



Oxysterols and apolipoproteins in multiple sclerosis: a 5 year follow-up study^S

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Abstract The purpose of this work was to investigate whether changes in oxysterol and apolipoprotein levels over 5 years are associated with disease course and disability progression in multiple sclerosis (MS). This study included 139 subjects [39 healthy controls (HCs), 61 relapsing-remitting MS (RR-MS) patients, and 39 progressive MS (P-MS) patients]. Oxysterols [24-hydroxycholesterol (24HC), 25-hydroxycholesterol (25HC), 27-hydroxycholesterol (27HC), 7 α -hydroxycholesterol (7 α HC), and 7-ketocholesterol (7KC)] were measured at baseline and 5 years using a novel mass spectrometric method, and apolipoproteins were measured using immunoturbidometric diagnostic kits. Levels of 24HC ($P = 0.004$), 25HC ($P = 0.029$), and 27HC ($P = 0.026$) increased in P-MS patients. 7KC ($P = 0.047$) and 7 α HC ($P = 0.001$) levels decreased in RR-MS patients, and there were no changes in any oxysterols in HCs. In MS patients, ApoC-II (all $P \leq 0.01$) and ApoE (all $P \leq 0.01$) changes were positively associated with all oxysterol levels. Increases in 24HC ($P = 0.038$) and ApoB ($P = 0.038$) and decreases in 7KC ($P = 0.020$) were observed in RR-MS patients who converted to secondary P-MS (SP-MS) at follow-up and in SP-MS patients compared with RR-MS patients. **OX** Oxysterols and their associations with apolipoproteins differed between MS patients and HCs over 5 years.

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Multiple sclerosis (MS) is a chronic inflammatory demyelinating disease of the CNS that results in blood-brain barrier (BBB) breakdown, inflammation, and neurodegeneration, and causes physical and cognitive disability.

There is extensive evidence suggesting that higher levels of total cholesterol (TC) and LDL cholesterol are associated with increasing disability in MS (1–4). Cholesterol is required for myelin structure and proper functioning of neuronal, vascular, and immune cells in the CNS. Cholesterol and lipoproteins do not cross the BBB, so the brain is dependent on de novo cholesterol synthesis.

Oxysterols are oxygenated cholesterol metabolites that traverse the BBB rapidly (5) and act as signaling mediators between the periphery and the CNS (6) to enable the maintenance of cholesterol homeostasis in the brain. Oxysterols are ligands for LXR (7, 8) and are involved in

Abbreviations: BBB, blood-brain barrier; DMT, disease-modifying therapy; EDSS, Expanded Disability Status Scale; HC, healthy control; 24HC, 24-hydroxycholesterol; 25HC, 25-hydroxycholesterol; 27HC, 27-hydroxycholesterol; 7 α HC, 7 α -hydroxycholesterol; 7KC, 7-ketocholesterol; MS, multiple sclerosis; P-MS, progressive multiple sclerosis; RR-MS, relapsing-remitting multiple sclerosis; SP-MS, secondary progressive multiple sclerosis; TC, total cholesterol.

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regulating the biosynthesis, cellular efflux, and elimination of cholesterol (9–13). We investigated three side-chain oxysterols [24-hydroxycholesterol (24HC), 25-hydroxycholesterol (25HC), and 27-hydroxycholesterol (27HC)] and two B-ring oxysterols [7 α -hydroxycholesterol (7 α HC) and 7-ketocholesterol (7KC)]. 24HC is produced exclusively in the brain (11) and is the primary regulator of cholesterol synthesis and the principal mechanism for cholesterol elimination in the brain (14–16). 25HC is produced in activated macrophages and mediates inflammatory signaling. 7 α HC is the product of the rate-limiting step in the bile acid pathway of cholesterol elimination (17), and it is a surrogate marker for bile acid synthesis and cholesterol excretion (17). 27HC is produced by CYP27A1 in the acidic pathway (18). 7KC results from oxidative stress and can induce apoptosis (19), inflammation in endothelial cells, and neuronal injury in the brain (20). van de Kraats et al. (21) found that MS patients had lower levels of serum 24HC and 27HC compared with healthy controls (HCs), and higher 24HC levels were associated with decreased normalized brain volume measures. Our group also found that 24HC, 27HC, and 7 α HC (all $P < 0.015$) were lower in MS patients compared with HCs, and 7KC was higher in progressive MS (P-MS) compared with relapsing-remitting MS (RR-MS) ($P < 0.001$) (22).

Apolipoproteins contribute to maintaining peripheral cholesterol homeostasis and are surrogates for oxysterol signaling. ApoA-I acts as an acceptor of cholesterol following efflux from cells (23). Serum ApoB is strongly associated with multiple oxysterols, including 24HC, 25HC, 27HC, 7 α HC, and 7KC (22, 24), and is associated with inflammatory markers (25). In this study, we will focus on ApoE and ApoC-II as the primary biomarkers for LXR signaling. ApoC-II shuttles between lipoproteins and is important for the removal of cholesterol from tissues, and its levels are unaffected by inflammation (26). ApoE, which is modulated by 24HC, plays an important role in maintaining cholesterol homeostasis in the CNS by accepting cholesterol from astrocytes and shuttling it to neurons (7).

The associations of oxysterol changes with MS disease progression have not been extensively investigated in longitudinal studies. The goals of this study were to investigate oxysterol and apolipoprotein levels in 5 year follow-up samples and assess whether changes in these lipid mediators were associated with MS disease course conversion and disability progression.

