Journal Club

Editor's Note: These short reviews of recent *JNeurosci* articles, written exclusively by students or postdoctoral fellows, summarize the important findings of the paper and provide additional insight and commentary. If the authors of the highlighted article have written a response to the Journal Club, the response can be found by viewing the Journal Club at www.jneurosci.org. For more information on the format, review process, and purpose of Journal Club articles, please see http://jneurosci.org/content/preparing-manuscript#journalclub.

SIRT1: A Novel Way to Target Tau?

Hunter S. Futch^{1,2,3} and ©Cara L. Croft^{1,2,3}

¹Department of Neuroscience, University of Florida, Gainesville, Florida 32610, ²Center for Translational Research in Neurodegenerative Disease, University of Florida, Gainesville, Florida 32610, and ³McKnight Brain Institute, University of Florida, Gainesville, Florida 32610 Review of Min et al.

ert and Spillantini, 2017), and therapeu-

tics that can halt this propagation or target

NFTs might therefore be useful to treat

AD and other tauopathies (Golde et al.,

tion of tau increases its propensity to

Much evidence indicates that acetyla-

Tau is a microtubule-associated protein that forms pathogenic inclusions in several neurodegenerative diseases, including Alzheimer's disease (AD). Under physiological conditions, dynamic posttranslational modifications of tau regulate cytoskeletal stability. Under pathological conditions, tau undergoes numerous post-translational modifications that may precede aggregation, including hyperphosphorylation, truncation, and abnormal acetylation. Neurofibrillary tangles (NFTs) are intraneuronal inclusions of tau aggregated into an amyloid structure and are one of the main histopathological characteristics of AD. Importantly, the density of NFTs in the AD brain is correlated with neuronal death and cognitive decline in the disease (Iqbal et al., 2016). Although the precise mechanisms of initial tau seeding into an amyloid structure are unknown, aggregated tau fibrils can conformationally template soluble tau and therefore "propagate" tau aggregation throughout the brain via anatomically connected pathways (Ayers et al., 2018). This spreading of pathological tau throughout the AD brain is thought to be important in disease progression (Goed-

aggregate, influences other post-translational modifications, and can reduce degradation of phosphorylated tau (Min et al., 2010; Cook et al., 2014b). In addition, tau acetylation promotes the seeding of tau *in vitro* and may contribute to tau propagation (Trzeciakiewicz et al., 2017). Studies of human AD brain found colocalization of tau acetylated on lysine residue 280 (K280) with phosphorylated tau in NFTs. Increased amounts of acetylated K280-positive NFTs were directly associated with more advanced AD stage (Irwin et al., 2012). Together, these data suggest

of tau (Lysines 259, 290, 321, and 353) has been found to inhibit aggregation, possibly by preventing phosphorylation at these sites (Cook et al., 2014a). Therefore, any viable therapeutic approach will need to deacetylate tau preferentially at the residues (K280 and K174) that promote ag-

that promoting tau deacetylation might

be an effective strategy for inhibiting tau

aggregation and propagation in tauopa-

thies. However, acetylation on four resi-

dues in the microtubule binding domain

When considering possible therapeutic strategies that target tau acetylation, the most logical direct approaches would be

gregation (Min et al., 2015).

through inhibition of tau acetylation or by promotion of tau deacetylation (Cook et al., 2014b). One way to increase tau deacetylation is by targeting sirtuins. Sirtuins are a class of deacetylases that have widespread effects on cellular physiology through deacetylation of histones and other proteins. Sirtuin 1 (SIRT1) can deacetylate tau; and notably, its levels are reduced in AD brain tissue, with levels progressively decreasing as the disease advances (Julien et al., 2009; Jęśko et al., 2017). SIRT1 can deacetylate tau residues 160–182 and 264–287 in vitro (Min et al., 2010). In addition, SIRT1 deacetylates components of the autophagy pathway and inhibits mammalian target of rapamycin signaling, both of which could lead to increased breakdown of intracellular aggregates, such as tau (Ng and Tang, 2013).

