

Serum R-Spondin 1 Is a New Surrogate Marker for Obesity and Insulin Resistance

Yea Eun Kang, Ji Min Kim, Hyon-Seung Yi, Kyong Hye Joung, Ju Hee Lee, Hyun Jin Kim, Bon Jeong Ku

Department of Internal Medicine, Chungnam National University College of Medicine, Daejeon, Korea

Background: Recent *in vivo* studies indicated that R-spondin 1 (RSPO1) regulates food intake and increases insulin secretion, but its role in humans remains unknown. This study investigated the association between serum levels of RSPO1 and diverse metabolic parameters in humans.

Methods: The study population consisted of 43 subjects with newly diagnosed diabetes mellitus, and 79 non-diabetic participants. Serum levels of RSPO1 were measured using the enzyme-linked immunosorbent assay. The relationships between circulating RSPO1 and diverse metabolic parameters were analyzed.

Results: Circulating RSPO1 levels increased to a greater extent in the obese group than in the lean group. Moreover, serum levels of RSPO1 were higher in the insulin-resistant group than in the insulin-sensitive group. Serum levels of RSPO1 were significantly correlated with a range of metabolic parameters including body mass index, fasting C-peptide, homeostasis model assessment of insulin resistance index, and lipid profile. Moreover, levels were significantly associated with insulin resistance and obesity in non-diabetic subjects.

Conclusion: This study demonstrated the association between serum levels of RSPO1 and a range of metabolic parameters in humans. Serum levels of RSPO1 are significantly related to obesity and insulin resistance, although the precise mechanisms remain unknown.


Keywords: Insulin resistance; Obesity; RSPO1 protein, human


INTRODUCTION

Wnt signaling plays essential roles during embryonic development and has been implicated in both physiological and pathologic conditions [1,2]. Binding of Wnt ligands to the receptor complex results in stabilization and nuclear translocation of β -catenin, binding to the transcriptional repressor, and activation of target genes [3,4]. Previously, many researchers focused the link of Wnt signaling and metabolic disease, and most of observations suggested that the upregulation of Wnt signaling blocked adipose tissue expansion [5-8]. However, a

recent study indicated that WNT5A expression was significantly positively correlated with obesity [9,10]. Despite inconsistent reports, there is increasing research interest regarding which components of the Wnt signaling pathway play specific roles in the regulation of metabolism.

The secreted R-spondin proteins 1-4 (RSPO1-4) were identified as endogenous ligands to potentiate Wnt signaling [11,12]. These proteins show 40% to 60% identity to each other, and have similar structures as a thrombospondin-like domain [13]. RSPOs can bind leucine-rich repeat-containing G protein-coupled receptors 4-6 (LGR4-6), group B of the LGR family,

Corresponding authors: Bon Jeong Ku  <https://orcid.org/0000-0002-3414-8949>
Department of Internal Medicine, Chungnam National University College of Medicine,
282 Munhwa-ro, Jung-gu, Daejeon 35015, Korea
E-mail: bonjeong@cnu.ac.kr

Hyun Jin Kim  <https://orcid.org/0000-0002-6760-4963>
Department of Internal Medicine, Chungnam National University College of Medicine,
282 Munhwa-ro, Jung-gu, Daejeon 35015, Korea
E-mail: kimhj43@cnuh.co.kr

Received: Apr. 18, 2018; Accepted: Jun. 22, 2018

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Copyright © 2019 Korean Diabetes Association <http://e-dmj.org>

and potentiate Wnt signaling [14]. A recent study indicated that *Lgr4* and its ligands, RSPO1 and RSPO3, are expressed in hypothalamic energy homeostatic areas of male rats [15]. Interestingly, expression levels of RSPO1 and RSPO3 were down-regulated by fasting and increased by insulin, which suggests that RSPOs play roles as anorexigenic factors to regulate energy homeostasis [15]. RSPO1 was also detected in murine islets, and stimulated insulin secretion in a glucose-dependent manner in both the MIN6 cell line and in mouse islets [16]. However, there have been no studies regarding the roles of RSPOs in human patients with metabolic diseases. A recent study revealed a role of LGR4, the receptor for RSPOs, in human metabolic disease. Mice with ablation of *Lgr4*, encoding the specific receptor for RSPO1, showed reduced adiposity and improved glucose metabolism through upregulation of energy expenditure via white-to-brown fat transition [17]. The authors also explored the association between human *LGR4* variant and activated point mutation (A750T), with human obesity [17]. These observations regarding LGR4 in human metabolic disease were not consistent with previous reports focusing on the role of RSPO1 in metabolic disease, although RSPO1 binds to LGR4 with greater affinity than other RSPOs [14]. Moreover, data on the role of RSPO1 in human subjects with metabolic disease are limited. To investigate the potential role and value of RSPO1 in metabolic disease, we measured and compared plasma levels of RSPO1 between subjects with and without diabetes.