METHODS

Study population

Study setting and design. This study included samples and clinical data from a single-center longitudinal prospective study of clinical, genetic, and environmental risk factors in MS at the MS Center of the State University of New York at Buffalo. The study includes patients with MS and clinically isolated syndrome, HCs, and controls with other neurological diseases.

Patients and controls provided blood samples, underwent neurological examination, and responded to a structured question-

naire administered in-person at baseline and at 5 year follow-up visits.

Informed consent. The University at Buffalo Human Subjects Institutional Review Board approved the study protocol, and all participants provided written informed consent. The study was conducted in accordance with the principles of the Declaration of Helsinki.

Inclusion and exclusion criteria. This substudy was limited to HCs and MS patients 18–65 years of age with oxysterol and apolipoprotein measures available at baseline and 5 years. MS disease course at baseline and follow-up were reported by MS specialists based on patients' clinical characteristics and published disease course classification (27).

HCs needed to meet the health-screen requirements and had to have a normal physical and neurological examination. They were recruited from respondents to a local advertisement.

Children younger than 18 years of age, adults over 65 years of age, clinically isolated syndrome, neuromyelitis optica, or other neurological diseases were excluded from this substudy. P-MS consisted of patients with secondary P-MS (SP-MS) or primary P-MS.

A conversion to SP-MS categorical variable was defined to classify MS patients into the three groups: *i*) RR-MS at baseline and remained RR-MS at follow-up (RR-MS); *ii*) RR-MS at baseline and converted to SP-MS by the 5 year follow-up visit (RR-MS to SP-MS); and *iii*) SP-MS at baseline and follow-up (SP-MS).

Oxysterol assays

The plasma total plasma oxysterol assay (inclusive of free and esterified oxysterols) included room temperature saponification, solid phase extraction, and a modification of the LC-mass spectrometry analysis conditions previously described (28).

Sample preparation. Plasma samples were stored at -80°C until use without prior freeze thaw cycles. Baseline and 5 year follow-up samples were grouped together by a technician separate from the analytical laboratory and analysts were blind to case/control and baseline/follow-up status.

In-house quality control materials consisting of -80°C frozen aliquots of pooled human plasma and lipid-stripped human blood plasma spiked with authentic oxysterol standards demonstrated -80°C stability ($<15\%$ deviation from nominal) for 4.5 years.

Samples, matrix calibrators, and quality control materials were prepared as previously described (28). In brief, 200 μl of sample were vortex-mixed with 100 μl of deuterated internal standard mix [150 ng/ml each of vitamin D3 (d3), 22HC (d7), 7 α HC (d7), 7KC (d7)] and 10 μl of 50 mg/dl ethanolic butylated hydroxytoluene and saponified using 875 μl of 0.5 M ethanolic KOH for 3 h at room temperature under argon. The pH was neutralized using 20 μl of 85% phosphoric acid; 1 ml of water was added, the vial centrifuged, and the supernatant was loaded under gravity onto a HyperSep C18 solid phase extraction cartridge (200 mg, 3 ml) that had been conditioned with 1 ml of hexane:isopropanol (50:50 v/v), 1 ml of methanol, and 2 ml of water, sequentially. Polar lipids were washed off the SPE column using 4 ml of methanol:water (75:25 v/v). Nonpolar sterols (including cholesterol and oxysterols) were eluted using 2 ml of hexane:isopropanol (50:50 v/v). The eluate was evaporated under nitrogen, reconstituted in 300 μl of methanol:water (90:10, v/v), and 75 μl injected for LC-mass spectrometry analysis.

LC-mass spectrometry analysis. Oxysterols were analyzed on a Shimadzu Scientific (Columbia, MD) LCMS-2010A mass spectrometer system with atmospheric pressure chemical ionization

interface in positive ion mode. Mobile phase composition was 100% methanol to pump A and methanol:water (50:50 v/v) to pump B, both containing 0.1% formic acid. The chromatographic conditions were optimized from our originally reported method (28). Oxysterols were separated on a Supelcosil LC-18-S, 10 cm × 3.0 mm, 3 μm column (Sigma-Aldrich, St. Louis, MO). Column oven temperature was set at 10°C and flow rate was at 0.75 ml/min. The mobile phase gradient was as follows: 80% A for 10 min, linear increase to 100% A over 5 min and held for 13 min followed by re-equilibration at 80% A for 6 min; total run time was 34 min. Temperature settings for the mass spectrometer were: interface at 400°C, CDL at 230°C, and heat block at 200°C. Nitrogen was used as a nebulizing gas for the ion source at a flow rate of 2.5 l/min. Data were acquired in a time segmented-single ion monitoring manner to achieve maximum sensitivity. The *m/z* ratios used for quantifying oxysterols were: *m/z* 367.30 for 24HC, 25HC, and 7αHC; *m/z* 385.30 for 27-OHC; *m/z* 401.40 for 7KC; *m/z* 374.30 for 22-OHC(d7); *m/z* 374.30 for 7αHC(d7); and *m/z* 408.40 for 7KC(d7). Calibrator standards were included in every run and the oxysterol concentrations (in picograms per milliliter) were calculated using calibrator standards.

Lipids and apolipoproteins

EDTA plasma samples for lipid and apolipoprotein analyses were obtained in the nonfasted state. The methods used for lipid and apolipoprotein analyses were previously published (29). Analyses were blinded to the clinical status of samples.

