

HHS Public Access

Author manuscript Curr HIV/AIDS Rep. Author manuscript; available in PMC 2020 June 01.

Published in final edited form as:

Curr HIV/AIDS Rep. 2019 June ; 16(3): 214–223. doi:10.1007/s11904-019-00442-9.

Lipidome abnormalities and cardiovascular disease risk in HIV infection

Emily Bowman, PhD1 and **Nicholas T. Funderburg, PhD**¹

¹School of Health and Rehabilitation Sciences, Division of Medical Laboratory Science, Ohio State University College of Medicine, Columbus, Ohio, USA

Abstract

Purpose of review: Human immunodeficiency virus (HIV) infection, and its treatment with antiretroviral therapy (ART), are associated with lipid abnormalities that may enhance cardiovascular disease risk (CVD).

Recent Findings: Chronic inflammation persists in HIV+ individuals, and complex relationships exist among lipids and inflammation, as immune activation may be both a cause and a consequence of lipid abnormalities in HIV infection. Advances in mass spectrometry-based techniques now allow for detailed measurements of individual lipid species; improved lipid measurement might better evaluate CVD risk compared to the prognostic value of traditional assessments.

Summary: Lipidomic analyses have begun to characterize dynamic changes in lipid composition during HIV infection and following treatment with ART, and further investigation may identify novel lipid biomarkers predictive of adverse outcomes. Developing strategies to improve management of comorbidities in the HIV+ population is important, and statin therapy and lifestyle modifications, including diet and exercise, may help to improve lipid levels and mitigate CVD risk.

Keywords

lipidome; free fatty acids; HIV; cardiovascular disease; inflammation

Corresponding Author: Nicholas Funderburg, 453 W. 10th Ave., 535A Atwell Hall, Columbus OH, 43210, Phone: 614 366-7667,
Nicholas.Funderburg@osumc.edu, Emily Bowman, 460 W. 12th Ave, 740 Biomedical Research Tower, Columbu 614-292-1483, Emily.Bowman@osumc.edu.

Publisher's Disclaimer: This Author Accepted Manuscript is a PDF file of a an unedited peer-reviewed manuscript that has been accepted for publication but has not been copyedited or corrected. The official version of record that is published in the journal is kept up to date and so may therefore differ from this version.

Conflict of Interest

Dr. Bowman declares that she has no conflict of interest.

Dr. Funderburg serves as a consultant for Gilead.

Human and Animal rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors

Introduction

Antiretroviral therapy (ART) has prolonged the lives of individuals living with HIV; however, morbidity and mortality rates remain elevated compared to the general population. Management of comorbidities has become increasingly important in individuals chronically infected with HIV. HIV-infected (HIV+) individuals are at increased risk for cardiovascular disease (CVD), which persists despite virologic suppression with ART (1, 2). Further, HIV infection (3), ART (4), and chronic immune activation (5) can all alter lipid and metabolic profiles (6, 7). Thus, monitoring and controlling lipid levels is crucial for HIV+ individuals, likely even more so than for the HIV uninfected population. Modern mass spectrometrybased techniques enable comprehensive lipid analyses in which concentration and composition of individual lipid species can be evaluated (the lipidome). This review focuses on the effects of HIV infection and its treatment, on the lipidome, and the relationships among lipid abnormalities and enhanced CVD risk in HIV+ individuals (Figure 1).

HIV infection and ART treatment both alter traditional lipid measurements

Lipids have diverse biological roles, including signal transduction, protein trafficking, and regulation of membrane permeability (8). The physiological importance of lipids is underscored by the numerous diseases associated with lipid abnormalities, including CVD, diabetes, obesity, non-alcoholic fatty liver disease (NAFLD), non-alcoholic steatohepatitis (NASH), and Alzheimer's disease (9–14). Dyslipidemia is often observed in HIV+ individuals, and is associated with reduced levels of high-density (HDL) cholesterol, and increased low-density (LDL) lipoprotein, total (TC) cholesterol, and triglycerides (TG) (3). Similar lipid profiles have been linked to the development of atherosclerosis in the general population (15).

HIV-associated dyslipidemia was observed prior to the advent of ART (3), however, specific combinations of ART drugs also have varying effects on lipid metabolism (4, 16), likely contributing to increased risk of CVD (17). ART-associated lipid abnormalities are most evident with protease inhibitor (PI) use, particularly when combined with inhibitors of cytochrome p450 3A4, but other drug classes, including nucleoside reverse transcriptase inhibitors (NRTIs), may also have deleterious effects (17, 18). Even with the most 'lipid friendly' ART regimens, alterations in lipid concentration and composition are frequently observed (4).

Interventions to increase HDL concentrations in humans have failed to reduce clinical cardiovascular events, suggesting that overall HDL levels are not always reflective of efficient function (19–21). HDL has several cardioprotective and anti-inflammatory roles, including promoting cholesterol efflux from macrophages in the vessel wall via reverse cholesterol transport, preventing oxidative modification of LDL, and supporting endothelial cell repair (22–25). Importantly, the cholesterol efflux capacity of HDL particles was shown to be inversely related to CVD risk, and this was independent of total HDL concentration (26). The anti-atherogenic function of HDL is often compromised during HIV infection and in inflammatory environments (27, 28). Exposure to Toll-like receptor (TLR) ligands, such as lipopolysaccharide (LPS), can impair the efficacy of reverse cholesterol transport (29).

Bowman and Funderburg **Page 3** Page 3

This is particularly significant in HIV infection, as reduced gut barrier function and increased plasma levels of microbial products have been reported in HIV+ individuals (30, 31).

Furthermore, monocytes in HIV infection readily form cholesterol-laden foam cells (32, 33), and impaired HDL-mediated reverse cholesterol transport may be a contributing factor. HIV can directly block ATP-binding cassette transporter A1 (ABCA-1) –mediated cholesterol efflux to HDL particles, resulting in intracellular accumulation of lipids and enhanced foam cell formation (34, 35). HIV RNA levels also correlate inversely with HDL concentrations (33). Although, acute inflammatory responses, independent of HIV infection, may similarly alter HDL levels and impair cholesterol efflux from macrophages (25). Low HDL levels have been observed in individuals with acute infections, systemic lupus erythematosus, and rheumatoid arthritis, and cholesterol efflux is impaired in animal models of sepsis (29, 36– 38). Modulation of lipid metabolism during chronic infection may be, in part, a non-specific consequence of inflammation.

Conventional lipid assessments performed routinely in clinics may not adequately assess perturbations in overall lipid profiles or sufficiently inform clinicians to CVD risk in HIV+ individuals (39). Alterations in lipid particle composition and size tend to correlate more strongly with CVD risk than traditional lipid measurements in the HIV+ population (40, 41). HDL and LDL particles are heterogeneous in size; large HDL is more cardioprotective (42), whereas small, dense LDL particles are associated with increased clinical and subclinical presentation of CVD (43, 44). In a study examining ART-treated HIV+ individuals with benign traditional lipid panels, these individuals actually had pro-atherogenic lipid profiles with elevated small LDL particle numbers, and reduced large HDL particles and impaired cholesterol efflux capacity (45). Advanced lipid phenotyping by nuclear magnetic resonance spectroscopy may capture lipid-induced CVD progression better than traditional lipid measurements.

