here, or at the physiological pH in the brain (Martin, 1992).

Although the role of aluminum in AD is uncertain (McLachlan et al., 1992; Perl and Good, 1992; Marksbery and Ehmann, 1994), circumstantial evidence implicates aluminum in the pathogenesis of AD. This evidence includes (1) the detection of aluminum in some AD lesions as well as in tangle-bearing neurons (Perl and Brody, 1980; Garruto et al., 1984; Good et

al., 1992); (2) epidemiological data linking aluminum levels in the water supply to the prevalence of AD (Martyn et al., 1989); (3) the modest therapeutic response to treatment of AD patients with DFO (Crapper-McLachlan et al., 1991); (4) the ability of aluminum to induce phosphorylation of normal  $\tau$  (Yamamoto et al., 1990; Guy et al., 1991; Abdel-Ghany et al., 1993) and a small number of other CNS proteins (Johnson and Jope, 1988; Johnson et al., 1990); and (5) the ability of aluminum to promote aggregation of A $\beta$  (Mantyh et al., 1993). Although aluminum does not induce normal CNS  $\tau$  to form PHF $\tau$ -like structures *in vitro* (Scott et al., 1993), we have demonstrated here that aluminum interacts with PHF $\tau$  and alters its physical and chemical properties. We also show that aluminum enhances the longevity of PHF $\tau$  *in vivo* and extends the longevity of PHF $\tau$ -induced co-deposits of A $\beta$  and plaque and tangle-associated proteins. Further, it also induces rodent neurons to acquire PHF $\tau$ -like immunoreactivity. The evidence here that aluminum enhances the resistance of PHF $\tau$  to *in vivo* proteolysis is compatible with the notion that aluminum may play a role in the pathogenesis of AD. If this indeed is the case, then our data imply that aluminum may serve as a cofactor in the formation of PHF $\tau$ -rich neurofibrillary lesions as well as in the convergence of PHF $\tau$ , A $\beta$ , ubiquitin, ACT, and ApoE in amyloid-rich SPs.

One of the key events in the formation of PHF $\tau$  involves the hyperphosphorylation of normal adult CNS  $\tau$ , which renders these abnormal proteins incapable of binding to microtubules (Bramblett et al., 1993). Recently, it was shown that a PHF $\tau$ -like species could be generated from normal adult recombinant human CNS  $\tau$  *in vitro* using purified mitogen-activated protein kinases, glycogen synthase kinase-3, and proline-directed kinases (Drewes et al., 1992; Goedert et al., 1992; Hanger et al., 1992; Mandelkow et al., 1992; Vulliet et al., 1992) as well as by specific inhibitors of protein phosphatase 2A, and protein phosphatase 2B (Goedert et al., 1992; Harris et al., 1993). Whether or not aluminum plays a direct role in the generation of PHF $\tau$  is unknown, but aluminum has been shown to modulate the phosphorylation and dephosphorylation of  $\tau$  proteins (Guy et al., 1991; Abdel-Ghany et al., 1993). Further, an inhibitory effect of aluminum on the dephosphorylation of  $\tau$  mediated by protein phosphatase 2A has been documented although there was no indication that aluminum was directly bound to this enzyme (Yamamoto et al. 1990). Additionally, aluminum is able to modify the conformation of proteins by inducing  $\beta$ -pleated sheet structure (Hollosi et al., 1992). Although neurons near the injection sites of PHF $\tau$  plus aluminum acquired PHF $\tau$ -like epitopes (as recognized by AT8, T3P, and PHF1 antibodies as well as by the Tau1 MAb following alkaline phosphatase treatment), the mechanism whereby this occurs is not clear, and previous studies of injections of aluminum alone into the brains of experimental animals produced accumulations of neurofilaments but no PHF $\tau$  inclusions (Selkoe et al., 1979; Munoz-Garcia et al., 1986; Troncoso et al., 1986; Wisniewski et al., 1986; Kowall et al., 1989). Further, patients with elevated aluminum levels due to dialysis develop an encephalopathy, but this is not due to the accumulation of AD-like pathology (Candy et al., 1992), and the accidental implantation of metallic aluminum in the human brain has not led to the induction of AD lesions (Foncin and El Hachimi, 1986). Nonetheless, the reproducible effect of injections of PHF $\tau$  plus aluminum on  $\tau$  immunoreactivity in rodent neurons does suggest that aluminum in combination with PHF $\tau$  may modulate the phosphorylation state of CNS  $\tau$  *in vivo*. However, further studies are needed to

determine if and how aluminum modulates the phosphorylation state of  $\tau$  in the rodent and human CNS.

These and other questions concerning the pathogenesis of plaques, tangles, neuron loss, and other molecular lesions in the AD brain have yet to be fully answered even though there has been dramatic progress in the past decade toward elucidating the molecular composition of plaques and tangles (Goedert, 1993; Selkoe, 1993; Trojanowski et al., 1993b, 1994). In the absence of a complete animal model of the authentic human AD phenotype, we expect that experimental paradigms such as the one described here should prove effective in research efforts to arrive at a closer understanding of the mechanisms leading to the formation of complex AD lesions such as plaques and tangles.

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