METHODS

Subjects

The initial study population consisted of 134 participants from the outpatient clinic of the Division of Endocrinology and Metabolism of Chungnam National University Hospital (Daejeon, Korea) between January 2014 and December 2017. The inclusion criteria were as follows: age >18 years, absence of any clinical signs of infection or inflammation, systolic blood pressure (SBP) <140 mm Hg and diastolic blood pressure (DBP) <100 mm Hg, no alcohol or drug abuse, no history of diagnosis with diabetes mellitus (DM), and no pregnancy. Twelve participants were excluded because of liver cirrhosis ($n=1$), malignancies (prostate cancer, $n=1$; liver cancer, $n=2$; and pituitary adenoma, $n=1$), and clinical signs of infection or inflammation (pneumonia, $n=1$; acute gastroenteritis, $n=2$; and fever of unknown origin, $n=4$). Finally, 43 participants with

newly diagnosed type 2 DM and 79 control non-diabetic participants were included in this study. In our study, the participants were divided into three groups: non-obese subjects ($n=45$), obese subjects ($n=34$), and patients with type 2 DM ($n=43$). Our experimental protocol was performed in accordance with the Declaration of Helsinki and The Institutional Review Board of Chungnam National University Hospital (CNUH 2014-12) approved the protocol for this research, and written informed consent was obtained from all the subjects.

Clinical parameters

All participants underwent physical examinations on the day on which the study commenced. Height, body weight, SBP, and DBP were recorded. Blood pressure was measured after 10 minutes of rest from the right arm of each patient in the seated position using an Omron IntelliSense Automatic Blood Pressure Monitor (Omron, Kyoto, Japan). Height and body weight were determined without shoes in the morning. Body mass index (BMI) was calculated as weight in kilograms (kg) divided by height in meters squared (m^2).

Measurement of biochemical parameters

All blood samples were collected in the morning after an overnight fast of >8 hours into tubes containing ethylenediaminetetraacetic acid (EDTA) [18]. We measured the levels of fasting glucose, C-peptide, insulin, triglycerides (TGs), total cholesterol, low density lipoprotein cholesterol (LDL-C), high density lipoprotein cholesterol (HDL-C), apolipoprotein B, aspartate aminotransferase, alanine aminotransferase (ALT), blood urea nitrogen, creatinine, high-sensitivity C-reactive protein (hsCRP), and glycosylated hemoglobin (HbA1c). Blood chemistry and lipid profiles were measured using a blood chemistry analyzer (Hitachi 747; Hitachi, Tokyo, Japan). HbA1c was measured by high-performance liquid chromatography (BioRad, Hercules, CA, USA). Insulin and C-peptide were measured using a radioimmunoassay (Roche, Penzberg, Germany). We also performed the 75 g oral glucose tolerance test. The homeostasis model assessment of insulin resistance (HOMA-IR) was calculated as follows: fasting insulin level ($\mu\text{IU/mL}$) \times fasting glucose level (in mM)/22.5. The homeostasis model assessment of β -cell index (HOMA- β) was calculated as follows: fasting insulin level ($\mu\text{IU/mL}$) \times 20/fastest glucose level (in mM) -3.5 [19,20]. In addition, we defined obesity as BMI ≥ 25 , calculated as weight in kilograms divided by height in meters squared, following the WHO Asia-Pacific

classification guidelines [21]. DM was defined according to the American Diabetes Association criteria [22].

Measurements of serum levels of RSPO1

Serum levels of RSPO1 were measured using a commercially available quantitative sandwich enzyme-linked immunosor-

bent assay (ELISA) kit (Catalog No. DY4645-05; R&D Systems, Minneapolis, MN, USA). The ELISA quantitatively immunoassayed human RSPO1 levels. The intra-assay coefficient of variation (CV) of the ELISA kit was 5.7% and the inter-assay CV of the ELISA Kit was 9.5% in our laboratory.

Table 1. Characteristics of participants and the association of serum levels of RSPO1 with obesity and DM