Few studies have investigated the complex relationships among the initiation of ART, lipid particle size and function, and CVD outcomes in HIV infection. In the AIDS Clinical Trials Group (ACTG) A5248 trial (46–49), treatment naïve HIV+ individuals initiating a 'lipid friendly' Raltegravir-based ART regimen were followed longitudinally, and after 48 weeks of ART, LDL levels were increased, but HDL composition and efflux capacity were improved (47). These findings would not have been evident when monitoring traditional lipid panels alone. Measurements of cholesterol particle size, composition and function will likely improve the identification of HIV+ individuals with elevated CVD risk. Lo and colleagues also reported improvement in cholesterol efflux capacity following ART initiation, and this improvement was independently associated with a reduction in HIV viremia (50). Further studies, comparing the effects various ART regimens on lipid particles and cholesterol efflux should be considered.

Dyslipidemia and chronic immune activation in HIV infection

Chronic immune activation is a characteristic of ART-treated HIV infection, and is likely driven by multiple factors (51). Persistent inflammation underlies the development of many

Bowman and Funderburg **Page 4** Page 4

diseases, including atherosclerosis (52), and the inflammatory environment in HIV infection may accelerate progression of CVD. Further, there is a complex relationship between inflammation and the lipidome, as inflammatory processes can alter lipid metabolism, but many lipid species may also exacerbate persistent inflammation (5). Thus, immune activation may be both a cause and a consequence of lipid abnormalities in HIV infection. Similarly, other chronic inflammatory diseases, including rheumatoid arthritis, systemic lupus erythematosus, and psoriasis have also been linked to atherogenic lipid profiles (53).

Increased plasma levels of IL-6, C-reactive protein (CRP), and the coagulation marker Ddimer independently predict cardiovascular events and overall mortality in ART-treated HIV + individuals (40, 54, 55). Importantly, these markers were more strongly predictive of mortality in HIV+ individuals than in HIV- individuals (56–58), indicating that persistent inflammation plays a more critical role in morbidity and mortality in the context of HIV infection than it does for the general population. The Strategies for Management of Antiretroviral Therapy (SMART) study also demonstrated that lower total, large, and small HDL concentrations were associated with a higher risk for CVD in HIV+ individuals (40). Moreover, this work identified an inverse relationship between HDL particle numbers and IL-6 and D-dimer levels. In patients initiating ART, HDL concentration increased, yet, the degree to which HDL levels were improved was dependent on levels of inflammation present at baseline (59). In a separate study of ART-treated HIV+ individuals, metabolic factors such as LDL and ApoA1, a major protein component of HDL, correlated even more strongly with CVD risk than did inflammatory biomarkers (CRP, IL-6, TNF α) (60). There is likely a complex relationship among HIV-associated dyslipidemia, activation of inflammatory pathways, and CVD risk.

Detailed lipidomics analyses have identified lipid profiles associated with CVD

In contrast to basic lipid panels routinely performed in the clinic, detailed lipidomic analysis employs techniques to characterize lipid content in its entirety. Advances in multiple approaches for quantitative mass spectrometry (MS)-based lipid analyses (liquid chromatography, shotgun lipidomics, ion-mobility (61)) allow for more sensitive and extensive assessments of the complete lipidome (62–64), and have enabled the identification of over 40 different lipid classes and thousands of individual lipid species. Lipidomics has broad applications, including providing an important tool for identifying lipid biomarkers relevant to human disease.

Novel lipid biomarkers are predictive of NAFLD, systemic lupus erythematous, preeclampsia, and certain cancers (65–70). Importantly, lipidomic profiling has also outperformed traditional lipid panels for prediction of CVD risk (39, 71, 72). The prospective population-based Bruneck study identified 135 plasma lipid species from 8 different lipid classes (phosphatidylcholine (PC), lysophosphatidylcholine (LPC), cholesterol ester (CE), sphingomyelin (SM), phosphatidylserine (PS), phosphatidylethanolamine (PE), lysophosphatidylethanolamine (LPE), triacylglycerol/ triglycerides (TAG)) that were associated significantly with CVD events (73). Moreover,

Bowman and Funderburg **Page 5** Page 5

there was significant overlap among plasma lipids associated with CVD and lipids previously determined to be enriched in plaques (74), suggesting that circulating lipids reflect disease progression in the atherosclerotic plaque. Of particular importance, the CVDassociated lipids, TAG(54:2), CE(16:1), and PE(36:5), outperformed traditional lipid panels for CVD risk stratification, and in network analyses, were highly interconnected with other lipids predictive of CVD. Furthermore, substitution of standard lipid measurements of Framingham Risk Score (HDL, TC) with TAG(54:2), CE(16:1), and PE(36:5) significantly improved 10-year risk classification (73).

Elevated TAG levels have been linked previously to insulin resistance and increased diabetes risk (75), and were implicated as an independent risk factor for CVD (76). TAG concentrations increase during HIV infection (28), and are related to poor control of virus and higher circulating TNF-α levels, which may interfere further with lipid metabolism (72, 77). More recent lipidomics analyses, however, have implicated specific TAG species as being more predictive of CVD compared to total TAG levels (71, 73–75). The fatty acid composition of TAGs is also important, as TAGs containing myristic acid (14:0), palmitic acid (16:0), stearic acid (18:0), myristoleic acid (14:1), palmitoleic acid (16:1), and oleic acid (18:1) are more closely associated with CVD than other fatty acids (73). Similar trends among the fatty acid composition of TAGs, CEs, SMs, and free fatty acids (FFA) were also linked to increased CVD risk in both the Bruneck study and TwinsUK cohort (78).

In a study of HIV- individuals with asymptomatic or symptomatic CVD, 150 lipid species were uniquely enriched in atherosclerotic plaques, and further, distinct lipid signatures distinguished stable and unstable plaques (74). Additionally, in a cohort of patients with coronary artery disease (CAD), lipidomic analyses more accurately characterized stable and unstable CAD than did traditional lipid measurements, with specific Ceramides (CER) having even more predictive potential than LDL levels (79). CERs have diverse proinflammatory properties. In mouse models, CERs induce NF-κB activation and inflammatory cytokine production (80), and CER levels are associated with chronic heart failure and all-cause mortality in humans (79, 81–83).

Integration of lipidomics data with other–omics strategies will likely enhance understanding of the mechanisms underlying the pathogenesis of CVD and allow for dynamic metabolic pathway reconstruction. An analysis of the Malmo Diet and Cancer (MDC) study cohort demonstrated significant relationships among plasma levels of specific lipid species that correlated with CVD events and validated CVD-associated gene variants (71). Several of these gene variants were also directly involved in coding for lipid biosynthesis enzymes. Bridging lipidomics with genomics data may identify important links between lipids and genetic susceptibility for CVD.

The altered lipidome in HIV infection

Recent lipidomics analyses have begun to elucidate lipidome abnormalities characteristic of HIV infection. Wong et al reported altered levels of 7 lipid classes and 83 individual lipid species that were associated with HIV infection (72). Further analyses identified associations among diacylglycerols (DAGs), phosphatidylinositol (PI), phosphatidylglycerol (PG), TAGs,

Bowman and Funderburg **Page 6** Page 6

CERs, PEs, LPEs, and CEs with CVD risk in HIV+ individuals. Individual DAG and TAG lipid biomarkers were most strongly associated with elevated risk for future CVD events, and outperformed clinical measurements for risk assessment. Similar risk-associated profiles have been related to CVD and diabetes risk in HIV- populations (11, 84, 85). Currently, no studies have explored changes in the lipidome pre- and directly post HIV infection. HIV itself can modulate levels of fatty acid synthase (86), an enzyme important in de novo fatty acid synthesis, and as a consequence, may alter cell associated and plasma lipid profiles. One could speculate that the changes in the lipidome induced by HIV replication during acute or chronic infection may differ from the lipidome within that individual following viral suppression by ART.

