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Accelerating Research on Biological Aging and Mental Health: Current Challenges and Future Directions

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Abstract

Aging is associated with complex biological changes that can be accelerated, slowed, or even temporarily reversed by biological and non-biological factors. This article focuses on the link between biological aging, psychological stressors, and mental illness. Rather than comprehensively reviewing this rapidly expanding field, we highlight challenges in this area of research and propose potential strategies to accelerate progress in this field. This effort requires the interaction of scientists across disciplines - including biology, psychiatry, psychology, and

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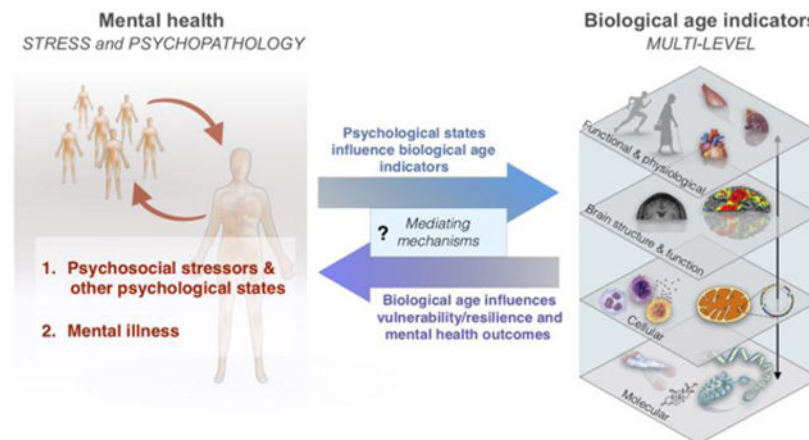
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epidemiology; and across levels of analysis that emphasize different outcome measures - functional capacity, physiological, cellular, and molecular. Dialogues across disciplines and levels of analysis naturally lead to new opportunities for discovery but also to stimulating challenges. Some important challenges consist of 1) establishing the best objective and predictive biological age indicators or combinations of indicators, 2) identifying the basis for inter-individual differences in the rate of biological aging, and 3) examining to what extent interventions can delay, halt or temporarily reverse aging trajectories. Discovering how psychological states influence biological aging, and vice versa, has the potential to create novel and exciting opportunities for healthcare and possibly yield insights into the fundamental mechanisms that drive human aging.

Graphical abstract



Keywords

biological age; psychopathology; DNA methylation; brain; mitochondria; telomere length

INTRODUCTION

Aging is the strongest risk factor for many chronic illnesses, loss of functional capacity, and mortality (Fernandes et al., 2016). It is associated with complex biological changes, but there is no consensus on the very definition of aging, nor on the best methods to quantify it biologically (Xia et al., 2017). Chronological age is based on the passage of time and is invariable. But biological age may fall behind or else outpace chronological age – it is modifiable. Based on specific molecular and other measures discussed below, the rate of biological aging has been reported to vary substantially between individuals (Cole et al., 2018; Xia et al., 2017), although the causes of such inter-individual differences are mostly unclear. In particular, a major gap in knowledge is reflected in our ignorance of the mechanisms for the transduction of psychological states, and of psychopathology, into changes in biological aging (Figure 1). How do “mind” states influence biological aging and vice versa?

This psycho-biological problem is a unique opportunity to make scientific progress on two main fronts: First, it is an opportunity to develop new measurements and technical

approaches to capture meaningful, valid, and reproducible measures of biological aging. Second, this interdisciplinary problem requires dialogue across research and clinical domains. We see the intersection of experiential, psychological, and biological aging processes as a platform for the development of new (and possibly radically different) concepts and measures that will most faithfully capture human health and the aging process. Currently, although we have some quantifiable measures of biological aging in humans - *biological age indicators* - we still know little about their causal role in the aging process, and about their modifiability by psychological states and psychopathology.

One important shared goal towards enhancing well-being across the lifespan is to understand aging as dynamic trajectories determined by a variety of factors. Some determinants of biological aging are pre-programmed (“intrinsic”; e.g., genetic), while others are affected by the environment (“extrinsic”; e.g., diet, adversity) (de Magalhães, 2012). Most definitions of biological aging include loss of function, increased propensity to certain diseases, and closer proximity to death (de Magalhães and Passos, 2018). Certain objective biological measures (or “clocks”) may also track biological aging. Development and validation of biological age indicators and clarification of their mediators and moderators are high priorities, since they may lead to a better understanding of the underpinnings of healthy and unhealthy aging trajectories. These indicators may also present proximal outcomes, or “early warning signs” that portend disease development and may provide a more sensitive platform to detect – and intervene upon – meaningful interactions between psychological, social, and bio-behavioral factors that influence aging trajectories and health outcomes.

Biological age indicators currently being investigated include telomere length (TL), epigenetic changes, alterations of mitochondrial function and mitochondrial DNA (mtDNA), age-related brain structure and function, and transcriptomic, metabolomic, and proteomic changes, among others (see, Cole et al., 2018; Jylhava et al., 2017; Xia et al., 2017 for recent reviews). Current topics of investigation include the nature of the inter-relationship of these biological age indicators, whether they measure the same or different aspects of biological aging, whether they are causally involved in the aging process, whether they have a causal role in disease and disorders, and the best ways to assess them. The possibility that the aging process is accelerated by chronic psychological stress and that it plays a role in the pathophysiology of some mental illnesses has been supported by observations that chronically stressed or psychiatrically ill individuals are at increased risk of acquiring specific age-related diseases and have a reduced life expectancy (Epel and Prather, 2018; Lindqvist et al., 2015; Penninx et al., 2013; Verhoeven et al., 2014; Walker et al., 2015). But certain conceptual and methodological obstacles are impeding growth in this field and hinder replication of findings across laboratories.

Rather than comprehensively reviewing this rapidly expanding field, here we focus on highlighting various challenges and arising opportunities for this interdisciplinary endeavor. We conclude by proposing strategies to accelerate the progress of this field towards a predictive science that can enhance our understanding of the psychobiological factors that influence the aging process and lifespan.

1. The concept of biological aging: Definitions and obstacles

Chronological age is strictly quantitative and requires no more than a calendar to measure. Biological age is more elusive as it reflects the functional and biological condition of an individual. The difference between biological age and chronological age can indicate whether the individual's biological state is "older" or "younger" than would be expected for a given chronological age. This is often referred to as "accelerated" or "slowed" aging, respectively. However, cross-sectional assessments of biological age do not allow to determine aging rates, or to distinguish between "accelerated" and "premature" or "advanced" aging (Figure 2). The rate of increase in biological aging over time may also exhibit nonlinear behavior, particularly in early life where the measured rate of aging may be more rapid than across adult life (Cole et al., 2018; Lohr et al., 2015).

Since it is not possible to directly assess the total biological state of a person, biological age indicators serve as proxies. Biological age *indicators* are functional, anatomical, biochemical, cellular or molecular measures that are correlated with age and that may reflect the health status of specific cell types and/or organ systems. The term *biomarker*, in contrast to indicators as defined here, is best used in the context of specific disease or health outcomes. By definition, biomarkers must exhibit both sensitivity and specificity in relation to the outcome we design them to predict (Abi-Dargham and Horga, 2016). For aging, a still broadly defined process compared to a disease that can be ascertained with certainty, the term indicator is rendered more appropriate. Although some biological age indicators have undoubtedly established their sensitivity to chronological age, few have convincingly demonstrated their specificity - that changes in their value occurs specifically in response to the aging process and not in response to other pathophysiological process. Some biological age indicators are indeed modified by disease states independent of aging, and some as discussed below may notably be sensitive to psychological states, namely stress and psychopathology.

2. Biological age indicators

Aging is a multifaceted and complex process that manifests across multiple levels. In recent decades, measurements spanning each of these levels have been developed, reflecting our prevailing reductionist scientific approach to biomedical sciences. Here, rather than providing an exhaustive overview that can be found in recent reviews (Engelfriet et al., 2013; Horvath and Raj, 2018; Jylhava et al., 2017; López-Otín et al., 2013; Wagner et al., 2016; Xia et al., 2017), we provide a selective overview of aging indicators commonly studied in relation to mental health (Table 1). This table, which is illustrative rather than comprehensive, also includes some emerging biological age indicators that reflect the development of omics technologies and of computational approaches to integrate multiple metrics into composite indices, as also discussed in Section 5. Due to space limitation, we do not cover self-reported age (Stephan et al., 2018), self-reported perception of aging (Levy et al., 2002), more specific brain measures (Cole, 2018), or inflammatory markers (Franceschi et al., 2000), which have also been associated with lifespan. We then discuss the practical limitations and conceptual challenges commonly associated with these measurements.

2.1. General limitations common to biological age indicators—There are limitations inherent to existing biological age indicators: they rely on specific organs or tissue types, can be confounded by cell type heterogeneity, and represent static measures of dynamic states (Figure 3). These limitations apply to most molecular biological age indicators and should represent the foundation from which we design research projects, and interpret findings. However, these limitations are often not well understood and, instead, only considered post-hoc once data is collected and is being analyzed. Here, to be consistent with the logic whereby limitations inform research design, method development, and data interpretation, we discuss these challenges prior to the literature review. Recommendations to overcome some of these limitations are also discussed in Section 5.

2.1.1. Organs and tissue types: A common limitation of biological age indicators is that they are generally measured in one particular tissue and then used as a general age estimator for the person from whom the sample was obtained. However, it is unlikely that every tissue - peripheral and central - are entirely synchronized and that one tissue accurately reflects the biological age of all other tissues. Tissues from multiple organs have been found to age at different rates in terms of epigenetic age (Horvath et al., 2015). With age, mtDNA mutations also accumulate differently between brain regions (Bender et al., 2006; Corral-Debrinski et al., 1992; Picard and Hirano, 2016) and even between cells of a given organ (Vincent et al., 2018). Within the brain, TL also varies between different cortical areas (Mamdani et al., 2015). This limitation may also apply to measures of functional capacity. For example, there is relatively poor agreement between muscle strength measured from handgrip or knee extension, suggesting that the most commonly used metric of muscle strength - handgrip strength - is not a proxy for overall muscle strength (Yeung et al., 2018). Developing approaches to measure and perhaps capitalize on the heterogeneous nature of aging dynamics across tissues requires further research.

2.1.2. Cellular composition and heterogeneity: Tissues such as the brain, heart, muscles, and the blood are composed of multiple different cell types. Although all cell types have the same genome, they show unique epigenetic, morphological, functional, and molecular differences relevant to biological age indicators. For example, different blood cell types have different epigenomes (Reinius et al., 2012), telomerase activity and TL (Boeck et al., 2018b, 2018d; Lin et al., 2010), and mitochondrial respiratory capacity (Chacko et al., 2013). These intrinsic differences are inevitable and may introduce bias and confound findings when assessed individually. Similar limitations of cellular heterogeneity also apply to saliva, skin, and any tissue such as placenta, brain, and others. However, in many cases, the relative cellular composition of these tissues is poorly characterized or methods may not be available to effectively disentangle cellular composition effects, relative to the true biological aging signal.

2.1.3. Static indicators of dynamic processes: Most biological age indicators reflect the current state of the organism, and the biological age of the sampled tissue and cells, at the moment of collection. In many cases, it is unknown how dynamic these markers are. In other words, how much they change from day-to-day, across the day (i.e., diurnal variation), or sometimes even within minutes, as is the case for neuroendocrine mediators and blood-

based metabolites. It is generally assumed that most biological age indicators (and the specific measures that compose some of them) are largely stable, changing slowly over the course of years, but that assumption has gone untested or proven false for most indicators listed in Table 1. Unrecognized, unmeasured, or uncontrolled variability of biological age indicators due to regular or irregular changes over time has two undesirable effects: it introduces noise that cannot be accounted for, and possibly limits our interpretation of the downstream result. Biological age indicators may follow different trajectories over time and may theoretically be differentially sensitive to behaviors such as sleep, exercise, diet, meditation, and others. Studies with frequently – over hours, days, months, and years – repeated measures of biological age indicators will be necessary to establish the temporal kinetics for existing and new biological age indicators.

2.2 Specific limitations to measurements of biological aging

2.2.1 Limitations of functional capacity and physiological measures: A major advantage of functional capacity and physiological measures is the high efficiency in terms of costs and collection as well as their integrative informativeness, specifically compared to blood-based, molecular and DNA-based measurements. However, the predictive value of objective functional and physiological measures on mortality has mostly been reported in older populations (Cooper et al. 2010) and in middle-aged populations (Rantanen et al., 2000). Except for some data reporting associations between handgrip strength and mortality in male adolescents (Ortega et al., 2012), whether functional capacity measures such as walking speed are sensitive to aging and predictive of morbidity and mortality in younger populations largely remain to be established.

2.2.2 Limitations of brain function- and structure-based measures: Normal aging is accompanied by brain atrophy and loss of brain tissue volume, which can be quantified non-invasively with magnetic resonance imaging (MRI). Voxel-based morphometry and surface-based analysis are two commonly used image preprocessing techniques, which may yield divergent results (Chung et al., 2017; Clarkson et al., 2011). Moreover, the macroscopic volumetric changes observed in T1- and T2-weighted MR imaging reflect microscopic changes at the tissue and cellular levels, and in many circumstances possibly represent an aggregate of multiple cellular mechanisms related to synapses, neurons, and glial cells (Tardif et al., 2016). Thus, what changes in brain volume represent is not fully understood.

Age-related differences in brain function can also be detected with functional connectivity (e.g., Geerligts et al., 2015) and novel analytics on brain response are also available. For example, Garrett and colleagues (2010) showed that the age-predictive power of the brain's signal variability was five times higher than that of the conventional method of assessing the average signal across time. But functional MRI data has low signal-to-noise ratio, and movement artifacts are one source of such noise. Because individuals of different ages might move differently in response to assessments, movement artifacts are a possible confounder in many study designs. Implementing methods to systematically review individual participants' images and manually separating noise from signal (Griffanti et al., 2017) could be a useful technique to minimize artifacts. There is evidence that manually cleaned BOLD-fMRI data, compared to data preprocessed with conventional automatic methods, better

predicts chronological age (Garrett et al., 2010), emphasizing the importance of data quality and pre-processing procedures in conclusions derived from brain-based age indicators.

A popular approach in neuroscience is to use statistical approaches to translate complex whole-brain multivariate patterns of aging into a single outcome (Luders et al., 2016), the so-called “brain age” (see Cole and Franke, 2017 for details). Brain age algorithms (Aycheh et al., 2018; Ball et al., 2017; Cole et al., 2016; Gaser et al., 2013; Liem, 2016; Schnack et al., 2016) generate accurate individual age predictions in healthy controls, but show greater prediction errors when applied to patient groups (Cole et al., 2018). Within this framework, neuropathology may be reflected by the trajectory of aberrant normal aging, rather than a *different* deteriorating pattern of pathology. Gutierrez Becker and colleagues (2018) show that Gaussian Process uncertainty in age estimation may yield a better separation between cases and healthy individuals than the prediction error. Nevertheless, brain age models have high reliability in terms of test-retest performance at both same and different scanners (Cole et al., 2016; Franke and Gaser, 2012), and have shown biologically meaningful associations with health, clinical, and neuropsychiatric phenotypes (Cole et al., 2018).

2.2.3. Limitations of cellular measures: Although cellular measures of aging have been used widely in the laboratory setting, they are seldom applied to human (clinical, epidemiological) research. For instance, replicative senescence, a cellular measure of aging, involves monitoring cells grown in culture, and counting the number of cells over time (e.g., Lawless et al., 2010). This enables the investigator to count the number of times cells divide (i.e., total population doublings or “hayflick limit”), and determine the time required per cell division (i.e., population doubling time), which increases as cells age and divide more slowly. It should be noted that although senescence - defined as the loss of the ability of a cell to grow or divide - is associated with aging, it is not equivalent and can be dissociated from chronological age. Indeed, other factors such as irradiation can specifically induce senescence, even in chronologically young cells, thus reflecting biological aging.

The assessment of cellular bioenergetics, particularly mitochondrial content and functions, represents another domain of cellular aging measures. These indicators reflect the ability of cells to generate energy through oxygen-dependent mechanisms (for a review, see Picard and McEwen, 2018a). Respiration can be measured in whole cells (Boeck et al., 2018d) where it mostly reflects cellular energy demand, or in permeabilized cells (Ehinger et al., 2015) and isolated mitochondria where the intrinsic function of the organelle can be directly assessed independent of cellular contributions (Tyrrell et al., 2015). A major limitation of functional measures on intact cells and mitochondria is that measurements must be performed rapidly after blood sampling (within minutes to hours), which limits throughput and increases technical variability between samples, requiring exceptional standardization of procedures.

Other approaches relying on lysates (homogenized cells or mitochondria) from frozen samples allow measurement of enzymatic activity (e.g., telomerase, mitochondrial respiratory chain complexes) for multiple samples at once or in large batches (Picard et al., 2018). An ubiquitous limitation to all measures of biological *activity* (as opposed to inert molecules) is the degradation of cellular and enzymatic activities over time when samples

are stored under suboptimal conditions. An important unknown in this field is the degree to which storage conditions, and especially the length of time a sample resides in a freezer, contributes to changes in assay results. This issue is worthy of applied investigation, since there is commonly a trade-off between freezing samples for shorter periods of time vs. freezing samples for longer periods of time to minimize inter-assay variability in assaying sequential frozen batches. In contrast to molecular analytes that are mostly or fully preserved at -80°C , samples destined for functional measurements should be stored in liquid nitrogen ($< -150^{\circ}\text{C}$).

2.2.4 Limitations of molecular measures: A large fraction of the most widely used biological age indicators are molecular in nature. They include DNA methylation (DNAm), metabolites, proteins, TL, mtDNAcn, circulating cell-free mtDNA (ccf-mtDNA), mtDNA damage, and others. One important consideration to all molecular measures is that inadequate handling of fresh samples can alter the concentration of various analytes, particularly metabolites. For example, whereas DNA markers are believed to be quite stable over minute to days, blood glucose concentration decreases within minutes when the blood is left at room temperature (Chan et al., 1989), owing to metabolic activities of white and red blood cells. The same must also apply to other metabolites that are detected by metabolomics. Gene expression assessed from messenger RNA transcript levels is also subject to rapid degradation and special care must be applied to blood destined to transcriptomic analyses (Kågedal et al., 2005). These effects are minimized by rapid separation of the liquid and cellular components of whole blood by centrifugation immediately after blood draw, refrigeration (immediate storage of samples on wet ice, 4°C), and subsequently freezing biological samples in a timely fashion.

Below we discuss specific molecular biological age indicators that have been subject of considerable research in relation to stress and psychopathology. Although exciting new findings from proteomic (Menni et al., 2015; Tanaka et al., 2018) and metabolomic (Chaleckis et al., 2016; Hertel et al., signatures of aging are beginning to arise, they have not been examined in relation to psychological factors. In this section, we focus our discussion on DNAm, TL, and mtDNAcn.

2.2.4.1. DNA methylation and epigenetic age: To date, several epigenetic age estimators have been developed from e.g. whole blood (Hannum et al., 2013; Levine et al., 2018; Weidner et al., 2014), neonatal cord blood and blood spots (Knight et al., 2017, and skin and blood cells (Horvath et al., 2018). Many potential confounders may cause technical variation in DNAm studies, of which population stratification and genetic ancestry are major contributors (Heijmans and Mill, 2012; Ratanatharathorn et al., 2017; Susser et al., 2016). Therefore, there is also reason to assume that genetic variation impacts epigenetic age estimates, particularly considering recent studies that report strong genetic links (Lu et al., 2018). Other potential confounders include smoking (Elliott et al., 2014), sex, and prenatal factors (Simpkin et al., 2016). Technically, DNAm arrays can show large variations between individual arrays and batches, and methods have been designed to statistically correct for these prior to analyses (Akulenko et al., 2016; Price and Robinson, 2018).

While the validity of different methylation-based predictors is questioned, applications of the original pan-tissue Horvath clock (Horvath, 2013) have been successful across tissue types and proven accurate even in embryonic brain samples (Spiers et al., 2015). The latest developed skin and blood predictor seems to be even more robust across tissue types (Horvath et al., 2018). More recently, methods using as little as 3-10 CpG sites from blood samples also accurately predict age (Li et al., 2018; Weidner et al., 2014) and mortality risk scores (Gao et al., 2018).

Whether statistical adjustment for cell type composition should be uniformly applied to whole blood-derived DNA to achieve optimal age prediction is the subject of ongoing debate. Whereas it has been argued that the Horvath method incorporates the estimation of cell type composition from blood and may not require cell type adjustment, some have shown that intrinsic (without adjustment for cell type composition) and extrinsic (with adjustment for cell type composition) age estimates may have a different biological meaning (Chen et al., 2017). Also important to mention here is that other DNAm-based indicators of aging have been developed that are trained on phenotypic markers of age (DNAm PhenoAge) rather than chronological age, leading to improved predicted risk of mortality (Levine et al., 2018). Briefly, phenotypic age is a combination of chronological age and nine disease-related biomarkers selected based on their association with mortality. The sensitivity of epigenetic predictors to psychosocial stress and psychopathology remains a gap of knowledge.