Variable	All participant (n=122)	Non-DM patients (n=79)	DM patients (n=43)	P value ^a	Non-obese subjects (n=45)	Obese subjects (n=34)	DM patients (n=43)	P value ^b
Age, yr	50.6±11.8	50.5±12.0	50.7±11.6	0.912	51.3±9.1	49.3±15.2	50.7±11.6	0.767
Sex, male/female	54/68	34/45	20/23	0.712	17/28	17/17	20/23	0.520
BMI, kg/m ²	24.9±4.0	24.6±3.5	25.4±5.0	0.329	22.3±1.9	27.0±2.3 ^c	25.4±5.0 ^c	<0.001
Body weight, kg	65.9±13.4	64.8±12.5	67.5±14.7	0.319	58.8±8.9	71.9±11.4 ^c	67.5±14.7 ^c	<0.001
Height, cm	160.8±17.7	159.5±21.7	162.7±8.7	0.287	161.2±7.8	158.5±9.9	162.7±8.7	0.548
SBP, mm Hg	126.2±12.5	124.7±13.6	128.6±10.3	0.124	122.5±11.3	128.1±16.3	128.6±10.3	0.070
DBP, mm Hg	77.0±9.0	75.4±8.8	79.4±8.7	0.027	73.8±8.3	77.8±9.3	79.4±8.7 ^c	0.018
Fasting glucose, mg/dL	126.5±54.2	99.1±10.3	176.7±65.4	<0.001	98.9±10.6	99.4±10.0	176.7±65.4 ^{cd}	<0.001
Fasting insulin, µIU/mL	10.6±5.2	9.8±4.6	12.1±6.0	0.034	8.1±3.2	12.1±5.3 ^c	12.1±6.0 ^c	<0.001
Fasting C-peptide, ng/mL	0.8±0.4	0.7±0.4	0.9±0.4	0.016	0.6±0.3	0.9±0.5 ^c	0.9±0.4 ^c	<0.001
Post-load 2-hr glucose, mg/dL	184.9±119.4	119.9±31.8	320.1±121.7	<0.001	117.3±33.9	123.4±28.6	320.1±121.7 ^{cd}	<0.001
Post-load 2-hr insulin, µIU/mL	54.1±54.1	53.1±55.2	56.4±52.3	0.755	43.9±34.4	65.2±73.3	56.4±52.3	0.216
Post-load 2-hr C-peptide, ng/mL	3.1±1.6	3.1±1.4	3.0±2.0	0.652	2.8±1.2	3.5±1.6	3.0±2.0	0.218
HbA1c, %	6.4±2.0	5.4±0.3	8.1±2.5	<0.001	5.4±0.4	5.4±0.3	8.1±2.5 ^{cd}	<0.001
HOMA-IR	3.3±2.2	2.4±1.2	5.0±2.5	<0.001	2.0±0.7	3.1±1.2 ^c	5.0±2.5 ^{cd}	<0.001
HOMA-β	29.8±17.6	32.1±16.7	25.4±18.6	0.053	25.9±11.1	40.4±19.2 ^c	25.4±18.6 ^{cd}	<0.001
TG, mg/dL	135.6±97.9	126.8±94.0	151.6±104.0	0.202	113.4±67.6	145.7±120.6	151.6±104.0	0.152
TC, mg/dL	198.3±40.0	195.8±34.9	203.1±47.9	0.339	192.0±35.7	200.1±33.6	203.1±47.9	0.398
LDL-C, mg/dL	124.0±35.9	121.0±31.9	129.3±42.2	0.235	116.3±31.7	127.8±31.4	129.3±42.2	0.193
HDL-C, mg/dL	54.2±14.0	55.9±13.8	51.0±14.0	0.075	58.1±15.2	52.6±11.0	51.0±14.0 ^c	0.048
Apo-lipoprotein A, mg/dL	131.8±29.2	128.8±25.2	135.4±33.7	0.500	130.8±22.3	125.9±30.5	135.4±33.7	0.749
Apo-lipoprotein B, mg/dL	101.2±30.7	86.6±21.2	118.4±31.6	0.001	85.6±23.8	88.0±17.9	118.4±31.6 ^c	0.004
AST, U/L	23.2±10.9	21.8±7.5	25.6±15.1	0.069	21.3±6.8	22.6±8.4	25.6±15.1	0.169
ALT, U/L	25.8±17.2	23.6±16.3	30.0±18.4	0.063	20.8±12.0	27.4±20.3	30.0±18.4 ^c	0.036
hsCRP, mg/L	1.6±4.3	1.1±1.8	4.0±9.5	0.021	0.9±1.7	1.4±1.8	4.0±9.5	0.062
RSPO1, pg/mL	51.4±13.2	58.0±29.8	56.8±16.2	0.784	51.4±13.0	66.8±41.6 ^c	56.8±16.2	0.030

Values are presented as mean ± standard deviation or numbers of participants.

RSPO1, R-spondin 1; DM, diabetes mellitus; BMI, body mass index; SBP, systolic blood pressure; DBP, diastolic blood pressure; HbA1c, glycosylated hemoglobin; HOMA-IR, homeostasis model assessment of insulin resistance; HOMA-β, homeostasis model assessment of β-cell; TG, triglyceride; TC, total cholesterol; LDL-C, low density lipoprotein cholesterol; HDL-C, high density lipoprotein cholesterol; AST, aspartate aminotransferase; ALT, alanine aminotransferase; hsCRP, high-sensitivity C-reactive protein.

^aP value from unpaired *t*-test for continuous parametric variables and Mann-Whitney *U* test for nonparametric variables between non-DM subjects and DM patients, ^bP value from analysis of variance (ANOVA) analysis for continuous parametric variables and Mann-Whitney *U* test for nonparametric variables among non-obese subjects, obese subjects, and DM patients, ^cP<0.05 obese subjects or DM patients versus non-obese subjects, ^dP<0.05 obese subjects versus DM patients.

Statistical analyses

Continuous variables are expressed as the mean \pm standard deviation (SD), and the significance of between-group differences was evaluated using the Student *t*-test or Mann-Whitney *U* test. Categorical variables are expressed as percentages, and between-group differences were compared to the chi-square test. To evaluate associations between serum levels of RSPO1 and other variables, we subjected the data to Pearson's and partial correlation analyses. A two-tailed $P < 0.05$ was considered statistically significant. All statistical analyses were performed with SPSS version 22.0 (IBM Co., Armonk, NY, USA).

RESULTS

Baseline characteristics of the participants and comparison of serum levels of RSPO1 in relation to obesity, insulin resistance, and DM

We enrolled both healthy control subjects and drug-naïve patients with newly diagnosed DM to evaluate the role of serum RSPO1 in humans. We measured serum levels of RSPO1 in all participants, and the clinical parameters of the 122 participants enrolled in the study are presented in Table 1. The proportion of male participants was 44.3%. The study population had a mean age of 50.6 ± 11.8 years, mean BMI of 24.9 ± 4.0 kg/m², mean fasting serum glucose level of 126.5 ± 54.2 mg/dL, and mean HbA1c of $6.4\% \pm 2.0\%$. We compared the serum levels of RSPO1 in relation to obesity, insulin resistance, and DM (Fig. 1). Serum levels of RSPO1 were significantly different between the lean group (BMI < 25 kg/m²) and obese group (BMI ≥ 25 kg/m²) with values of 53.0 ± 12.2 and 66.2 ± 39.0 pg/mL, re-