Immunoturbidometric diagnostic reagent kits, calibrators, and controls (Kamiya Biomedical, Thousand Oaks, CA) were used for the apolipoprotein (ApoA-I, ApoA-II, ApoB, ApoC-II, and ApoE) assays. The coefficient of variation of these assays is <5%.

Data analysis

The SPSS (IBM Inc., Armonk, NY, version 24.0) statistical program was used.

Oxysterol concentrations were log-transformed to reduce skew. Apolipoprotein concentrations were normally distributed and not log-transformed.

Paired *t*-tests were used to investigate changes in 24HC, 25HC, 27HC, 7αHC, 7KC, ApoA-I, ApoA-II, ApoB, ApoC-II, and ApoE levels from baseline to follow-up in RR-MS, P-MS, and HC groups alone. Additionally, we used repeated measures analyses to investigate associations between change in oxysterol or apolipoprotein level over 5 years with disease course at baseline (either HC vs. MS or RR-MS vs. P-MS) following adjustment for age, gender, and BMI.

The independent samples *t*-test was used to assess differences in baseline and follow-up levels of 24HC, 25HC, 27HC, 7αHC, 7KC, ApoA-I, ApoA-II, ApoB, ApoC-II, and ApoE between HCs and MS patients and between RR-MS and P-MS patients.

Linear regression was used to investigate the associations between change in TC and change in each of the apolipoproteins (ApoA-I, ApoA-II, ApoB, ApoC-II, or ApoE) with the change in each of the oxysterol levels (24HC, 25HC, 27HC, 7αHC, or 7KC) over 5 years. The change in TC or change in the apolipoprotein level of interest was the dependent variable; and age, gender, BMI, and the change in the oxysterol level of interest were treated as covariates.

Linear regression was used to investigate associations between changes in oxysterol and apolipoprotein levels from baseline to 5 years with the conversion to SP-MS variable and SP-MS subgroups variable. In these analyses, the change in oxysterol or apolipoprotein variable of interest from baseline to 5 years was the depen-

dent variable; and the predictor variables included age, gender, baseline BMI, baseline oxysterol, or apolipoprotein of interest and either the conversion to SP-MS variable or the SP-MS subgroups variable.

RESULTS

Demographic and clinical characteristics

Baseline and follow-up clinical and demographic characteristics of the entire study sample and for HC, RR-MS, and P-MS groups are summarized in **Table 1**. The higher average age and median Expanded Disability Status Scale (EDSS) scores observed in P-MS patients are representative of the progressive disease course. The majority of subjects in the HC (55%), RR-MS (53%), and P-MS (54%) groups had a BMI of 25 kg/m³ or more at baseline. At baseline, the percentages of RR and P-MS patients on disease-modifying therapies (DMTs) were 80% and 77%, respectively. At baseline, the percentages of HC, RR-MS, and P-MS patients on statins were 8, 12, and 13%, respectively.

Oxysterol changes at 5 year follow-up

Figure 1 summarizes the changes in oxysterol (24HC, 25HC, 27HC, 7KC, and 7αHC) levels from baseline to 5 years in HC, RR-MS, and P-MS patients.

There were no significant changes in any oxysterol levels between baseline and 5 years in HCs. We compared oxysterol levels in the entire MS patient group to the HC group and found that follow-up 7αHC levels were 14.5 ng/ml lower in MS patients compared with HCs (*P* = 0.012). No other differences were observed between baseline or follow-up levels of oxysterols in the MS patient group compared with HCs.

In RR-MS patients, 7KC levels decreased by 1.2 ng/ml (*P* = 0.047), and 7αHC levels decreased by 15.8 ng/ml (*P* = 0.001) from baseline to 5 years. No significant changes in 24HC, 25HC, or 27HC were observed in the RR-MS patients.

In P-MS patients, 24HC levels increased by 7.9 ng/ml (*P* = 0.004), 25HC levels increased by 1.3 ng/ml (*P* = 0.029), and 27HC levels increased by 33.2 ng/ml (*P* = 0.026) from baseline to 5 years. There were no significant changes in 7KC or 7αHC from baseline to 5 years in the P-MS group.

Follow-up levels of 25HC were 1.4 ng/ml higher in P-MS patients compared with RR-MS patients (*P* = 0.008). Follow-up levels of 7KC were 2.6 ng/ml higher in P-MS patients compared with RR-MS patients (*P* = 0.023). There were no other differences in baseline or follow-up oxysterols between RR-MS and P-MS patients.

In repeated measures analyses that adjusted for age, gender, BMI, and RR-MS versus P-MS diagnosis at baseline, the greater decreases in 7KC and 7αHC over time in RR-MS compared with P-MS patients, and the greater increases in 25HC over time in P-MS patients compared with RR-MS patients remained significant (data not shown).

