

Fusion of the Genes *WWTR1* and *FOSB* in Pseudomyogenic Hemangioendothelioma

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Abstract. *Background/Aim:* Pseudomyogenic hemangioendothelioma is a rare endothelial tumor. Previous genetic investigations have shown that the tumors carry either a *SERPINE1-FOSB* or an *ACTB-FOSB* fusion gene. The aim of the study was to identify *FOSB* fusions linked with pseudomyogenic hemangioendothelioma. *Materials and Methods:* RNA sequencing, reverse transcription polymerase chain reaction (RT-PCR) and Sanger sequencing analyses were performed on a pseudomyogenic hemangioendothelioma. *Results:* An in-frame fusion was found between exon 4 of *WWTR1* from 3q25 and exon 2 of *FOSB* from 19q13. The fusion gene not only places *FOSB* under the control of the *WWTR1* promoter, but is predicted to encode a chimeric *WWTR1-FOSB* transcription factor. *Conclusion:* *FOSB* may be fused with *SERPINE1*, *ACTB*, or *WWTR1* in pseudomyogenic hemangioendotheliomas. The resulting overexpression of *FOSB* fusion is a potentially useful marker that could be helpful in the diagnosis of these tumors.

Pseudomyogenic hemangioendothelioma is a rare endothelial neoplasm that is multifocal in two-thirds of cases and often occurs within a limited region (1, 2). It is more frequently found in young males and primarily in soft tissue, with only 20% presenting with secondary manifestations in bone. About 60% of the lesions occur in the lower limbs. Half of the patients experience pain. Histologically, the tumors are

ill-defined, composed of sheets and cords of spindle cells with abundant, eosinophilic cytoplasm. The nucleus is vesicular, with small nucleoli. There are few mitotic figures and little atypia. Immunohistochemistry (IHC) shows diffuse positive expression of AE1/AE3 and ERG and, in 30% of cases, positive staining for CD31 and smooth muscle actin (SMA). The tumors are negative for Desmin, S100 and CD34 (1, 2). Cytogenetic information on pseudomyogenic hemangioendothelioma is restricted to 4 tumors all of which carried a t(7;19)(q22;q13) chromosomal translocation (3, 4). The translocation resulted in the fusion of the *SERPINE1* gene from 7q22 with *FOSB* gene from 19q13 resulting in overexpression of *FOSB* (4). In two recent studies, fusion of *ACTB* from 7p22 with *FOSB* was described in pseudomyogenic hemangioendotheliomas lacking *SERPINE1* rearrangement (5, 6). Agaram *et al.* (5) have studied 15 pseudomyogenic hemangioendotheliomas and, by using fluorescence *in situ* hybridization and ARCHER FusionPlex analysis, found that seven of the tumors had an *ACTB-FOSB* fusion whereas eight of them had a *SERPINE1-FOSB* fusion. Zhu *et al.* have detected *ACTB-FOSB* fusion in two pseudomyogenic hemangioendotheliomas using target RNA sequencing (6).

We here present another pseudomyogenic hemangioendothelioma that carried none of the above-mentioned fusions, but instead a novel *WWTR1-FOSB* fusion gene.

Materials and Methods

Ethics statement. The study was approved by the regional ethics committee (Regional komité for medisinsk forskningsetikk Sør-Øst, Norge, <http://helseforskning.etikkom.no>). Written informed consent was obtained from the patient for publication of the case details. The ethics committee's approval included a review of the consent procedure. All patient information has been de-identified.

Case description. The patient was a 33-year-old woman who had a history of back pain for over 6 years. A sudden worsening of her symptoms led to radiological examination which revealed a lytic lesion in the upper lateral region of the sacral bone, presumably of

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Key Words: Pseudomyogenic hemangioendothelioma, *WWTR1*, *FOSB*, *WWTR1-FOSB* fusion gene, RNA sequencing.

Table I. Detection of *WWTR1-FOSB* fusion in the RNA sequencing data of pseudomyogenic hemangioendothelioma. The retrieved sequences from the fastq file of the RNA sequencing using the “grep” command and the search term “GAGTGCGCCGGTCTCGGGGA” which is the first 20 nt in the exon 2 of *FOSB* corresponding to nt 719-738 in the *FOSB* reference sequence with the accession number NM_006732.2.