spectively (Fig. 1A). In addition, we divided all participants into an insulin-resistant group (HOMA-IR ≥ 2.5) and insulin-sensitive (HOMA-IR < 2.5) group, and compared the serum levels of RSPO1 between the two groups. Serum levels of RSPO1 were significantly higher in the insulin-resistant group (63.4 ± 32.6 pg/mL) than in the insulin-sensitive group (50.4 ± 9.8 pg/mL) (Fig. 1B). To investigate the exact relationship of serum levels of RSPO1 and diabetes, we also compared the metabolic parameters and serum levels of RSPO1 between non-DM subjects and DM patients (Table 1). Although fasting glucose levels, fasting insulin levels, fasting C-peptide levels, HOMA-IR index, and HOMA- β index was significantly increased in DM patients compared to in non-DM subjects, there were no significant differences between non-diabetic participants (58.0 ± 29.8 pg/mL) and diabetic participants (56.8 ± 16.2 pg/mL) (Fig. 1C). Moreover, we compared diverse metabolic parameters among non-obese group, obese group, and DM patients (Table 1). Interestingly, serum levels of RSPO1 were significantly increased in obese subjects compared to in non-obese subjects. However, there was no association of circulating RSPO1 levels with diabetes (Table 1).

Relationships between circulating RSPO1 levels and metabolic parameters

The relationships between metabolic parameters and serum levels of RSPO1 were investigated using correlation analyses. First, we investigated the association between serum levels of RSPO1 and a range of clinical parameters including BMI, fasting glucose, fasting C-peptide, HbA1c, HOMA-IR, HOMA- β , lipid profile, liver enzymes, and hsCRP in all participants (Ta-

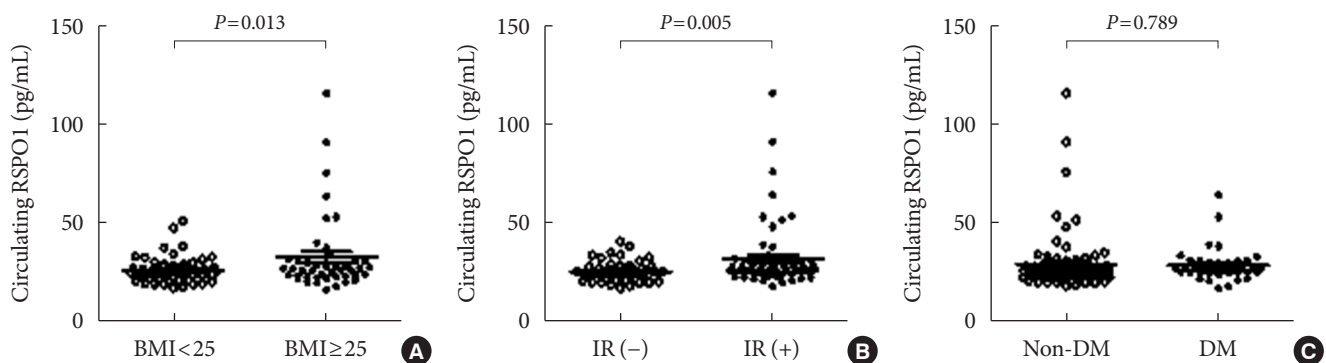


Fig. 1. Comparison of R-spondin 1 (RSPO1) levels in human in relation to diverse metabolic parameters. (A) Comparison of RSPO1 levels between lean group and obese group. (B) Comparison of RSPO1 levels between insulin-sensitive group and insulin resistant group (IR). (C) Comparison of RSPO1 levels between non-diabetic participants and diabetic participants. BMI, body mass index; DM, diabetes mellitus.

Table 2. Correlation analysis between serum RSPO1 level and diverse parameters in relation to obesity and DM

Variable	All participants (n=123)		Non-DM subjects (n=79)		Non-obese subjects (n=45)		Obese subjects (n=34)		DM patients (n=43)	
	<i>r</i> ^a	<i>P</i> value	<i>r</i> ^a	<i>P</i> value	<i>r</i> ^a	<i>P</i> value	<i>r</i> ^a	<i>P</i> value	<i>r</i> ^a	<i>P</i> value
BMI	0.244	0.010 ^b	0.343	0.005 ^b	0.242	0.014 ^b	0.407	0.017 ^b	0.084	0.593
Fasting glucose	0.108	0.237	-0.058	0.612	0.161	0.291	0.056	0.751	0.035	0.825
Fasting insulin	0.155	0.088	0.256	0.023 ^b	0.033	0.831	0.265	0.129	-0.156	0.319
Fasting C-peptide	0.244	0.007 ^c	0.266	0.019 ^b	0.062	0.688	0.347	0.048 ^b	0.016	0.920
Post-load 2-hr glucose	0.194	0.039 ^b	0.102	0.378	0.002	0.989	-0.067	0.928	0.029	0.863
Post-load 2-hr insulin	0.157	0.093	0.185	0.103	0.144	0.345	-0.201	0.254	0.085	0.623
Post-load 2-hr C-peptide	0.167	0.079	0.175	0.131	<0.001	0.998	0.294	0.097	0.164	0.345
HbA1c	0.098	0.285	0.025	0.829	0.032	0.836	0.005	0.980	-0.165	0.634
HOMA-IR	0.311	0.001 ^c	0.367	0.001 ^b	0.203	0.180	0.274	0.117	0.001	0.995
HOMA-β	0.100	0.271	0.238	0.035 ^b	0.087	0.568	0.184	0.298	-0.087	0.579
TG	0.198	0.031 ^b	0.205	0.074	0.013	0.934	0.295	0.102	0.052	0.745
TC	0.115	0.213	0.096	0.406	0.039	0.799	0.002	0.991	0.048	0.762
LDL-C	0.153	0.099	0.119	0.306	0.074	0.630	0.011	0.953	0.107	0.501
HDL-C	-0.199	0.032 ^b	-0.253	0.027 ^b	-0.153	0.315	-0.306	0.094	0.106	0.509
Apo-lipoprotein A	-0.222	0.187	-0.134	0.573	-0.366	0.244	-0.024	0.955	-0.405	0.107
Apo-lipoprotein B	0.456	0.005 ^c	0.374	0.104	0.200	0.534	0.714	0.047 ^b	0.171	0.513
AST	0.106	0.248	0.214	0.060	-0.001	0.995	0.327	0.064	-0.187	0.236
ALT	0.234	0.010 ^b	0.268	0.018 ^b	0.052	0.708	0.377	0.031 ^b	-0.050	0.755
hsCRP	0.320	0.004 ^c	0.300	0.015 ^b	0.505	0.104	0.354	0.044 ^b	-0.155	0.614