In an article recently published in The Journal of Neuroscience, Min et al. (2018) confirmed that SIRT1 deacetylates tau at several residues in vitro as reported previously. In addition, the authors use the PS19 mouse model of tauopathy that overexpresses a human tau variant that contains a serine residue instead of a proline at amino acid 301 (P301S) and is associated with frontotemporal dementia with parkinsonism, a primary tauopathy (Sperfeld et al., 1999; Yoshiyama et al., 2007). The authors generated a line of these transgenic tau mice with brainspecific deletion of the SIRT1 gene to investigate the absence of SIRT1 on tau acetylation and disease progression in vivo.

Received May 12, 2018; revised July 3, 2018; accepted July 12, 2018. The authors declare no competing financial interests.

Correspondence should be addressed to either Hunter S. Futch or Dr. Cara L. Croft, University of Florida, Department of Neuroscience, 1275 Center Drive, BMS J492, Box 100159 Gainesville, FL 32610, E-mail: hsfutch@ufl.edu or cara.croft@ufl.edu.

DOI:10.1523/JNEUROSCI.1201-18.2018

Copyright © 2018 the authors 0270-6474/18/387755-03\$15.00/0

Brain-specific deletion of SIRT1 increased acetylation of tau at K174 in vivo. In addition, SIRT1 deletion modestly reduced synaptophysin levels, which is suggestive of reduced synaptic connectivity, a feature of AD (Spires-Jones and Hyman, 2014). Reduction of SIRT1 also led to increased amounts of total tau and acetylated tau at the postsynaptic compartment; increased synaptic tau has been shown to contribute to tau propagation in AD (DeVos et al., 2018). Lastly, brain SIRT1 deletion increased the mortality rate in PS19 mice, suggesting that reduction of SIRT1 in the setting of tauopathy is detrimental to survival.

The authors went on to investigate the effects of SIRT1 on the propagation of tau aggregates. To do so, they injected tau fibrils into the hippocampus on one side of the brain and virally expressed SIRT1 in the contralateral hippocampus via injection of recombinant adeno-associated virus. SIRT1 overexpression reduced the spread of tau aggregates across the midline, reducing tau staining by 50%. This result is somewhat surprising given that their previous experiment showed that SIRT1 deletion in the PS19 model did not affect the overall levels of tau aggregation. This discrepancy could be attributed to recent findings that show the PS19 mouse model can display widespread variability in tau aggregation and neurodegeneration (Woerman et al., 2017). Alternatively, this finding may indicate that tau acetylation is more relevant to tau propagation than to aggregation.

Finally, Min et al. (2018) showed that brain-specific deletion of SIRT1 had deleterious effects on learning in both nontransgenic and tau transgenic mice, indicating that SIRT1 is important for normal cognition. This is not wholly surprising, as SIRT1 affects numerous pathways other than tau deacetylation. Most importantly, as a histone deacetylase, SIRT1 regulates DNA coiling and genomewide transcriptional activity. Reported effects of SIRT1 on normal cellular physiology include upregulation of mitochondrial transcriptional programs that prevent oxidative stress via increased mitogenesis. In addition, SIRT1 can increase neurotrophic signals, such as BDNF (Jęśko et al., 2017). In this paper, SIRT1 deletion negatively impacted longevity and learning ability, suggesting that decreasing SIRT1 expression (such as that observed in AD brain) may be deleterious. Indeed, due to the extensive influences of SIRT1 on cellular physiology, it would be necessary to establish that SIRT1 overexpression will not dysregulate normal brain function if one were to propose it as a potential therapeutic.