Sequence	Gene
CTACGCCTGTGTGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCCA	<i>FOSB</i>
GTGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGA	<i>FOSB</i>
CCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATCACAACAGCCA	<i>FOSB</i>
CTTTGTTTGTGTGTCTACGCCTGTGTGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGG	<i>FOSB</i>
TGTGTCTACGCCTGTGTGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGT	<i>FOSB</i>
CACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATCACAACAG	<i>FOSB</i>
CATGAGGCAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATCACAACAG	WWTR1-FOSB
GTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATC	<i>FOSB</i>
TGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATCACAAC	<i>FOSB</i>
AGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATCACAACAGCCAGGACCTCC	<i>FOSB</i>
TGGAGAGAGAAAGGATTGCAATGCGCCAAGAGGAGCTCATGAGGCAGGAGTGCGCCGGTCTCGGGGAAATGCCCG	WWTR1-FOSB
GCCTGTGTGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGT	<i>FOSB</i>
CTCCAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATCACAACAGCCA	<i>FOSB</i>
GTGTCTTTGTTGTGTGTCTACGCCTGTGTGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGC	<i>FOSB</i>
GGAGCTCATGAGGCAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATAC	WWTR1-FOSB
TGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGAT	<i>FOSB</i>
CGCCTGTGTGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGG	<i>FOSB</i>
GTCACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATCACAAC	<i>FOSB</i>
TGTCGGTGTCTTTGTTGTGTGTCTACGCCTGTGTGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGA	<i>FOSB</i>
GTGTCCGGTGTCTTTGTTGTGTGTCTACGCCTGTGTGTATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGG	<i>FOSB</i>
TATGTGTACACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATCA	<i>FOSB</i>
AGGATTGCAATGCGCCAAGAGGAGCTCATGAGGCAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTG	WWTR1-FOSB
GTCACCCGTAGGAGTGCGCCGGTCTCGGGGAAATGCCCGGTTCCCTTCGTGCCACGGTACCGCGATCACAAC	<i>FOSB</i>

a benign, fibrous nature, in addition to a prolapse in the lumbar spine. The lesion in the sacrum had presumably brought about the longstanding dumb, continuous pain. Magnetic resonance imaging (MRI) and positron emission tomography (PET) examinations showed also a second lytic lesion in the lumbar spine. Examination of an initial core needle biopsy showed a spindle cell lesion without evidence of atypia leading to a tentative diagnosis of benign fibrous histiocytoma. In a second core needle biopsy, areas with giant cells were observed. Based on the evidence that a variant of giant cell tumor was present, the patient treated with Denosumab. After initial alleviation of symptoms, the pain came back after four injections of Denosumab. A new PET scan showed minimal reduction in signal intensity and no reduction in tumor size. The treatment was discontinued and an open biopsy was performed, which again revealed a spindle cell lesion without atypia (Figure 1A-B). Immunohistochemistry (IHC) showed positivity for SMA, high molecular weight cytokeratins AE1/AE3 (Figure 1C), ERG (Figure 1D), and CD31 (Figure 1E). After consulting Professor Nielsen at Massachusetts General Hospital, the patient was diagnosed with a pseudomyogenic hemangioendothelioma. IHC for *FOSB* was then performed which showed strong nuclear positivity (Figure 1F).

Chromosome banding analysis. Fresh tissue from a representative area of the tumor was received and isolated cells were short-term cultured and analysed cytogenetically, as part of our diagnostic routine, as described elsewhere (7).

RNA sequencing. Total RNA was extracted from frozen (−80°C) tumor tissue using miRNeasy Mini Kit (Qiagen Nordic, Oslo, Norway) and one µg of total RNA was sent to the Genomics Core Facility at the

Norwegian Radium Hospital, Oslo University Hospital (<http://genomics.no/oslo/>) for high-throughput paired-end RNA-sequencing according to the Illumina TruSeq Stranded mRNA protocol. The “grep” command was used to search the fastq files of the sequence data for *FOSB* sequence. The principle of this approach has been described in detail elsewhere (7, 8). The search term was the 20-nucleotide-sequence (nt) “GAGTGCGCCGGTCTCGGGGA” which is the first 20 nt in exon 2 of *FOSB* corresponding to nt 719-738 in the *FOSB* reference sequence with accession number NM_006732.2.