RSPO1, R-spondin 1; DM, diabetes mellitus; BMI, body mass index; HbA1c, glycosylated hemoglobin; HOMA-IR, homeostasis model assessment of insulin resistance; HOMA-β, homeostasis model assessment of β-cell; TG, triglyceride; TC, total cholesterol; LDL-C, low density lipoprotein cholesterol; HDL-C, high density lipoprotein cholesterol; AST, aspartate aminotransferase; ALT, alanine aminotransferase; hsCRP, high-sensitivity C-reactive protein.

^aCoefficients (*r*) were calculated using Spearman's method, ^b*P*<0.05, ^c*P*<0.01.

ble 2). Interestingly, increased serum levels of RSPO1 were positively correlated with BMI ($r=0.244$, $P=0.010$), fasting C-peptide ($r=0.244$, $P=0.007$), post-prandial glucose ($r=0.194$, $P=0.039$), HOMA-IR ($r=0.311$, $P=0.001$), TG ($r=0.198$, $P=0.031$), apolipoprotein B ($r=0.456$, $P=0.005$), ALT ($r=0.234$, $P=0.010$), and hsCRP ($r=0.320$, $P=0.004$) (Table 2). In addition, there was a significant negative correlation between RSPO1 level and HDL-C ($r=-0.199$, $P=0.032$). However, RSPO1 level was not associated with serum fasting glucose ($r=-0.108$, $P=0.237$) or HbA1C ($r=0.098$, $P=0.285$) levels (Table 2). Most of the parameters significantly related to serum levels of RSPO1 in correlation analyses were factors associated with DM including obesity, insulin resistance, and insulin secretion. Therefore, we investigated the correlations of serum levels of RSPO1 with a range of metabolic parameters in rela-

tion to the obesity in non-diabetic patients. Interestingly, RSPO1 levels were positively correlated with BMI ($r=0.343$, $P=0.005$), fasting C-peptide ($r=0.266$, $P=0.019$), fasting insulin ($r=0.256$, $P=0.023$), HOMA-IR ($r=0.367$, $P=0.001$), HOMA-β ($r=0.238$, $P=0.035$), ALT ($r=0.268$, $P=0.018$), and hsCRP ($r=0.300$, $P=0.015$) in non-diabetic participants. In addition, they were significantly negatively correlated with HDL-C ($r=-0.253$, $P=0.027$) in non-diabetic participants (Table 2). Additionally, we performed the correlation analysis in relation to obesity in non-diabetic participants. Non-obese subjects revealed the positive correlation of serum levels of RSPO1 and BMI ($r=0.242$, $P=0.014$); however, other parameters showed no significance with serum levels of RSPO1. Obese subjects revealed the positive correlation of serum levels of RSPO1 and BMI ($r=0.407$, $P=0.017$), fasting C-peptide

Table 3. Comparison of metabolic parameters of study population by serum RSPO1 levels in non-DM participants

Variable	Low RSPO1 group (n=39)	High RSPO1 group (n=40)	P value
Age, yr	51.9±12.0	49.2±12.1	0.327
BMI, kg/m ²	23.3±2.7	25.6±3.4	0.003
Body weight, kg	63.0±11.3	67.3±13.4	0.159
Height, cm	162.4±8.2	157.9±29.4	0.421
SBP, mm Hg	124.0±12.8	125.4±14.7	0.691
DBP, mm Hg	75.1±8.2	75.7±9.6	0.813
Fasting glucose, mg/dL	99.9±9.8	98.4±10.9	0.516
Fasting insulin, µU/mL	9.4±4.9	10.3±4.5	0.390
Fasting C-peptide, ng/mL	0.6±0.3	0.8±0.4	0.128
Post-load 2-hr glucose, mg/dL	119.0±32.2	120.7±31.7	0.816
Post-load 2-hr insulin, µU/mL	48.5±57.6	57.5±53.2	0.476
Post-load 2-hr C-peptide, ng/mL	2.9±1.4	3.3±1.4	0.249
HbA1c, %	5.4±0.4	5.4±0.3	0.842
HOMA-IR	2.1±0.9	2.8±1.2	0.007
HOMA-β	30.3±17.7	33.9±15.6	0.335
TG, mg/dL	110.3±64.5	142.9±114.3	0.127
TC, mg/dL	190.2±30.7	201.2±38.1	0.169
LDL-C, mg/dL	115.8±28.6	126.3±34.4	0.154
HDL-C, mg/dL	58.1±12.3	23.7±15.1	0.166
AST, U/L	20.3±5.2	23.4±9.1	0.063
ALT, U/L	19.2±9.0	28.0±20.4	0.017
hsCRP, mg/L	2.9±1.4	3.3±1.4	0.387
RSPO1, pg/mL	44.6 ± 3.6	71.0±37.6	<0.001

Values are presented as mean±standard deviation. P value from unpaired *t*-test for continuous parametric variables and Mann-Whitney *U* test for nonparametric variables.