Few studies have investigated dynamic changes to lipidome composition following initiation of ART. In a substudy of the ACTG A5248 trial (46–49), plasma concentrations and fatty acid composition of over 1,300 different lipid species across 14 lipid classes were profiled in longitudinal samples from treatment naïve HIV+ individuals initiating a Raltegravir-based ART regimen and following 48 weeks of ART (87). There were broad alterations in lipidome composition after 48 weeks of ART compared to HIV+ individuals at baseline, and when compared to levels in cross-sectional samples obtained from age and sex matched HIV- individuals. Multiple individual lipid species previously linked to CVD-risk in the HIV- population were also increased in HIV+ individuals (88). Notably, the concentration of LPC was increased significantly in HIV+ individuals at baseline, and remained elevated after ART treatment. Increased LPC levels have also been observed in CVD and diabetes (14, 89, 90). Furthermore, the fatty acid composition of LPC particles plays an important role in function. LPCs containing saturated fatty acids (SaFAs) are pro-inflammatory, whereas polyunsaturated fatty acid (PUFA)-containing LPCs are anti-inflammatory (14). The A5248 study also demonstrated that levels of SaFAs, including palmitic acid (16:0) and stearic acid (18:0), were enriched among samples from the HIV population at baseline and after ART administration, and these SaFAs were directly related to levels of immune activation (sCD14, sTNF-R1, IL-6). Elevated circulating SaFAs are associated with greater risk of CVD in HIV- populations (91), and may similarly promote chronic inflammation and the development of comorbidities in HIV+ individuals (87, 92).

In vitro exposure of myeloid cells to SaFAs induces inflammasome activation, TLR signaling, and secretion of inflammatory cytokines (IL-6, TNF-α, and IL-1β) (92–95). In contrast, polyunsaturated fatty acids (PUFAs) inhibit inflammation (14, 96–98), and may protect against the development of diabetes, obesity, NAFLD, and NASH (10–13). Moreover, depletion of PUFAs has been associated with hepatic triglyceride accumulation and endoplasmic reticulum stress (99, 100). By interacting with PPAR transcription factors, PUFAs modulate fatty acid oxidation pathways and the inflammatory mediators, NF-kB, AP-1, NFAT, and STATs (101). Furthermore, LDL particles composed of PUFA-containing cholesterol esters are thought to be less atherogenic; LDL particles enriched with SaFAs tend to be larger and more readily bind arterial proteoglycans, leading to atherosclerotic lesion formation (102). In the A5248 trial, PUFA levels were reduced in HIV+ individuals at baseline, but improved following 48 weeks of ART (87). Imbalanced proportions of SaFAs and PUFAs may contribute to chronic inflammation and directly alter progression of

diseases, including CVD and HIV infection. The precise mechanisms by which lipidome perturbations mediate CVD development need to be explored.

As part of the Alternative Antiretroviral strategies: a comparison of three Initial Regimens (ALTAIR) trial, plasma lipidomic analyses were performed on a subset of treatment-naïve HIV+ individuals randomized to one of three initial ART regimens (efavirenz-, ritonavirboosted atazanavir-, or zidovudine/abacavir-based regimens) (103). Following 48 weeks of ART, numerous lipid alterations were observed, and changes in lipid levels differed by treatment group. In the efavirenz cohort, concentrations of PI, PC, and sphingolipids were increased compared to baseline, whereas monohexoslyceramide and G_{M3} ganglioside classes were decreased with atazanavir/r, and there were no significant changes in lipid class concentrations with zidovudine/abacavir (103). Overall, consistently elevated lipid concentrations were measured in individuals taking efavirenz compared to the atazanavir/r and zidovudine/abacavir populations. Previous studies have suggested that efavirenz-induced dyslipidemia does not alter LDL/HDL ratio, and is therefore not particularly atherogenic (104), however, analyses of the United States veterans affairs database have recently identified a potential link between efavirenz treatment and increased cardiovascular events (105). Distinct efavirenz-induced lipidome alterations, particularly increased sphingolipid levels, which are predictive of symptomatic CAD (79), may better explain clinical outcomes in HIV+ individuals on this ART regimen.

The mechanisms by which various ART drugs affect lipid metabolism and contribute to dyslipidemia are different (4), therefore it is reasonable to assume characteristic lipidome alterations in HIV+ individuals will differ depending on specific ART regimen. Further indepth studies are warranted to characterize unique ART-associated lipid profiles in treated HIV infection, and the clinical relevance of these lipid alterations. There is also a significant gap in knowledge regarding the dynamics of age-related effects on lipidome composition, and comprehensive longitudinal aging studies should be performed to identify lipid perturbations associated with age. Previously, increased incidence of insulin resistance and triglyceride accumulation was observed in elderly study populations (106, 107). Lipid abnormalities may play a particularly important role in comorbidity risk in the aging HIV population. Future lipidomics analyses should also explore the contributions of lifestyle factors, such as diet and smoking status, infection with copathogens, and the composition of the microbiome on lipid profiles in HIV infection.

Strategies to improve lipids in HIV+ individuals

The potential link between dyslipidemia and increased risk for CVD in chronic HIV infection has led to strategies for modulating lipid levels and mitigating inflammation in HIV+ individuals. Multiple studies have examined the efficacy of statin (3-hydroxy-3 methylglutaryl coenzyme A reductase inhibitors) usage in improving lipid profiles in HIV+ individuals, and have reported beneficial effects on inflammatory markers and CVD risk with statin use (108–111). In the Stopping Atherosclerosis and Treating Unhealthy Bone with Rosuvastatin in HIV (SATURNHIV) trial, ART-treated HIV+ individuals with normal LDL levels, but increased biomarkers of immune activation, had reductions in T cell and monocyte activation markers, reduced vascular inflammation, and improved renal function

Bowman and Funderburg **Page 8**

following statin treatment (110–112). Plasma levels of sCD14, a biomarker linked to morbidity and mortality in HIV infection (113), were also decreased by statin treatment. In a separate study, atorvastatin treatment resulted in reduced non-calcified plaque volumes in HIV+ individuals (114). Additionally, statin use has been associated with decreased levels of oxidized LDL (OxLDL), a principle component in atherosclerotic lesions that is associated with plaque instability (109, 110, 114–116). Changes in OxLDL levels were directly related to decreases in sCD14, tissue factor expression on monocytes, and improved carotid intimamedia thickness (cIMT) measurements (115).

In a recent analysis, current lipid guidelines in place for the general population would not have recommended statins to the majority (~74%) of HIV+ individuals that did, in fact, have subclinical high-risk plaques (117). Additional factors may need to be considered when identifying subgroups of the HIV+ population that could benefit from statin use. The Randomized Trial to Prevent Vascular Events in HIV (REPRIEVE) study is currently evaluating the efficacy of statin use in preventing CVD events in HIV+ individuals that would be considered low risk based on current guidelines (118, 119). Future studies should also take into account varying effects of different statins on specific lipid profiles. Comparative lipidomic analyses have demonstrated unique lipid species alterations induced differentially by rosuvastatin and atorvastatin, despite similar overall LDL lowering effects (120).