TABLE 1. Demographic and clinical characteristics by disease course at baseline visit

	HC	RR-MS	P-MS
Sample size, number	39	61	39
Percent female	28 (71.8%)	43 (70.5%)	29 (74.4%)
Age, years			
Baseline	46.4 ± 12.7	44.7 ± 11.1	56.0 ± 6.1
Follow-up	52.2 ± 12.7	50.7 ± 11.2	62.2 ± 6.0
Disease duration, years			
Baseline	—	13.2 ± 8.7	21.3 ± 10.4
Follow-up	—	19.2 ± 8.9	27.3 ± 10.2
BMI, kg/m ^{2a}			
Baseline BMI	27.0 ± 6.1	28.1 ± 6.3	26.0 ± 3.9
Normal/overweight/obese	45%/31%/24%	47%/25%/28%	46%/44%/10%
Follow-up BMI	26.6 ± 6.7	28.8 ± 7.9	26.5 ± 4.7
Normal/overweight/obese	55%/24%/21%	33%/35%/32%	39%/41%/21%
EDSS, median (IQR) ^b			
Baseline	—	2.5 (2.0)	5.0 (3.0)
Follow-up	—	2.5 (1.5)	6.0 (2.8)
TC, mg/dl			
Baseline	207 ± 42	210 ± 39	214 ± 35
Follow-up	235 ± 43	227 ± 38	244 ± 41
DMT at baseline ^c			
Interferon	—	19 (32%)	14 (36%)
Glatiramer acetate	—	13 (22%)	10 (26%)
Natalizumab	—	14 (23%)	2 (5%)
Other	—	1 (2%)	3 (8%)
None	—	12 (20%)	9 (23%)
DMT at follow-up			
Interferon	—	16 (27%)	13 (33%)
Glatiramer acetate	—	18 (30%)	13 (33%)
Natalizumab	—	2 (3%)	1 (3%)
Orals	—	10 (17%)	2 (5%)
Other	—	7 (12%)	2 (5%)
None	—	6 (10%)	8 (21%)
Statins			
Baseline	8%	12%	13%
Follow-up	10%	20%	13%

All continuous variables (age, BMI, disease duration) are mean ± standard deviation.

^aEDSS in median [interquartile range (IQR)].

^bThe BMI category variable was defined according to National Institutes of Health guidelines (https://www.nhlbi.nih.gov/health/educational/lose_wt/BMI/bmi_dis.htm): normal weight (BMI <25 kg/m²), overweight (BMI ≥25 to <30 kg/m²), and obese (BMI ≥30 kg/m²). BMI is missing for one HC and one RR-MS subject at baseline.

^cDMT data for two RR-MS subjects are missing at baseline and follow-up. DMT data for one P-MS subject is missing at baseline. The interferon category includes the products AVONEX[®], REBIF[®], BETASERON[®], EXTAVIA[®], and PLEGRIDY[™], whereas orals include fingolimod, dimethyl fumarate, and teriflunomide.

These results suggest that MS disease course at baseline (RR-MS or P-MS) is associated with changes in oxysterol levels over 5 years.

Apolipoprotein changes at 5-year follow-up

Figure 2 summarizes the changes in apolipoproteins (ApoA-I, ApoA-II, ApoB, ApoC-II, and ApoE) levels from baseline to 5 years in the HCs, RR-MS patients, and P-MS patients.

In HCs, ApoA-I levels increased by 10.9 mg/ml ($P = 0.012$), ApoA-II levels increased by 2.0 mg/ml ($P = 0.041$), ApoB levels increased by 7.1 mg/ml ($P = 0.049$), ApoC-II levels increased by 0.55 mg/ml ($P = 0.001$), and ApoE levels increased by 0.37 mg/ml ($P = 0.044$) over 5 years.

In the RR-MS group, only ApoA-I levels increased by 12.5 mg/ml ($P < 0.001$) from baseline to 5 years. No significant changes from baseline to follow-up were observed for ApoA-II, ApoB, ApoC-II, or ApoE levels in the RR-MS group.

In the P-MS group, ApoC-II levels increased by 0.6 mg/ml ($P = 0.014$), and ApoB levels increased by 6.2 mg/ml ($P = 0.011$) from baseline to 5 years. No significant changes were observed in ApoA-I, ApoA-II, and ApoE levels in the P-MS group.

In repeated measures analyses that corrected for age, gender, BMI, and HC versus MS diagnosis at baseline, we found that the increases in ApoC-II levels from baseline to 5 years in HCs compared with MS patients remained significant (data not shown).

Interdependence of apolipoproteins, cholesterol, and oxysterol changes over 5 years

We investigated the associations, if any, between apolipoprotein, cholesterol, and oxysterol changes, because oxysterols are known to induce ApoE and ApoC-II biosynthesis via transcriptional activation of LXR (30).

In the HC group, greater increases in 24HC (partial correlation $r_p = 0.66$, $P < 0.001$) and 27HC ($r_p = 0.60$, $P = 0.001$) were associated with greater increases in TC,