Reverse transcription (RT) and genomic PCR analyses. One µg of total RNA was reverse-transcribed in a 20 µl reaction volume using iScript Advanced cDNA Synthesis Kit for RT-qPCR according to the manufacturer’s instructions (Bio-Rad, Oslo, Norway). One µl of the synthesized cDNA was used as template in subsequent PCR assays. PCR amplifications were performed in a 25 µl reaction volume which contained 12.5 µl Premix Ex Taq™ DNA Polymerase Hot Start Version (Takara Bio Europe/SAS, Saint-Germain-en-Laye, France), template (1 µl cDNA), and 0.4 µM of each of the forward and reverse primers. The forward primer was WWTR1-996F1: TGA GTA TGC CCA ATG CGC TGA CCA corresponding to position 996-1019 in the *WWTR1* reference sequence with accession number NM_015472.4. The reverse primer was FOSB-817R1: TGG GAC TGG GCC ATG GAA GAG ATG corresponding to position 840-817 in the *FOSB* reference sequence with accession number NM_006732.2. PCR amplifications were run on a C-1000 Thermal cycler (Bio-Rad) and the cycling was: an initial denaturation at 94°C for 30 sec, followed by 35 cycles of 7 sec at 98°C and 2 min at 68°C, and a final extension for 5 min at 72°C. Three µl of the PCR