Lifestyle intervention studies, including changes in diet and exercise, may also prove useful in favorably modulating lipid levels and improving CVD outcomes in the HIV+ population. The American Heart Association (AHA) has long recommended reducing dietary saturated fat intake to protect against CVD (121). In randomized, controlled trials, reductions in SaFA consumption and subsequent increases in PUFA intake reduced the incidence of CVD in the general population (91, 122). Furthermore, several clinical studies have reported promising beneficial effects of PUFA supplementation on hypertriglyceridemia, hypertension, inflammation, and insulin sensitivity (123–126), however, there is some disagreement concerning the association of fatty acid intake and clinical outcomes (91, 127). In a randomized placebo-controlled trial, oral supplementation of the PUFAs, EPA (20:5) and DHA (22:6), reduced inflammation and soluble TNFR1 levels in HIV+ individuals (128). Studies that examine relationships among dietary and lifestyle interventions, the lipidome, inflammation, and comorbidities in HIV+ individuals should be pursued.

Conclusions

Chronic HIV infection and its treatment are associated with altered lipid profiles and increased risk for CVD. These lipid abnormalities are complicated further by persistent inflammation in treated HIV+ individuals that may also accelerate progression of CVD. Characterizing alterations in the lipidome of HIV+ individuals will likely help to identify metabolic abnormalities and elucidate determinants of enhanced CVD risk. Developing strategies to improve clinical management of CVD in the HIV+ population will be important, as guidelines in place for the general population may not adequately address the needs of HIV+ individuals. Further lipidomic analyses may provide novel drug targets and

References

- 1. Triant VA, Lee H, Hadigan C, Grinspoon SK. Increased acute myocardial infarction rates and cardiovascular risk factors among patients with human immunodeficiency virus disease. J Clin Endocrinol Metab. 2007;92(7):2506–12. [PubMed: 17456578]
- 2. Shah ASV, Stelzle D, Lee KK, Beck EJ, Alam S, Clifford S, et al. Global Burden of Atherosclerotic Cardiovascular Disease in People Living With HIV. Circulation. 2018;138(11):1100–12. [PubMed: 29967196] • This work provides a thorough description of CVD burden in the global HIV+ population, and discusses the importance of improving strategies for risk stratification and treatment.
- 3. Grunfeld C, P M, Doerrler W, Shigenaga J, Jensen P, Feingold KR. Lipids, Lipoproteins, Triglyceride Clearance, and Cytokines in Human Immunodeficiency Virus Infection and the Acquired Immunodeficiency Syndrome. J Clin Endocrinol Metab. 1992;74(5):1045–52. [PubMed: 1373735]
- 4. Lake JE, Currier JS. Metabolic disease in HIV infection. The Lancet Infectious Diseases. 2013;13(11):964–75. [PubMed: 24156897]
- 5. Tall AR, Yvan-Charvet L. Cholesterol, inflammation and innate immunity. Nat Rev Immunol. 2015;15(2):104–16. [PubMed: 25614320]
- 6. Grinspoon S, Carr A. Cardiovascular Risk and Body-Fat Abnormalities in HIV-Infected Adults. New England Journal of Medicine. 2005;352(1):48–62. [PubMed: 15635112]
- 7. Rose H, Hoy J, Woolley I, Tchoua U, Bukrinsky M, Dart A, et al. HIV infection and high density lipoprotein metabolism. Atherosclerosis. 2008;199(1):79–86. [PubMed: 18054941]
- 8. Harayama T, Riezman H. Understanding the diversity of membrane lipid composition. Nat Rev Mol Cell Biol. 2018;19(5):281–96. [PubMed: 29410529]
- 9. Liu Q, Zhang J. Lipid metabolism in Alzheimer's disease. Neurosci Bull. 2014;30(2):331–45. [PubMed: 24733655]
- 10. Cassol E, M V, Holman A, Kamat A, Morgello S, Gabuzda D. Plasma metabolomics identifies lipid abnormalities linked to markers of inflammation, microbial translocation, and hepatic function in HIV patients receiving protease inhibitors. BMC Infectious Diseases. 2013;13:203–20. [PubMed: 23641933]
- 11. Hodge AM, E D, O'Dea K, Sinclair AJ, Makrides M, Gibson RA, Giles GG. Plasma phospholipid and dietary fatty acids as predictors of type 2 diabetes: interpreting the role of linoleic acid. Am J Clin Nutr. 2007;86:189–97. [PubMed: 17616780]
- 12. Suvitaival T, Bondia-Pons I, Yetukuri L, Poho P, Nolan JJ, Hyotylainen T, et al. Lipidome as a predictive tool in progression to type 2 diabetes in Finnish men. Metabolism. 2018;78:1–12. [PubMed: 28941595]
- 13. Tavares De Almeida I, Cortez-Pinto H, Fidalgo G, Rodrigues D, Camilo ME. Plasma total and free fatty acids composition in human non-alcoholic steatohepatitis. Clinical Nutrition. 2002;21(3): 219–23. [PubMed: 12127930]
- 14. Akerele OA, Cheema SK. Fatty acyl composition of lysophosphatidylcholine is important in atherosclerosis. Med Hypotheses. 2015;85(6):754–60. [PubMed: 26604024]
- 15. Stamler J, D M, Garside DB, Dyer AR, Greenland P, Neaton JD. Relationship of Baseline Serum Cholesterol Levels in 3 Large Cohorts of Younger Men to Long-term Coronary, Cardiovascular, and All-Cause Mortality and to Longevity. JAMA. 2000;284(3):311–8. [PubMed: 10891962]
- 16. Willig AL, Overton ET. Metabolic consequences of HIV: pathogenic insights. Curr HIV/AIDS Rep. 2014;11(1):35–44. [PubMed: 24390642]
- 17. Estrada V, P J. Dyslipidemia Related to Antiretroviral Therapy. AIDS Rev. 2011;13:49–56. [PubMed: 21412389]