2.2.4.2. Telomere length: Issues related to TL assessments have been presented elsewhere (Aubert et al., 2012) and will not be discussed in detail here. In general, we will note that multiple different assays exist, which vary in their cost, required volume, applicability on frozen samples, and throughput. Technically, these are important considerations that impact the feasibility of clinical and epidemiological studies. Moreover, because of the relatively inexpensive and high-throughput capacity of qPCR-based methods, TL has frequently been measured on total DNA extracted from cell mixtures, which can be derived from a variety of sources such as buccal swabs (which include both epithelial cells and leukocytes) and whole blood (which include a variety of leukocytes). For these measurements, and as for mtDNA measurements below, the limitations presented in Section 2.1 are particularly important.

2.2.4.3. mtDNA copy number and circulating cell-free mtDNA: Counting the number of mtDNA molecules per cell, or mtDNA_{cn}, can indirectly provide an indication of the bioenergetic state of the cell. The mtDNA_{cn} measurements are based on either qPCR (e.g., Tyrka et al., 2016) or derived from whole exome or genome sequencing data (e.g., Cai et al., 2015) where “counts” of both the mitochondrial and nuclear genomes are estimated. The ratio of mtDNA and nuclear DNA (nDNA) is then multiplied by 2 to account for the diploid nature of the nuclear genome and taken as mtDNA_{cn} (Malik et al., 2011). Here, given that cells with different metabolic demand can differ by as much as an order of magnitude in their content of mtDNA, cell type differences may have a particularly profound effect on this measure. When applied to a homogenous cell population, mtDNA_{cn} can provide valuable information. However, most reported studies with mtDNA_{cn} have relied on whole blood DNA, which is confounded by cell type heterogeneity, and by the presence of platelets.

Platelets do not have nDNA, but have mtDNA, which artificially inflates mtDNA copy number in whole blood preparations (Hurtado-Roca et al., 2016; Urata et al., 2008). In tissues with less heterogeneity than blood, such as skeletal muscle, some have observed no difference in mtDNAcn between young and old individuals (Miller et al., 2003). The rate of decline in mtDNAcn per year also varies widely between studies, possibly as a result of differences in methodology and tissue source.

Similarly, measures of ccf-mtDNA are sensitive to cellular contamination and biological sample used. Serum (post-coagulation fraction of whole blood) may contain substantially more ccf-mtDNA than plasma (liquid fraction collected with an anticoagulant) (Xia et al., 2009). Sufficient centrifugation speed and time are required to successfully eliminate cells, particularly platelets, that could artificially inflate serum or plasma ccf-mtDNA (Nakahira et al., 2013). In studies where blood samples were not centrifuged at sufficient speeds, ccf-mtDNA levels are reportedly higher, making platelet contamination the most likely contributor to measured mtDNA levels and thus complicating interpretation of these results.

2.3. Associations among biological age indicators—While there is preliminary evidence for some cross-correlations among the different biological aging indicators, few have been examined in relation to other indicators. This highlights the need for examining multiple markers in an integrative study, as e.g. Belsky et al. (2018) recently showed low agreement between eleven quantifications of biological aging, with only modest associations to e.g. physical functioning and cognitive decline. Other studies suggest that TL is correlated to mtDNAcn (Tyrka et al., 2015), but the direction of stress and psychopathology effects with mtDNAcn or mitochondrial content (citrate synthase) and TL may vary (Boeck et al., 2018d; Cai et al., 2015; Picard et al., 2018; Tyrka et al., 2016). TL is not correlated with epigenetic age (Breitling et al., 2016; Han et al., 2018; Marioni et al., 2016), although cell type composition adjustments may reveal a modest association (Chen et al., 2017). Both epigenetic age and TL seem uncorrelated to brain age, and no associations were found between brain predicted age difference (brain-PAD) and epigenetic predicted age difference (Cole et al., 2017). The correlations between the Hannum and Horvath clocks vary from relatively strong ($r=0.76$) to low ($r=0.37$) in independent studies (Belsky et al., 2018; Chen et al., 2016), and both clocks showed modest correlations (0.10-0.33) to the transcriptomic age indicator by Peters et al. (2015). The microRNA age indicator of Huan et al. (2018) was modestly correlated to epigenetic age ($r=0.3$) and microRNA expression ($r=0.2$). Cross-correlations between metabolomic/proteomic aging and other biological aging indicators remain to be explored.

3. Do psychological stress and psychopathology influence biological aging?

Nearly 15 years have elapsed since Epel and colleagues first described the association of psychosocial stress with short leukocyte telomeres in a sample of healthy premenopausal mothers of a chronically ill child and mothers of healthy children (Epel et al., 2004). The association of shortened telomeres with stress exposure has since been replicated in a wide variety of studies, and this observation stimulated several related lines of research examining the relationship between various forms of stress exposure or perceived stress and TL across the lifespan. Several excellent qualitative reviews describe and critically review this

literature (Entringer et al., 2018; Epel and Prather, 2018; Price et al., 2013; Shalev et al., 2013; Shields and Slavich, 2017) and meta-analyses now quantify the magnitude of these associations and identify potential moderators of effects (e.g., Hanssen et al., 2017; Mathur et al., 2016; Ridout et al., 2018, 2016; Schutte and Malouff, 2015, 2014). More recently, an appreciation of the role of mitochondria in the acute stress response and chronic allostatic load (Picard et al., 2014; Picard and McEwen, 2018a, 2018b) has led to investigations of the association of psychological states, stress exposure in relation to mitochondrial functions and mtDNA (Boeck et al., 2016; Cai et al., 2015; Picard et al., 2018; Tyrka et al., 2016). Rather than being a comprehensive review, this section provides a brief overview of this field, emphasizing recent developments.

3.1. What is (psychological) stress?—Broadly defined, “stress” is the condition of being subjected to a stimulus (i.e., stressor) that invokes a response requiring the use of resources to adapt or cope (Monroe, 2008; Shields and Slavich, 2017). “Stress” may refer to particular life events (e.g., job loss, death of a loved one, assault), contexts that are experienced as stressful or contain numerous stressors (e.g., poverty, neighborhood violence, famine) or the psychological or biological response to such an event or exposure (i.e., stress response, perceived stress) (Epel et al., 2018). Characteristics of the stressor(s) and the stress response may be important determinants of the biological response or adaptation and account for heterogeneity in the literature on stress and aging.

While some stressors occur in isolation, it is important to recognize that stress-inducing contexts, exposures, and perceptions of stress often covary, for example when families living in poverty experience neighborhood violence and feel unsafe. In addition, the level of perceived stress may vary substantially within a group exposed to the same stressor. Determinants of psychological and biological stress responses include the nature of the stressor in terms of type, scope, severity, chronicity, and how predictable and controllable the stressor is. Individual and social characteristics influencing the level of perceived stress include social, financial, cognitive, emotional, and behavioral resources for coping with, controlling, avoiding, and compensating for stressors (Epel et al., 2018).

3.2 Stress and biological aging: Evidence for a stress-aging axis involving telomeres and mitochondria—The literature on the association of stressors and perceived stress on TL is now sufficiently large that a number of meta-analyses have been conducted on the topic. Meta-analyses of the association between TL and childhood psychosocial stressors document significant effects that vary from small to medium in size (Hanssen et al., 2017; Li et al., 2017; Ridout et al., 2018). Moderator analyses suggest larger effects for studies that examine more severe exposures (Hanssen et al., 2017) and those that include wide range of adversity types (Ridout et al., 2018). In addition to cross-sectional investigations, a longitudinal study (Shalev et al., 2012) found that exposure to violence over a 5-year period in childhood predicted greater TL attrition, suggesting the possibility of a causal relationship. The biological mechanisms whereby adverse or positive experiences exert their lasting health effects remain mostly unknown. Effects on the germline and stem cells reserves, metabolic reprogramming, and rewiring of neural networks and brain circuitry are among many areas that deserve further research.

Turning to stressors that occur in adulthood, a significant association between perceived stress and shorter TL has been documented in meta-analyses, though this effect ranged from very small (Mathur et al., 2016) to modest (Schutte and Malouff, 2016) in size. Several studies have also shown associations of shorter TL with measures of severe or cumulative stress exposure in adulthood (for reviews see Shalev et al, 2013; Epel & Prather, 2018; Oliveira et al., 2016). Although some studies suggest that childhood adversity may account for associations between TL and adult stressors (Puterman et al., 2016; Revesz et al., 2016), there is also evidence that stressors experienced in adulthood prospectively predict telomere attrition (Puterman et al., 2015; Van Ockenburg et al., 2015).

Given the growing literature demonstrating the central role that mitochondria play in the stress response and the aging process, recent studies have examined the association of early life stress with measures of mitochondrial function or mtDNAcn. Although mtDNAcn is not a measure of mitochondrial function and is impossible to interpret on its own, it is easily measured from stored DNA and has been measured in different studies. For example, childhood trauma or adversity, as well as adult psychopathology have been linked to higher mtDNAcn (Cai et al., 2015; Tyrka et al., 2016). In a small study of postpartum women, early life adversity was associated with greater cellular respiration reflecting increased cellular energy demand, which in turn was positively correlated with levels of pro-inflammatory cytokines and childhood maltreatment (Boeck et al., 2016). In a study of caregiving stress, caregivers were found to have reductions in a functional index of mitochondrial health (MHI) in blood leukocytes. Mitochondrial health was operationalized as a multivariate index designed to reflect functional capacity on a “per mitochondrion” basis. In this first study of MHI in mixed human leukocytes, the index included biochemical enzymatic activities for three mitochondrial enzymes and mtDNAcn. Using this composite index as an outcome, this study found that positive mood was associated with higher MHI and was a mediator of the association between caregiving and MHI (Picard et al., 2018). Another study found that suicide attempters have significantly higher plasma levels of ccf-mtDNA (Lindqvist et al., 2016) and another study found elevated ccf-mtDNA levels in individuals with major depressive disorder (MDD) (Lindqvist et al., 2018).

A limitation of this body of research is that certain behavioral, psychiatric and medical conditions frequently co-occur with stress exposures and covary with TL and other biological processes central to aging, and thus may have confounding effects. These include smoking, obesity, dietary influences, anxiety, depressed mood, post-traumatic stress disorder (PTSD), medications, and cardiometabolic conditions, among others. These influences are not uniformly assessed, excluded, or statistically controlled. A meta-analysis on the association of early adversity and TL identified that the magnitude of the effect was smaller in studies that included participants with medical or psychiatric conditions, and participants on medications (Ridout et al., 2018). This finding suggests that the relationship between these conditions and shortened telomeres might obscure the effect of stress exposure, or, alternatively, that the psychiatric and medical conditions may be primarily responsible for some of the telomere effect (Epel & Prather, 2018). Thus, the complex inter-relationships among these exposures, behavioral factors, and health conditions should be carefully considered when designing studies and analyzing and interpreting results.

3.3. Psychopathology and biological aging—Psychiatric disorders are associated with increased risk of aging-related medical conditions, including cardiovascular disease, stroke, dementia, diabetes, and obesity (Penninx et al., 2013; Viron and Stern, 2010), and early mortality (Walker et al., 2015). While part of the association may be explained by differences in health behaviors, because individuals with psychiatric disorders are more likely to smoke, drink alcohol, eat poorly, and exercise less than others (van Gool et al., 2007), associations between psychiatric disorder status and medical morbidity remain significant after adjusting for these factors. This has led to the hypothesis that psychiatric conditions may induce or result from accelerated or premature biological aging. As reviewed in this review, there are multiple biological age indicators, including a range of cellular and molecular measures such as TL, mitochondrial dysfunction, oxidative stress, gene expression, and others (López-Otín et al., 2013). Notably, inflammation is also a widely-used indicator of biological aging (Baylis et al., 2013), coined as “inflammaging” by Franceschi et al. (2000). However, because of its elaborate discussion elsewhere (Franceschi et al., 2018b; Fulop et al., 2018), as well as in respect to mental health (Diniz and Vieira, 2018), we do not discuss it here. Overall, the following section will review the most frequently studied biological age indicators in epidemiological psychiatric research, including TL, epigenetic age, brain age, and to a lesser extent pro-inflammatory cytokines.

3.3.1. Associations of psychiatric disorders and biological age indicators: Simon and colleagues (Simon et al., 2006) were the first to report a relationship between psychiatric disorders and shorter telomeres in a sample that included MDD, bipolar disorder (BD) and anxiety disorder patients. Since then, a large number of studies have been conducted in an assortment of psychiatric disorders. MDD is among the most frequently studied disorders in this context, possibly as a consequence of its relatively well-documented associations with dysregulated physical health (Penninx et al., 2013). Several meta-analyses, the largest one containing >34,000 subjects from 38 studies, summarized the results and provided consistent evidence of an inverse association between TL and depression, generally with small to medium effect sizes (Darrow et al., 2016; Ridout et al., 2016; Schutte and Malouff, 2015). Similar meta-analytic results were found for anxiety disorders (N>19,000) (Malouff and Schutte, 2017), and PTSD (N>3,800) (Li et al., 2017). BD, schizophrenia and other psychotic disorders have been less extensively examined. A meta-analysis including 1,100 subjects from 7 studies found no difference in TL between BD cases and controls (Colpo et al., 2015). Two meta-analyses on schizophrenia of 1,200 and 1,600 subjects, respectively, found small effects for TL differences (Polho et al., 2015; Rao et al., 2016).

The epigenetic age indicator is most frequently examined in individuals with PTSD. A metaanalysis using data from 9 cohorts (combined N=2,186) found significant, albeit small, associations of greater epigenetic age with traumatic stress, but not with PTSD diagnosis (Wolf et al., 2018). Other studies also found such relations using the Horvath predictor (Boks et al., 2015; Mehta et al., 2018; Zannas et al., 2015), consistent with enrichment for glucocorticoid response elements (Zannas et al., 2015). Two recent studies for the first time showed “older” epigenetic age in MDD patients versus controls (Han et al., 2018; Whalley et al., 2017), while this is not seen in schizophrenia (McKinney et al., 2017b; Voisey et al., 2017).

Furthermore, chronic, low-grade inflammation that increases on average with age is captured in the term “inflammaging” - a pro-inflammatory state proposed to contribute to the pathogenesis of age-related diseases (Franceschi et al., 2018a). An increase in the inflammatory response, together with microglial activation, in turn, can contribute to psychiatric diseases, such as MDD, schizophrenia, BD, and autism (Khandaker et al., 2015; Martínez-Cengotitabengoa et al., 2016; Réus et al., 2015). The mechanisms responsible for increased inflammation in mood disorders remains poorly understood. Recent evidence suggests that ccf-mtDNA could contribute to this pro-inflammatory state, although evidence is mixed (Kageyama et al., 2018; Lindqvist et al., 2018, 2016). Related to the bacterial origin of mitochondria, the mtDNA is immunogenic. Released mtDNA molecules thereby act as damage associated molecular patterns (DAMPs) recognized by toll-like receptors on immune cells and trigger immune cell activation (West and Shadel, 2017). Inflammaging could in part be due to increased ccf-mtDNA in older individuals (Pinti et al., 2014).

Increased plasma levels of ccf-mtDNA have been reported in suicidal and depressed patients (Lindqvist et al., 2016). Worse response to an antidepressant was associated with increasing ccf-mtDNA levels over the treatment course, and ccf-mtDNA was correlated with antioxidant enzyme glutathione peroxidase, possibly as a result of a compensatory response to cellular oxidative stress (Lindqvist et al., 2018). An experimental study using psychological stress induction in healthy middle-aged individuals also demonstrated that an acute bout of psychological stress may be sufficient to elicit a 2-3 fold increase in serum ccf-mtDNA within 30 minutes, suggesting that ccf-mtDNA is dynamically regulated (Trumpff et al., 2019). Consistent with previous findings linking ccf-mtDNA levels to cortisol levels following a dexamethasone suppression test (Lindqvist et al., 2016), glucocorticoid stimulation of human cells (fibroblasts) induced the release of mtDNA by mitochondria within minutes (Trumpff et al., 2019). Thus, the causes of elevated ccf-mtDNA in certain psychiatric conditions remain unknown, although clinical and cellular studies suggest that canonical neuroendocrine stress mediators, including but not limited to glucocorticoids, may be implicated.

In addition, associations have been reported between “older” brain age and psychiatric disorders such as borderline personality disorder (Nenadic et al., 2017), schizophrenia (Koutsouleris et al., 2014; Nenadic et al., 2017; Schnack et al., 2016), and first-episode psychosis (Kolenic et al., 2018), as compared to younger in BD (Nenadic et al., 2017). One relatively small study by Koutsouleris et al. (2014) showed a higher brain-PAD of +4.0 years in MDD (N=104). However, a preliminary study by Kaufmann et al. (2018) finds increased brain age in schizophrenia (Cohen’s $d=0.55$) and BD ($d=0.30$), but not MDD (N=211, $d=0.10$). In addition, these authors suggest that the brain age gap is a genetically modulated trait that is heritable and overlaps with polygenic architecture observed in common brain disorders. There is also preliminary evidence of an association between psychiatric pathology and glycomic-based biological age indicators, which represent sugar-based modifications of proteins, RNA, and DNA molecules. Two studies have shown altered protein N-glycosylation profiles in female patients with MDD (Boeck et al., 2018c) and in PTSD (Moreno-Villanueva et al., 2013), indicative of advanced aging at the glycomic level. Associations with other omics-based indicators (transcriptomics, proteomics, metabolomics) and their interrelation remain to be explored.

3.3.2. Biological aging and psychopathology: The chicken or the egg?: While robust cross-sectional associations between psychiatric disorders and biological aging have been documented - at least for TL - the nature and direction of these associations remain unclear. Longitudinal studies have found mixed effects (see, e.g. Boks et al., 2015; Maniates et al., 2018; Shalev et al., 2014; Vance et al., 2018; Verhoeven et al., 2016). It is currently unknown whether 1) psychopathology-associated physiological disturbances accelerate biological aging, 2) premature biological aging antedates and is a vulnerability factor that causes psychopathology, or alternatively, 3) psychopathology and biological aging processes share underlying etiological roots, such as shared genetic risks, and happen to be correlated without a causal link between them. Recent studies using genomic and causal inference tools are being developed to overcome this limitation of observational studies (e.g., Wium-Andersen et al., 2017). This challenge, among others, as well as recommendations to move the field towards a predictive science are discussed in details in Section 5.

4. Clinical implications

4.1. Disorder-specific or transdiagnostic phenomenon?—In a large meta-analysis considering multiple psychiatric disorders and TL, including depressive and anxiety disorders, PTSD, bipolar and psychotic disorder, no difference in effect sizes between disorders was found (Darrow et al., 2016). This indicates that different DSM-based diagnoses may not be associated with meaningful differences in biological aging (Lindqvist et al., 2015). Furthermore, several studies showed that short TL is associated with the same physiological dysregulations that are found in some but not all persons with psychiatric disorders. These include increased inflammation, oxidative stress markers, dysregulated HPA-axis, and metabolic dysregulations (Lee et al., 2011; Monickaraj et al., 2012; Wikgren et al., 2012; Zhang et al., 2016). While the degree to which telomeres are causally related to these mechanisms is unknown, this is suggestive of pathways through which telomere shortening and psychiatric disorders are interrelated, that are not limited to one diagnostic category. The current evidence suggests that short TL may be a non-specific biological marker for conditions in which people experience chronic psychological or physiological stress, rather than being a marker of a specific psychiatric condition. Similarly, the downstream biological ramifications of different disorders also overlap, with alterations at the organs and systems level (e.g. brain aging patterns, Cole, 2018). This evidence leaves us to consider that these indicators are not disease-specific nor suitable as diagnostic tools, but rather general indicators of psychopathology or abnormal mental states.

4.2. Biological age indicators as predictors of treatment outcome—Only a small number of studies have investigated whether biological aging indicators predict antidepressant treatment response. The first study suggesting such a link showed in a small sample of previously unmedicated MDD subjects that low baseline telomerase activity, and a greater increase in telomerase activity during eight weeks of selective serotonin reuptake inhibitors (SSRI) treatment were associated to superior clinical outcome (Wolkowitz et al., 2012). However, this study lacked a control condition, leaving open the possibility that naturally different clinical trajectories contributed to these effects. Nevertheless, these findings suggested that depressed patients with relatively low baseline telomerase activity may most benefit from therapies that may secondarily induce telomerase activation, and that

telomerase activation may represent a mechanism of antidepressant action, consistent with several animal studies (reviewed in Bersani et al., 2015).