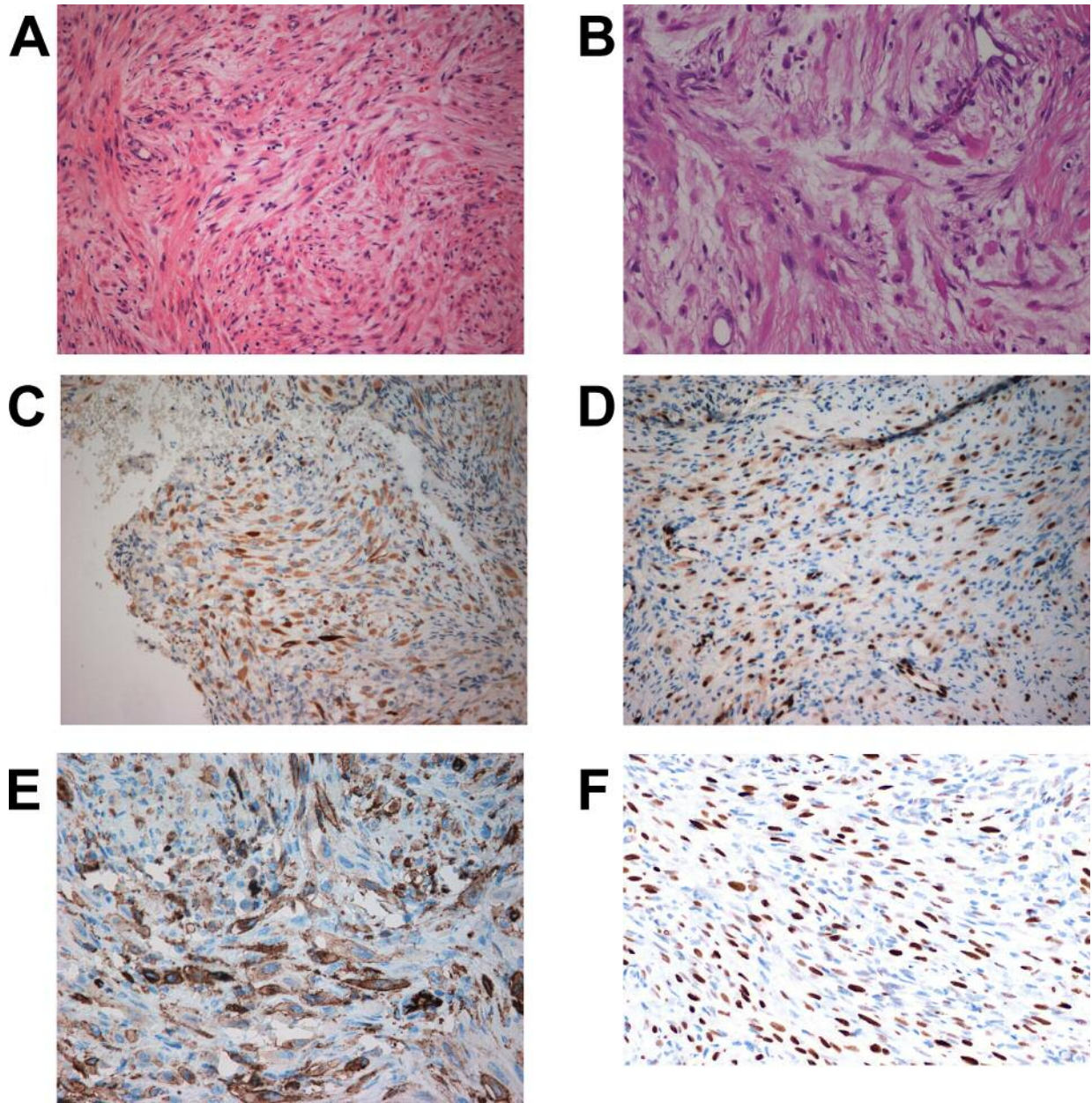


Figure 1. Microscopic examination of the pseudomyogenic hemangioendothelioma. A) H&E-stained section showing a tumor with a spindle cell lesion without atypia, $\times 20$. B) H&E-stained section, $\times 40$. C) Immunorexpression of cytokeratin AE1/AE3, $\times 20$. D) Immunorexpression of ERG, $\times 20$. E) Immunorexpression of CD31, $\times 20$. F) Immunorexpression of FOSB showing strong nuclear positivity, $\times 20$.

products were stained with GelRed (Biotium, VWR International, Oslo, Norway), analyzed by electrophoresis through 1.0 % agarose gel, and photographed. The remaining PCR products were purified using the MinElute PCR Purification Kit (Qiagen) and analysed by direct sequencing using the dideoxy procedure with the BigDye terminator v1.1 cycle sequencing kit (ThermoFisher Scientific, Waltman, MA, USA) on the Applied Biosystems Model 3500 Genetic Analyzer sequencing system. The BLAST software (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) was used for computer analysis of sequence data.

Results

The G-banding analysis revealed a normal karyotype 46, XX in all examined 25 metaphases (data not shown).

Using the “grep” command and a search term corresponding to the first 20 nt in exon 2 of *FOSB* on the raw RNA sequence data, which were in the text-based fastq format, 23 unique sequences were extracted (Table I). Alignment of each of them with the human genome using BLAT on the genome browser