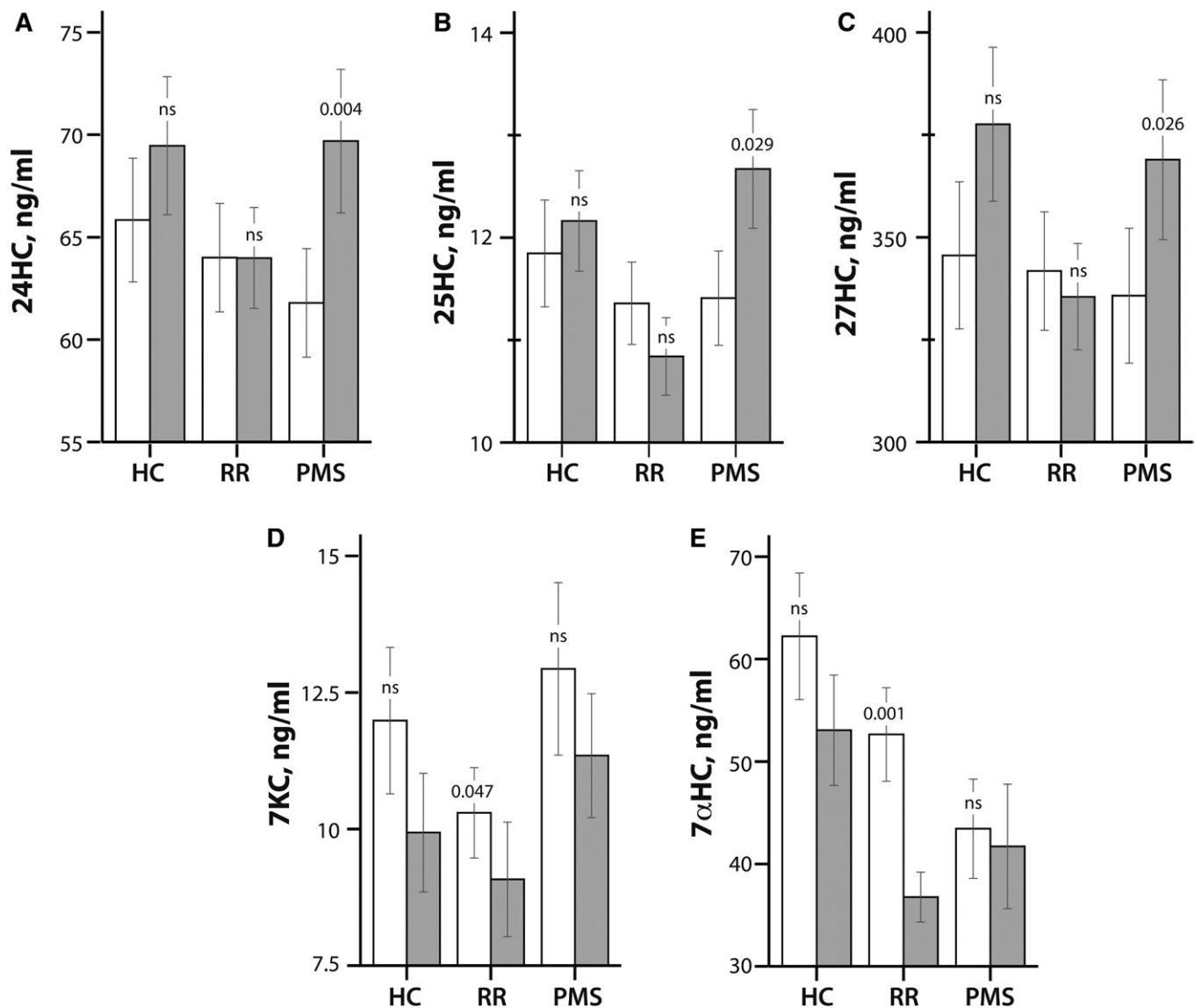


Fig. 1. Oxysterol levels in HCs, RR-MS patients (RR), and P-MS patients (PMS) at baseline (white bars) and 5 years (gray bars). The values of 24HC (A), 25HC (B), 27HC (C), 7KC (D), and 7 α HC (E) are shown in nanograms per milliliter by disease course (HC, RR-MS, and P-MS). The bars compare mean oxysterol values shown on the y axis by disease course shown on the x axis. The error bars indicate the standard error of the mean. The *P*-values from paired *t*-tests are provided for the changes in oxysterol levels from baseline to 5 years within each disease course. ns, not significant.

whereas in the MS group, greater increases in 24HC ($r_p = 0.40$) and 27HC ($r_p = 0.52$) and also 25HC ($r_p = 0.51$), 7 α HC ($r_p = 0.29$), and 7KC ($r_p = 0.31$) levels from baseline to follow-up were associated with increases in TC (all $P \leq 0.011$).

In HCs, changes in 25HC were positively associated with changes in ApoA-I ($P = 0.041$). In MS patients, changes in 7 α HC levels were positively associated with changes in ApoA-I over 5 years ($P = 0.043$). There were no significant associations between changes in any of the oxysterol levels and the changes in ApoA-II in HCs. In MS patients, changes in 24HC ($P = 0.008$), 27HC ($P = 0.005$), and 7 α HC ($P = 0.018$) were positively associated with changes in ApoA-II over 5 years. None of the other oxysterol changes were associated with ApoA-I changes in HCs or with ApoA-I and ApoA-II changes in MS patients.

In HCs, changes in 24HC ($P < 0.001$), 25HC ($P = 0.002$), and 27HC ($P < 0.001$) levels were positively associated with increases in ApoB over 5 years. ApoB changes were not associated with 7 α HC or 7KC changes in HCs. In MS patients, changes in 24HC, 25HC, 27HC, 7 α HC, and 7KC levels were positively associated with ApoB (all $P \leq 0.008$).

In HCs, only changes in 27HC levels were positively associated with changes in ApoC-II ($P = 0.009$). Changes in the other oxysterols were not associated with ApoC-II changes over time in HCs. There were no significant associations between changes in any oxysterol levels and the changes in ApoE in HCs. In MS patients, changes in 24HC, 25HC, 27HC, 7 α HC, and 7KC levels were all positively associated with ApoC-II (all $P \leq 0.01$) and ApoE (all $P \leq 0.01$) changes over the 5 year study period.