Subsequently, one human study found that shorter leukocyte TL predicted worse antidepressant response to an SSRI (Hough et al., 2016). To the extent that accelerated biological aging is associated with antidepressant response, this effect does not seem likely to be confined to a specific treatment modality or drug. Shorter TL may also predict worse antidepressant response to pioglitazone, which also has antidiabetic effects (Rasgon et al., 2016). Turning to other disorders, in a group of bipolar and schizophrenia/psychosis patients, lithium non-responders had shorter telomeres than responders (Martinsson et al., 2013), and similar associations were found linking short TL to poor treatment response (Li et al., 2015; Yu et al., 2008). No studies have investigated the association between TL and response to psychotherapy (e.g., cognitive behavioral therapy), although preliminary evidence suggests a positive correlation between mindfulness/meditation practices and telomere biology, including increased telomerase activity (Conklin et al., 2018; Schutte and Malouff, 2014). Furthermore, an increased inflammatory response may hamper the responsiveness to mood disorder treatments, and higher baseline inflammation may lead to treatment resistance (Réus et al., 2015; Strawbridge et al., 2015). Changes in ccf-mtDNA levels were also found to be associated with SSRI treatment response, with the non-responders showing an increase in ccf-mtDNA and responders not changing (Lindqvist et al., 2018). A shared genetic disposition for inflammation, psychopathology, and treatment responsiveness has also been suggested (Zwicker et al., 2018). Furthermore, inflammatory indicators may potentially offer personalized antidepressant recommendations, and could eventually guide the development of novel antidepressant treatments (Jha and Trivedi, 2018). Overall, the link between biological age and treatment responsiveness is a young field requiring further research.

4.3 Can biological aging be reversed with treatment?—One of the most relevant clinical questions in the field of biological aging and psychiatry is whether accelerated biological aging is a permanent imprint or a reversible process. While this may differ between indicators of biological aging, reversibility is at least possible to some extent for some indicators. For TL, the main mechanism of restoration is likely telomerase activation. Animal and in-vitro research provided evidence that telomerase-associated recovery of TL is to some extent possible (Batista et al., 2011; Jaskelioff et al., 2011). Several intervention studies have attempted to influence TL in humans (see Verhoeven et al., 2014 for an overview). For example, recent small controlled studies have shown elongation of telomeres in response to highly controlled aerobic exercise verified with actigraphy (Puterman et al., 2018), losing and maintaining a weight loss of 10% or greater (Mason et al., 2018), and meditation-based interventions (Conklin et al., 2018). As mentioned above, for the field of psychiatry telomerase activation may be a mechanism of antidepressant action. Although no strong conclusions can be drawn due to a lack of well-powered clinical studies, there are several potential mechanisms by which psychiatric medications might modulate telomerase activity or TERT expression, including via increased brain-derived neurotrophic factor (BDNF) expression (Bersani et al., 2015). Increased telomerase activity may, in turn, induce clinical effects by promoting cellular survival and/or functioning. It should however be noted

that increased telomerase activity can prevent cell senescence, an anti-cancer mechanism, and thus excessive telomerase activity is also associated with increased risk of cancer. Thus, while telomerase activation may hold the promise of reducing risk of aging-related disease, the risk of less common but very serious cancer outcomes must be carefully weighed (Blackburn et al., 2015).

Cross-sectional studies show that certain behaviors may provide some protection against brain aging. Higher physical activity levels are associated with lower brain age (Steffener et al., 2016), and “younger brains” are seen in those that learn to play an instrument (Rogenmoser et al., 2018) and those who have practiced meditation for long periods (Luders et al., 2016). It remains to be elucidated whether brain age is responsive to intervention, but a randomized controlled trial (RCT) showed that ibuprofen temporarily reduced brain-PAD by 1.1 years in healthy individuals, likely due to its acute anti-inflammatory effects (Le et al., 2018). Physical activity may slow age-related DNAm changes in humans (Ren et al., 2012; Voisin et al., 2015). Longitudinal data shows that increasing BMI is associated with increasing epigenetic age (Quach et al., 2017), but in another report, epigenetic age from the liver was not “decelerated” after successful weight loss over a 9-month period through bariatric surgery (Horvath et al., 2014). Quach et al. (2017) also found that consumption of fish, fruits, and vegetables, as well as effects of moderate alcohol, education, and income and exercise induced anti-aging effects based on epigenetic age (Hannum clock). However, these findings are cross-sectional observations rather than longitudinal effects from RCTs. Nonetheless, a recent RCT suggests that vitamin D supplementation may decrease epigenetic aging based on the Horvath, but not Hannum epigenetic clock (Chen et al., 2018).

More controlled intervention studies are needed to determine whether biological aging indicators are truly modifiable in response to exercise, nutritional and/or pharmacological interventions. A common problem to observational studies is that behaviors tend to correlate, making it difficult to evaluate the specific influence of a given intervention or behavior in isolation. Individuals who exercise more tend to practice meditation more frequently, eat more plant-based and vegetarian diets, consume less illicit substances, etc. An additional problem with observational studies is that it is impossible to establish the direction of effects; while exercise may attenuate indicators of aging, biologically younger individuals may be more able and inclined to exercise.

5. Key challenges and priorities for future research

There are key challenges to accurately measure and interpret biological age indicators and to further our understanding of stress and biological aging. A partial list of six major challenges related to priority areas for the field, as well as recommendations to overcome them is presented in this section. We also summarize essential steps and propose a minimum standard for the design, collection, processing, analysis, and reporting of data involving biological age indicators (Figure 4).

Challenge 1. Correlation is not causation.—It is hazardous to infer causality from cross-sectional correlational data (Simons, 2015). For example, in the case of TL, it is possible that telomere shortening reflects states of stress, or responds to somatic or

psychiatric illness, or at least to biochemical abnormalities associated with these states. It is also possible, however, that telomere shortening precedes somatic or psychiatric illness (Gotlib et al., 2014) or even underlies biological changes that causes these conditions. Biological age indicators could be entirely independent from directly assessing biological age mechanisms that drive the aging process. Changes in biological age indicators could be the “canary in the coal mine” (Effros, 2009), representing factors *associated* with aging rather than aging itself.

Recommendation 1: Collect data longitudinally, consider experimentation, and choose prediction over explanation.

This is a three-part recommendation. First, future studies should include longitudinal designs to increase the reliability and accuracy of measuring aging (Moffitt et al., 2017). Longitudinal measurements will also be important in determining which variables are critical for the maintenance of successful aging throughout the lifespan (e.g. absolute levels, change, variability), the “recipe” of which may vary between individuals (Sanders et al., 2012). Moreover, it will be critical to determine the optimal intervals of time between repeated assessments that are needed to detect meaningful changes in specific biological age indicators (see Figure 3). This information about the timing and spacing of repeated measures, and real-life constraints, should then be used to inform the design and choice of outcome measures when evaluating the effectiveness of interventions aiming to influence biological aging.

Second, experimental approaches utilizing cellular (or animal) models allow the direct manipulation of a specific (set of) variable(s). Thus, if we assume that a given stressor or predictor can be modeled accurately *in vitro*, experimental designs can provide direct causal evidence that a given factor is necessary and sufficient to produce a given outcome of interest. For this approach to empirically support a biological interaction between stressors and biological aging, the biological age indicator also needs to be detectable and meaningful *in vitro*, such as epigenetic age (e.g., Horvath et al., 2018). In cases where experimental demonstration is not possible, statistical methods such as causal inference (Bind et al., 2017) and genomic methods including Mendelian randomization (Burgess et al., 2012; Wium-Andersen et al., 2017) can substantially reinforce our confidence regarding the direction of effects.

Third, in some cases, the number of predictors one wishes to consider is very large, either because there is no prior knowledge of their relative importance, or because the problem is truly complex – such as human biological aging. In such cases, the number of predictors can be large relative to the number of individuals, providing insufficient power for traditional inference-based statistics. In such cases, machine learning-based predictive modeling may be advisable to discover and validate predictive relationships between variables. Whereas statistics draw population inferences from a sample, machine learning finds generalizable predictive patterns (Bzdok et al., 2018). Using predictive modeling approaches that identify and validate combinations of predictors in relation to a particular health outcome can increase the likelihood that the identified predictors of biological age are robust, specific, and generalizable. An excellent article on the value of prediction over explanation in the psychological sciences is Yarkoni and Westfall (2017). Regardless of the analytical approach taken, we should emphasize the value of converging evidence collected using different

methods, measuring multiple (related and unrelated) predictors in parallel, and assessing multiple outcomes (Munafò and Davey Smith, 2018)

Challenge 2. Single biological age indicators are not correlated and may be better integrated.—Not all measurements of biological aging are equally useful or inter-related, and it remains to be elucidated if and how different indicators relate to one another and which biological determinants are consistent across measures. As previously noted (Cole et al., 2018), no single biological age indicator can currently fully capture the complexity of the aging process, nor predict future health outcomes or lifespan with sufficient accuracy. There is therefore a need for combined indices that logically integrate multiple indicators (Figure 5), hopefully resulting in accurate integrative panels that outperform single measurements of biological aging, also previously suggested by Xia et al. (2017). Nevertheless, integration of indicators cannot compensate for inadequately powered studies, which require large sample sizes to ensure high generalizability.

Recommendation 2: Combine machine learning and other artificial intelligence techniques to create composite indices and panels of biological age indicators relevant to mental health.: To date, several efforts to develop combined indices have been presented. For example, Marioni et al. (2016) showed an additive effect of combining TL and epigenetic age in explaining the proportion of age variance of their model. Similarly, Cole et al. (2017) explained significantly more variance in the prediction of mortality by combining brain-PAD and the Horvath epigenetic predicted age difference, than either indicator alone. Other examples include multivariate indicators of aging incorporating multiple physiological and functional measures (Belsky et al., 2015) and indices integrating multiple enzymatic and molecular measures of mitochondrial content and function in blood leukocytes as the MHI in association to psychological states (i.e., positive mood) (Picard et al., 2018). These constitute early attempts to reverse our reductionist inclinations and to move towards integrative metrics that will hopefully lead to improved prediction.

Challenge 3. Biological aging may be tissue- and cell-type specific.—Related to the above, biological age indicators derived from specific cell or tissue types may not generalize to other cells or tissues. Biomarkers assessed in blood, for example, represents the average of multiple heterogeneous cell types. But cellular or organismal health may be more closely related to or reflected by indicators within individual cell types or those with the most extreme values (e.g., Flow-FISH, Aubert et al., 2012). Furthermore, TL differs across leukocyte cell types, such as naïve vs mature T cells (Chou and Effros, 2013; Lin et al., 2016), and differs in different regions of the brain (Mamdani et al., 2015). Moreover, TL of particular cell types can be differentially vulnerable to attrition or affected by stress-related pathology (Boeck et al., 2018b).

Recommendation 3: Purify cell types using established molecular markers.: Purification of cell types can be accomplished by a variety of methods (flow cytometry, magnetic-activated cell sorting), yielding living cells amenable to downstream molecular and cellular analyses (Lin et al., 2010). Under certain conditions where it is not possible to isolate specific cell subtypes, it may be difficult to interpret certain indicators that exhibit large cell

type-specific values (such as mtDNA_{cn} in blood). In some limited cases where a lot of information is available for adjustment, such as for DNAm measured on bead chips (100,000's of data points), it is possible to use statistical approaches, i.e. reference-based (Hattab et al., 2017) or reference-free (Houseman et al., 2014), to infer underlying cell type proportions (Titus et al., 2017) and adjust results accordingly. Another interesting application is demonstrated in a preliminary study by Chan et al. (2018) that uses deconvolution approaches to show novel MDD-methylation associations in individual sub-populations of neurons/glia from bulk brain, as well as in granulocytes/T-cells/B-cells/monocytes from bulk blood data. In addition, creative ways to harvest other cell types from tissues other than blood could yield increasingly meaningful biological age indicators. For example, the DNAm-based skin & blood clock by (Horvath et al., 2018) is robust across tissues (e.g. fibroblasts, buccal, endothelial, saliva samples) and can, therefore, be applied to many organs, as well as ex vivo. Thus, this approach provides extended information on synchronized biological aging, independent of sampling source.

Challenge 4. Within-group variance may be larger than between-group variance.—Although group differences in biological age indicators have been reported in several psychiatric illnesses (Darrow et al., 2016; Lindqvist et al., 2015), these reflect average group differences. However, there is often considerable within-groups variability and considerable overlap between groups, making it very difficult to use most biological age indicators as diagnostic aids. In addition, even specific psychopathological diagnoses (e.g., MDD or schizophrenia) often include individuals that vary widely in their symptoms and presentation, making the search for predictors of mental illnesses defined diagnostically somewhat elusive (Cuthbert and Insel, 2013). As Wolfers et al. (2018) suggest, the complex, highly polygenic and multifaceted causes of severe mental disorders may only be fully understood by mapping patients' individual signatures, rather than studying the average patient. Population-based normal ranges have yet to be reliably determined, so biological age indicators may be more useful in detecting within-subject changes over time rather than in comparing individuals or in establishing actuarial norms. Nonetheless, a recent study utilizing flow-FISH assay techniques reported reproducible and definable upper and lower normal boundaries for TL in a hospital population (Alder et al., 2018), indicating that standardization of these measures may be achievable. Within-group variance is often due to measurable individual differences in behavior, health condition, and lifestyle.

Recommendation 4: Visualize your data, carefully assess known influences, move beyond group-based analyses, and use within-person modeling approaches.: To visualize data where there are multiple measures over time per individual, we advocate for spaghetti plots (example with epigenetic age and TL can be found in Marioni et al. (2016), and with cortisol trajectories in Ram and Grimm (2009)). Standard measures of variance can also be used to model population variability at different timepoints and to compare sub-groups, and more sophisticated mathematical approaches can be useful to generate individualized phenotypes (Hertel et al., 2018). Data reduction approaches such as principal component analysis (PCA), partial least square discriminant analysis (PLS-DA), and t-distributed stochastic neighbor embedding (t-SNE) are also useful to visualize high-dimensional data in two or three dimensions and to assess whether subsets of individuals in

the sample naturally cluster together (Maaten and Hinton, 2008; Xia et al., 2015). This kind of approach can provide evidence of shared phenotypes or trajectories that would otherwise remain undetectable by standard uni- or multivariate analyses. Other statistical approaches to identify different subgroups exhibiting different trajectories in biological aging or in clinical course include growth mixture modeling (Ram and Grimm, 2009), and random coefficients linear regression models to examine within-person changes over time (Diaz et al., 2018).

Challenge 5. Large sample sizes are needed to detect small effect sizes.—

Related to the point above, group differences between individuals with certain psychiatric illnesses vs. healthy individuals, even if statistically significant, often have small effect sizes that require large sample sizes to be demonstrated (Darrow et al., 2016; Han et al., 2018; Verhoeven et al., 2014; Whalley et al., 2017). Similarly, even using predictive modeling and machine learning approaches, small samples sizes are more susceptible to overfitting (Yarkoni and Westfall, 2017). It would be informative to pool databases across studies and to check for consistency and predictive accuracy across studies, albeit at the cost of increasing heterogeneity of the samples studied and of the laboratory methods used. (Schnack and Kahn, 2016) argue that larger sample sizes will have more generalization power. This is important for creating robust “canonical” publicly available prediction models (e.g. Horvath’s epigenetic clock) that can be readily applied to smaller studies that cannot permit partitioning their data into a training and validation set.

Recommendation 5: Collaborate and harmonize data collection and analysis protocols to facilitate data pooling worldwide.:

Certain types of data may be available through crowdsourcing, a data collection process with remarkable scalability that may enable identification of robust small effect size effects (Mohammadi, 2015). Large-scale collaborative initiatives like the Enhancing Neuroimaging Genetics through Meta-Analysis consortium (ENIGMA) for imaging and genetics data may be useful examples (Thompson et al., 2014). When not possible to replicate findings on sufficiently large independent datasets, use cross-validation methods to avoid overfitting (Yarkoni and Westfall, 2017).

Challenge 6. Non-uniform laboratory assays and storage conditions.—

Different assay methodologies can yield relatively different results (e.g. qPCR vs. Southern blot for TL assessment, Aviv et al., 2011), and most assays such as Flow-FISH assays and qPCR ascertain relative TL rather than absolute length. These issues have been discussed in detail elsewhere (Aubert et al., 2012), and the relative merits of the different assays have been compared. A more mundane but important and under-appreciated caveat is that methodological differences can yield spurious results (e.g., length of time a specimen was kept in a freezer, freezer temperature, whether whole blood or intact cells vs. cellular lysate were frozen, specific batch of reagents used, assay technique, method of DNA extraction [for TL] (Dagnall et al., 2017; Khincha et al., 2017; Shiels, 2010). An important issue that has not yet been experimentally assessed in multi-year human studies is the relative merit of assaying batches of samples at short intervals (e.g., every year) to minimize freezer time, compared to keeping all samples frozen until the end of the entire multi-year study so that all samples can be assayed simultaneously using identical procedures and reagents.

Recommendation 6: Harmonize measurements and storage conditions - colder is

better.: Follow guidelines, where available, to ensure that samples are collected and measured with the highest standards. Efforts are currently underway to systematically compare methods available to measure TL and will hopefully produce specific guidelines that can be implemented at a large scale. Figure 4 summarizes some measures that can be considered in the design, collection, pre-processing, analysis, and reporting of data to give researchers the ability to critically evaluate published results and hopefully harmonize methods and datasets.

6. Summary

Much progress has been made in the last decade towards developing objective biological age indicators but several key challenges and opportunities remain. Multiple new indicators spanning physiological and functional capacity, brain function and structure, and cellular and molecular levels of analysis have recently emerged, particularly under the force of ‘omics’ technologies. Most indicators show good to excellent correlations with chronological age, and some have the ability to predict age-related outcomes such as mortality with moderate accuracy (see Table 1). However, some have not yet been prospectively studied in large cohorts so their predictive power in relation to health outcomes remains unknown. In fact, many proposed biological age indicators still require replication and validation in larger independent datasets. As a whole, the development of multi-systemic biological age indicators has demonstrated that aging occurs not at a single level in a specific cell type, but rather manifests somewhat differently in various organs and tissues, cell types, and across a number of levels (see Figure 1). Rapidly developing methods show that biological age indicators gain in precision and prediction accuracy by leveraging biologically-informed approaches to integrate individual indicators into more powerful indices and panels of measures.

In relation to psychological stress and mental health, much work also remains to establish to what extent and how psychological states influence the aging process. Equally important to mapping these psycho-biological processes is to understand the “reverse” causal link whereby accelerated biological aging may impact physiological vulnerability and resilience to life stressors and psychopathology. A shared objective for our field is to disentangle these associations and identify the causal pathways that drive aging and mental health trajectories. To do so, we need to address number of key challenges that, if adequately met, will yield exciting opportunities to advance our understanding of human health.

To achieve this objective, it will also be essential for clinicians, psychologists, epidemiologists, and behavioral scientists to enter in a dialogue with experimental biologists. Such dialogue immediately opens new questions that would not otherwise arise within the silos of individual disciplines, departments, and laboratories. For example, the joining of ideas around psychosocial stress and cellular aging (Epel et al. 2004), age-related elevation in circulating DNA (Teo et al. 2018), and of psychoneuroimmunology and the pro-inflammatory effects of psychological stress (Marsland et al. 2017) has led to the idea that acute psychological stress may rapidly trigger immunogenic mtDNA release into the circulation (Trumpff et al. 2019). If met with sufficient enthusiasm and resources, these

jointly-created interdisciplinary questions can subsequently push the development of new laboratory methods, statistical and analytical tools, and new theoretical models. The interdisciplinary field of human psychobiology is replete with opportunities for innovation and discoveries.

Overall, we believe that developing, validating, and studying predictive biological age indicators – and defining how psychological states or psychiatric illnesses influence them – will fill important knowledge gaps linking stress and mental illness to aging. Achieving this goal will have at least three main positive consequences for the biomedical sciences generally. *First*, it will enable the stratification of individuals (and group of individuals) based on their current health state and future disease risk, including symptoms, disorders. This notion converges with the precision medicine agenda that aims to identify individualized predictors of future mental health and disease states (Insel and Cuthbert, 2015; Abi-Dargham and Horga, 2016). Achieving this goal could benefit clinical practice by providing clear and objective guidelines to direct health-promoting interventions and treatment delivery in a personalized way, specifically one that is most aligned with the individual's needs (Picard et al., 2013). *Second*, achieving this goal will provide objective and sensitive methods to evaluate the effectiveness of health-promoting interventions. The efficacy of interventions aimed at decreasing or even reversing advanced or premature aging, as well as interventions aimed at enhancing well-being and other health outcomes would be more effectively assessed using precise and sensitive biological age indicators. *Finally*, mapping stress-sensitive biological age indicators will also generate knowledge about human aging that can be taken back to the laboratory bench to orient basic biological science. Specifically, understanding the basis of normal and abnormal aging processes may identify new physiological targets for therapeutic intervention. Thus, an unintended consequence of improving existing biological age indicators and developing new end points for clinical and epidemiological human research may, in the end, create leaps in understanding about the fundamental biological mechanisms that explain why we age in the first place.