RSPO1, R-spondin 1; DM, diabetes mellitus; BMI, body mass index; SBP, systolic blood pressure; DBP, diastolic blood pressure; HbA1c, glycosylated hemoglobin; HOMA-IR, homeostasis model assessment of insulin resistance; HOMA-β, homeostasis model assessment of β-cell; TG, triglyceride; TC, total cholesterol; LDL-C, low density lipoprotein cholesterol; HDL-C, high density lipoprotein cholesterol; AST, aspartate aminotransferase; ALT, alanine aminotransferase; hsCRP, high-sensitivity C-reactive protein.

($r=0.347$, $P=0.048$), Apo-lipoprotein B ($r=0.714$, $P=0.047$), and ALT ($r=0.337$, $P=0.031$). We also investigated the correlations of serum levels of RSPO1 with a range of metabolic parameters in relation to the presence of DM. However, there

were no significant relationships between RSPO1 levels and metabolic parameters in diabetic patients.

Comparison of metabolic parameters between the low and high RSPO1 groups among non-diabetic participants

Serum levels of RSPO1 were associated with obesity and insulin resistance, but the results of correlation analyses revealed significant associations of RSPO1 with metabolic parameters only in non-diabetic patients. To determine the contribution of serum RSPO1 to insulin resistance, we compared a range of metabolic parameters between the low and high RSPO1 groups of non-diabetic subjects. The participants were divided into two groups according to the median RSPO1 level in total patients. The clinical characteristics of the two groups are shown in Table 3. BMI and HOMA-IR index were significantly higher in the high RSPO1 group than in the low RSPO1 group (Table 3). In addition, ALT levels were significantly elevated in the high RSPO1 group compared to the low RSPO1 group.

DISCUSSION

We identified novel associations between serum RSPO1 and human obesity and insulin resistance. We explored the potential involvement of upregulation of circulating RSPO1 in metabolic diseases in humans. In addition, we demonstrated that serum levels of RSPO1 were positively correlated with obesity and insulin resistance in non-diabetic subjects.

Our results are not consistent with the previously described mechanism of action of RSPO1 *in vitro* and *in vivo* [16,23]. The major source of RSPOs is unknown [24], but there has been a great deal of research interest in the action of RSPOs in enhancing Wnt signaling, which plays important roles in the development and pathogenesis of various diseases [25]. A previous study in transgenic mice expressing human RSPO1 suggested that human RSPO1 is a thrombospondin domain-containing protein expressed in epithelial cells in various tissues [26]. The authors reported that transgenic mice overexpressing RSPO1 exhibited proliferation of intestinal crypt epithelial cells through upregulation of Wnt signaling. Interestingly, RSPO1 was expressed in epithelial cells of other organs such as renal tubules, adrenal glands, and pancreas. Recent studies have suggested that RSPOs stimulate Wnt-dependent stem cells to maintain tissue homeostasis [27]. Moreover, previous studies have demonstrated the importance of RSPOs in multiple cancer types and have suggested the potential utility of the

therapeutic targeting of tumor-derived RSPO regulating Wnt signaling and tumorigenesis [28,29]. However, there have been few reports on the RSPO-LGR signaling axis in metabolic disease.

Here, we demonstrated that serum levels of RSPO1 were positively correlated with fasting C-peptide level, a marker of insulin secretion, and that RSPO1 levels were positively correlated with obesity and insulin resistance in human subjects. Serum levels of RSPO1 were significantly increased not only in the obese group but also in the insulin-resistant group compared to the controls. Furthermore, these results were observed in non-diabetic participants, while there were no such correlations of RSPO1 with metabolic parameters in patients with DM. Our observations suggested that, in humans, serum RSPO1 plays distinct roles in the maintenance of insulin sensitivity in non-diabetic subjects, but not in diabetic patients with disruption of glucose homeostasis.

We examined the reasons for the links between serum levels of RSPO1 and obesity and insulin resistance in humans. Although the source of RSPOs is unknown, an *in vivo* study showed that RSPO1 was expressed in hypothalamic energy homeostatic areas [15]. Under fasting conditions, hypothalamic expression levels of RSPO1 were significantly decreased, and increased in a manner dependent on insulin secretion in response to elevated glucose levels [15]. Few studies have examined the role of RSPO1 in β -cell physiology. A previous report regarding the action of RSPO1 in metabolic homeostasis showed that it is expressed in both MIN6 β -cells and mouse islets, and that it stimulates the proliferation and insulin secretion by β -cells [16]. A recent study by Huch et al. [23] also suggested that the Wnt-RSPO1 axis is activated upon pancreas injury via upregulation of the expansion of pancreas progenitor cells. These reports supported the suggestion that RSPO1 regulates insulin secretion. Our results indicated positive correlations of serum levels of RSPO1 with insulin secretion, insulin resistance, and obesity in nondiabetic participants, but no such correlations were observed in diabetic patients. These differences in patterns of RSPO1 effects in humans compared to mice may be due to a compensatory mechanism to maintain glucose homeostasis. In non-diabetic patients, increases in adiposity may induce increases in serum RSPO1, which acts as an anorexigenic hormone and insulin secretory ligands by a compensatory mechanism to maintain glucose homeostasis. However, in diabetic conditions, the compensatory mechanism of action of RSPO1 may be downregulated in response to ex-

remely high glucose levels, as there are other many regulators of Wnt signaling in adipogenesis [30].