- 18. Crane HM, Grunfeld C, Willig JH, Mugavero MJ, Van Rompaey S, Moore R, et al. Impact of NRTIs on lipid levels among a large HIV-infected cohort initiating antiretroviral therapy in clinical care. AIDS. 2011;25(2):185–95. [PubMed: 21150555]
- 19. Schwartz GG, Olsson AG, Abt M, Ballantyne CM, Barter PJ, Brumm J, et al. Effects of dalcetrapib in patients with a recent acute coronary syndrome. N Engl J Med. 2012;367(22):2089–99. [PubMed: 23126252]
- 20. Barter PJ, Caulfield M, Eriksson M, Grundy SM, Kastelein JJP, Komajda M, et al. Effects of Torcetrapib in Patients at High Risk for Coronary Events. New England Journal of Medicine. 2007;357(21):2109–22. [PubMed: 17984165]
- 21. Niacin in Patients with Low HDL Cholesterol Levels Receiving Intensive Statin Therapy. New England Journal of Medicine. 2011;365(24):2255–67. [PubMed: 22085343]
- 22. Rye KA, Barter PJ. Cardioprotective functions of HDLs. J Lipid Res. 2014;55(2):168–79. [PubMed: 23812558]
- 23. Tso C, Martinic G, Fan WH, Rogers C, Rye KA, Barter PJ. High-density lipoproteins enhance progenitor-mediated endothelium repair in mice. Arterioscler Thromb Vasc Biol. 2006;26(5): 1144–9. [PubMed: 16528007]
- 24. Triolo M, Annema W, Dullaart RPF, Tietge UJF. Assessing the functional properties of highdensity lipoproteins: an emerging concept in cardiovascular research. Biomarkers in Medicine. 2013;7(3):457–72. [PubMed: 23734809]
- 25. Khovidhunkit W, Memon RA, Feingold KR, Grunfeld C. Infection and Inflammation-Induced Proatherogenic Changes of Lipoproteins. The Journal of Infectious Diseases. 2000;181(Supplement_3):S462–S72. [PubMed: 10839741]
- 26. Qiu C, Zhao X, Zhou Q, Zhang Z. High-density lipoprotein cholesterol efflux capacity is inversely associated with cardiovascular risk: a systematic review and meta-analysis. Lipids Health Dis. 2017;16(1):212. [PubMed: 29126414]
- 27. Dullaart RP, Annema W, Tio RA, Tietge UJ. The HDL anti-inflammatory function is impaired in myocardial infarction and may predict new cardiac events independent of HDL cholesterol. Clin Chim Acta. 2014;433:34–8. [PubMed: 24613518]
- 28. Grunfeld C Dyslipidemia and its Treatment in HIV Infection. Topics in HIV medicine : a publication of the International AIDS Society, USA. 2010;18(3):112–8.
- 29. McGillicuddy FC, de la Llera Moya M, Hinkle CC, Joshi MR, Chiquoine EH, Billheimer JT, et al. Inflammation impairs reverse cholesterol transport in vivo. Circulation. 2009;119(8):1135–45. [PubMed: 19221221]
- 30. Brenchley JM, Douek DC. HIV infection and the gastrointestinal immune system. Mucosal Immunol. 2008;1(1):23–30. [PubMed: 19079157]
- 31. Brenchley JM, Price DA, Schacker TW, Asher TE, Silvestri G, Rao S, et al. Microbial translocation is a cause of systemic immune activation in chronic HIV infection. Nat Med. 2006;12(12):1365– 71. [PubMed: 17115046]
- 32. Maisa A, Hearps AC, Angelovich TA, Pereira CF, Zhou J, Shi MD, et al. Monocytes from HIVinfected individuals show impaired cholesterol efflux and increased foam cell formation after transendothelial migration. AIDS. 2015;29(12):1445–57. [PubMed: 26244384]
- 33. Feeney ER, McAuley N, O'Halloran JA, Rock C, Low J, Satchell CS, et al. The expression of cholesterol metabolism genes in monocytes from HIV-infected subjects suggests intracellular cholesterol accumulation. J Infect Dis. 2013;207(4):628–37. [PubMed: 23204179]
- 34. Asztalos BF, Mujawar Z, Morrow MP, Grant A, Pushkarsky T, Wanke C, et al. Circulating Nef induces dyslipidemia in simian immunodeficiency virus-infected macaques by suppressing cholesterol efflux. J Infect Dis. 2010;202(4):614–23. [PubMed: 20617930]
- 35. Mujawar Z, Rose H, Morrow MP, Pushkarsky T, Dubrovsky L, Mukhamedova N, et al. Human immunodeficiency virus impairs reverse cholesterol transport from macrophages. PLoS Biol. 2006;4(11):e365. [PubMed: 17076584]
- 36. Sammalkorpi K, Valtonen V, Kerttula Y, Nikkilä E, Taskinen M-R. Changes in serum lipoprotein pattern induced by acute infections. Metabolism - Clinical and Experimental. 1988;37(9):859–65. [PubMed: 3419323]

- 37. Sarkissian T, Beyene J, Feldman B, McCrindle B, Silverman ED. Longitudinal examination of lipid profiles in pediatric systemic lupus erythematosus. Arthritis Rheum. 2007;56(2):631–8. [PubMed: 17265498]
- 38. Mostaza JM, Camino N, Gerique JG, Peña R, Baquero M, Lahoz C. C-reactive protein levels and prevalence of chronic infections in subjects with hypoalphalipoproteinemia. Metabolism. 2005;54(1):33–7.
- 39. Friis-Moller N, Ryom L, Smith C, Weber R, Reiss P, Dabis F, et al. An updated prediction model of the global risk of cardiovascular disease in HIV-positive persons: The Data-collection on Adverse Effects of Anti-HIV Drugs (D:A:D) study. Eur J Prev Cardiol. 2016;23(2):214–23. [PubMed: 25882821]
- 40. Duprez DA, Kuller LH, Tracy R, Otvos J, Cooper DA, Hoy J, et al. Lipoprotein particle subclasses, cardiovascular disease and HIV infection. Atherosclerosis. 2009;207(2):524–9. [PubMed: 19515371]
- 41. Bucher HC, Richter W, Glass TR, Magenta L, Wang Q, Cavassini M, et al. Small Dense Lipoproteins, Apolipoprotein B, and Risk of Coronary Events in HIV-Infected Patients on Antiretroviral Therapy: The Swiss HIV Cohort Study. JAIDS Journal of Acquired Immune Deficiency Syndromes. 2012;60(2):135–42. [PubMed: 22156913]
- 42. Mora S, Otvos JD, Rifai N, Rosenson RS, Buring JE, Ridker PM. Lipoprotein particle profiles by nuclear magnetic resonance compared with standard lipids and apolipoproteins in predicting incident cardiovascular disease in women. Circulation. 2009;119(7):931–9. [PubMed: 19204302]
- 43. Gazi IF, Tsimihodimos V, Tselepis AD, Elisaf M, Mikhailidis DP. Clinical importance and therapeutic modulation of small dense low-density lipoprotein particles. Expert Opin Biol Ther. 2007;7(1):53–72. [PubMed: 17150019]
- 44. St-Pierre AC, Cantin B, Dagenais GR, Mauriege P, Bernard PM, Despres JP, et al. Low-density lipoprotein subfractions and the long-term risk of ischemic heart disease in men: 13-year follow-up data from the Quebec Cardiovascular Study. Arterioscler Thromb Vasc Biol. 2005;25(3):553–9. [PubMed: 15618542]
- 45. Munger AM, Chow DC, Playford MP, Parikh NI, Gangcuangco LM, Nakamoto BK, et al. Characterization of lipid composition and high-density lipoprotein function in HIV-infected individuals on stable antiretroviral regimens. AIDS Res Hum Retroviruses. 2015;31(2):221–8. [PubMed: 25416403]
- 46. Andrade A, Rosenkranz SL, Cillo AR, Lu D, Daar ES, Jacobson JM, et al. Three distinct phases of HIV-1 RNA decay in treatment-naive patients receiving raltegravir-based antiretroviral therapy: ACTG A5248. J Infect Dis. 2013;208(6):884–91. [PubMed: 23801609]
- 47. Funderburg NT, Xu D, Playford MP, Joshi AA, Andrade A, Kuritzkes DR, et al. Treatment of HIV infection with a raltegravir-based regimen increases LDL levels, but improves HDL cholesterol efflux capacity. Antivir Ther. 2017;22(1):71–5. [PubMed: 27740536] • This study examines longitudinal changes in lipid composition and function in HIV+ individuals initiating a Raltegravir-based ART regimen. These analyses may better assess CVD risk than traditional lipid measurements.
- 48. McCausland MR, Juchnowski SM, Zidar DA, Kuritzkes DR, Andrade A, Sieg SF, et al. Altered Monocyte Phenotype in HIV-1 Infection Tends to Normalize with Integrase-Inhibitor-Based Antiretroviral Therapy. PLoS One. 2015;10(10):e0139474. [PubMed: 26430882]
- 49. Funderburg NT, Andrade A, Chan ES, Rosenkranz SL, Lu D, Clagett B, et al. Dynamics of Immune Reconstitution and Activation Markers in HIV+ Treatment-Naïve Patients Treated with Raltegravir, Tenofovir Disoproxil Fumarate and Emtricitabine. PLOS ONE. 2013;8(12):e83514. [PubMed: 24367599]
- 50. Lo J, Rosenberg ES, Fitzgerald ML, Bazner SB, Ihenachor EJ, Hawxhurst V, et al. High-density lipoprotein-mediated cholesterol efflux capacity is improved by treatment with antiretroviral therapy in acute human immunodeficiency virus infection. Open forum infectious diseases. 2014;1(3):ofu108–ofu. [PubMed: 25734176]
- 51. Lederman MM, Funderburg NT, Sekaly RP, Klatt NR, Hunt PW. Residual immune dysregulation syndrome in treated HIV infection. Adv Immunol. 2013;119:51–83. [PubMed: 23886064]
- 52. Inflammation GKH, Atherosclerosis, and Coronary Artery Disease. N Engl J Med. 2005;352:1685–95. [PubMed: 15843671]