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LIST OF ABBREVIATIONS

BD	Bipolar Disorder
BDNF	Brain-derived Neurotrophic Factor
Brain-PAD	Brain-predicted age difference
ccf-mtDNA	Circulating cell-free mitochondrial DNA
DNAm	DNA methylation

MDD	Major Depressive Disorder
MHI	Mitochondrial Health Index
mtDNA	Mitochondrial DNA
mtDNAcn	Mitochondrial DNA copy number
nDNA	Nuclear DNA
PTSD	Post-traumatic Stress Disorder
SCZ	Schizophrenia
SSRI	Selective Serotonin Reuptake Inhibitor
TL	Telomere length

REFERENCES

- Abi-Dargham A, Horga G, 2016 The search for imaging biomarkers in psychiatric disorders. *Nat. Med.* 22,1248–1255. [PubMed: 27783066]
- Akulenko R, Merl M, Helms V, 2016 BEclear: Batch Effect Detection and Adjustment in DNA Methylation Data. *PLoS One* 11, e0159921. [PubMed: 27559732]
- Alder JK, Hanumanthu VS, Strong MA, DeZern AE, Stanley SE, Takemoto CM, Danilova L, Applegate CD, Bolton SG, Mohr DW, Brodsky RA, Casella JF, Greider CW, Jackson JB, Armanios M, 2018 Diagnostic utility of telomere length testing in a hospital-based setting. *Proc. Natl. Acad. Sci. U. S. A.* 115, E2358–E2365. [PubMed: 29463756]
- Aubert G, Hills M, Lansdorp PM, 2012 Telomere length measurement-caveats and a critical assessment of the available technologies and tools. *Mutat. Res.* 730, 59–67. [PubMed: 21663926]
- Aviv A, Hunt SC, Lin J, Cao X, Kimura M, Blackburn E, 2011 Impartial comparative analysis of measurement of leukocyte telomere length/DNA content by Southern blots and qPCR. *Nucleic Acids Res.* 39, e134. [PubMed: 21824912]
- Aycheh HM, Seong J-K, Shin J-H, Na DL, Kang B, Seo SW, Sohn K-A, 2018 Biological Brain Age Prediction Using Cortical Thickness Data: A Large Scale Cohort Study. *Front. Aging Neurosci.* 10, 1385.
- Ball G, Adamson C, Beare R, Seal ML, 2017 Modelling neuroanatomical variation during childhood and adolescence with neighbourhood-preserving embedding. *Sci. Rep.* 7, 17796. [PubMed: 29259302]
- Batista LFZ, Pech MF, Zhong FL, Nguyen HN, Xie KT, Zaug AJ, Crary SM, Choi J, Sebastiano V, Cherry A, Giri N, Wernig M, Alter BP, Cech TR, Savage SA, Reijo Pera RA, Artandi SE, 2011 Telomere shortening and loss of self-renewal in dyskeratosis congenita induced pluripotent stem cells. *Nature* 474, 399–402. [PubMed: 21602826]
- Baylis D, Bartlett DB, Patel HP, Roberts HC, 2013 Understanding how we age: insights into inflammaging. *Longev Healthspan* 2, 8. [PubMed: 24472098]
- Belsky DW, Caspi A, Houts R, Cohen HJ, Corcoran DL, Danese A, Harrington H, Israel S, Levine ME, Schaefer JD, Sugden K, Williams B, Yashin AI, Poulton R, Moffitt TE, 2015 Quantification of biological aging in young adults. *Proc. Natl. Acad. Sci. U. S. A.* 112, E4104–10. [PubMed: 26150497]
- Belsky DW, Moffitt TE, Cohen AA, Corcoran DL, Levine ME, Prinz JA, Schaefer J, Sugden K, Williams B, Poulton R, Caspi A, 2018 Eleven Telomere, Epigenetic Clock, and Biomarker-Composite Quantifications of Biological Aging: Do They Measure the Same Thing? *Am. J. Epidemiol.* 187, 1220–1230. [PubMed: 29149257]
- Bender A, Krishnan KJ, Morris CM, Taylor GA, Reeve AK, Perry RH, Jaros E, Hershenson JS, Betts J, Klopstock T, Taylor RW, Turnbull DM, 2006 High levels of mitochondrial DNA deletions in

substantia nigra neurons in aging and Parkinson disease. *Nat. Genet.* 38, 515–517. [PubMed: 16604074]

- Bersani FS, Lindqvist D, Mellon SH, Penninx BWJH, Verhoeven JE, Revesz D, Reus VI, Wolkowitz OM, 2015 Telomerase activation as a possible mechanism of action of psychopharmacological interventions. *Drug Discov. Today.* 20, 1305–1309. [PubMed: 26166813]
- Bind M-A, VanderWeele TJ, Schwartz JD, Coull BA, 2017 Quantile causal mediation analysis allowing longitudinal data. *Stat. Med.* 36, 4182–4195. [PubMed: 28786129]
- Blackburn EH, Epel ES, Lin J, 2015 Human telomere biology: A contributory and interactive factor in aging, disease risks, and protection. *Science* 350, 1193–1198. [PubMed: 26785477]
- Boeck C, Gump AM, Calzia E, Radermacher P, Waller C, Karabatsiakis A, Kolassa I-T, 2018a The association between cortisol, oxytocin, and immune cell mitochondrial oxygen consumption in postpartum women with childhood maltreatment. *Psychoneuroendocrinology* 96, 69–77. [PubMed: 29908404]
- Boeck C, Koenig AM, Schury K, Geiger ML, Karabatsiakis A, Wilker S, Waller C, Gündel H, Fegert JM, Calzia E, Kolassa I-T, 2016 Inflammation in adult women with a history of child maltreatment: The involvement of mitochondrial alterations and oxidative stress. *Mitochondrion* 30, 197–207. [PubMed: 27530300]
- Boeck C, Krause S, Karabatsiakis A, Schury K, Gündel H, Waller C, Kolassa I-T, 2018b History of child maltreatment and telomere length in immune cell subsets: Associations with stress- and attachment-related hormones. *Dev. Psychopathol.* 30, 539–551. [PubMed: 28803568]
- Boeck C, Pfister S, Bürkle A, Vanhooren V, Libert C, Salinas-Manrique J, Dietrich DE, Kolassa I-T, Karabatsiakis A, 2018c Alterations of the serum N-glycan profile in female patients with Major Depressive Disorder. *J. Affect. Disord.* 234, 139–147. [PubMed: 29529546]
- Boeck C, Salinas-Manrique J, Calzia E, Radermacher P, von Arnim CAF, Dietrich DE, Kolassa I-T, Karabatsiakis A, 2018d Targeting the association between telomere length and immuno-cellular bioenergetics in female patients with Major Depressive Disorder. *Sci. Rep.* 8, 9419. [PubMed: 29925891]
- Bohannon RW, 2008 Hand-grip dynamometry predicts future outcomes in aging adults. *J. Geriatr.Phys. Ther.* 31,3–10. [PubMed: 18489802]
- Boks MP, van Mierlo HC, Rutten BPF, Radstake TRDJ, De Witte L, Geuze E, Horvath S, Schalkwyk LC, Vinkers CH, Broen JCA, Vermetten E, 2015 Longitudinal changes of telomere length and epigenetic age related to traumatic stress and post-traumatic stress disorder. *Psychoneuroendocrinology* 51,506–512. [PubMed: 25129579]
- Breitling LP, Saum K-U, Perna L, Schöttker B, Holleczer B, Brenner H, 2016 Frailty is associated with the epigenetic clock but not with telomere length in a German cohort. *Clin. Epigenetics* 8, 21. [PubMed: 26925173]
- Burgess S, Butterworth A, Malarstig A, Thompson SG, 2012 Use of Mendelian randomisation to assess potential benefit of clinical intervention. *BMJ* 345, e7325. [PubMed: 23131671]
- Bzdok D, Altman N, Krzywinski M, 2018 Points of significance: statistics versus machine learning. *Nat. Methods* 1–7.
- Cai N, Chang S, Li Y, Li Q, Hu J, Liang J, Song L, Kretschmar W, Gan X, Nicod J, Rivera M, Deng H, Du B, Li K, Sang W, Gao J, Gao S, Ha B, Ho H-Y, Hu C, Hu J, Hu Z, Huang G, Jiang G, Jiang T, Jin W, Li G, Li K, Li Y, Li Y, Li Y, Lin Y-T, Liu L, Liu T, Liu Y, Liu Y, Lu Y, Lv L, Meng H, Qian P, Sang H, Shen J, Shi J, Sun J, Tao M, Wang G, Wang G, Wang J, Wang L, Wang X, Wang X, Yang H, Yang L, Yin Y, Zhang J, Zhang K, Sun N, Zhang W, Zhang X, Zhang Z, Zhong H, Breen G, Wang J, Marchini J, Chen Y, Xu Q, Xu X, Mott R, Huang G-J, Kendler K, Flint J, 2015 Molecular signatures of major depression. *Curr. Biol.* 25, 1146–1156. [PubMed: 25913401]
- Cawthon RM, 2002 Telomere measurement by quantitative PCR 30, 1–6.
- Chacko BK, Kramer PA, Ravi S, Johnson MS, Hardy RW, Ballinger SW, Darley-USmar VM, 2013 Methods for defining distinct bioenergetic profiles in platelets, lymphocytes, monocytes, and neutrophils, and the oxidative burst from human blood. *Lab. Invest.* 93, 690–700. [PubMed: 23528848]

- Chaleckis R, Murakami I, Takada J, Kondoh H, Yanagida M, 2016 Individual variability in human blood metabolites identifies age-related differences. *Proc. Natl. Acad. Sci. U. S. A.* 113, 4252–4259. [PubMed: 27036001]
- Chan AY, Swaminathan R, Cockram CS, 1989 Effectiveness of sodium fluoride as a preservative of glucose in blood. *Clin. Chem.* 35, 315–317. [PubMed: 2914384]
- Chan RF, Turecki G, Shabalin AA, Guintivano J, Zhao M, Xie LY, van Grootheest G, Kaminsky ZA, Dean B, Penninx BWJH, Aberg KA, van den Oord EJCG, 2018 Cell-type-specific methylome-wide association studies implicate neurodegenerative processes and neuroimmune communication in major depressive disorder. *bioRxiv*.
- Chen BH, Carty CL, Kimura M, Kark JD, Chen W, Li S, Zhang T, Kooperberg C, Levy D, Assimes T, Absher D, Horvath S, Reiner AP, Aviv A, 2017 Leukocyte telomere length, T cell composition and DNA methylation age. *Aging* 9, 1983–1995. [PubMed: 28930701]
- Chen BH, Marioni RE, Colicino E, Peters MJ, Ward CK, Tsai PC, Roetker NS, Just AC, Demerath EW, Bressler J, Fornage M, Studenski S, Vandiver AR, Tanaka T, Kiel DP, Liang L, Vokonas P, Lunetta KL, Murabito JM, Bandinelli S, Dena G, Melzer D, Nalls M, Pilling LC, Price TR, Andrew B, Gieger C, Holle R, Kretschmer A, Kronenberg F, Visscher PM, Shah S, Wray NR, Mcrae AF, Levine ME, Lu AT, Tsao PS, Hou L, Manson JE, Levy D, Baccarelli A, Meurs JV, Bell JT, 2016 DNA methylation-based measures of biological age: meta-analysis predicting time to death. *Aging* 8, 1844–1859. [PubMed: 27690265]
- Chen L, Dong Y, Bhagatwala J, Raed A, Huang Y, Zhu H, 2018 Effects of Vitamin D3 supplementation on epigenetic aging in overweight and obese African Americans with suboptimal vitamin D status: a randomized clinical trial. *J. Gerontol. A Biol. Sci. Med. Sci.* <https://doi.org/10.1093/gerona/gly223>
- Chou JP, Effros RB, 2013 T cell replicative senescence in human aging. *Curr. Pharm. Des.* 19, 1680–1698. [PubMed: 23061726]
- Christiansen L, Lenart A, Tan Q, Vaupel JW, Aviv A, Mcgue M, Christensen K, 2016 DNA methylation age is associated with mortality in a longitudinal Danish twin study. *Aging Cell* 15, 149–154. [PubMed: 26594032]
- Chung S, Wang X, Lui YW, 2017 Influence of T1-Weighted Signal Intensity on FSL Voxel-Based Morphometry and FreeSurfer Cortical Thickness. *AJNR Am. J. Neuroradiol.* 38, 726–728. [PubMed: 28034997]
- Clarkson MJ, Cardoso MJ, Ridgway GR, Modat M, Leung KK, Rohrer JD, Fox NC, Ourselin S, 2011 A comparison of voxel and surface based cortical thickness estimation methods. *Neuroimage* 57, 856–865. [PubMed: 21640841]
- Cole JH, 2018 Neuroimaging Studies Illustrate the Commonalities Between Ageing and Brain Diseases. *Bioessays* 1700221, 1700221.
- Cole JH, Franke K, 2017 Predicting Age Using Neuroimaging: Innovative Brain Ageing Biomarkers. *Trends Neurosci.* 40, 681–690. [PubMed: 29074032]
- Cole JH, Marioni RE, Harris SE, Deary IJ, Cole JH, 2018 Brain age and other bodily “ages”: implications for neuropsychiatry. *Mol. Psychiatry*, 10.1038/s41380-018-0098-1
- Cole JH, Poudel RPK, Tsagkrasoulis D, Caan MWA, Steves C, Spector TD, Montana G, 2016 Predicting brain age with deep learning from raw imaging data results in a reliable and heritable biomarker.
- Cole JH, Ritchie SJ, Bastin ME, Valdés Hernández MC, Muñoz Maniega S, Royle N, Corley J, Pattie A, Harris SE, Zhang Q, Wray NR, Redmond P, Marioni RE, Starr JM, Cox SR, Wardlaw JM, Sharp DJ, Deary IJ, 2017 Brain age predicts mortality. *Mol. Psychiatry* 1–8. [PubMed: 27994236]
- Colpo GD, Leffa DD, Kohler CA, Kapczinski F, Quevedo J, Carvalho AF, 2015 Is bipolar disorder associated with accelerating aging? A meta-analysis of telomere length studies. *J. Affect. Disord.* 186, 241–248. [PubMed: 26253905]
- Conklin QA, Crosswell AD, Saron CD, Epel ES, 2018 Meditation, Stress Processes, and Telomere Biology. *Current Opinion in Psychology.* <https://doi.org/10.1016/j.copsyc.2018.11.009>
- Cooper R, Kuh D, Hardy R, Mortality Review Group, FALCon and HALCyon Study Teams, 2010 Objectively measured physical capability levels and mortality: systematic review and meta-analysis. *BMJ* 341, C4467. [PubMed: 20829298]

- Corral-Debrinski M, Horton T, Lott MT, Shoffner JM, Beal MF, Wallace DC, 1992 Mitochondrial DNA deletions in human brain: regional variability and increase with advanced age. *Nat. Genet.* 2, 324–329. [PubMed: 1303288]
- Cuthbert BN, Insel TR, 2013 Toward the future of psychiatric diagnosis: the seven pillars of RDoC. *BMC Med.* 11, 126. [PubMed: 23672542]
- Dagnall CL, Hicks B, Teshome K, Hutchinson AA, Gadalla SM, Khincha PP, Yeager M, Savage SA, 2017 Effect of pre-analytic variables on the reproducibility of qPCR relative telomere length measurement. *PLoS One* 12, e0184098. [PubMed: 28886139]
- Darrow SM, Verhoeven JE, Révész D, Lindqvist D, Penninx BWJH, Delucchi KL, Wolkowitz OM, Mathews CA, 2016 The Association Between Psychiatric Disorders and Telomere Length. *Psychosom. Med.* 78, 776–787. [PubMed: 27359174]
- de Magalhães JP, 2012 Programmatic features of aging originating in development: aging mechanisms beyond molecular damage? *FASEB J.* 26, 4821–4826. [PubMed: 22964300]
- de Magalhães JP, Passos JF, 2018 Stress, cell senescence and organismal ageing. *Mech. Ageing Dev.* 170, 2–9. [PubMed: 28688962]
- Diaz KM, Thanataveerat A, Parsons FE, Yoon S, Cheung YK, Alcántara C, Duran AT, Ensari I, Krupka DJ, Schwartz JE, Burg MM, Davidson KW, 2018 The Influence of Daily Stress on Sedentary Behavior: Group and Person (N of 1) Level Results of a 1-Year Observational Study. *Psychosom. Med.* 80, 620–627. [PubMed: 29846309]
- Diniz BS, Vieira EM, 2018 Stress, Inflammation, and Aging: An Association Beyond Chance. *Am. J. Geriatr. Psychiatry* 26, 964–965. [PubMed: 30064870]
- Effros RB, 2009 Kleemeier Award Lecture 2008—The Canary in the Coal Mine: Telomeres and Human Healthspan. *J. Gerontol. A Biol. Sci. Med. Sci.* 64A, 511–515.
- Ehinger JK, Morota S, Hansson MJ, Paul G, Elmér E, 2015 Mitochondrial dysfunction in blood cells from amyotrophic lateral sclerosis patients. *J. Neurol.* 262, 1493–1503. [PubMed: 25893255]
- Elliott HR, Tillin T, McArdle WL, Ho K, Duggirala A, Frayling TM, Davey Smith G, Hughes D, Chaturvedi N, Relton CL, 2014 Differences in smoking associated DNA methylation patterns in South Asians and Europeans. *Clin. Epigenetics* 6, 4. [PubMed: 24485148]
- Engelfriet PM, Jansen EHJM, Picavet HSJ, Dollé MET, 2013 Biochemical markers of aging for longitudinal studies in humans. *Epidemiol. Rev.* 35, 132–151. [PubMed: 23382477]
- Entringer S, de Punder K, Buss C, Wadhwa PD, 2018 The fetal programming of telomere biology hypothesis: an update. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 373 <https://doi.org/10.1098/rstb.2017.0151>
- Epel ES, Blackburn EH, Lin J, Dhabhar FS, Adler NE, Morrow JD, Cawthon RM, 2004 Accelerated telomere shortening in response to life stress. *Proc. Natl. Acad. Sci. U. S. A.* 101, 17312–17315. [PubMed: 15574496]
- Epel ES, Crosswell AD, Mayer SE, Prather AA, Slavich GM, Puterman E, Mendes WB, 2018 More than a feeling: A unified view of stress measurement for population science. *Front. Neuroendocrinol.* 49, 146–169. [PubMed: 29551356]
- Epel ES, Prather AA, 2018 Stress, Telomeres, and Psychopathology: Toward a Deeper Understanding of a Triad of Early Aging. *Annu. Rev. Clin. Psychol.* 14, 371–397. [PubMed: 29494257]
- Fernandes M, Wan C, Tacutu R, Barardo D, Rajput A, Wang J, Thoppil H, Thornton D, Yang C, Freitas A, de Magalhães JP, 2016 Systematic analysis of the gerontome reveals links between aging and age-related diseases. *Hum. Mol. Genet.* 25, 4804–4818. [PubMed: 28175300]
- Franceschi C, Bonafè M, Valensin S, Olivieri F, De Luca M, Ottaviani E, De Benedictis G, 2000 Inflamm-aging: an evolutionary perspective on immunosenescence. *Ann. N. Y. Acad. Sci.* 908, 244–254. [PubMed: 10911963]
- Franceschi C, Garagnani P, Parini P, Giuliani C, Santoro A, 2018a Inflammaging: a new immune-metabolic viewpoint for age-related diseases. *Nat. Rev. Endocrinol.* 10.1038/s41574-018-0059-4
- Franceschi C, Zaikin A, Gordleeva S, Ivanchenko M, Bonifazi F, Storci G, Bonafè M, 2018b Inflammaging 2018: An update and a model. *Semin. Immunol* <https://doi.org/10.1016/j.smim.2018.10.008>
- Franke K, Gaser C, 2012 Longitudinal Changes in Individual *BrainAGE* in Healthy Aging, Mild Cognitive Impairment, and Alzheimer's Disease | Data used in preparation of this article were

obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.edu). *GeroPsych* 25, 235–245.