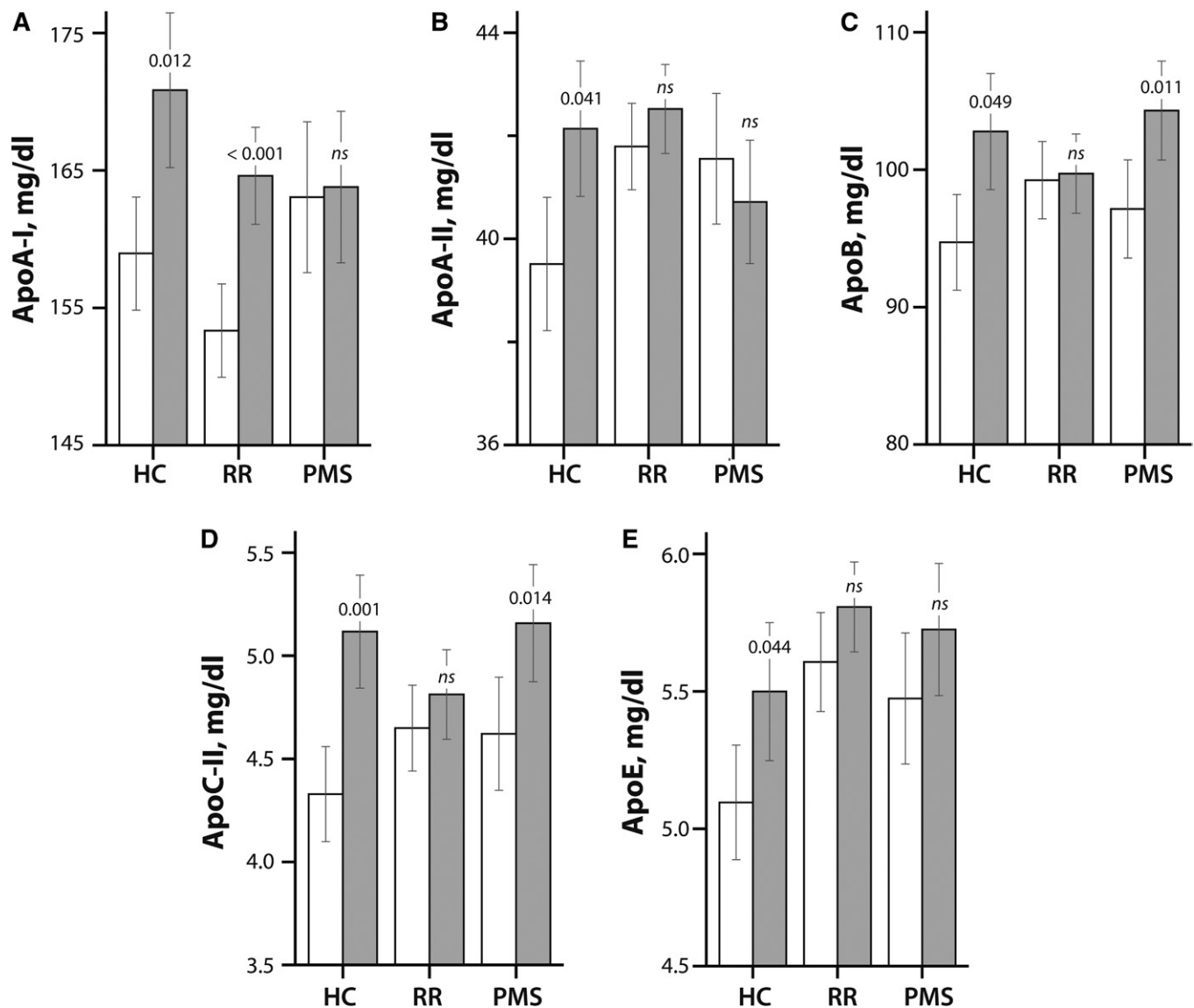


Fig. 2. Apolipoprotein levels in HCs, RR-MS patients (RR), and P-MS patients (PMS) at baseline (white bars) and at 5 year follow-up (gray bars). The values for ApoA-I (A), ApoA-II (B), ApoB (C), ApoC-II (D), and ApoE (E) are shown in milligrams per deciliter. The bars compare mean apolipoprotein values on the y axis by disease course shown on the x axis. The error bars indicate the standard error of the mean. The *P*-values are provided from paired *t*-tests assessing the differences between baseline and 5 year apolipoprotein levels within each disease course.

These results indicate that increases in ApoC-II and ApoE are associated with increases in oxysterols in MS patients, but not in HCs.

Oxysterol and apolipoprotein changes in RR-MS patients converting to SP-MS

Next, we investigated whether changes in oxysterol and apolipoprotein levels from baseline to 5 years differed in the subset of patients who converted from RR-MS at baseline to SP-MS at 5 years. There were 52 patients with RR-MS at baseline and 5 years, 31 patients with SP-MS at baseline and 5 years, and 9 patients with RR-MS at baseline that progressed to SP-MS at 5 years.

The RR-MS patients who converted to SP-MS had a higher average EDSS score at both baseline and follow-up visits and a lower relapse rate compared with RR-MS patients (Fig. 3), which is a clinical profile consistent with

the conversion of the MS disease course from RR-MS to SP-MS.

Greater increases in 24HC ($P = 0.038$, Fig. 4A) and ApoB ($P = 0.038$, Fig. 4D) and greater decreases in 7KC ($P = 0.020$, Fig. 4C) were observed in the RR-MS patients who converted to SP-MS and SP-MS patients compared with the RR-MS patients. However, greater decreases in 25HC ($P = 0.020$, Fig. 4B) were seen in the RR-MS patients, and the RR-MS patients who converted to SP-MS compared with the SP-MS patients. These results suggest that 24HC, 7KC, and ApoB are associated with progression from RR-MS to SP-MS.

