

SCIENTIFIC REPORTS

OPEN

MicroRNA signature refine response prediction in CML

Raquel Alves^{1,2,3}, Ana Cristina Gonçalves^{1,2,3}, Joana Jorge^{1,2,3}, Gilberto Marques⁴, Dino Luís⁵, André B. Ribeiro^{1,2,5}, Paulo Freitas-Tavares⁵, Bárbara Oliveiros^{2,6}, António M. Almeida^{7,8} & Ana Bela Sarmiento-Ribeiro^{1,2,3,5}

Received: 11 November 2018

Accepted: 19 June 2019

Published online: 04 July 2019

microRNAs (miRs) dysregulation have emerged as a crucial step in tumorigenesis, being related with cancer development, progression and response to treatment. In chronic myeloid leukaemia (CML), the resistance to tyrosine kinase inhibitors (TKI) is responsible for treatment failure and could be linked to changes in miRs expression. This work aimed to correlate the expression levels of 3 miRs, miR-21, miR-26b and miR-451, with response to TKI treatment in CML patients. miR-451 levels at diagnosis were significantly higher in patients with optimal response after 6 and 12 months of therapy. Conversely, patients without optimal response had highest levels of miR-21. miR-21 and miR-451 appear to be good biomarkers of response, able to predict optimal TKI responders ($p < 0.05$). Using the combined profile of both miRs, we create a predictive model of optimal response after one year of treatment. This study highlights the role of miR-21 and miR-451 expression levels at diagnosis in predicting which patients achieve the optimal response.

MicroRNAs (miRs) are a group of small single-stranded non-coding RNAs (≈ 22 nucleotides of length) that act as epigenetic regulators of specific targets by modulating gene expression via translational repression or mRNA cleavage¹. Although protein inhibition is the expected result of miR function, this is not always an inhibitory process. miRs can indirectly activate the expression of some mRNAs by degradation of their natural inhibitors^{1,2}. These small RNAs regulate around 80% of the transcriptome and play a key role in multiple cellular processes, like proliferation, development, differentiation and apoptosis².

The dysregulation of miRs expression patterns can have numerous implications including the promotion of tumorigenesis³. miRs expression pattern differs according to tissues, cell types and developmental stages, and could act as oncomiRs, promoting processes that favour cancer cells, or as tumour suppressor miRs, regulating the expression of oncogenes^{4,5}. Aberrant expression of miRs has been reported in solid tumours and haematological neoplasia, as chronic myeloid leukaemia (CML)^{6,7}. The presence of *BCR-ABL1* is the distinctive molecular characteristic of CML and the target for treatment with tyrosine kinase inhibitors (TKI)⁸. Resistance to imatinib and other TKIs has been recognised as the major challenge for CML treatment and monitoring, since some of the imatinib-resistant patients had no mutations on *BCR-ABL1* oncogene⁹. Since miRs are potent regulators, they may be implicated in the acquisition of drug resistance because they could regulate not only *BCR-ABL1* but also modulate the expression of genes involved in drug transporter or activation of essential signalling pathways^{6,10}. In CML the most deregulated miRs studied include miR-10a, miR-130b, miR-150 and miR-203, but several others miRs emerge in the complex network of miR regulation^{11,12}.

Overexpression of the oncomiR miR-21 has been associated with the development of neoplasias where it targets many tumour suppressor genes related to proliferation, apoptosis and survival¹³. One example of miR-21 targets is PTEN, a negative regulator of the PI3K/AKT pathway, which is often described as deregulated in cancer. In addition, levels of miR-21 were correlated with acquisition of resistance to multiple drugs, as gemcitabine and TKIs^{13,14}. PTEN is also a target of miR-26b, but the role of this miR in cancer is still controversial, being

¹Laboratory of Oncobiology and Hematology and University Clinic of Hematology/Faculty of Medicine, University of Coimbra (FMUC), Coimbra, Portugal. ²Coimbra Institute for Clinical and Biomedical Research (iCIBR) - Group of Environment Genetics and Oncobiology (CIMAGO), FMUC, Coimbra, Portugal. ³Center for Neuroscience and Cell Biology (CNC.IBILI), University of Coimbra, Coimbra, Portugal. ⁴Clinical Pathology Service, Centro Hospitalar Universitário de Coimbra (CHUC), Coimbra, Portugal. ⁵Clinical Hematology Department, CHUC, Coimbra, Portugal. ⁶Laboratory of Biostatistics and Medical Informatics, Faculty of Medicine, University of Coimbra, Coimbra, Portugal. ⁷Hospital da Luz, Lisbon, Portugal. ⁸CIIS (Centro de Investigação Interdisciplinar em Saúde) Universidade Católica Portuguesa de Lisboa, Lisbon, Portugal. Correspondence and requests for materials should be addressed to A.B.S.-R. (email: absarmiento@fmed.uc.pt)