- Fries GR, Bauer IE, Scaini G, Wu MJ, Kazimi IF, Valvassori SS, Zunta-Soares G, Walss-Bass C, Soares JC, Quevedo J, 2017 Accelerated epigenetic aging and mitochondrial DNA copy number in bipolar disorder. *Transl. Psychiatry* 7 10.1038/s41398-017-0048-8
- Fulop T, Larbi A, Dupuis G, Le Page A, Frost EH, Cohen AA, Witkowski JM, Franceschi C, 2018 Immunosenescence and inflamm-aging as two sides of the same coin: friends or foes? *Front. Immunol.* 8, 1960. [PubMed: 29375577]
- Gao X, Zhang Y, Mons U, Brenner H, 2018 Leukocyte telomere length and epigenetic-based mortality risk score: associations with all-cause mortality among older adults. *Epigenetics*. <https://doi.org/10.1080/15592294.2018.1514853>
- Garrett DD, Kovacevic N, McIntosh AR, Grady CL, 2010 Blood oxygen level-dependent signal variability is more than just noise. *J. Neurosci.* 30, 4914–4921. [PubMed: 20371811]
- Gaser C, Franke K, Klöppel S, Koutsouleris N, Sauer H, 2013 BrainAGE in Mild Cognitive Impaired Patients: Predicting the Conversion to Alzheimer's Disease. *PLoS One* 8 10.1371/journal.pone.0067346
- Geerligs L, Renken RJ, Saliassi E, Maurits NM, Lorist MM, 2015 A Brain-Wide Study of Age-Related Changes in Functional Connectivity. *Cereb. Cortex* 25, 1987–1999. [PubMed: 24532319]
- Gonzalez-Freire M, Scalzo P, D'Agostino J, Moore ZA, Diaz-Ruiz A, Fabbri E, Zane A, Chen B, Becker KG, Lehrmann E, Zukley L, Chia CW, Tanaka T, Coen PM, Bernier M, de Cabo R, Ferrucci L, 2018 Skeletal muscle ex vivo mitochondrial respiration parallels decline in vivo oxidative capacity, cardiorespiratory fitness, and muscle strength: The Baltimore Longitudinal Study of Aging. *Aging Cell* 17 10.1111/accel.12725
- Gotlib IH, LeMoult J, Colich NL, Foland-Ross LC, Hallmayer J, Joormann J, Lin J, Wolkowitz OM, 2014 Telomere length and cortisol reactivity in children of depressed mothers. *Mol. Psychiatry* 1–6. [PubMed: 24362539]
- Greco M, Villani G, Mazzucchelli F, Bresolin N, Papa S, Attardi G, 2003 Marked aging-related decline in efficiency of oxidative phosphorylation in human skin fibroblasts. *FASEB J.* 17, 1706–1708. [PubMed: 12958183]
- Griffanti L, Douaud G, Bijsterbosch J, Evangelisti S, Alfaro-Almagro F, Glasser MF, Duff EP, Fitzgibbon S, Westphal R, Carone D, Beckmann CF, Smith SM, 2017 Hand classification of fMRI ICA noise components. *Neuroimage* 154, 188–205. [PubMed: 27989777]
- Gutierrez Becker B, Klein T, Wachinger C, 2018 Gaussian process uncertainty in age estimation as a measure of brain abnormality. *Neuroimage* 175, 246–258. [PubMed: 29627589]
- Hajek T, Franke K, Kolenic M, Capkova J, Matejka M, Propper L, Uher R, Stopkova P, Novak T, Paus T, Kopecek M, Spaniel F, Alda M, 2017 Brain Age in Early Stages of Bipolar Disorders or Schizophrenia. *Schizophr. Bull.* 10.1093/schbul/sbx172
- Han LKM, Aghajani M, Clark SL, Chan RF, Hattab MW, Shabalin AA, Zhao M, Kumar G, Xie LY, Jansen R, Milaneschi Y, Dean B, Aberg KA, van den Oord EJCG, Penninx WJH, 2018 Epigenetic Aging in Major Depressive Disorder. *Am. J. Psychiatry* appi.app.2018.1.
- Hannum G, Guinney J, Zhao L, Zhang L, Hughes G, Sadda S, Klotzle B, Bibikova M, Fan JB, Gao Y, Deconde R, Chen M, Rajapakse I, Friend S, Ideker T, Zhang K, 2013 Genome wide Methylation Profiles Reveal Quantitative Views of Human Aging Rates. *Mol. Cell* 49, 359–367. [PubMed: 23177740]
- Hanssen LM, Schutte NS, Malouff JM, Epel ES, 2017 The Relationship Between Childhood Psychosocial Stressor Level and Telomere Length: A Meta-Analysis. *Health Psychol Res* 5, 6378. [PubMed: 28603779]
- Hattab MW, Shabalin AA, Clark SL, Zhao M, Kumar G, Chan RF, Xie LY, Jansen R, Han KM, Magnusson PKE, van Grootheest G, Hultman CM, Penninx BWJH, Aberg KA, van den Oord EJCG, 2017 Correcting for cell-type effects in DNA methylation studies: reference-based method outperforms latent variable approaches in empirical studies. *Genome Biol.* 18 <https://doi.org/10.1186/s13059-017-1149-7>

- Hatton SN, Franz CE, Elman JA, Panizzon MS, Hagler DJ Jr, Fennema-Notestine C, Eyer T, McEvoy LK, Lyons MJ, Dale AM, Kremen WS, 2018 Negative fateful life events in midlife and advanced predicted brain aging. *Neurobiol. Aging* 67, 1–9. [PubMed: 29609076]
- Heijmans BT, Mill J, 2012 Commentary: The seven plagues of epigenetic epidemiology. *Int. J. Epidemiol.* 41,74–78. [PubMed: 22269254]
- Hertel J, Frenzel S, König J, Wittfeld K, Fuellen G, Holtfreter B, Pietzner M, Friedrich N, Nauck M, Völzke H, Kocher T, Grabe HJ, 2018 The informative error: A framework for the construction of individualized phenotypes. *Stat. Methods Med. Res.* 962280218759138.
- Hertel J, Friedrich N, Wittfeld K, Pietzner M, Budde K, Van der Auwera S, Lohmann T, Teumer A, Volzke H, Nauck M, Grabe HJ, 2016 Measuring Biological Age via Metabonomics: The Metabolic Age Score. *J. Proteome Res.* 15, 400–410. [PubMed: 26652958]
- Horvath S, 2013 DNA methylation age of human tissues and cell types. *Genome Biol.* 14, R115. [PubMed: 24138928]
- Horvath S, Erhart W, Brosch M, Ammerpohl O, Schönfels W von, Ahrens M, Heits N, Bell JT, Tsai P-C, Spector TD, Deloukas P, Siebert R, Sipos B, Becker T, Röcken C, Schafmayer C, Hampe J, 2014 Obesity accelerates epigenetic aging of human liver. *Proceedings of the National Academy of Sciences* 201412759.
- Horvath S, Mah V, Lu AT, Woo JS, Choi O-W, Jasinska AJ, Riancho JA, Tung S, Coles NS, Braun J, Vinters HV, Coles LS, 2015 The cerebellum ages slowly according to the epigenetic clock. *Aging* 7, 294–306. [PubMed: 26000617]
- Horvath S, Oshima J, Martin GM, Lu AT, Quach A, Cohen H, Felton S, Matsuyama M, Lowe D, Kabacik S, Wilson JG, Reiner AP, Maierhofer A, Flunked J, Aviv A, Hou L, Baccarelli AA, Li Y, Stewart JD, Whitsel EA, Ferrucci L, Matsuyama S, Raj K, 2018 Epigenetic clock for skin and blood cells applied to Hutchinson Gilford Progeria Syndrome and ex vivo studies. *Aging* 10, 1758–1775. [PubMed: 30048243]
- Horvath S, Raj K, 2018 DNA methylation-based biomarkers and the epigenetic clock theory of ageing. *Nat. Rev. Genet.* 19, 371–384. [PubMed: 29643443]
- Hough CM, Bersani FS, Mellon SH, Epel ES, Reus VI, Lindqvist D, Lin J, Mahan L, Rosser R, Burke H, Coetzee J, Nelson JC, Blackburn EH, Wolkowitz OM, 2016 Leukocyte telomere length predicts SSRI response in major depressive disorder: A preliminary report. *Mol Neuropsychiatry* 2, 88–96. [PubMed: 27429957]
- Houseman EA, Molitor J, Marsit CJ, 2014 Reference-free cell mixture adjustments in analysis of DNA methylation data. *Bioinformatics* 30, 1431–1439. [PubMed: 24451622]
- Huan T, Chen G, Liu C, Bhattacharya A, Rong J, Chen BH, Seshadri S, Tanriverdi K, Freedman JE, Larson MG, Others, 2018 Age-associated micro RNA expression in human peripheral blood is associated with all-cause mortality and age-related traits. *Aging Cell* 17, e12687.
- Hummel EM, Hesses E, Müller S, Beiter T, Fisch M, Eibl A, Wolf OT, Giebel B, Platen P, Kumsta R, Moser DA, 2018 Cell-free DNA release under psychosocial and physical stress conditions. *Transl. Psychiatry* 8, 236. [PubMed: 30374018]
- Hurtado-Roca Y, Ledesma M, Gonzalez-Lazaro M, Moreno-Loshuertos R, Fernandez-Silva P, Enriquez JA, Laclaustra M, 2016 Adjusting MtDNA Quantification in Whole Blood for Peripheral Blood Platelet and Leukocyte Counts. *PLoS One* 11, e0163770. [PubMed: 27736919]
- Insel TR, Cuthbert BN, 2015 Brain disorders? Precisely. *Science* 348, 499–500. [PubMed: 25931539]
- Jaskelioff M, Muller FL, Paik J-HH, Thomas E, Jiang S, Adams AC, Sahin E, Kost-Alimova M, Protopopov A, Cadinanos J, Horner JW, Maratos-Flier E, DePinho R. a., Cadinanos J, Horner JW, Maratos-Flier E, DePinho R. a., 2011 Telomerase reactivation reverses tissue degeneration in aged telomerase-deficient mice. *Nature* 469, 102–106. [PubMed: 21113150]
- Jha MK, Trivedi MH, 2018 Personalized Antidepressant Selection and Pathway to Novel Treatments: Clinical Utility of Targeting Inflammation. *Int. J. Mol. Sci* 19 10.3390/ijms19010233
- Jylhava J, Pedersen NL, Hagg S, 2017 Biological Age Predictors. *EBioMedicine* 21,29–36. [PubMed: 28396265]
- Kågedal B, Lindqvist M, Farnebäck M, Lenner L, Peterson C, 2005 Failure of the PAXgene Blood RNA System to maintain mRNA stability in whole blood. *Clin. Chem. Lab. Med.* 43, 1190–1192. [PubMed: 16232084]

- Kageyama Y, Kasahara T, Kato M, Sakai S, Deguchi Y, Tani M, Kuroda K, Hattori K, Yoshida S, Goto Y, Kinoshita T, Inoue K, Kato T, 2018 The relationship between circulating mitochondrial DNA and inflammatory cytokines in patients with major depression. *J. Affect. Disord.* 233, 15–20. [PubMed: 28633757]
- Karabatsiakos A, Böck C, Salinas-Manrique J, Kolassa S, Calzia E, Dietrich DE, Kolassa I-T, 2014 Mitochondrial respiration in peripheral blood mononuclear cells correlates with depressive subsymptoms and severity of major depression. *Transl. Psychiatry* 4, e397.
- Kaufmann T, Meer DVD, Doan NT, Schwarz E, Martina J, Alnæs D, Barch DM, 2018 Genetics of brain age suggest an overlap with common brain disorders.
- Khandaker GM, Cousins L, Deakin J, Lennox BR, Yolken R, Jones PB, 2015 Inflammation and immunity in schizophrenia: implications for pathophysiology and treatment. *Lancet Psychiatry* 2, 258–270. [PubMed: 26359903]
- Khinchu PP, Dagnall CL, Hicks B, Jones K, Aviv A, Kimura M, Katki H, Aubert G, Giri N, Alter BP, Savage SA, Gadalla SM, 2017 Correlation of Leukocyte Telomere Length Measurement Methods in Patients with Dyskeratosis Congenita and in Their Unaffected Relatives. *Int. J. Mol. Sci* 18 10.3390/ijms18081765
- Knight AK, Conneely KN, Smith AK, 2017 Gestational age predicted by DNA methylation: potential clinical and research utility. *Epigenomics.* 10.2217/epi-2016-0157
- Kolenic M, Franke K, Hlinka J, Matejka M, Capkova J, Pausova Z, Uher R, Alda M, Spaniel F, Hajek T, 2018 Obesity, dyslipidemia and brain age in first-episode psychosis. *J. Psychiatr. Res.* 99, 151–158. [PubMed: 29454222]
- Koutsouleris N, Davatzikos C, Borgwardt S, Gaser C, Bottlender R, Frodl T, Falkai P, Riecher- Rössler A, Moller HJ, Reiser M, Pantelis C, Meisenzahl E, 2014 Accelerated brain aging in schizophrenia and beyond: A neuroanatomical marker of psychiatric disorders. *Schizophr. Bull.* 40, 1140–1153. [PubMed: 24126515]
- Krišti J, Vu kovi F, Menni C, Klari L, Keser T, Beceheli I, Pu i -Bakovi M, Novokmet M, Mangino M, Thaqi K, Rudan P, Novokmet N, Sarac J, Missoni S, Kol i I, Polašek O, Rudan I, Campbell H, Hayward C, Aulchenko Y, Valdes A, Wilson JF, Gornik O, Primorac D, Zoldoš V, Spector T, Lauc G, 2014 Glycans are a novel biomarker of chronological and biological ages. *J. Gerontol. A Biol. Sci. Med. Sci.* 69, 779–789. [PubMed: 24325898]
- Lawless C, Wang C, Jurk D, Merz A, von Zglinicki T, Passos JF, 2010 Quantitative assessment of markers for cell senescence. *Exp. Gerontol.* 45, 772–778. [PubMed: 20117203]
- Lee M, Martin H, Firpo MA, Demerath EW, 2011 Inverse association between adiposity and telomere length: The Fels Longitudinal Study. *Am. J. Hum. Biol.* 23, 100–106. [PubMed: 21080476]
- Lee M-R, Jung SM, Bang H, Kim HS, Kim YB, 2018 The association between muscular strength and depression in Korean adults: a cross-sectional analysis of the sixth Korea National Health and Nutrition Examination Survey (KNHANES VI) 2014. *BMC Public Health* 18, 1123. [PubMed: 30219042]
- Lemke MR, Wendorff T, Mieth B, Buhl K, Linnemann M, 2000 Spatiotemporal gait patterns during overground locomotion in major depression compared with healthy controls. *J. Psychiatr. Res.* 34, 277–283. [PubMed: 11104839]
- Le TT, Kuplicki R, Yeh HW, Aupperle RL, Khalsa SS, Simmons WK, Paulus MP, 2018 Effect of Ibuprofen on BrainAGE: A Randomized, Placebo-Controlled, Dose-Response Exploratory Study. *Biological Psychiatry: Cognitive Neuroscience and Neuroimaging* 1–8.
- Lever-van Milligen BA, Lamers F, Smit JH, Penninx BW, 2017 Six-year trajectory of objective physical function in persons with depressive and anxiety disorders. *Depress. Anxiety* 34, 188–197. [PubMed: 27701790]
- Levine ME, Lu AT, Quach A, Chen BH, Assimes TL, Bandinelli S, Hou L, Baccarelli AA, Stewart JD, Li Y, Whitsel EA, Wilson JG, Reiner AP, Aviv A, Lohman K, Liu Y, Ferrucci L, Horvath S, 2018 An epigenetic biomarker of aging for lifespan and healthspan. *Aging* 10, 573–591. [PubMed: 29676998]
- Levy BR, Slade MD, Kunkel SR, Kasl SV, 2002 Longevity increased by positive self-perceptions of aging. *J. Pers. Soc. Psychol.* 83, 261–270. [PubMed: 12150226]
- Liem F, 2016 Predicting brain-age from multimodal imaging data captures cognitive impairment.

- Lindqvist D, Epel ES, Mellon SH, Penninx BW, Revesz D, Verhoeven JE, Reus VI, Lin J, Mahan L, Hough CM, Rosser R, Bersani FS, Blackburn EH, Wolkowitz OM, 2015 Psychiatric disorders and leukocyte telomere length: Underlying mechanisms linking mental illness with cellular aging. *Neurosci. Biobehav. Rev.* 55, 333–364. [PubMed: 25999120]
- Lindqvist D, Fernström J, Grudet C, Ljunggren L, Träskman-Bendz L, Ohlsson L, Westrin Å, 2016 Increased plasma levels of circulating cell-free mitochondrial DNA in suicide attempters: associations with HPA-axis hyperactivity. *Transl. Psychiatry* 6, e971. [PubMed: 27922635]
- Lindqvist D, Wolkowitz OM, Picard M, Ohlsson L, Bersani FS, Fernström J, Westrin Å, Hough M, Lin J, Reus VI, Epel ES, Mellon SH, 2018 Circulating cell-free mitochondrial DNA, but not leukocyte mitochondrial DNA copy number, is elevated in major depressive disorder. *Neuropsychopharmacology* 43, 1557–1564. [PubMed: 29453441]
- Lin J, Cheon J, Brown R, Coccia M, Puterman E, Aschbacher K, Sinclair E, Epel E, Blackburn H, 2016 Systematic and Cell Type-Specific Telomere Length Changes in Subsets of Lymphocytes. *J Immunol Res* 2016, 5371050. [PubMed: 26977417]
- Lin J, Epel E, Cheon J, Kroenke C, Sinclair E, Bigos M, Wolkowitz O, Mellon S, Blackburn E, 2010 Analyses and comparisons of telomerase activity and telomere length in human T and B cells: insights for epidemiology of telomere maintenance. *J. Immunol. Methods* 352, 71–80. [PubMed: 19837074]
- Li X, Li W, Xu Y, 2018 Human Age Prediction Based on DNA Methylation Using a Gradient Boosting Regressor. *Genes* 9 10.3390/genes9090424
- Li X, Wang J, Zhou J, Huang P, Li J, 2017 The association between post-traumatic stress disorder and shorter telomere length: A systematic review and meta-analysis. *J. Affect. Disord.* 218, 322–326. [PubMed: 28486180]
- Li Z, He Y, Wang D, Tang J, Chen X, 2017 Association between childhood trauma and accelerated telomere erosion in adulthood: A meta-analytic study. *J. Psychiatr. Res.* 93, 64–71. [PubMed: 28601667]
- Li Z, Hu M, Zong X, He Y, Wang D, Dai L, Dong M, Zhou J, Cao H, Lv L, Chen X, Tang J, 2015 Association of telomere length and mitochondrial DNA copy number with risperidone treatment response in first-episode antipsychotic-naïve schizophrenia. *Sci. Rep.* 5, 18553. [PubMed: 26680692]
- Lohr JB, Palmer BW, Eidt CA, Aailaboyina S, Mausbach BT, Wolkowitz OM, Thorp SR, Jeste DV, 2015 Is Post-Traumatic Stress Disorder Associated with Premature Senescence? A Review of the Literature. *Am. J. Geriatr. Psychiatry* 23, 709–725. [PubMed: 25959921]
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G, 2013 The hallmarks of aging. *Cell* 153, 1194–1217. [PubMed: 23746838]
- Lu AT, Xue L, Salfati EL, Chen BH, Ferrucci L, Levy D, Joeannes R, Murabito JM, Kiel DP, Tsai P-C, Yet I, Bell JT, Mangino M, Tanaka T, McRae AF, Marioni RE, Visscher PM, Wray NR, Deary IJ, Levine ME, Quach A, Assimes T, Tsao PS, Absher D, Stewart JD, Li Y, Reiner AP, Hou L, Baccarelli AA, Whitsel EA, Aviv A, Cardona A, Day FR, Wareham NJ, Perry JRB, Ong KK, Raj K, Lunetta KL, Horvath S, 2018 GWAS of epigenetic aging rates in blood reveals a critical role for TERT. *Nat. Commun.* 9, 387. [PubMed: 29374233]
- Luders E, Cherbuin N, Gaser C, 2016 Estimating brain age using high-resolution pattern recognition: Younger brains in long-term meditation practitioners. *Neuroimage* 134, 508–513. [PubMed: 27079530]
- van der Maaten L, Hinton G, 2008 Visualizing Data using t-SNE. *J. Mach. Learn. Res.* 9, 2579–2605.
- Malik AN, Shahni R, Rodriguez-de-Ledesma A, Laftah A, Cunningham P, 2011 Mitochondrial DNA as a non-invasive biomarker: accurate quantification using real time quantitative PCR without co-amplification of pseudogenes and dilution bias. *Biochem. Biophys. Res. Commun.* 412, 1–7. [PubMed: 21703239]
- Malouff JM, Schutte NS, 2017 A meta-analysis of the relationship between anxiety and telomere length. *Anxiety Stress Coping* 30, 264–272. [PubMed: 27844481]
- Mamdani F, Rollins B, Morgan L, Myers RM, Barchas JD, Schatzberg a. F., Watson SJ, Akil H, Potkin SG, Bunney WE, Vawter MP, Sequeira P. a., 2015 Variable telomere length across post-mortem