Similar to the pleotropic effects that SIRT1 has on normal cellular processes, SIRT1 can affect tau through several pathways, including regulation of autophagy (Ng and Tang, 2013), by promoting proteasomal degradation, through posttranslational modifications and at a transcriptional level. SIRT1 inhibits mammalian target of rapamycin signaling and thereby enhances the autophagy system; stimulation of autophagy has been proposed as a potential therapeutic strategy across neurodegenerative diseases, as nearly all of the pertinent protein aggregates, including tau, can be degraded via autophagy pathways (Menzies et al., 2017). SIRT1 can promote proteasomal degradation of tau via deacetylation, which increases available sites for the ubiquitination of tau and therefore increases targeting of tau to the proteasome (Min et al., 2010). SIRT1 overexpression can also reduce tau phosphorylation (Corpas et al., 2017), and SIRT1 deletion increases tau phosphorylation (Min et al., 2010). There is a tremendous amount of evidence supporting the dephosphorylation of tau as a therapeutic strategy in AD, and SIRT1 overexpression could target tau in this manner. It should also be discussed that human tau undergoes alternative splicing to produce six distinct isoforms, containing either three (3R) or four (4R) microtubule binding domains due to exclusion or inclusion of exon 10, respectively. Increased 4R tau can increase tau phosphorylation and aggregation, and SIRT1 activity can drive exclusion of exon 10 during tau splicing (Yin et al., 2018). This is an alternative mechanism through which SIRT1 affects tau at a transcriptional level and is unique to previously proposed mechanisms that act at a protein level. In summary, these are all potential mechanisms where SIRT1 overexpression could beneficially affect tau aggregation, degradation, and propagation in AD.

In conclusion, this work by Min et al. (2018) contributes to a nascent field investigating whether SIRT1 activation or overexpression could be beneficial in tauopathies. Moving forward, as neuro-degeneration is a key feature of AD, it will be valuable to determine whether tau acetylation impacts neuronal loss and whether SIRT1 overexpression is able to prevent neurotoxicity. Further experiments should also determine which of the mechanisms discussed enables SIRT1 overexpression to prevent tau propaga-

tion. While there are always concerns about off-target effects caused by overexpression of a protein with wide-ranging functions such as SIRT1, it has been previously reported that hippocampal overexpression of SIRT1 via lentiviral transduction had positive or nondeleterious effects on cognition, anxiety, locomotor activity, and memory in 10-month-old nontransgenic and 3xTg-AD mice (Corpas et al., 2017). Together, these data warrant further exploration of SIRT1 overexpression as a therapeutic strategy in AD.

References

Ayers JI, Giasson BI, Borchelt DR (2018) Prionlike spreading in tauopathies. Biol Psychiatry 83:337–346. CrossRef Medline

Cook C, Carlomagno Y, Gendron TF, Dunmore J, Scheffel K, Stetler C, Davis M, Dickson D, Jarpe M, DeTure M, Petrucelli L (2014a) Acetylation of the KXGS motifs in tau is a critical determinant in modulation of tau aggregation and clearance. Hum Mol Genet 23: 104–116. CrossRef Medline

Cook C, Stankowski JN, Carlomagno Y, Stetler C, Petrucelli L (2014b) Acetylation: a new key to unlock tau's role in neurodegeneration. Alzheimers Res Ther 6:29. CrossRef Medline

Corpas R, Revilla S, Ursulet S, Castro-Freire M, Kaliman P, Petegnief V, Giménez-Llort L, Sarkis C, Pallàs M, Sanfeliu C (2017) SIRT1 overexpression in mouse hippocampus induces cognitive enhancement through proteostatic and neurotrophic mechanisms. Mol Neurobiol 54:5604–5619. CrossRef Medline

DeVos SL, Corjuc BT, Oakley DH, Nobuhara CK, Bannon RN, Chase A, Commins C, Gonzalez JA, Dooley PM, Frosch MP, Hyman BT (2018) Synaptic tau seeding precedes tau pathology in human Alzheimer's disease brain. Front Neurosci 12:267. CrossRef Medline

Goedert M, Spillantini MG (2017) Propagation of tau aggregates. Mol Brain 10:18. CrossRef Medline

Golde TE, Petrucelli L, Lewis J (2010) Targeting Aβ and tau in Alzheimer's disease, an early interim report. Exp Neurol 223:252–266. CrossRef Medline

Iqbal K, Liu F, Gong CX (2016) Tau and neurodegenerative disease: the story so far. Nat Rev Neurol 12:15–27. CrossRef Medline

Irwin DJ, Cohen TJ, Grossman M, Arnold SE, Xie SX, Lee VM, Trojanowski JQ (2012) Acetylated tau, a novel pathological signature in Alzheimer's disease and other tauopathies. Brain 135:807–818. CrossRef Medline