- 53. Liao KP, Playford MP, Frits M, Coblyn JS, Iannaccone C, Weinblatt ME, et al. The association between reduction in inflammation and changes in lipoprotein levels and HDL cholesterol efflux capacity in rheumatoid arthritis. J Am Heart Assoc. 2015;4(2).
- 54. Kuller LH, Tracy R, Belloso W, De Wit S, Drummond F, Lane HC, et al. Inflammatory and coagulation biomarkers and mortality in patients with HIV infection. PLoS Med. 2008;5(10):e203. [PubMed: 18942885]
- 55. Triant VA, Meigs JB, Grinspoon SK. Association of C-reactive protein and HIV infection with acute myocardial infarction. J Acquir Immune Defic Syndr. 2009;51(3):268–73. [PubMed: 19387353]
- 56. Wikby A, Nilsson BO, Forsey R, Thompson J, Strindhall J, Lofgren S, et al. The immune risk phenotype is associated with IL-6 in the terminal decline stage: findings from the Swedish NONA immune longitudinal study of very late life functioning. Mech Ageing Dev. 2006;127(8):695–704. [PubMed: 16750842]
- 57. Harris TB, Ferrucci L, Tracy RP, Corti MC, Wacholder S, Ettinger WH, et al. Associations of elevated Interleukin-6 and C-Reactive protein levels with mortality in the elderly. The American Journal of Medicine. 1999;106(5):506–12. [PubMed: 10335721]
- 58. Reuben DB, Cheh AI, Harris TB, Ferrucci L, Rowe JW, Tracy RP, et al. Peripheral Blood Markers of Inflammation Predict Mortality and Functional Decline in High-Functioning Community-Dwelling Older Persons. Journal of the American Geriatrics Society. 2002;50(4):638–44. [PubMed: 11982663]
- 59. Baker JV, Neuhaus J, Duprez D, Cooper DA, Hoy J, Kuller L, et al. Inflammation predicts changes in high-density lipoprotein particles and apolipoprotein A1 following initiation of antiretroviral therapy. AIDS. 2011;25(17):2133–42. [PubMed: 21857489]
- 60. Piconi S, Parisotto S, Rizzardini G, Passerini S, Meraviglia P, Schiavini M, et al. Atherosclerosis is associated with multiple pathogenic mechanisms in HIV-infected antiretroviral-naive or treated individuals. AIDS. 2013;27(3):381–9. [PubMed: 23079800]
- 61. Han X Lipidomics for studying metabolism. Nat Rev Endocrinol. 2016;12(11):668–79. [PubMed: 27469345]
- 62. Watson AD. Thematic review series: systems biology approaches to metabolic and cardiovascular disorders. Lipidomics: a global approach to lipid analysis in biological systems. J Lipid Res. 2006;47(10):2101–11. [PubMed: 16902246]
- 63. Weir JM, Wong G, Barlow CK, Greeve MA, Kowalczyk A, Almasy L, et al. Plasma lipid profiling in a large population-based cohort. J Lipid Res. 2013;54(10):2898–908. [PubMed: 23868910]
- 64. Yang K, Cheng H, Gross RW, Han X. Automated lipid identification and quantification by multidimensional mass spectrometry-based shotgun lipidomics. Anal Chem. 2009;81(11):4356– 68. [PubMed: 19408941]
- 65. Anand S, Young S, Esplin MS, Peaden B, Tolley HD, Porter TF, et al. Detection and confirmation of serum lipid biomarkers for preeclampsia using direct infusion mass spectrometry. J Lipid Res. 2016;57(4):687–96. [PubMed: 26891737]
- 66. Gorden DL, Myers DS, Ivanova PT, Fahy E, Maurya MR, Gupta S, et al. Biomarkers of NAFLD progression: a lipidomics approach to an epidemic. J Lipid Res. 2015;56(3):722–36. [PubMed: 25598080]
- 67. Hu C, Zhou J, Yang S, Li H, Wang C, Fang X, et al. Oxidative stress leads to reduction of plasmalogen serving as a novel biomarker for systemic lupus erythematosus. Free Radic Biol Med. 2016;101:475–81. [PubMed: 27836780]
- 68. Perrotti F, Rosa C, Cicalini I, Sacchetta P, Del Boccio P, Genovesi D, et al. Advances in Lipidomics for Cancer Biomarkers Discovery. Int J Mol Sci. 2016;17(12).
- 69. Li J, Ren S, Piao HL, Wang F, Yin P, Xu C, et al. Integration of lipidomics and transcriptomics unravels aberrant lipid metabolism and defines cholesteryl oleate as potential biomarker of prostate cancer. Sci Rep. 2016;6:20984. [PubMed: 26865432]
- 70. Chen X, Chen H, Dai M, Ai J, Li Y, Mahon B, et al. Plasma lipidomics profiling identified lipid biomarkers in distinguishing early-stage breast cancer from benign lesions. Oncotarget. 2016;7(24):36622–31. [PubMed: 27153558]