- human brain regions and specific reduction in the hippocampus of major depressive disorder. *Transl. Psychiatry* 5, e636. [PubMed: 26371764]
- Maniates H, Stoop TB, Miller MW, Halberstadt L, Wolf EJ, 2018 Stress-Generative Effects of Posttraumatic Stress Disorder: Transactional Associations Between Posttraumatic Stress Disorder and Stressful Life Events in a Longitudinal Sample. *J. Trauma. Stress* 31, 191–201. [PubMed: 29630742]
- Marioni RE, Harris SE, Shah S, Mcrae AF, Zglinicki TV, Martin-ruiz C, Wray NR, Visscher PM, Deary IJ, 2016 The epigenetic clock and telomere length are independently associated with chronological age and mortality. *Int. J. Epidemiol.* 1–9. [PubMed: 27433568]
- Marioni RE, Shah S, McRae AF, Chen BH, Colicino E, Harris SE, Gibson J, Henders AK, Redmond P, Cox SR, Pattie A, Corley J, Murphy L, Martin NG, Montgomery GW, Feinberg AP, Fallin MD, Multhaup ML, Jaffe AE, Joehanes R, Schwartz J, Just AC, Lunetta KL, Murabito JM, Starr JM, Horvath S, Baccarelli AA, Levy D, Visscher PM, Wray NR, Deary IJ, 2015 DNA methylation age of blood predicts all-cause mortality in later life. *Genome Biol.* 16, 25. [PubMed: 25633388]
- Marsland AL, Walsh C, Lockwood K, John-Henderson NA, 2017 The effects of acute psychological stress on circulating and stimulated inflammatory markers: A systematic review and meta-analysis. *Brain, Behavior, and Immunity*, 64, 208–219.
- Martinez-Cengotitabengoa M, Carrascón L, O'Brien JT, Díaz-Gutiérrez M-J, Bermúdez-Ampudia C, Sanada K, Arrasate M, González-Pinto A, 2016 Peripheral Inflammatory Parameters in Late-Life Depression: A Systematic Review. *Int. J. Mol. Sci.* 17 10.3390/ijms17122022
- Martinsson L, Wei Y, Xu D, Melas PA, Mathé AA, Schalling M, Lavebratt C, Backlund L, 2013 Long-term lithium treatment in bipolar disorder is associated with longer leukocyte telomeres. *Transl. Psychiatry* 3, e261. [PubMed: 23695236]
- Mason AE, Hecht FM, Daubenmier JJ, Sbarra DA, Lin J, Moran PJ, Schleicher SG, Acree M, Prather AA, Epel ES, 2018 Weight Loss Maintenance and Cellular Aging in the Supporting Health Through Nutrition and Exercise Study. *Psychosom. Med.* 80, 609–619. [PubMed: 29901486]
- Mathur MB, Epel E, Kind S, Desai M, Parks CG, Sandler DP, Khazeni N, 2016 Perceived stress and telomere length: A systematic review, meta-analysis, and methodologic considerations for advancing the field. *Brain Behav. Immun.* 54, 158–169. [PubMed: 26853993]
- McKinney BC, Lin H, Ding Y, Lewis DA, Sweet RA, 2017a DNA methylation age is not accelerated in brain or blood of subjects with schizophrenia. *Schizophr. Res* <https://doi.org/10.1016/j.schres.2017.09.025>
- McKinney BC, Lin H, Ding Y, Lewis DA, Sweet RA, 2017b DNA methylation evidence against the accelerated aging hypothesis of schizophrenia. *npj Schizophrenia* 3, 13. [PubMed: 28560259]
- Mehta D, Bruenig D, Lawford B, Harvey W, Carrillo-Roa T, Morris CP, Jovanovic T, Young RM, Binder EB, Voisey J, 2018 Accelerated DNA methylation aging and increased resilience in veterans: The biological cost for soldiering on. *Neurobiol Stress* 8, 112–119. [PubMed: 29888306]
- Mengel-From J, Thinggaard M, Dalgård C, Kyvik KO, Christensen K, Christiansen L, 2014 Mitochondrial DNA copy number in peripheral blood cells declines with age and is associated with general health among elderly. *Hum. Genet.* 133, 1149–1159. [PubMed: 24902542]
- Menni C, Kiddle SJ, Mangino M, Viñuela A, Psatha M, Steves C, Sattler M, Buil A, Newhouse S, Nelson S, Williams S, Voyle N, Soininen H, Kloszewska I, Mecocci P, Tsolaki M, Vellas B, Lovestone S, Spector TD, Dobson R, Valdes AM, 2015 Circulating Proteomic Signatures of Chronological Age. *J. Gerontol. A Biol. Sci. Med. Sci.* 70, 809–816. [PubMed: 25123647]
- Michalak J, Troje NF, Fischer J, Vollmar P, Heidenreich T, Schulte D, 2009 Embodiment of Sadness and Depression—Gait Patterns Associated With Dysphoric Mood. *Psychosom. Med.* 71, 580. [PubMed: 19414617]
- Miller FJ, Rosenfeldt FL, Zhang C, Linnane AW, Nagley P, 2003 Precise determination of mitochondrial DNA copy number in human skeletal and cardiac muscle by a PCR-based assay: lack of change of copy number with age. *Nucleic Acids Res.* 31, e61–e61. [PubMed: 12771225]
- Moffitt TE, Belsky DW, Danese A, Poulton R, Caspi A, 2017 The Longitudinal Study of Aging in Human Young Adults: Knowledge Gaps and Research Agenda. *J. Gerontol. A Biol. Sci. Med. Sci.* 72, 210–215. [PubMed: 28087676]

- Mohammadi D, 2015 ENIGMA: crowdsourcing meets neuroscience. *Lancet Neurol.* 14, 462–463. [PubMed: 25814394]
- Monickaraj F, Gokulakrishnan K, Prabu P, Sathishkumar C, Anjana RM, Rajkumar JS, Mohan V, Balasubramanyam M, 2012 Convergence of adipocyte hypertrophy, telomere shortening and hypoadiponectinemia in obese subjects and in patients with type 2 diabetes. *Clin. Biochem.* 45, 1432–1438. [PubMed: 22827962]
- Monroe SM, 2008 Modern approaches to conceptualizing and measuring human life stress. *Annu. Rev. Clin. Psychol.* 4, 33–52. [PubMed: 17716038]
- Moreno-Villanueva M, Morath J, Vanhooren V, Elbert T, Kolassa S, Libert C, Biirke A, Kolassa I-T, 2013 N-glycosylation profiling of plasma provides evidence for accelerated physiological aging in post-traumatic stress disorder. *Transl. Psychiatry* 3, e320. [PubMed: 24169639]
- Munafò MR, Davey Smith G, 2018 Robust research needs many lines of evidence. *Nature* 553, 399–401. [PubMed: 29368721]
- Nakahira K, Kyung S-Y, Rogers AJ, Gazourian L, Youn S, Massaro AF, Quintana C, Osorio JC, Wang Z, Zhao Y, Lawler LA, Christie JD, Meyer NJ, Me Causland FR, Waikar SS, Waxman AB, Chung RT, Bueno R, Rosas IO, Fredenburgh LE, Baron RM, Christiani C, Hunninghake GM, Choi AMK, 2013 Circulating mitochondrial DNA in patients in the ICU as a marker of mortality: derivation and validation. *PLoS Med.* 10, e1001577; discussion e1001577. [PubMed: 24391478]
- Nenadic I, Dietzek M, Langbein K, Sauer H, Gaser C, 2017 BrainAGE score indicates accelerated brain aging in schizophrenia, but not bipolar disorder. *Psychiatry Research: Neuroimaging.* <https://doi.org/10.1016/j.pscychresns.2017.05.006>
- Ortega FB, Silventoinen K, Tynelius P, Rasmussen F, 2012 Muscular strength in male adolescents and premature death: cohort study of one million participants. *BMJ* 345, e7279. [PubMed: 23169869]
- Park DI, Štambuk J, Razdorov G, Pu i -Bakovi M, Martins-de-Souza D, Lauc G, Turck CW, 2018 Blood plasma/IgG N-glycome biosignatures associated with major depressive disorder symptom severity and the antidepressant response. *Sci. Rep.* 8, 179. [PubMed: 29317657]
- Peng W, Zhao J, Dong X, Banazadeh A, Huang Y, Hussien A, Mechref Y, 2018 Clinical application of quantitative glycomics. *Expert Rev. Proteomics* 15, 1007–1031. [PubMed: 30380947]
- Penninx BWJH, Milaneschi Y, Lamers F, Vogelzangs N, 2013 Understanding the somatic consequences of depression: biological mechanisms and the role of depression symptom profile. *BMC Med.* 11, 129. [PubMed: 23672628]
- Peters MJ, Joehanes R, Pilling LC, Schurmann C, Conneely KN, Powell J, Reinmaa E, Sutphin GL, Zhernakova A, Schramm K, Wilson YA, Kobes S, Tukiainen T, NABEC/UKBEC Consortium, Ramos YF, Goring HHH, Fornage M, Liu Y, Gharib SA, Stranger BE, De Jager PL, Aviv A, Levy D, Murabito JM, Munson PJ, Huan T, Hofman A, Uitterlinden G, Rivadeneira F, van Rooij J, Stolk L, Broer L, Verbiest MMPJ, Jhamai M, Arp P, Metspalu A, Tserel L, Milani L, Samani NJ, Peterson P, Kasela S, Codd V, Peters A, Ward-Caviness CK, Herder C, Waldenberger M, Roden M, Singmann P, Zeilinger S, Illig T, Homuth G, Grabe H-J, Volzke H, Steil L, Kocher T, Murray A, Melzer D, Yaghootkar H, Bandinelli S, Moses EK, Kent JW, Curran JE, Johnson MP, Williams-Blangero S, Westra H-J, McRae AF, Smith JA, Kardia SLR, Hovatta I, Perola M, Ripatti S, Salomaa V, Henders AK, Martin NG, Smith AK, Mehta D, Binder EB, Nylocks KM, Kennedy EM, Klengel T, Ding J, Suchy-Dacey AM, Enquobahrie DA, Brody J, Rotter JI, Chen Y.-D.L., Houwing-Duistermaat J, Kloppenburg M, Slagboom PE, Helmer Q, den Hollander W, Bean S, Raj T, Bakshi N, Wang QP, Oyston LJ, Psaty BM, Tracy RP, Montgomery GW, Turner ST, Blangero J, Meulenberg I, Ressler KJ, Yang J, Franke L, Kettunen J, Visscher PM, Neely GG, Korstanje R, Hanson RL, Prokisch H, Ferrucci L, Esko T, Teumer A, van Meurs JBJ, Johnson AD, 2015 The transcriptional landscape of age in human peripheral blood. *Nat. Commun.* 6, 8570. [PubMed: 26490707]
- Picard M, Hirano M, 2016 Disentangling (Epi) Genetic and Environmental Contributions to the Mitochondrial 3243A> G Mutation Phenotype: Phenotypic Destiny in Mitochondrial Disease? *JAMA Neurol.*
- Picard M, Juster R-P, McEwen BS, 2014 Mitochondrial allostatic load puts the “glue” back in glucocorticoids. *Nat. Rev. Endocrinol.* 10, 303–310. [PubMed: 24663223]
- Picard M, Juster R-P, Sabiston CM, 2013 Is the whole greater than the sum of the parts? Self-rated health and transdisciplinarity. *Health* 5, 24.

- Picard M, McEwen BS, 2018a Psychological Stress and Mitochondria: A Systematic Review. *Psychosom. Med.* 80, 141–153. [PubMed: 29389736]
- Picard M, McEwen BS, 2018b Psychological Stress and Mitochondria: A Conceptual Framework. *Psychosom. Med.* 80, 126–140. [PubMed: 29389735]
- Picard M, Prather AA, Puterman E, Cuillerier A, Coccia M, Aschbacher K, Burelle Y, Epel ES, 2018 A Mitochondrial Health Index Sensitive to Mood and Caregiving Stress. *Biol. Psychiatry* 84, 9–17. [PubMed: 29525040]
- Pinti M, Cevenini E, Nasi M, De Biasi S, Salvioli S, Monti D, Benatti S, Gibellini L, Cotichini R, Stazi MA, Trenti T, Franceschi C, Cossarizza A, 2014 Circulating mitochondrial DNA increases with age and is a familiar trait: Implications for “inflamm-aging.” *Eur. J. Immunol.* 44, 1552–1562. [PubMed: 24470107]
- Polho GB, De-Paula VJ, Cardillo G, dos Santos B, Kerr DS, 2015 Leukocyte telomere length in patients with schizophrenia: A meta-analysis. *Schizophr. Res.* 165, 195–200. [PubMed: 25975826]
- Price EM, Robinson WP, 2018 Adjusting for Batch Effects in DNA Methylation Microarray Data, a Lesson Learned. *Front. Genet* 9, 83. [PubMed: 29616078]
- Price LH, Kao H-T, Burgers DE, Carpenter LL, Tyrka AR, 2013 Telomeres and early-life stress: an overview. *Biol. Psychiatry* 73, 15–23. [PubMed: 22831981]
- Puterman E, Weiss J, Lin J, Schilf S, Slusher AL, Johansen KL, Epel ES, 2018 Aerobic exercise lengthens telomeres and reduces stress in family caregivers: A randomized controlled trial - Curt Richter Award Paper 2018. *Psychoneuroendocrinology* 98, 245–252. [PubMed: 30266522]
- Quach A, Levine ME, Tanaka T, Lu AT, Chen BH, Ferrucci L, Ritz B, Bandinelli S, Neuhauser ML, Beasley JM, Snetselaar L, Wallace RB, Tsao PS, Absher D, Assimes TL, Stewart JD, Li Y, Hou L, Baccarelli AA, Whitsel EA, Horvath S, 2017 Epigenetic clock analysis of diet, exercise, education, and lifestyle factors. *Aging* 9, 419–446. [PubMed: 28198702]
- Ram N, Grimm KJ, 2009 Growth Mixture Modeling: A Method for Identifying Differences in Longitudinal Change Among Unobserved Groups. *Int. J. Behav. Dev.* 33, 565–576. [PubMed: 23885133]
- Rantanen T, Harris T, Leveille SG, Visser M, Foley D, Masaki K, Guralnik JM, 2000 Muscle strength and body mass index as long-term predictors of mortality in initially healthy men. *J. Gerontol. A Biol. Sci. Med. Sci.* 55, M168–73. [PubMed: 10795731]
- Rao S, Kota LN, Li Z, Yao Y, Tang J, Mao C, Jain S, Xu Y, Xu Q, 2016 Accelerated leukocyte telomere erosion in schizophrenia: Evidence from the present study and a meta-analysis. *J. Psychiatr. Res.* 79, 50–56. [PubMed: 27174400]
- Rasgon N, Lin KW, Lin J, Epel E, Blackburn E, 2016 Telomere length as a predictor of response to Pioglitazone in patients with unremitted depression: a preliminary study. *Transl. Psychiatry* 6, e709. [PubMed: 26731446]
- Ratanatharathorn A, Boks MP, Maihofer AX, Aiello AE, Amstadter AB, Ashley-Koch AE, Baker G, Beckham JC, Bromet E, Dennis M, Garrett ME, Geuze E, Guffanti G, Hauser MA, Kilaru V, Kimbrel NA, Koenen KC, Kuan P-F, Logue MW, Luft BJ, Miller MW, Mitchell C, Nugent NR, Ressler KJ, Rutten BPF, Stein MB, Vermetten E, Vinkers CH, Youssef NA, VA Mid-Atlantic MIRECC Workgroup, PGC PTSD Epigenetics Workgroup, Uddin M, Nievergelt CM, Smith AK, 2017 Epigenome-wide association of PTSD from heterogeneous cohorts with a common multi-site analysis pipeline. *Am. J. Med. Genet. B Neuropsychiatr. Genet* 174, 619–630. [PubMed: 28691784]
- Reinius LE, Acevedo N, Joerink M, Pershagen G, Dahlén S-E, Greco D, Soderhall C, Scheynius A, Kere J, 2012 Differential DNA methylation in purified human blood cells: implications for cell lineage and studies on disease susceptibility. *PLoS One* 7, e41361. [PubMed: 22848472]
- Ren H, Collins V, Clarke SJ, Han J-S, Lam P, Clay F, Williamson LM, Andy Choo KH, 2012 Epigenetic changes in response to tai chi practice: a pilot investigation of DNA methylation marks. *Evid. Based. Complement. Alternat. Med.* 2012, 841810. [PubMed: 22719790]
- Rééus GZ, Fries GR, Stertz L, Badawy M, Passos IC, Barichello T, Kapczinski F, Quevedo J, 2015 The role of inflammation and microglial activation in the pathophysiology of psychiatric disorders. *Neuroscience* 300, 141–154. [PubMed: 25981208]

- Ridout KK, Levandowski M, Ridout SJ, Gantz L, Goonan K, Palermo D, Price LH, Tyrka AR, 2018 Early life adversity and telomere length: a meta-analysis. *Mol. Psychiatry* 23, 858–871. [PubMed: 28322278]
- Ridout KK, Ridout SJ, Price LH, Sen S, Tyrka AR, 2016 Depression and telomere length: A meta-analysis. *J. Affect. Disord.* 191,237–247. [PubMed: 26688493]
- Rogenmoser L, Kernbach J, Schlaug G, Gaser C, 2018 Keeping brains young with making music. *Brain Struct. Funct.* 223, 297–305. [PubMed: 28815301]
- Sanders JL, Boudreau RM, Newman AB, Newman AB, Newman AB, 2012 Understanding the Aging Process Using Epidemiologic Approaches, in: Newman AB, Cauley JA (Eds.), *The Epidemiology of Aging*. Springer Netherlands, Dordrecht, pp. 187–214.
- Sasaki H, Kasagi F, Yamada M, Fujita S, 2007 Grip strength predicts cause-specific mortality in middle-aged and elderly persons. *Am. J. Med.* 120, 337–342. [PubMed: 17398228]
- Sayer AA, Kirkwood TBL, 2015 Grip strength and mortality: a biomarker of ageing? *Lancet* 386, 226–227. [PubMed: 25982159]
- Schnack HG, Kahn RS, 2016 Detecting neuroimaging biomarkers for psychiatric disorders: Sample size matters. *Front. Psychiatry* 7 10.3389/fpsyt.2016.00050
- Schnack HG, van Haren NEM, Nieuwenhuis M, Hulshoff Pol HE, Cahn W, Kahn RS, 2016 Accelerated Brain Aging in Schizophrenia: A Longitudinal Pattern Recognition Study. *Am. J. Psychiatry* 173, 607–616. [PubMed: 26917166]
- Schutte NS, Malouff JM, 2016 The relationship between perceived stress and telomere length: A meta-analysis. *Stress Health* 32, 313–319. [PubMed: 25393133]
- Schutte NS, Malouff JM, 2015 The association between depression and leukocyte telomere length: A meta-analysis. *Depress. Anxiety* 32, 229–238. [PubMed: 25709105]
- Schutte NS, Malouff JM, 2014 A meta-analytic review of the effects of mindfulness meditation on telomerase activity. *Psychoneuroendocrinology* 42, 45–48. [PubMed: 24636500]
- Shalev I, Entringer S, Wadhwa PD, Wolkowitz OM, Puterman E, Lin J, Epel ES, 2013 Stress and telomere biology: a lifespan perspective. *Psychoneuroendocrinology* 38, 1835–1842. [PubMed: 23639252]
- Shalev I, Moffitt TE, Braithwaite AW, Danese A, Fleming NI, Goldman-Mellor S, Harrington HL, Houts RM, Israel S, Poulton R, Robertson SP, Sugden K, Williams B, Caspi A, 2014 Internalizing disorders and leukocyte telomere erosion: a prospective study of depression, generalized anxiety disorder and post-traumatic stress disorder. *Mol. Psychiatry* 19, 1163–1170. [PubMed: 24419039]
- Shalev I, Moffitt TE, Sugden K, Williams B, Houts RM, Danese A, Mill J, Arseneault L, Caspi A, 2012 Exposure to violence during childhood is associated with telomere erosion from 5 to 10 years of age : a longitudinal study. *Mol. Psychiatry* 18, 576–581. [PubMed: 22525489]
- Shields GS, Slavich GM, 2017 Lifetime Stress Exposure and Health: A Review of Contemporary Assessment Methods and Biological Mechanisms. *Soc. Personal. Psychol. Compass* 11 <https://doi.org/10.1111/spc3.12335>
- Shiels PG, 2010 Improving precision in investigating aging: why telomeres can cause problems. *J. Gerontol. A Biol. Sci. Med. Sci.* 65, 789–791. [PubMed: 20538902]
- Simon NM, Smoller JW, McNamara KL, Maser RS, Zalta AK, Pollack MH, Nierenberg AA, Fava M, Wong KK, 2006 Telomere shortening and mood disorders: preliminary support for a chronic stress model of accelerated aging. *Biol. Psychiatry* 60, 432–435. [PubMed: 16581033]
- Simons MJP, 2015 Questioning causal involvement of telomeres in aging. *Ageing Res. Rev.* 24, 191–196. [PubMed: 26304838]
- Simpkin AJ, Hemani G, Suderman M, Gaunt TR, Lyttleton O, Mcardle WL, Ring SM, Sharp GC, Tilling K, Horvath S, Kunze S, Peters A, Waldenberger M, Ward-Caviness C, Nohr A, Sørensen TIA, Relton CL, Smith GD, 2016 Prenatal and early life influences on epigenetic age in children: a study of mother-offspring pairs from two cohort studies. *Hum. Mol. Genet.* 25, 191–201. [PubMed: 26546615]
- Spiers H, Hannon E, Schalkwyk LC, Smith R, Wong CCY, O'Donovan MC, Bray NJ, Mill J, 2015 Methylomic trajectories across human fetal brain development. *Genome Res.* 25, 338–352. [PubMed: 25650246]