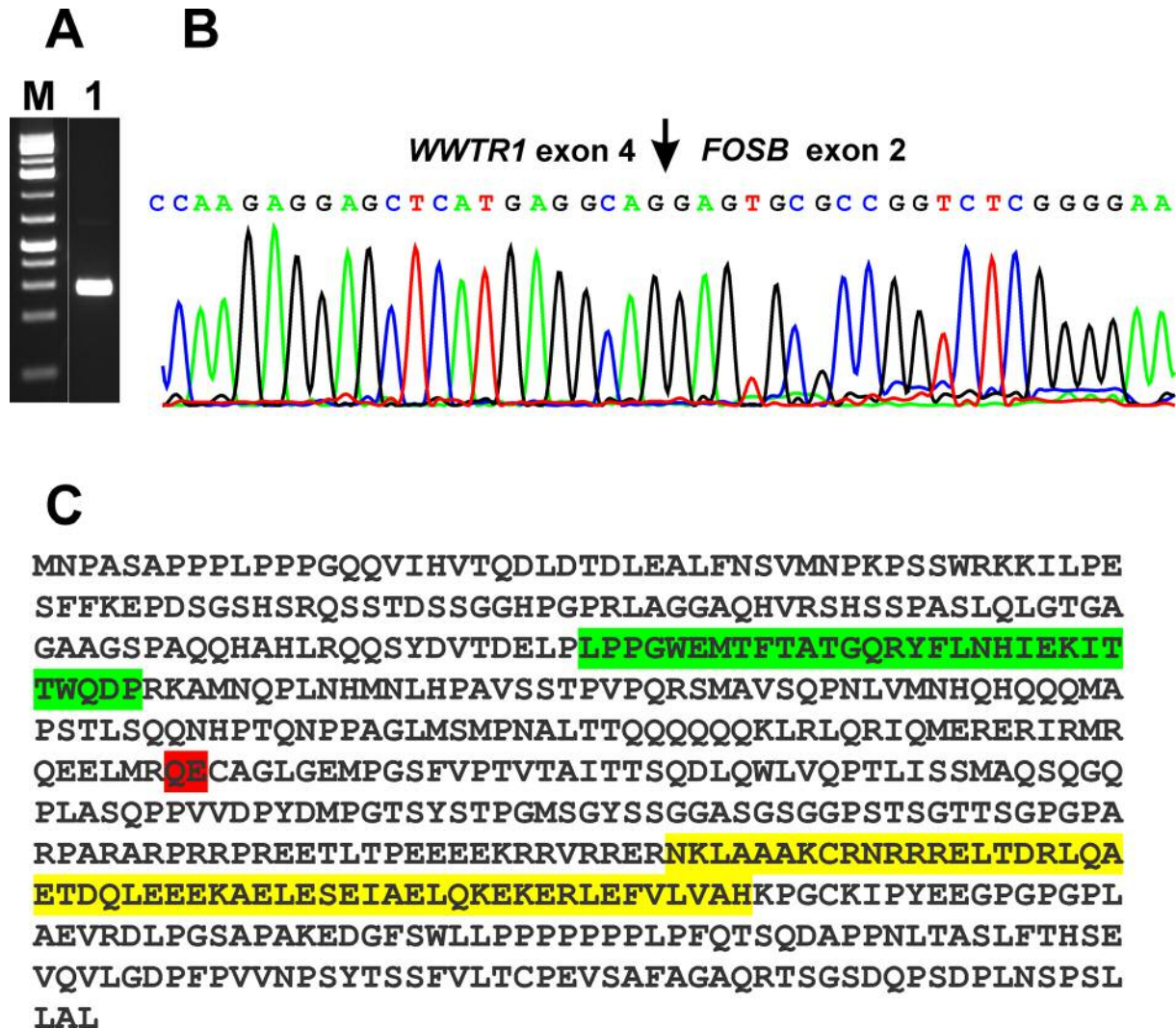


Figure 2. RT-PCR and Sanger sequencing of the PCR products in pseudomyogenic hemangioendothelioma. A) Gel electrophoresis showing the amplified WWTR1-FOSB cDNA fragment. M, GeneRuler 1 Kb DNA ladder (Thermo Scientific). Lane 1, amplification using the forward primer WWTR1-996F1 and the reverse primer FOSB-817R1. B) Partial sequence chromatograms of the cDNA amplified fragment showing the fusion (arrow) of WWTR1 with FOSB. C) The putative WWTR1-FOSB fusion protein. The WW domain from WWTR1 is in green, the bZIP region from FOSB is in yellow, the WWTR1-FOSB junction is in red.

(<https://genome-euro.ucsc.edu/index.html>) and BLAST on the NCBI database (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>) showed that 19 sequences were from FOSB whereas 4 sequences were hybrids containing sequences from exon 4 of WWTR1 (sequence with accession number NM_015472) and exon 2 of FOSB (sequence with accession number NM_006732) (Table I).

RT-PCR with the primer set WWTR1-996F1/FOSB-817R2 amplified a cDNA fragment strongly suggesting the presence of a WWTR1-FOSB fusion transcript in the examined tumor (Figure 2A, lane 1). Sequencing of the

amplified cDNA fragment showed that it consisted of a WWTR1-FOSB chimeric fragment in which exon 4 of WWTR1 (nucleotide 1322 accession number NM_012330 version 3) was fused in-frame to exon 2 of FOSB (Figure 2B). Thus, taking into account the relevant reference sequences NM_015472/NP_056287 for WWTR1 and NM_006732/NP_006723 for FOSB, the fusion WWTR1-FOSB appears to code for a 553 amino acid residue chimeric WWTR1-FOSB protein consisting of the first 257 and the last 296 (position 43-338) amino acid residues of the WWTR1 and FOSB proteins, respectively (Figure 2C).

Discussion

In the present study, a *WWTR1-FOSB* fusion gene was identified in a pseudomyogenic hemangioendothelioma, the third fusion gene to be detected in tumors of this type. One consequence of the fusion is that it places coding regions of the *FOSB* gene under the control of the *WWTR1* promoter. Replacement of the native *FOSB* promoter with the promoter of the fusion partner has been a theme common to all reported *FOSB* fusions described (4-6, 9). For example, in the first reported *SERPINE1-FOSB* fusion gene in pseudomyogenic hemangioendothelioma, the breakpoint in *SERPINE1* was in the non-coding region of exon 1. Therefore, *SERPINE1* provided a strong promoter for the expression of its fusion partner, *FOSB* (4). Similarly, in the *ZFP36-FOSB* fusion gene, exon 1 of *ZFP36* was fused in-frame to exon 2 of *FOSB* and contributed only eight amino acid residues to the *ZFP36-FOSB* chimeric protein (9). Thus, the role of *ZFP36* in the fusion with *FOSB* was to provide a promoter for regulating the expression of *FOSB*.