DISCUSSION

We measured oxysterol and apolipoprotein levels from baseline to 5 years in MS patients and HCs, and investigated

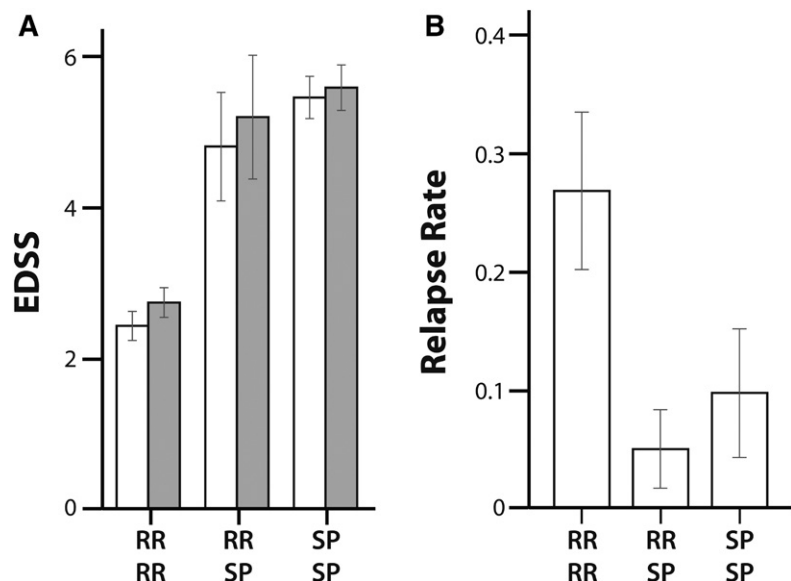


Fig. 3. EDSS score at baseline (white bars) and 5 years (gray bars) (A) and relapse rate over 5 years (B) in patients who were diagnosed with RR-MS at baseline and remained diagnosed with RR-MS at 5 years (RR RR), patients who switched from a RR-MS diagnosis at baseline to SP-MS at 5 years (RR SP), and patients who were diagnosed with SP-MS at baseline and remained SP-MS at 5 years (SP SP). The error bars indicate the standard error of the mean.

whether these changes were associated with disease course conversion and disability progression. 24HC, 25HC, and 27HC levels increased in P-MS patients, whereas 7KC and 7 α HC decreased in RR-MS patients over 5 years. However, no changes in oxysterol levels were observed in HCs. Interestingly, all of the apolipoprotein levels investigated increased in HCs, while ApoB increased and ApoC-II decreased in P-MS patients and only ApoA-I increased in RR-MS patients. Increases in many of the oxysterols measured were associated with increases in ApoB over 5 years in both MS patients and HCs. However, increasing levels of almost every oxysterol were associated with increases in ApoA-II, ApoC-II, and ApoE levels in MS patients but not HCs. Changes in oxysterol and apolipoprotein levels were observed with conversion of RR-MS patients to SP-MS at follow-up.

We compared the oxysterol and apolipoprotein profiles in the patients who progressed from RR-MS at baseline to SP-MS at 5 years with those of the patients who remained RR-MS at 5 years and the patients who remained SP-MS at 5 years. There were nine patients who converted from RR-MS at baseline to SP-MS at follow-up. These patients exhibited EDSS scores and relapse rates qualitatively similar to those of the patients who were diagnosed with SP-MS at baseline and follow-up. Additionally, the changes in 24HC, 7KC, and ApoA-I over 5 years in the nine RR-MS patients who converted to SP-MS by follow-up are similar to the changes seen in the SP-MS patients rather than the RR-MS patients.

A strength of this study was its longitudinal follow-up analysis that included an extensive oxysterol and apolipoprotein panel assayed in paired baseline and follow-up samples. However, the limitations of this study warrant discussion. There were a relatively small number of patients who converted from RR-MS at baseline to SP-MS at follow-up, which limits the statistical power in this study. Furthermore, the diagnosis of SP-MS is subjective and is arrived at retrospectively (31). Our study would also be strengthened with the inclusion of CSF measures of oxysterols and apoli-

poproteins, which would provide additional information on the cross-talk between cholesterol in the CNS and periphery via the oxysterol network and apolipoprotein signaling.

In our previous study (22), we showed how baseline oxysterol levels differed by disease course. This study expands on our previous work by including 5 year follow-up data, which allowed us to investigate long-term changes in oxysterols and apolipoproteins. The results from this study, which are concordant with our previous results, indicate that the RR-MS and P-MS disease courses are associated with changes in different oxysterol levels. RR-MS patients exhibited decreases in sterol B-ring oxysterols (7 α HC and 7KC) in contrast to P-MS patients who exhibited increases in side-chain oxysterols (24HC, 25HC, and 27HC), all ligands of the LXR receptor.

From the analytical standpoint, 7 α HC and 7 β HC are generally considered difficult to resolve chromatographically. We included a deuterated 7 α HC internal standard and further optimized the chromatographic conditions for our previously published assay method (28) to ensure separation of 7 α HC from 7 β HC. While we used -80°C frozen samples that had not been previously thawed, we did not include an antioxidant, such as dibutylhydroxytoluene, to minimize autoxidation during -80°C storage. While BHT addition did not improve bioanalytical stability of oxysterols in a 2 year standardization study (32), it should be considered for future studies.