- Stavem K, Aaser E, Sandvik L, Bjørnholt JV, Erikssen G, Thaulow E, Erikssen J, 2005 Lung function, smoking and mortality in a 26-year follow-up of healthy middle-aged males. *Eur. Respir. J.* 25, 618–625. [PubMed: 15802334]
- Steffener J, Habeck C, O'Shea D, Razlighi Q, Bherer L, Stern Y, 2016 Differences between chronological and brain age are related to education and self-reported physical activity. *Neurobiol. Aging* 40, 138–144. [PubMed: 26973113]
- Stephan Y, Sutin AR, Terracciano A, 2018 Subjective Age and Mortality in Three Longitudinal Samples. *Psychosom. Med.* 80, 659–664. [PubMed: 29864106]
- Strawbridge R, Arnone D, Danese A, Papadopoulos A, Herane Vives A, Cleare AJ, 2015 Inflammation and clinical response to treatment in depression: A meta-analysis. *Eur. Neuropsychopharmacol.* 25, 1532–1543. [PubMed: 26169573]
- Susser E, Verhulst S, Kark JD, Factor-Litvak PR, Keyes K, Magnus P, Aviv A, 2016 Non-Dynamic Association of Depressive and Anxiety Disorders With Leukocyte Telomere Length? *Am. J. Psychiatry* 173, 1147.
- Tanaka T, Biancotto A, Moaddel R, Moore AZ, Gonzalez-Freire M, Aon MA, Candia J, Zhang P, Cheung F, Fantoni G, CHI consortium, Semba RD, Ferrucci L, 2018 Plasma proteomic signature of age in healthy humans. *Aging Cell* e12799. [PubMed: 29992704]
- Tardif CL, Gauthier CJ, Steele CJ, Bazin P-L, Schäfer A, Schaefer A, Turner R, Villringer A, 2016 Advanced MRI techniques to improve our understanding of experience-induced neuroplasticity. *Neuroimage* 131,55–72. [PubMed: 26318050]
- Teo YV, Capri M, Morsiani C, Pizza G, Faria AMC, Franceschi C, Neretti N, 2018 Cell-free DNA as a biomarker of aging. *Aging Cell*, e12890 Doi: 10.1111/ace1.12890 [PubMed: 30575273]
- Thompson PM, Stein JL, Medland SE, Hibar DP, Vasquez AA, Renteria ME, Toro R, Jahanshad N, Schumann G, Franke B, Wright MJ, Martin NG, Agartz I, Alda M, Alhusaini S, Almasy L, Almeida J, Alpert K, Andreasen NC, Andreassen OA, Apostolova LG, Appel K, Armstrong NJ, Aribisala B, Bastin ME, Bauer M, Beard CE, Bergmann O, Binder B, Blangero J, Bockholt HJ, Bøen E, Bois C, Boomsma DI, Booth T, Bowman IJ, Bralten J, Brouwer RM, Brunner HG, Brohawn DG, Buckner RL, Buitelaar J, Bulayeva K, Bustillo JR, Calhoun VD, Cannon DM, Cantor RM, Carless MA, Caseras X, Cavalleri GL, Chakravarty MM, Chang KD, Ching CRK, Christoforou A, Cichon S, Clark VP, Conrod P, Coppola G, Crespo-Facorro B, Curran JE, Czisch M, Deary IJ, de Geus EJC, den Braber A, Delvecchio G, Depondt C, de Haan L, de Zubicaray GI, Dima D, Dimitrova R, Djurovic S, Dong H, Donohoe G, Duggirala R, Dyer TD, Ehrlich S, Ekman CJ, Elvsåshagen T, Emsell L, Erk S, Espeseth T, Fagerness J, Fears S, Fedko I, Fernández G, Fisher SE, Foroud T, Fox PT, Francks C, Frangou S, Frey EM, Frodl T, Frouin V, Garavan H, Giddaluru S, Glahn DC, Godlewska B, Goldstein RZ, Gollub RL, Grabe HJ, Grimm O, Gruber O, Guadalupe T, Gur RE, Gur RC, Göring HHH, Hagenaars S, Hajek T, Hall GB, Hall J, Hardy J, Hartman CA, Hass J, Hatton SN, Haukvik UK, Hegenscheid K, Heinz A, Hickie IB, Ho B-C, Hoehn D, Hoekstra PJ, Hollinshead M, Holmes AJ, Homuth G, Hoogman M, Hong LE, Hosten N, Hottenga J-J, Hulshoff Pol HE, Hwang KS, Jack CR jr, Jenkinson M, Johnston C, Jönsson EG, Kahn RS, Kasperaviciute D, Kelly S, Kim S, Kochunov P, Koenders L, Krämer B, Kwok JBJ, Lagopoulos J, Laje G, Landen M, Landman BA, Lauriello J, Lawrie SM, Lee PH, Le Hellard S, Lemaitre H, Leonardo CD, Li C-S, Liberg B, Liewald DC, Liu X, Lopez LM, Loth E, Lourdasamy A, Luciano M, Macciardi F, Machielsen MWJ, Macqueen GM, Malt UF, Mandl R, Manoach DS, Martinot J-L, Matarin M, Mather KA, Mattheisen M, Mattingsdal M, Meyer-Lindenberg A, McDonald C, McIntosh AM, McMahon FJ, McMahon KL, Meisenzahl E, Melle I, Milanese Y, Mohnke S, Montgomery GW, Morris DW, Moses EK, Mueller BA, Muñoz Maniega S, Mühleisen TW, Müller-Myhsok B, Mwangi B, Nauck M, Nho K, Nichols TE, Nilsson L-G, Nugent AC, Nyberg L, Olvera RL, Oosterlaan J, Ophoff RA, Pandolfo M, Papalampropoulou-Tsiridou M, Pappmeyer M, Paus T, Pausova Z, Pearlson GD, Penninx W, Peterson CP, Pfennig A, Phillips M, Pike GB, Poline J-B, Potkin SG, Pütz B, Ramasamy A, Rasmussen J, Rietschel M, Rijpkema M, Risacher SL, Roffman JL, Roiz-Saàtianeaz R, Romanczuk-Seiferth N, Rose EJ, Royle NA, Rujescu D, Ryten M, Sachdev PS, Salami A, Satterthwaite TD, Savitz J, Saykin AJ, Scanlon C, Schmaal L, Schnack G, Schork AJ, Schulz SC, Schür R, Seidman L, Shen L, Shoemaker JM, Simmons A, Sisodiya SM, Smith C, Smoller JW, Soares JC, Sponheim SR, Sprooten E, Starr JM, Steen VM, Strakowski S, Strike L, Sussmann J, Sämman PG, Teumer A, Toga AW, Tordesillas-Gutierrez D, Trabzuni D, Trost S, Turner J, Van

den Heuvel M, van der Wee NJ, van Eijk K, van Erp TGM, van Haren NEM, van Eijl D, van Tol M-J, Valdés Hernández MC, Veltman DJ, Versace A, Völzke H, Walker R, Walter H, Wang L, Wardlaw JM, Weale ME, Weiner MW, Wen W, Westlye LT, Whalley HC, Whelan CD, White T., Winkler AM, Wittfeld K, Woldehawariat G, Wolf C, Zilles D, Zwiers MP, Thalamuthu A, Schofield PR, Freimer NB, Lawrence NS, Drevets W, Alzheimer's Disease Neuroimaging Initiative, EPIGEN Consortium, IMAGEN Consortium, Saguenay Youth Study (SYS) Group, 2014 The ENIGMA Consortium: large-scale collaborative analyses of neuroimaging and genetic data. *Brain Imaging Behav.* 8, 153–182. [PubMed: 24399358]

- Titus AJ, Gallimore RM, Salas LA, Christensen BC, 2017 Cell-type deconvolution from DNA methylation: a review of recent applications. *Hum. Mol. Genet.* 26, R216–R224. [PubMed: 28977446]
- Trumpff C, Marsland L, Basualto-Alarcon C, Martin JL, Carroll JE, Sturm G, Vincent AE, Mosharov EV, Gu Z, Kaufman BA, Picard M., 2019 Acute Psychological Stress Triggers Circulating Cell-Free Mitochondrial DNA. *Psychoneuroendocrinology* (under revision).
- Tyrka AR, Carpenter LL, Kao H-T, Porton B, Philip NS, Ridout SJ, Ridout KK, Price LH, 2015 Association of telomere length and mitochondrial DNA copy number in a community sample of healthy adults. *Exp. Gerontol.* 66, 17–20. [PubMed: 25845980]
- Tyrka AR, Parade SH, Price LH, Kao H-T, Porton B, Philip NS, Welch ES, Carpenter LL, 2016 Alterations of Mitochondrial DNA Copy Number and Telomere Length With Early Adversity and Psychopathology. *Biol. Psychiatry* 79, 78–86. [PubMed: 25749099]
- Tyrrell DJ, Bharadwaj MS, Van Horn CG, Kritchevsky SB, Nicklas BJ, Molina AJA, 2015 Respirometric Profiling of Muscle Mitochondria and Blood Cells Are Associated With Differences in Gait Speed Among Community-Dwelling Older Adults. *J. Gerontol. A Biol. Sci. Med. Sci.* 70, 1394–1399. [PubMed: 25030980]
- Urata M, Koga-Wada Y, Kayamori Y, Kang D, 2008 Platelet contamination causes large variation as well as overestimation of mitochondrial DNA content of peripheral blood mononuclear cells. *Ann. Clin. Biochem.* 45, 513–514. [PubMed: 18753426]
- Vance MC, Bui E, Hoepfner SS, Kovachy B, Prescott J, Mischoulon D, Walton ZE, Dong M, Nadal MF, Worthington JJ, Hoge EA, Cassano P, Orr EH, Fava M, de Vivo I, Wong K-K, Simon NM, 2018 Prospective association between major depressive disorder and leukocyte telomere length over two years. *Psychoneuroendocrinology* 90, 157–164. [PubMed: 29499556]
- van Gool CH, Kempen GJM, Bosma H, van Boxtel MPJ, Jolles J, van Eijk JTM, 2007 Associations between lifestyle and depressed mood: longitudinal results from the Maastricht Aging Study. *Am. J. Public Health* 97, 887–894. [PubMed: 16735630]
- Vanhooren V, Laroy W, Libert C, Chen C, 2008 N-glycan profiling in the study of human aging. *BioGerontology* 9, 351–356. [PubMed: 18431686]
- Verhoeven JE, Révész D, Epel ES, Lin J, Wolkowitz OM, Penninx BWJH, Révész D, Epel ES, Lin J, Wolkowitz OM, Penninx BWJH, 2014 Major depressive disorder and accelerated cellular aging: results from a large psychiatric cohort study. *Mo I. Psychiatry* 19, 895–901.
- Verhoeven JE, Révész D, Picard M, Epel EE, Wolkowitz OM, Matthews KA, Penninx BWJH, Puterman E, 2018 Depression, telomeres and mitochondrial DNA: Between- and within-person associations from a 10-year longitudinal study. *Mol. Psychiatry* 23, 850–857. [PubMed: 28348385]
- Verhoeven JE, Révész D, Wolkowitz OM, Penninx BWJH, 2014 Cellular aging in depression: Permanent imprint or reversible process?: An overview of the current evidence, mechanistic pathways, and targets for interventions. *Bioessays* 1–11. [PubMed: 24323915]
- Verhoeven JE, van Oppen P, Revesz D, Wolkowitz OM, Penninx BWJH, 2016 Depressive and Anxiety Disorders Showing Robust, but Non-Dynamic, 6-Year Longitudinal Association With Short Leukocyte Telomere Length. *Am. J. Psychiatry* 173, 617–624. [PubMed: 26940806]
- Vetter VM, Antje M, Karbasiyan M, Steinhagen-Thiessen E, Hopfenmiiller W, Demuth I, 2018 Epigenetic clock and relative telomere length represent largely different aspects of aging in the Berlin Aging Study II (BASE-II). *J. Gerontol. A Biol. Sci. Med. Sci.* 10.1093/gerona/gly184
- Vincent AE, Rosa HS, Pabis K, Lawless C, Chen C, Grinewald A, Rygiel KA, Rocha MC, Reeve AK, Falkous G, Perissi V, White K, Davey T, Petrof BJ, Sayer AA, Cooper C, Deehan D, Taylor RW,

- Turnbull DM, Picard M, 2018 Subcellular origin of mitochondrial DNA deletions in human skeletal muscle. *Ann. Neurol.* 10.1002/ana.25288
- Viron MJ, Stern TA, 2010 The impact of serious mental illness on health and healthcare. *Psychosomatics* 51,458–465. [PubMed: 21051676]
- Voisey J, Lawford BR, Morris CP, Wockner LF, Noble EP, Young RM, Mehta D, 2017 Epigenetic analysis confirms no accelerated brain aging in schizophrenia, *npj Schizophrenia* 3, 26. [PubMed: 28871179]
- Voisin S, Eynon N, Yan X, Bishop DJ, 2015 Exercise training and DNA methylation in humans. *Acta Physiol.* 213, 39–59.
- Wagner KH, Cameron-Smith D, Wessner B, Franzke B, 2016 Biomarkers of aging: From function to molecular biology. *Nutrients* 8, 8–10.
- Walker ER, McGee RE, Druss BG, 2015 Mortality in mental disorders and global disease burden implications: a systematic review and meta-analysis. *JAMA Psychiatry* 72, 334–341. [PubMed: 25671328]
- Wang Q, Zhan Y, Pedersen NL, Fang F, Hägg S, 2018 Telomere Length and All-Cause Mortality: A Meta-analysis. *Ageing Res. Rev.* 48, 11–20. [PubMed: 30254001]
- Weidner CI, Lin Q, Koch CM, Eisele L, Beier F, Ziegler P, Bauerschlag DO, Jöckel K-H, Erbel R, Mühlhausen TW, Zenke M, Brümmendorf TH, Wagner W, 2014 Aging of blood can be tracked by DNA methylation changes at just three CpG sites. *Genome Biol.* 15, R24. [PubMed: 24490752]
- West AP, Shadel GS, 2017 Mitochondrial DNA in innate immune responses and inflammatory pathology. *Nat. Rev. Immunol.* 17, 363–375. [PubMed: 28393922]
- Whalley HC, Gibson J, Marioni R, Walker RM, Clarke T-K, Howard DM, Adams MJ, Hall L, 2017 Accelerated epigenetic ageing in depression.
- Wikgren M, Maripuu M, Karlsson T, Nordfjäll K, Bergdahl J, Hultdin J, Del-Favero J, Roos G, Nilsson LG, Adolfsson R, Norrback KF, Nordfjäll K, Bergdahl J, Hultdin J, Del-Favero J, Roos G, Nilsson LG, Adolfsson R, Norrback KF, 2012 Short telomeres in depression and the general population are associated with a hypocortisolemic state. *Biol. Psychiatry* 71,294–300. [PubMed: 22055018]
- Wium-Andersen MK, Ørsted DD, Rode L, Bojesen SE, Nordestgaard BG, 2017 Telomere length and depression: prospective cohort study and Mendelian randomisation study in 67 306 individuals. *Br. J. Psychiatry* 210, 31–38. [PubMed: 27810892]
- Wolf EJ, Maniates H, Nugent N, Maihofer AX, Armstrong D, Ratanatharathorn A, Ashley-Koch AE, Garrett M, Kimbrel NA, Lori A, Va Mid-Atlantic Mirecc Workgroup, Aiello AE, Baker DG, Beckham JC, Boks MP, Galea S, Geuze E, Hauser MA, Kessler RC, Koenen KC, Miller MW, Ressler KJ, Risbrough V, Rutten BPF, Stein MB, Ursano RJ, Vermetten E, Vinkers H, Uddin M, Smith AK, Nievergelt CM, Logue MW, 2018 Traumatic stress and accelerated DNA methylation age: A meta-analysis. *Psychoneuroendocrinology* 92, 123–134. [PubMed: 29452766]
- Wolfers T, Doan NT, Kaufmann T, Alnæss D, Moberget T, Agartz I, Buitelaar JK, Ueland T, Melle I, Franke B, Andreassen OA, Beckmann CF, Westlye LT, Marquand AF, 2018 Mapping the Heterogeneous Phenotype of Schizophrenia and Bipolar Disorder Using Normative Models. *JAMA Psychiatry*, 10.1001/jamapsychiatry.2018.2467
- Wolkowitz OM, Mellon SH, Epel ES, Lin J, Reus VI, Rosser R, Burke H, Compagnone M, Nelson JC, Dhabhar FS, Blackburn EH, 2012 Resting leukocyte telomerase activity is elevated in major depression and predicts treatment response. *Mol. Psychiatry* 17, 164–172. [PubMed: 21242992]
- Xia J, Sinelnikov IV, Han B, Wishart DS, 2015 MetaboAnalyst 3.0—making metabolomics more meaningful. *Nucleic Acids Res.* 43, W251–W257. [PubMed: 25897128]
- Xia P, Radpour R, Zachariah R, Fan AXC, Kohler C, Hahn S, Holzgreve W, Zhong XY, 2009 Simultaneous quantitative assessment of circulating cell-free mitochondrial and nuclear DNA by multiplex real-time PCR. *Genet. Mol. Biol.* 32, 20–24. [PubMed: 21637641]
- Xia X, Chen W, McDermott J, Han J-DJ, 2017 Molecular and phenotypic biomarkers of aging. *F1000Res.* 6, 860. [PubMed: 28663789]
- Yarkoni T, Westfall J, 2017 Choosing Prediction Over Explanation in Psychology: Lessons From Machine Learning. *Perspect. Psychol. Sci.* 12, 1100–1122. [PubMed: 28841086]

- Yates T, Zaccardi F, Dhalwani NN, Davies MJ, Bakrania K, Celis-Morales CA, Gill JMR, Franks PW, Khunti K, 2017 Association of walking pace and handgrip strength with all-cause, cardiovascular, and cancer mortality: a UK Biobank observational study. *Eur. Heart J.* 38, 3232–3240. [PubMed: 29020281]
- Yeung SSY, Reijnierse EM, Trappenburg MC, Hogrel J-Y, McPhee JS, Piasecki M, Sipilä S, Salpakoski A, Butler-Browne G, Pääsuke M, Gapeyeva H, Narici MV, Meskers CGM, Maier AB, 2018 Handgrip Strength Cannot Be Assumed a Proxy for Overall Muscle Strength. *J. Am. Med. Dir. Assoc* 19, 703–709. [PubMed: 29935982]
- Yu W-Y, Chang H-W, Lin C-H, Cho C-L, 2008 Short telomeres in patients with chronic schizophrenia who show a poor response to treatment. *J. Psychiatry Neurosci.* 33, 244–247. [PubMed: 18592039]
- Yu X, Wang Y, Kristie J, Dong J, Chu X, Ge S, Wang H, Fang H, Gao Q, Liu D, Zhao Z, Peng H, Pucic Bakovic M, Wu L, Song M, Rudan I, Campbell H, Lauc G, Wang W, 2016 Profiling IgG N-glycans as potential biomarker of chronological and biological ages: A community- based study in a Han Chinese population. *Medicine* 95, e4112. [PubMed: 27428197]
- Zannas AS, Arloth J, Carrillo-Roa T, Iurato S, Röh S, Ressler KJ, Nemeroff CB, Smith AK, Bradley B, Heim C, Menke A, Lange JF, Brückl T, Ising M, Wray NR, Erhardt A, Binder EB, Mehta D, 2015 Lifetime stress accelerates epigenetic aging in an urban, African American cohort: relevance of glucocorticoid signaling. *Genome Biol.* 16, 266. [PubMed: 26673150]
- Zhang J, Rane G, Dai X, Shanmugam MK, Arfuso F, Sarny RP, Lai MKP, Kappei D, Kumar AP, Sethi G, 2016 Ageing and the telomere connection: An intimate relationship with inflammation. *Ageing Res. Rev.* 25, 55–69. [PubMed: 26616852]
- Zhang R, Wang Y, Ye K, Picard M, Gu Z, 2017 Independent impacts of aging on mitochondrial DNA quantity and quality in humans. *BMC Genomics* 18, 890. [PubMed: 29157198]
- Zwicker A, Fabbri C, Rietschel M, Hauser J, Mors O, Maier W, Zobel A, Farmer A, Aitchison KJ, McGuffin P, Lewis CM, Uher R, 2018 Genetic disposition to inflammation and response to antidepressants in major depressive disorder. *J. Psychiatr. Res.* 105, 17–22. [PubMed: 30130674]