In the present case, however, the fusion *WWTR1-FOSB* appears to code for a chimeric transcription factor containing the N-terminal 14-3-3 binding region and the WW domain from *WWTR1* (10, 11). It also contains the region required for transformation activity, the basic-leucine zipper (bZIP) domain, and the C-terminal transcriptional activation domain from *FOSB* (10, 11). A *WWTR1-FOSB* fusion gene was also reported previously, in an epithelioid hemangioma (9) where exon 3 of *WWTR1* was fused in-frame with exon 1 of *FOSB*, and the coding chimeric *WWTR1-FOSB* protein contained the above-mentioned functional regions.

The recently described *ACTB-FOSB* fusion gene in pseudomyogenic hemangioendothelioma also codes for a chimeric transcription factor (5, 6). The *ACTB-FOSB* protein contains the nucleotide-binding domain of the sugar kinase/HSP70/actin superfamily from *ACTB* and all the above-mentioned regions of *FOSB* (12). However, there seemed to be no clinicopathological differences between pseudomyogenic hemangioendothelioma carrying an *ACTB-FOSB* fusion and the tumors with *SERPINE1-FOSB*, except that *ACTB-FOSB* positive tumors were often solitary (5).

At the genomic level, *WWTR1* (on chromosome band 3q25) is transcribed in the direction from telomere to centromere whereas the transcription of *FOSB* (on chromosome band 19q13) proceeds in the opposite direction, from centromere to telomere. Hence, formation of a *WWTR1-FOSB* fusion should not be possible from a simple and balanced t(3;19)(q25;q13) translocation. An additional genomic aberration would be required, for example an inversion or an insertion in one of the derivative chromosomes, der(3) and der(19). The orientation of *WWTR1* relative to *FOSB* may also explain the possibly low frequency of *WWTR1-FOSB* fusions. The *SERPINE1-FOSB*

fusion gene is the result of a balanced t(7;19)(q22;q13) chromosomal aberration because *SERPINE1* and *FOSB* are transcribed in the direction from centromere to telomere (3, 4). The *ACTB-FOSB* fusion gene could also be the result of a balanced t(7;19)(p22;q13) since *ACTB* on 7p22 is transcribed from centromere to telomere, although such a translocation has not been reported yet in pseudomyogenic hemangioendotheliomas (5, 6).

The *SERPINE1-FOSB*, *ACTB-FOSB*, and *WWTR1-FOSB* fusion genes and the taking over of *FOSB* expression control by *SERPINE1*, *ACTB*, and *WWTR1* strong promoters makes *FOSB* a useful immunohistochemical diagnostic marker for pseudomyogenic hemangioendotheliomas as it was the case here (5, 13, 14). However, genomic analyses are needed to clarify which gene recombinations occur at which frequencies in these tumors, and also whether as yet unknown variants of the *FOSB*-activation theme exist.

Conflicts of Interest

The Authors declare that they have no potential conflicts of interest exist.

Authors' Contributions

IP designed the research, performed the molecular genetic analyses, interpreted the data, and wrote the manuscript. IL did the pathological evaluations. LG performed cytogenetic experiments and interpreted the data. SH evaluated the cytogenetics wrote the manuscript.

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References

- 1 Hornick JL and Fletcher CD: Pseudomyogenic hemangioendothelioma: a distinctive, often multicentric tumor with indolent behavior. *Am J Surg Pathol* 35: 190-201, 2011. PMID: 21263239. DOI: 10.1097/PAS.0b013e3181ff0901
- 2 Al-Qaderi A and Mansour AT: Pseudomyogenic Hemangioendothelioma. *Arch Pathol Lab Med*, 2018. PMID: 30576238. DOI: 10.5858/arpa.2017-0430-RS
- 3 Trombetta D, Magnusson L, von Steyern FV, Hornick JL, Fletcher CD and Mertens F: Translocation t(7;19)(q22;q13)-a recurrent chromosome aberration in pseudomyogenic hemangioendothelioma? *Cancer Genet* 204: 211-215, 2011. PMID: 21536240. DOI: 10.1016/j.cancergen.2011.01.002
- 4 Walther C, Tayebwa J, Lilljebjorn H, Magnusson L, Nilsson J, von Steyern FV, Ora I, Domanski HA, Fioretos T, Nord KH, Fletcher CD and Mertens F: A novel *SERPINE1-FOSB* fusion gene results in transcriptional up-regulation of *FOSB* in pseudomyogenic haemangioendothelioma. *J Pathol* 232: 534-540, 2014. PMID: 24374978. DOI: 10.1002/path.4322