Although, we did not evaluate the relationship between RSPO1 level and visceral adiposity, the serum levels of RSPO1 showed significant positive correlations with levels of TG, apolipoprotein B, and hsCRP. Moreover, serum levels of RSPO1 were negatively correlated with LDL-C level. There was no direct link between RSPO1 and adipogenesis, but our findings were supported by a previous report regarding the role of LGR4, the main receptor for RSPO1, in adipogenesis [17]. The authors suggested that LGR4 is necessary to maintain energy expenditure via remodeling of brown adipose tissue and explored the upregulation of LGR4 significantly related to human obesity [17]. They divided a large cohort into an obese group and a lean group based on BMI, and observed a greater increase in the prevalence of LGR4 somatic mutation in the obese group compared to the lean group study. Moreover, they showed that the levels of LGR4 expression in human intra-adipose tissue and subcutaneous tissue were increased to a greater extent in the obese group compared to the lean group. Our study did not validate the expression of LGR4 and RSPO1 in adipose tissue, but increases in serum RSPO1 may activate LGR4, a strong binding receptor for RSPO1, in adipose tissue, and may contribute to human obesity.

Our study had several limitations. First, our sample size was small and further large-scale studies will be necessary to validate the distinct role of RSPO1 in relation to glucose levels. Second, we only investigated plasma levels of RSPO1 as a regulator in Wnt signaling, and therefore measurements of other molecules affecting Wnt signaling will be necessary to determine the mechanism of action RSPO1 in regulating insulin resistance. In addition, we could not validate the expression of RSPO1 in multiple organs, including the pancreas, adipose tissue, and brain, so the major source of RSPO1 remains unclear. Finally, follow-up studies after body weight loss or regulation of glucose levels via drug treatments to examine the associations of RSPO1 with human adiposity and insulin resistance are necessary.

In conclusion, the results of this study demonstrated the associations of serum levels of RSPO1 with human obesity and insulin resistance. Although the precise contribution and mechanism are not yet clear, our results suggest that serum RSPO1 reflects insulin resistance in non-diabetic patients and may play a pivotal role in regulation of obesity-induced insulin resistance.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

AUTHOR CONTRIBUTIONS

Conception or design: Y.E.K., H.J.K., B.J.K.

Acquisition, analysis, or interpretation of data: J.M.K., H.S.Y., K.H.J.

Drafting the work or revising: Y.E.K., J.H.L.

Final approval of the manuscript: H.J.K., B.J.K.

ORCID

Yea Eun Kang <https://orcid.org/0000-0002-2012-3716>

Hyun Jin Kim <https://orcid.org/0000-0002-6760-4963>

Bon Jeong Ku <https://orcid.org/0000-0002-3414-8949>

ACKNOWLEDGMENTS

This research was supported by the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT & Future Planning (NRF-2017R1D1A1B03027820) and the Korean Government (NRF-2016R1D1A1A09916900).