Highlights

- Stress and mental illness are prospectively associated with biological aging
- Technical limitations can compromise the interpretation of biological age indicators
- Integrating biological age indicators into composite indices improves their usefulness
- Human psychobiology is an opportunity to elucidate basic aging processes
- We highlight key challenges and suggest recommendations to overcome them

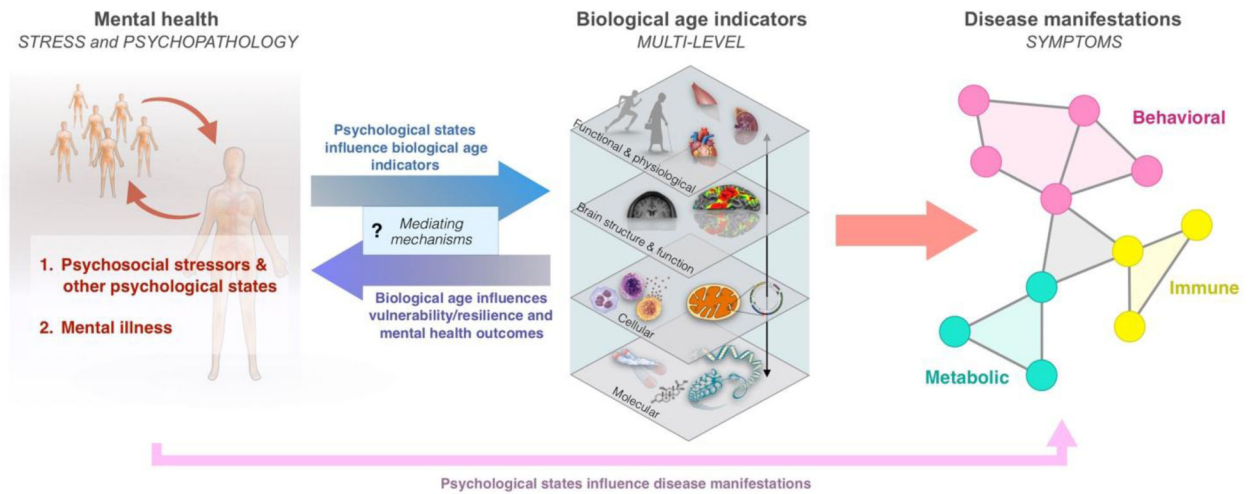


Figure 1. Integrative model for the transduction of mental health into biological aging and downstream disease manifestations.

(*Left*) Two main domains of mental health are considered: 1. Acute and chronic psychosocial stressors, which include distress and other subjective experiences; 2. Mental illness and clinical psychopathology (e.g., depression, anxiety, schizophrenia, bipolar disorder, etc). (*Middle*) These factors are transduced into biological age indicators, which span functional and physiological, brain structure and function, cellular, and molecular levels of analysis. In turn, the reverse association may transduce increased biological age into increased vulnerability and resilience to life stressors. The mechanisms responsible for the bi-directional flow of information between psychological states, psychopathology, and biological age indicators largely remain to be defined. (*Right*) Increased biological aging reflected in individual or combinations of biological age indicators manifest in symptoms across multiple interconnected systems, represented here as a functional network. Mental health domains can also directly contribute to disease manifestations (*bottom arrow*). Molecular indicators refer to components that are inert when in isolation (e.g. DNA, proteins) whereas cellular indicators refer to animated “living” components (e.g. breathing mitochondria, dividing/secretory cells).

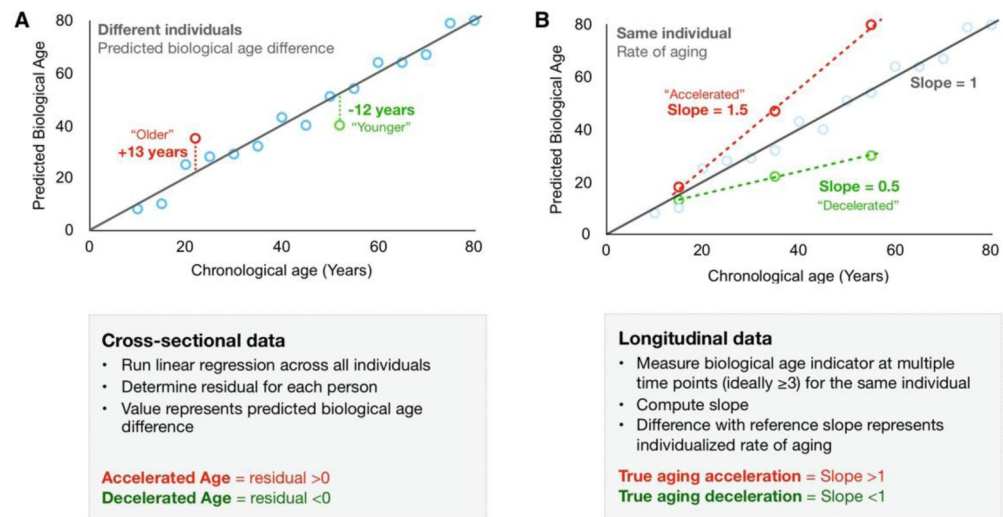


Figure 2. Computing age acceleration using cross-sectional and longitudinal data.

(A) From cross-sectional data, accelerated aging is established when biological age is over-predicted relative to the chronological age reference (group regression line). (B) In longitudinal data, the rate of aging is directly determined from multiple measurements in the same person (also same tissue and cell type). The slope for each individual can be compared to the theoretical slope of 1 to ascertain true aging acceleration or deceleration.

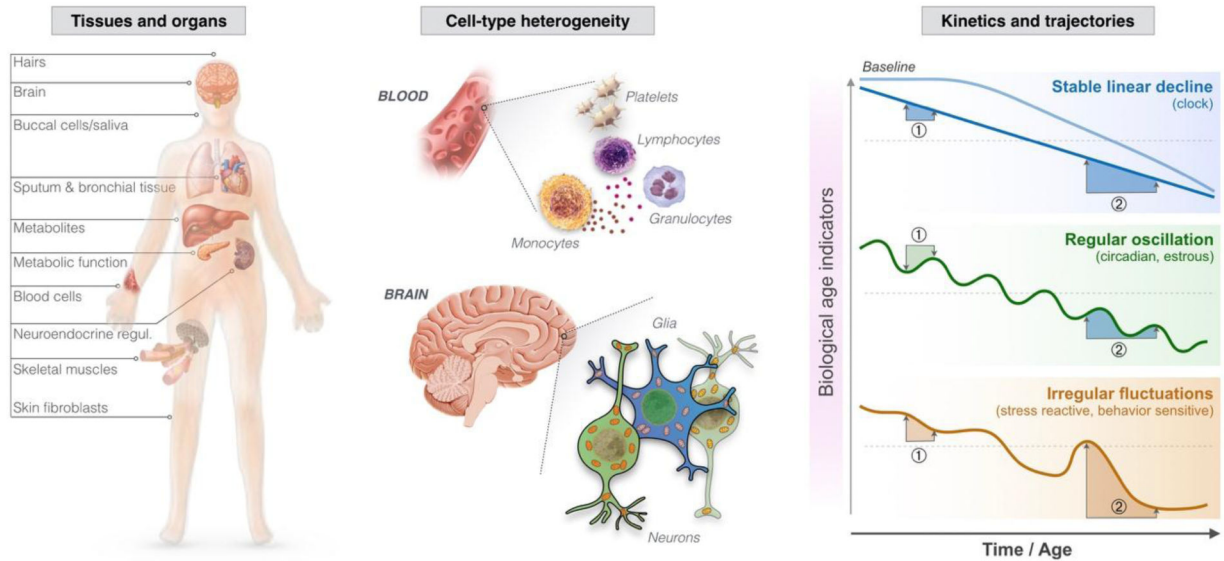


Figure 3. Major limitations of biological age indicators.

(*Left*) Biological age indicators are measured in samples from different tissues and organs of the body. (*Middle*) Individual tissues such as blood and the cortical regions of the brain also exhibit substantial heterogeneity marked by relative abundance of different cell-types. (*Right*) Three hypothetical kinetics for biological age indicators are shown: The first shows a stable decline, which is typically assumed, but not necessarily accurate for most biological age indicators. The second illustrates an indicator subject to either circadian regulation or monthly estrous cycle, thus exhibiting regular oscillations. One such example is cortisol. Knowledge of this oscillatory pattern can be used to adequately schedule time of sampling (e.g., morning or evening) and derive useful parameters (e.g., cortisol awakening response). The third indicator shows irregular fluctuations, which could arise from sensitivity to acute stress mediators or to behavior (exercise, sleep, or other). Note the two hypothetical pairs of assessments on each trajectory. Mis-timing of measurement ① for the regularly oscillating measure leads to pseudo-reversal of the biological aging indicator. Assessment ② shows an exaggerated decline in the irregular fluctuation.

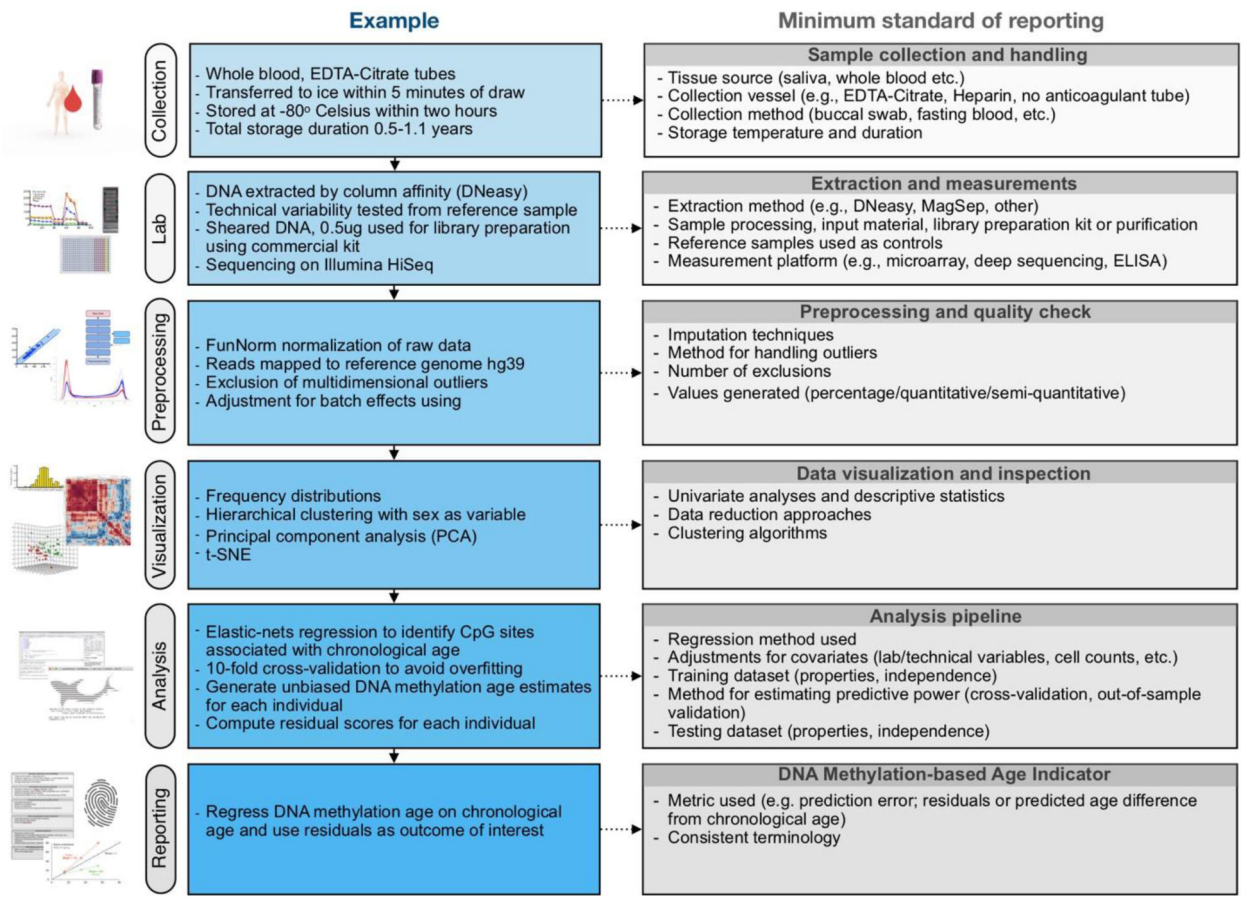


Figure 4. Overview of the biological age indicator prediction process and recommended minimum reporting guidelines.

(Left) Example workflow for calculating a epigenetic age indicator and age acceleration.

(Right) Recommended workflow that can inform study design, execution, analysis, and preparation of methods section in the resulting reports. Adherence to such standards for reporting results would facilitate harmonization of datasets across laboratories and cohort studies.

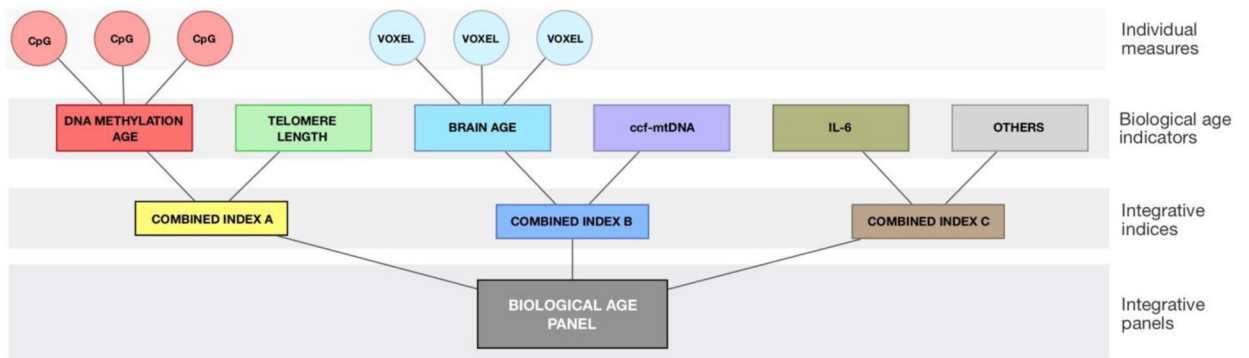


Figure 5. Topology of biological age indicators, upstream measures, and downstream integrative indices and panels.

Biologically-informed and functionally relevant composite indices integrating two or more indicators can be derived from individual indicators. Their added predictive power should be validated by either in-sample cross-validation or preferably out-of-sample validation.

Integrative biological age panels may eventually outperform single biological age indicators and indices due to the strength of their association, their construct stability, and/or their greater generalizability across individuals and independent samples.

Table 1.

Selective list of biological age indicators.

Category	Biological age indicator	Method	Correlation with chronological age	Evidence for mental health association	Representative survival or method-related references
Functional & physiological measures	Walking speed/gait	Timed distance walked Self-report	Inverse	Impaired in depression (Lemke et al., 2000; Michalak et al., 2009)	(Cooper et al., 2010 [†] ; Graham et al., 2008 [†] ; Yates et al., 2017)
	Hand-grip strength	Dynamometer	Inverse	Lower in depression (Lee et al., 2018; Lever-van Milligen et al., 2017)	(Bohannon, 2008 [†] ; Ortega et al., 2012; Sasaki et al., 2007; Sayer and Kirkwood, 2015)
	Lung function	Spirometer	Inverse	Impaired in depression (Lever-van Milligen et al., 2017)	(Stavem et al., 2005)
Brain measures	Brain age	T1-MRI	Positive/high	Higher in SCZ (Schmack et al., 2016), MDD, BPD, and psychosis (risk) (Kolenic et al., 2018; Koutsouleris et al., 2014) Inconsistent in BD (Hajek et al., 2017; Nenadic et al., 2017) Higher with more negative fateful life events (Hatton et al., 2018)	(Cole et al., 2018, 2017)
	Respiratory capacity	Mitochondrial enzymatic activities Respiratory capacity in fresh cells	Inverse	Enzymatic activities: correlated with previous day positive mood (Picard et al., 2018) Cellular respiration: associations with early life adversity (Boeck et al., 2018a) and MDD (Boeck et al., 2018d; Karabatsiakis et al., 2014)	(Gonzalez-Freire et al., 2018; Greco et al., 2003; Tyrrell et al., 2015)
Molecular measures	Telomere length	qPCR Southern blot Q-FISH	Inverse	Shorter in MDD (Schutte and Malouff, 2015), anxiety disorders (Malouff and Schutte, 2017), SCZ (Polho et al., 2015), PTSD (Li et al., 2017) N.S. in BD	(Aubert et al., 2012; Aviv et al., 2011; Cawthon, 2002; Wang et al., 2018 [†])
	Epigenetic age	Microarray and sequencing	Positive/high	Increased in response to traumatic stress (Wolf et al., 2018), life stress (Zannas et al., 2015), BD (Fries et al., 2017), MDD (Han et al., 2018; Whalley et al., 2017) N.S. in SCZ (McKinney et al., 2017a)	(Chen et al., 2016 [†] ; Christiansen et al., 2016; Hannum et al., 2013; Horvath, 2013; Marioni et al., 2015; Vetter et al., 2018)
Cellular measures	Transcriptomic age	RNA-seq	Positive/moderate to high	NA	(Huan et al., 2018; Peters et al., 2015)
	Proteomic age	Mass spectrometry	Positive/high	NA	(Menni et al., 2015; Tamaka et al., 2018)
	Metabolomic age	Mass spectrometry	Positive/high	NA	(Chaleckis et al., 2016; Hentel et al., 2016)
	Glycomics	Microarray and mass spectrometry	Positive/moderate to high	Altered in MDD (Boeck et al., 2018c; Park et al., 2018) GlycoAge Test was higher in PTSD (Moreno-Villanueva et al., 2013)	(Knšti et al., 2014; Peng et al., 2018; Vanhooren et al., 2008; Yu et al., 2016)
			Positive/moderate to high		

Category	Biological age indicator	Method	Correlation with chronological age	Evidence for mental health association	Representative survival or method-related references
	ccf-mtDNA	qPCR	Positive	Higher in plasma of suicidal patients (Lindqvist et al., 2016) and MDD (Lindqvist et al., 2018) Acutely elevated with induced psychological stress in plasma (Hummel et al., 2018) and serum (Trumpff et al., 2019)	(Pinti et al., 2014)
	mtDNAcn	qPCR (whole blood)	Inverse	Higher in BD (Fries et al., 2017) No evidence for an association with depression Increased in mixed psychiatric disorders	(Mengel-From et al., 2014; Verhoeven et al., 2018; Zhang et al., 2017)

Abbreviations: BD, bipolar disorder; BPD, borderline personality disorder; ccf-mtDNA, circulating cell-free mitochondrial DNA; MDD, major depressive disorder; MRI, magnetic resonance imaging; mtDNAcn, mitochondrial DNA copy number; NA, not applicable; N.S., not significant; PTSD, post-traumatic stress disorder; qPCR, quantitative polymerase chain reaction; Q-FISH, quantitative fluorescent in situ hybridization; SCZ, schizophrenia. Note that correlation coefficients (r) for biological age indicators and chronological age is dependent on the age range within each dataset and should not be used as a sole metric of accuracy. Criteria used for size of r: High: 0.7-0.9 (-0.7 to -0.9); Moderate: 0.5-0.7 (-0.5 to -0.7); Low: 0.3-0.5 (-0.3 to -0.5); Negligible: 0 - 0.3 (-0.3 to 0). Effect size reflects correlation coefficient reported in the original publications of the multivariate composites, indicated by †. Systematic reviews and meta-analytic studies are indicated by ‡.