- 5 Agaram NP, Zhang L, Cotzia P and Antonescu CR: Expanding the spectrum of genetic alterations in pseudomyogenic hemangioendothelioma with recurrent Novel *ACTB-FOSB* gene fusions. *Am J Surg Pathol* 42: 1653-1661, 2018. PMID: 30256258. DOI: 10.1097/PAS.0000000000001147
- 6 Zhu G, Benayed R, Ho C, Mullaney K, Sukhadia P, Rios K, Berry R, Rubin BP, Nafa K, Wang L, Klimstra DS, Ladanyi M and Hameed MR: Diagnosis of known sarcoma fusions and novel fusion partners by targeted RNA sequencing with identification of a recurrent *ACTB-FOSB* fusion in pseudomyogenic hemangioendothelioma. *Mod Pathol*, 2018. PMID: 30459475. DOI: 10.1038/s41379-018-0175-7
- 7 Panagopoulos I, Gorunova L, Bjerkehagen B and Heim S: The "grep" command but not FusionMap, FusionFinder or ChimeraScan captures the *CIC-DUX4* fusion gene from whole transcriptome sequencing data on a small round cell tumor with t(4;19)(q35;q13). *PLoS One* 9: e99439, 2014. PMID: 24950227. DOI: 10.1371/journal.pone.0099439
- 8 Panagopoulos I, Gorunova L, Bjerkehagen B and Heim S: Novel *KAT6B-KANSL1* fusion gene identified by RNA sequencing in retroperitoneal leiomyoma with t(10;17)(q22;q21). *PLoS One* 10: e0117010, 2015. PMID: 25621995. DOI: 10.1371/journal.pone.0117010
- 9 Antonescu CR, Chen HW, Zhang L, Sung YS, Panicek D, Agaram NP, Dickson BC, Krausz T and Fletcher CD: *ZFP36-FOSB* fusion defines a subset of epithelioid hemangioma with atypical features. *Genes Chromosomes Cancer* 53: 951-959, 2014. PMID: 25043949. DOI: 10.1002/gcc.22206
- 10 Kanai F, Marignani PA, Sarbassova D, Yagi R, Hall RA, Donowitz M, Hisaminato A, Fujiwara T, Ito Y, Cantley LC and Yaffe MB: TAZ: a novel transcriptional co-activator regulated by interactions with 14-3-3 and PDZ domain proteins. *EMBO J* 19: 6778-6791, 2000. PMID: 11118213. DOI: 10.1093/emboj/19.24.6778
- 11 Wisdom R and Verma IM: Proto-oncogene FosB: the amino terminus encodes a regulatory function required for transformation. *Mol Cell Biol* 13: 2635-2643, 1993. PMID: 8474434.
- 12 Hurley JH: The sugar kinase/heat shock protein 70/actin superfamily: implications of conserved structure for mechanism. *Annu Rev Biophys Biomol Struct* 25: 137-162, 1996. PMID: 8800467. DOI: 10.1146/annurev.bb.25.060196.001033
- 13 Hung YP, Fletcher CD and Hornick JL: FOSB is a Useful Diagnostic Marker for Pseudomyogenic Hemangioendothelioma. *Am J Surg Pathol* 41: 596-606, 2017. PMID: 28009608. DOI: 10.1097/PAS.0000000000000795
- 14 Sugita S, Hirano H, Kikuchi N, Kubo T, Asanuma H, Aoyama T, Emori M and Hasegawa T: Diagnostic utility of FOSB immunohistochemistry in pseudomyogenic hemangioendothelioma and its histological mimics. *Diagn Pathol* 11: 75, 2016. PMID: 27515856. DOI: 10.1186/s13000-016-0530-2

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