Jęśko H, Wencel P, Strosznajder RP, Strosznajder JB (2017) Sirtuins and their roles in brain aging and neurodegenerative disorders. Neurochem Res 42:876–890. CrossRef Medline

Julien C, Tremblay C, Emond V, Lebbadi M, Salem N Jr, Bennett DA, Calon F (2009) Sirtuin 1 reduction parallels the accumulation of tau in Alzheimer disease. J Neuropathol Exp Neurol 68:48–58. CrossRef Medline

Menzies FM, Fleming A, Caricasole A, Bento CF, Andrews SP, Ashkenazi A, Füllgrabe J, Jackson A, Jimenez Sanchez M, Karabiyik C, Licitra F,

- Lopez Ramirez A, Pavel M, Puri C, Renna M, Ricketts T, Schlotawa L, Vicinanza M, Won H, Zhu Y, et al. (2017) Autophagy and neuro-degeneration: pathogenic mechanisms and therapeutic opportunities. Neuron 93:1015–1034. CrossRef Medline
- Min SW, Cho SH, Zhou Y, Schroeder S, Haroutunian V, Seeley WW, Huang EJ, Shen Y, Masliah E, Mukherjee C, Meyers D, Cole PA, Ott M, Gan L (2010) Acetylation of tau inhibits its degradation and contributes to tauopathy. Neuron 67:953–966. CrossRef Medline
- Min SW, Chen X, Tracy TE, Li Y, Zhou Y, Wang C, Shirakawa K, Minami SS, Defensor E, Mok SA, Sohn PD, Schilling B, Cong X, Ellerby L, Gibson BW, Johnson J, Krogan N, Shamloo M, Gestwicki J, Masliah E, et al. (2015) Critical role of acetylation in tau-mediated neuro-degeneration and cognitive deficits. Nat Med 21:1154–1162. CrossRef Medline
- Min SW, Sohn PD, Li Y, Devidze N, Johnson JR, Krogan NJ, Masliah E, Mok SA, Gestwicki JE, Gan L (2018) SIRT1 deacetylates tau and reduces pathogenic tau spread in a mouse model of tauopathy. J Neurosci 38:3680–3688. CrossRef Medline
- Ng F, Tang BL (2013) Sirtuins' modulation of autophagy: sirtuins and autophagy. J Cell Physiol 228:2262–2270. CrossRef Medline
- Sperfeld AD, Collatz MB, Baier H, Palmbach M, Storch A, Schwarz J, Tatsch K, Reske S, Joosse M, Heutink P, Ludolph AC (1999) FTDP-17: an early-onset phenotype with parkinsonism and epileptic seizures caused by a novel mutation. Ann Neurol 46:708–715. CrossRef Medline
- Spires-Jones TL, Hyman BT (2014) The intersection of amyloid beta and tau at synapses in Alzheimer's disease. Neuron 82:756–771. CrossRef Medline
- Trzeciakiewicz H, Tseng JH, Wander CM, Mad-

- den V, Tripathy A, Yuan CX, Cohen TJ (2017) A dual pathogenic mechanism links tau acetylation to sporadic tauopathy. Sci Rep 7:44102. CrossRef Medline
- Woerman AL, Patel S, Kazmi SA, Oehler A, Freyman Y, Espiritu L, Cotter R, Castaneda JA, Olson SH, Prusiner SB (2017) Kinetics of human mutant tau prion formation in the brains of 2 transgenic mouse lines. JAMA Neurol 74:1464–1472. CrossRef Medline
- Yin X, Jiang X, Wang J, Qian S, Liu F, Qian W (2018) SIRT1 Deacetylates SC35 and Suppresses Its Function in Tau Exon 10 Inclusion. J Alzheimers Dis 61:561–570. CrossRef Medline
- Yoshiyama Y, Higuchi M, Zhang B, Huang SM, Iwata N, Saido TC, Maeda J, Suhara T, Trojanowski JQ, Lee VM (2007) Synapse loss and microglial activation precede tangles in a P301S tauopathy mouse model. Neuron 53: 337–351. CrossRef Medline