- 71. Fernandez C, Sandin M, Sampaio JL, Almgren P, Narkiewicz K, Hoffmann M, et al. Plasma lipid composition and risk of developing cardiovascular disease. PLoS One. 2013;8(8):e71846. [PubMed: 23967253]
- 72. Wong G, Trevillyan JM, Fatou B, Cinel M, Weir JM, Hoy JF, et al. Plasma lipidomic profiling of treated HIV-positive individuals and the implications for cardiovascular risk prediction. PLoS One. 2014;9(4):e94810. [PubMed: 24733512]
- 73. Stegemann C, Pechlaner R, Willeit P, Langley SR, Mangino M, Mayr U, et al. Lipidomics profiling and risk of cardiovascular disease in the prospective population-based Bruneck study. Circulation. 2014;129(18):1821–31. [PubMed: 24622385]
- 74. Stegemann C, Drozdov I, Shalhoub J, Humphries J, Ladroue C, Didangelos A, et al. Comparative lipidomics profiling of human atherosclerotic plaques. Circ Cardiovasc Genet. 2011;4(3):232–42. [PubMed: 21511877]
- 75. Rhee EP, Cheng S, Larson MG, Walford GA, Lewis GD, McCabe E, et al. Lipid profiling identifies a triacylglycerol signature of insulin resistance and improves diabetes prediction in humans. J Clin Invest. 2011;121(4):1402–11. [PubMed: 21403394]
- 76. Harchaoui KEL, Visser ME, Kastelein JJP, Stroes ES, Dallinga-Thie GM. Triglycerides and cardiovascular risk. Current cardiology reviews. 2009;5(3):216–22. [PubMed: 20676280]
- 77. Haugaard SB, Andersen O, Pedersen SB, Dela F, Fenger M, Richelsen B, et al. Tumor necrosis factor alpha is associated with insulin-mediated suppression of free fatty acids and net lipid oxidation in HIV-infected patients with lipodystrophy. Metabolism. 2006;55(2):175–82. [PubMed: 16423623]
- 78. Moayyeri A, Hammond CJ, Valdes AM, Spector TD. Cohort Profile: TwinsUK and healthy ageing twin study. Int J Epidemiol. 2013;42(1):76–85. [PubMed: 22253318]
- 79. Tarasov K, Ekroos K, Suoniemi M, Kauhanen D, Sylvänne T, Hurme R, et al. Molecular Lipids Identify Cardiovascular Risk and Are Efficiently Lowered by Simvastatin and PCSK9 Deficiency. The Journal of Clinical Endocrinology and Metabolism. 2014;99(1):E45–E52. [PubMed: 24243630]
- 80. Wu D, Ren Z, Pae M, Guo W, Cui X, Merrill AH, et al. Aging Up-Regulates Expression of Inflammatory Mediators in Mouse Adipose Tissue. The Journal of Immunology. 2007;179(7): 4829–39. [PubMed: 17878382]
- 81. Laaksonen R, Ekroos K, Sysi-Aho M, Hilvo M, Vihervaara T, Kauhanen D, et al. Plasma ceramides predict cardiovascular death in patients with stable coronary artery disease and acute coronary syndromes beyond LDL-cholesterol. Eur Heart J. 2016;37(25):1967–76. [PubMed: 27125947]
- 82. Yu J, Pan W, Shi R, Yang T, Li Y, Yu G, et al. Ceramide is upregulated and associated with mortality in patients with chronic heart failure. Can J Cardiol. 2015;31(3):357–63. [PubMed: 25746025]
- 83. Peterson LR, Xanthakis V, Duncan MS, Gross S, Friedrich N, Volzke H, et al. Ceramide Remodeling and Risk of Cardiovascular Events and Mortality. J Am Heart Assoc. 2018;7(10).
- 84. Ueeda M, D T, Takaya Y, Shinohata R, Katayma Y, Ohnishi N, Takaishi A, Miyoshi T, Hirohata S, Kusachi S. Serum N-3 Polyunsaturated Fatty Acid Levels Correlate With the Extent of Coronary Plaques and Calcifications in Patients With Acute Myocardial Infarction. Circ J. 2008;72:1836–43. [PubMed: 18812674]
- 85. Ellims AH, Wong G, Weir JM, Lew P, Meikle PJ, Taylor AJ. Plasma lipidomic analysis predicts non-calcified coronary artery plaque in asymptomatic patients at intermediate risk of coronary artery disease. Eur Heart J Cardiovasc Imaging. 2014;15(8):908–16. [PubMed: 24618657]
- 86. Kulkarni MM, Ratcliff AN, Bhat M, Alwarawrah Y, Hughes P, Arcos J, et al. Cellular fatty acid synthase is required for late stages of HIV-1 replication. Retrovirology. 2017;14(1):45. [PubMed: 28962653]
- 87. Belury MA, Bowman E, Gabriel J, Snyder B, Kulkarni M, Palettas M, et al. Prospective Analysis of Lipid Composition Changes with Antiretroviral Therapy and Immune Activation in Persons Living with HIV. Pathog Immun. 2017;2(3):376–403. [PubMed: 29098203] • This study examines longitudinal lipidome alterations in treatment-naïve HIV+ individuals at baseline, and after 48

weeks of a Raltegravir-based ART regimen, and the relationships among pro-inflammatory lipid species and immune activation in this population.

- 88. Funderburg NT, editor Prospective analysis of lipid compositional changes with antiretroviral therapy (ART) and immune activation in persons living with HIV. The 19th International Workshop on Co-morbidities and Adverse Drug Reactions in HIV; 2017 October 23–25; Milan, Italy.
- 89. Wells I, P G, Vincent JK. Lecithin: Cholesterol Acyltransferase and Lysolecithin in Coronary Atherosclerosis. Exp Mol Pathol. 1986;45:303–10. [PubMed: 3466803]
- 90. Rabini RA, Galassi R, Fumelli P, Dousset N, Solera ML, Valdiguie P, et al. Reduced Na(+) -K(+)- ATPase Activity and Plasma Lysophosphatidylcholine Concentrations in Diabetic Patients. Diabetes. 1994;43(7):915–9. [PubMed: 8013757]
- 91. Sacks FM, Lichtenstein AH, Wu JHY, Appel LJ, Creager MA, Kris-Etherton PM, et al. Dietary Fats and Cardiovascular Disease: A Presidential Advisory From the American Heart Association. Circulation. 2017;136(3):e1–e23. [PubMed: 28620111]
- 92. Robblee MM, Kim CC, Porter Abate J, Valdearcos M, Sandlund KL, Shenoy MK, et al. Saturated Fatty Acids Engage an IRE1alpha-Dependent Pathway to Activate the NLRP3 Inflammasome in Myeloid Cells. Cell Rep. 2016;14(11):2611–23. [PubMed: 26971994]
- 93. Wen H, Gris D, Lei Y, Jha S, Zhang L, Huang MT, et al. Fatty acid-induced NLRP3-ASC inflammasome activation interferes with insulin signaling. Nat Immunol. 2011;12(5):408–15. [PubMed: 21478880]
- 94. Suganami T, Nishida J, Ogawa Y. A paracrine loop between adipocytes and macrophages aggravates inflammatory changes: role of free fatty acids and tumor necrosis factor alpha. Arterioscler Thromb Vasc Biol. 2005;25(10):2062–8. [PubMed: 16123319]
- 95. Suganami T, Tanimoto-Koyama K, Nishida J, Itoh M, Yuan X, Mizuarai S, et al. Role of the Tolllike receptor 4/NF-kappaB pathway in saturated fatty acid-induced inflammatory changes in the interaction between adipocytes and macrophages. Arterioscler Thromb Vasc Biol. 2007;27(1):84– 91. [PubMed: 17082484]
- 96. Oh DY, Talukdar S, Bae EJ, Imamura T, Morinaga H, Fan W, et al. GPR120 is an omega-3 fatty acid receptor mediating potent anti-inflammatory and insulin-sensitizing effects. Cell. 2010;142(5):687–98. [PubMed: 20813258]
- 97. Yamada H, Yoshida M, Nakano Y, Suganami T, Satoh N, Mita T, et al. In vivo and in vitro inhibition of monocyte adhesion to endothelial cells and endothelial adhesion molecules by eicosapentaenoic acid. Arterioscler Thromb Vasc Biol. 2008;28(12):2173–9. [PubMed: 18948636]
- 98. Yan Y, Jiang W, Spinetti T, Tardivel A, Castillo R, Bourquin C, et al. Omega-3 fatty acids prevent inflammation and metabolic disorder through inhibition of NLRP3 inflammasome activation. Immunity. 2013;38(6):1154–63. [PubMed: 23809162]
- 99. Kim SJ, Zhang Z, Saha A, Sarkar C, Zhao Z, Xu Y, et al. Omega-3 and omega-6 fatty acids suppress ER- and oxidative stress in cultured neurons and neuronal progenitor cells from mice lacking PPT1. Neurosci Lett. 2010;479(3):292–6. [PubMed: 20561933]
- 100. Pachikian BD, Essaghir A, Demoulin JB, Neyrinck AM, Catry E, De Backer FC, et al. Hepatic n-3 polyunsaturated fatty acid depletion promotes steatosis and insulin resistance in mice: genomic analysis of cellular targets. PLoS One. 2011;6(8):e23365. [PubMed: 21853118]
- 101. Bensinger SJ, Tontonoz P. Integration of metabolism and inflammation by lipid-activated nuclear receptors. Nature. 2008;454(7203):470–7. [PubMed: 18650918]
- 102. Degirolamo C, Shelness GS, Rudel LL. LDL cholesteryl oleate as a predictor for atherosclerosis: evidence from human and animal studies on dietary fat. J Lipid Res. 2009;50 Suppl:S434–9. [PubMed: 19029117]
- 103. Trevillyan JM, Wong G, Puls R, Petoumenos K, Emery S, Mellett NA, et al. Changes in plasma lipidome following initiation of antiretroviral therapy. PLoS One. 2018;13(8):e0202944. [PubMed: 30157268] • This study describes ART-induced changes to the HIV+ lipidome, and compares lipid alterations unique to 3 distinct ART regimens.
- 104. Gotti D, Cesana BM, Albini L, Calabresi A, Izzo I, Focà E, et al. Increase in Standard Cholesterol and Large HDL Particle Subclasses in Antiretroviral-Naïve Patients Prescribed Efavirenz