REFERENCES

- Logan CY, Nusse R. The Wnt signaling pathway in development and disease. *Annu Rev Cell Dev Biol* 2004;20:781-810.
- Mammoto T, Ingber DE. Mechanical control of tissue and organ development. *Development* 2010;137:1407-20.
- Tolwinski NS, Wehrli M, Rives A, Erdeniz N, DiNardo S, Wieschaus E. Wg/Wnt signal can be transmitted through arrow/LRP5,6 and Axin independently of Zw3/Gsk3beta activity. *Dev Cell* 2003;4:407-18.
- Clevers H. Wnt/beta-catenin signaling in development and disease. *Cell* 2006;127:469-80.
- Saxena R, Gianniny L, Burt NP, Lyssenko V, Giuducci C, Sjogren M, Florez JC, Almgren P, Isomaa B, Orho-Melander M, Lindblad U, Daly MJ, Tuomi T, Hirschhorn JN, Ardlie KG, Groop LC, Altshuler D. Common single nucleotide polymorphisms in TCF7L2 are reproducibly associated with type 2 diabetes and reduce the insulin response to glucose in nondiabetic individuals. *Diabetes* 2006;55:2890-5.
- Jin T, Liu L. The Wnt signaling pathway effector TCF7L2 and type 2 diabetes mellitus. *Mol Endocrinol* 2008;22:2383-92.
- Wright WS, Longo KA, Dolinsky VW, Gerin I, Kang S, Bennett CN, Chiang SH, Prestwich TC, Gress C, Burant CF, Susulic VS, MacDougald OA. Wnt10b inhibits obesity in ob/ob and agouti mice. *Diabetes* 2007;56:295-303.
- Longo KA, Wright WS, Kang S, Gerin I, Chiang SH, Lucas PC, Opp MR, MacDougald OA. Wnt10b inhibits development of white and brown adipose tissues. *J Biol Chem* 2004;279:35503-9.
- Kanazawa A, Tsukada S, Sekine A, Tsunoda T, Takahashi A, Kashiwagi A, Tanaka Y, Babazono T, Matsuda M, Kaku K, Iwamoto Y, Kawamori R, Kikkawa R, Nakamura Y, Maeda S. Association of the gene encoding wingless-type mammary tumor virus integration-site family member 5B (WNT5B) with type 2 diabetes. *Am J Hum Genet* 2004;75:832-43.
- Fuster JJ, Zuriaga MA, Ngo DT, Farb MG, Aprahamian T, Yamaguchi TP, Gokce N, Walsh K. Noncanonical Wnt signaling promotes obesity-induced adipose tissue inflammation and metabolic dysfunction independent of adipose tissue expansion. *Diabetes* 2015;64:1235-48.
- Clevers H, Nusse R. Wnt/ β -catenin signaling and disease. *Cell* 2012;149:1192-205.
- Glinka A, Dolde C, Kirsch N, Huang YL, Kazanskaya O, Ingelfinger D, Boutros M, Cruciat CM, Niehrs C. LGR4 and LGR5 are R-spondin receptors mediating Wnt/ β -catenin and Wnt/PCP signalling. *EMBO Rep* 2011;12:1055-61.
- de Lau WB, Snel B, Clevers HC. The R-spondin protein family. *Genome Biol* 2012;13:242.
- Carmon KS, Gong X, Lin Q, Thomas A, Liu Q. R-spondins function as ligands of the orphan receptors LGR4 and LGR5 to regulate Wnt/beta-catenin signaling. *Proc Natl Acad Sci U S A* 2011;108:11452-7.
- Li JY, Chai B, Zhang W, Fritze DM, Zhang C, Mulholland MW. LGR4 and its ligands, R-spondin 1 and R-spondin 3, regulate food intake in the hypothalamus of male rats. *Endocrinology* 2014;155:429-40.
- Wong VS, Yeung A, Schultz W, Brubaker PL. R-spondin-1 is a novel beta-cell growth factor and insulin secretagogue. *J Biol Chem* 2010;285:21292-302.
- Wang J, Liu R, Wang F, Hong J, Li X, Chen M, Ke Y, Zhang X, Ma Q, Wang R, Shi J, Cui B, Gu W, Zhang Y, Zhang Z, Wang W, Xia X, Liu M, Ning G. Ablation of LGR4 promotes energy expenditure by driving white-to-brown fat switch. *Nat Cell Biol* 2013;15:1455-63.

18. Shin MY, Kim JM, Kang YE, Kim MK, Joung KH, Lee JH, Kim KS, Kim HJ, Ku BJ, Shong M. Association between growth differentiation factor 15 (GDF15) and cardiovascular risk in patients with newly diagnosed type 2 diabetes mellitus. *J Korean Med Sci* 2016;31:1413-8.
19. Kang YE, Kim JM, Joung KH, Lee JH, You BR, Choi MJ, Ryu MJ, Ko YB, Lee MA, Lee J, Ku BJ, Shong M, Lee KH, Kim HJ. The roles of adipokines, proinflammatory cytokines, and adipose tissue macrophages in obesity-associated insulin resistance in modest obesity and early metabolic dysfunction. *PLoS One* 2016;11:e0154003.
20. Kang YE, Choung S, Lee JH, Kim HJ, Ku BJ. The role of circulating Slit2, the one of the newly batokines, in human diabetes mellitus. *Endocrinol Metab (Seoul)* 2017;32:383-8.
21. Kanazawa M, Yoshiike N, Osaka T, Numba Y, Zimmet P, Inoue S. Criteria and classification of obesity in Japan and Asia-Oceania. *World Rev Nutr Diet* 2005;94:1-12.
22. American Diabetes Association. (2) Classification and diagnosis of diabetes. *Diabetes Care* 2015;38 Suppl:S8-16.
23. Huch M, Bonfanti P, Boj SF, Sato T, Loomans CJ, van de Wetering M, Sojoodi M, Li VS, Schuijers J, Gracanin A, Ringnalda F, Begthel H, Hamer K, Mulder J, van Es JH, de Koning E, Vries RG, Heimberg H, Clevers H. Unlimited in vitro expansion of adult bi-potent pancreas progenitors through the Lgr5/R-spondin axis. *EMBO J* 2013;32:2708-21.
24. Sato T, Clevers H. Growing self-organizing mini-guts from a single intestinal stem cell: mechanism and applications. *Science* 2013;340:1190-4.
25. Welters HJ, Kulkarni RN. Wnt signaling: relevance to beta-cell biology and diabetes. *Trends Endocrinol Metab* 2008;19:349-55.
26. Kim KA, Kakitani M, Zhao J, Oshima T, Tang T, Binnerts M, Liu Y, Boyle B, Park E, Emtage P, Funk WD, Tomizuka K. Mitogenic influence of human R-spondin1 on the intestinal epithelium. *Science* 2005;309:1256-9.
27. Clevers H, Loh KM, Nusse R. Stem cell signaling. An integral program for tissue renewal and regeneration: Wnt signaling and stem cell control. *Science* 2014;346:1248012.
28. Christodoulides C, Lagathu C, Sethi JK, Vidal-Puig A. Adipogenesis and WNT signalling. *Trends Endocrinol Metab* 2009;20:16-24.
29. Bennett CN, Ross SE, Longo KA, Bajnok L, Hemati N, Johnson KW, Harrison SD, MacDougald OA. Regulation of Wnt signaling during adipogenesis. *J Biol Chem* 2002;277:30998-1004.
30. Prestwich TC, Macdougald OA. Wnt/beta-catenin signaling in adipogenesis and metabolism. *Curr Opin Cell Biol* 2007;19:612-7.