Oxysterols exert their immunomodulatory effects on immune cells via LXR and LXR-independent EBI2-mediated signaling pathways (33). LXR signaling results in polarization of CD4 helper T cells into Th17 cells in humans (34) and suppresses T cell proliferation (35–37). LXR activation downregulates IL-17 production and Th17 cells, resulting in amelioration of experimental allergic encephalomyelitis, while LXR deficiency leads to exacerbation (34). Activation of LXR also inhibits production of pro-inflammatory cytokines. Finally, LXR agonists can regulate macrophage phagocytosis. Macrophages have been shown to accumulate

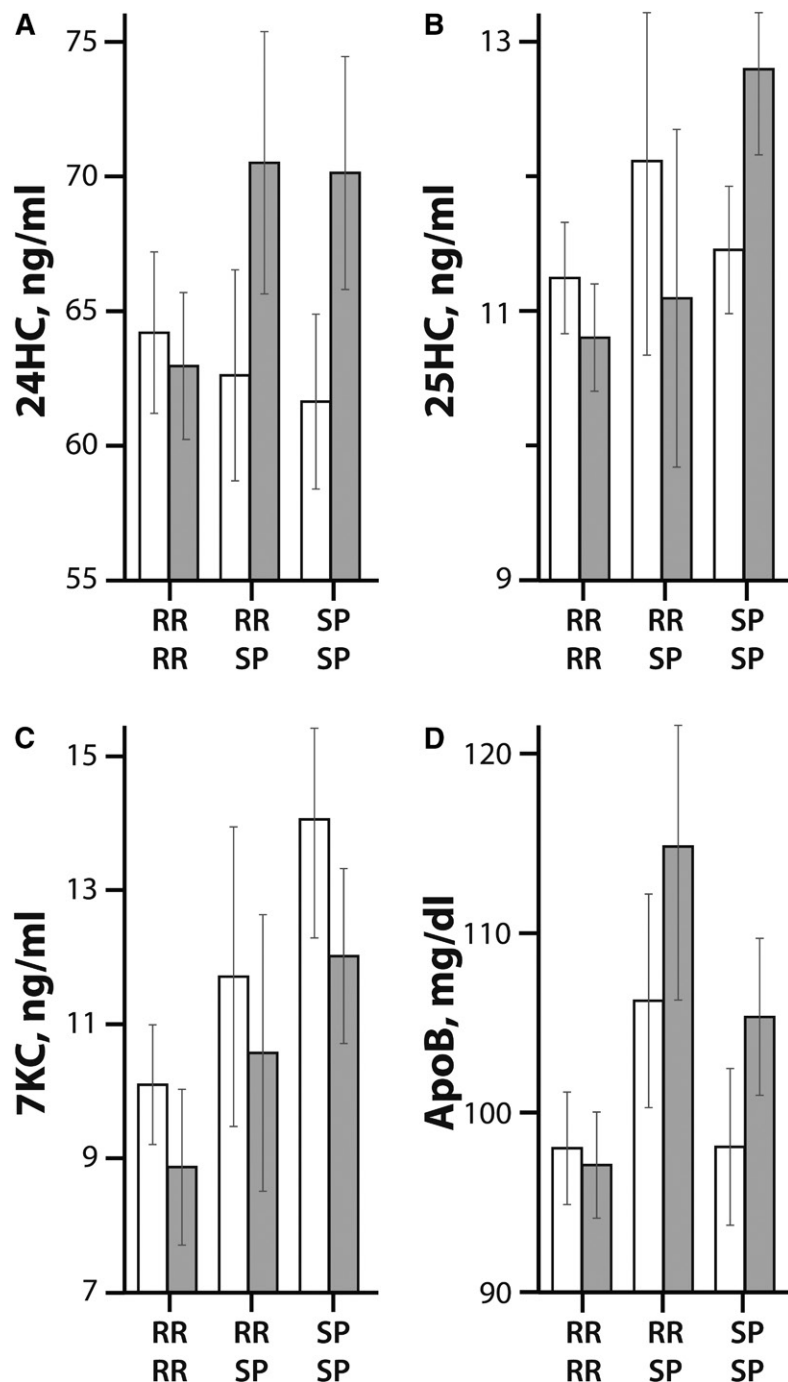


Fig. 4. Oxysterol and apolipoprotein levels at baseline (white bars) and 5 years (gray bars) by the disease course conversion status (RR RR, RR SP, and SP SP; as described in the legend to Fig. 3). The mean values for the oxysterols, 24HC (A), 25HC (B), 7KC (C), and ApoB (D) are shown in milligrams per deciliter by disease course conversion status. The error bars indicate standard error of the mean.

cholesterol from myelin debris, and these myelin-laden macrophages express LXR-induced genes and an increase in the efflux of cholesterol (38).

In conclusion, our results indicate differences in baseline and 5 year oxysterol levels between RR-MS patients, P-MS patients, and HCs. The direction and the extent of change in oxysterol and apolipoprotein levels also differ between the MS disease courses and HCs. The changes in oxysterols are associated with changes in apolipoprotein levels to a greater extent in MS patients than in HCs. Furthermore, there are distinct differences in oxysterol and apolipoprotein profiles in patients progressing to SP-MS. These findings may be useful for developing future studies that aim to delineate the mechanisms by which oxysterols,

as well as other cholesterol mediators, may contribute to MS pathogenesis.

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