Compared to Atazanavir/Ritonavir. HIV Clinical Trials. 2012;13(5):245–55. [PubMed: 23134625]

- 105. Desai M, Joyce V, Bendavid E, Olshen RA, Hlatky M, Chow A, et al. Risk of cardiovascular events associated with current exposure to HIV antiretroviral therapies in a US veteran population. Clin Infect Dis. 2015;61(3):445–52. [PubMed: 25908684]
- 106. Fink RI, Kolterman OG, Griffin J, Olefsky JM. Mechanisms of Insulin Resistance in Aging. The Journal of Clinical Investigation. 1983;71(6):1523–35. [PubMed: 6345584]
- 107. Petersen KF, Befroy D, Dufour S, Dziura J, Ariyan C, Rothman DL, et al. Mitochondrial Dysfunction in the Elderly: Possible Role in Insulin Resistance. Science. 2003;300(5622):1140– 2. [PubMed: 12750520]
- 108. Feinstein MJ, Achenbach CJ, Stone NJ, Lloyd-Jones DM. A Systematic Review of the Usefulness of Statin Therapy in HIV-Infected Patients. Am J Cardiol. 2015;115(12):1760–6. [PubMed: 25907504]
- 109. Eckard AR, Jiang Y, Debanne SM, Funderburg NT, McComsey GA. Effect of 24 weeks of statin therapy on systemic and vascular inflammation in HIV-infected subjects receiving antiretroviral therapy. J Infect Dis. 2014;209(8):1156–64. [PubMed: 24415784]
- 110. Funderburg NT, Jiang Y, Debanne SM, Labbato D, Juchnowski S, Ferrari B, et al. Rosuvastatin reduces vascular inflammation and T-cell and monocyte activation in HIV-infected subjects on antiretroviral therapy. J Acquir Immune Defic Syndr. 2015;68(4):396–404. [PubMed: 25514794]
- 111. Funderburg NT, Jiang Y, Debanne SM, Storer N, Labbato D, Clagett B, et al. Rosuvastatin treatment reduces markers of monocyte activation in HIV-infected subjects on antiretroviral therapy. Clin Infect Dis. 2014;58(4):588–95. [PubMed: 24253250]
- 112. Longenecker CT, Hileman CO, Funderburg NT, McComsey GA. Rosuvastatin preserves renal function and lowers cystatin C in HIV-infected subjects on antiretroviral therapy: the SATURN-HIV trial. Clin Infect Dis. 2014;59(8):1148–56. [PubMed: 25015912]
- 113. Sandler NG, Wand H, Roque A, Law M, Nason MC, Nixon DE, et al. Plasma levels of soluble CD14 independently predict mortality in HIV infection. J Infect Dis. 2011;203(6):780–90. [PubMed: 21252259]
- 114. Nou E, Lu MT, Looby SE, Fitch KV, Kim EA, Lee H, et al. Serum oxidized low-density lipoprotein decreases in response to statin therapy and relates independently to reductions in coronary plaque in patients with HIV. AIDS. 2016;30(4):583–90. [PubMed: 26558731]
- 115. Hileman CO, Turner R, Funderburg NT, Semba RD, McComsey GA. Changes in oxidized lipids drive the improvement in monocyte activation and vascular disease after statin therapy in HIV. AIDS. 2016;30(1):65–73. [PubMed: 26731754]
- 116. Nishi K, Itabe H, Uno M, Kitazato KT, Horiguchi H, Shinno K, et al. Oxidized LDL in Carotid Plaques and Plasma Associates With Plaque Instability. Arteriosclerosis, Thrombosis, and Vascular Biology. 2002;22(10):1649–54.
- 117. Zanni MV, Fitch KV, Feldpausch M, Han A, Lee H, Lu MT, et al. 2013 American College of Cardiology/American Heart Association and 2004 Adult Treatment Panel III cholesterol guidelines applied to HIV-infected patients with/without subclinical high-risk coronary plaque. AIDS. 2014;28(14):2061–70. [PubMed: 25265074]
- 118. Gilbert JM, Fitch KV, Grinspoon SK. HIV-Related Cardiovascular Disease, Statins, and the REPRIEVE Trial. Topics in antiviral medicine. 2016;23(4):146–9.
- 119. Mitka M Exploring statins to decrease hiv-related heart disease risk. JAMA. 2015;314(7):657–9. [PubMed: 26222872]
- 120. Bergheanu SC, Reijmers T, Zwinderman AH, Bobeldijk I, Ramaker R, Liem A-H, et al. Lipidomic approach to evaluate rosuvastatin and atorvastatin at various dosages: investigating differential effects among statins. Current Medical Research and Opinion. 2008;24(9):2477–87. [PubMed: 18655752]
- 121. Central Committee for M, Community Program of the American Heart A. Dietary fat and its relation to heart attacks and strokes. JAMA. 1961;175(5):389–91. [PubMed: 14447694]
- 122. Mozaffarian D, Micha R, Wallace S. Effects on coronary heart disease of increasing polyunsaturated fat in place of saturated fat: a systematic review and meta-analysis of randomized controlled trials. PLoS Med. 2010;7(3):e1000252. [PubMed: 20351774]

- 123. Calder PC. n−3 Polyunsaturated fatty acids, inflammation, and inflammatory diseases. The American Journal of Clinical Nutrition. 2006;83(6):1505S–19S. [PubMed: 16841861]
- 124. Davidson MH. Mechanisms for the hypotriglyceridemic effect of marine omega-3 fatty acids. Am J Cardiol. 2006;98(4A):27i–33i.
- 125. Appel LJ, Miller ER Iii, Seidler AJ, Whelton PK. Does supplementation of diet with 'fish oil' reduce blood pressure? A meta-analysis of controlled clinical trials. Archives of Internal Medicine. 1993;153(12):1429–38. [PubMed: 8141868]
- 126. Fedor D, Kelley DS. Prevention of insulin resistance by n-3 polyunsaturated fatty acids. Curr Opin Clin Nutr Metab Care. 2009;12(2):138–46. [PubMed: 19202385]
- 127. Chowdhury R, Warnakula S, Kunutsor S, et al. Association of dietary, circulating, and supplement fatty acids with coronary risk: A systematic review and meta-analysis. Annals of Internal Medicine. 2014;160(6):398–406. [PubMed: 24723079]
- 128. Hileman CO, Carman TL, Storer NJ, Labbato DE, White CA, McComsey GA. Omega-3 fatty acids do not improve endothelial function in virologically suppressed HIV-infected men: a randomized placebo-controlled trial. AIDS Res Hum Retroviruses. 2012;28(7):649–55. [PubMed: 21870979]

Bowman and Funderburg Page 17 and Page 17

Figure 1.

Lipid abnormalities and enhanced CVD risk in HIV+ individuals may be driven by HIV itself, its treatment with ART, or the immunologic consequences associated with chronic HIV-infection