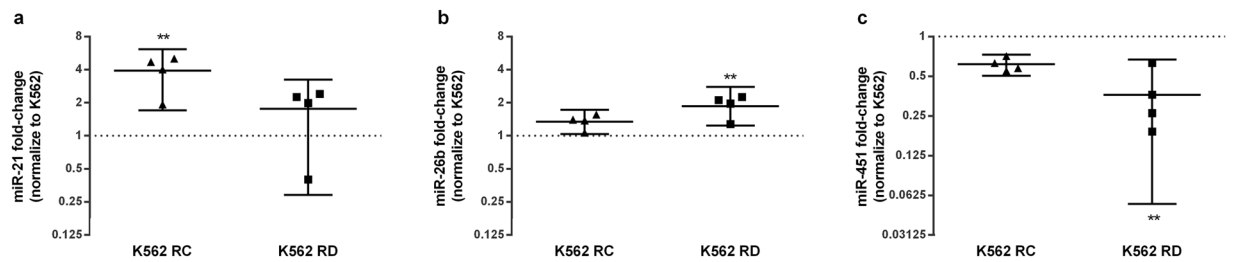


Figure 1. miRs expression profile of imatinib-resistant CML cell lines. Sensitive cell line, K562 cells, was used as reference to determine the fold-change of miR-21 (a), miR-26b (b) and miR-451 (c) of resistant cell lines, K562-RC and K562-RD cells. miR-21 was higher in K562-RC, while miR-26b was higher in K562-RD. Moreover, miR-451 was significantly down-regulated in K562-RD. The results are presented in mean with 95% CI of four independent samples and the expression of K562 cell line is represented by dot line. ** $p < 0.001$ compared with K562 cells.

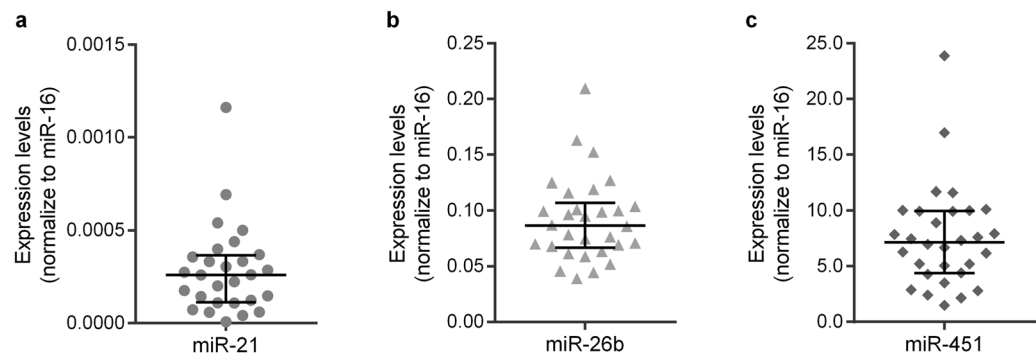


Figure 2. miRs expression levels in CML patients at diagnosis. The expression levels of miR-21 (a), miR-26b (b) and miR-451 (c) were normalised to miR-16 in CML patients at diagnosis time. At diagnosis, miR-451 was the miR with the highest expression and miR-21 with the lowest. The results are represented as median with interquartile range.

sometimes described as oncomiR and others as a tumour suppressor miR^{15,16}. Particularly relevant in CML are the miRs that target *ABL* and consequently *BCR-ABL1*, being some of the most deregulated ones. miR-451, miR-203 and miR-320a are described as downregulated in CML, since they directly target *ABL* and *BCR-ABL1* acting as tumour suppressors, by reducing BCR-ABL oncoprotein expression^{12,17}.

Furthermore, miRs expression is a dynamic process which reflects changes at cellular levels not only at the time of neoplasia development but also in progression or response to treatment. These characteristics made the expression levels of miR a potential biomarker for diagnosis, prognosis and treatment response^{18,19}.

In this work, we analysed the expression levels of miR-21, miR-26b and miR-451 in CML patients and *in vitro* models, and correlated them with TKI response levels. The potential of each miR as a predictive biomarker was explored, with a particular emphasis for the combinatorial miR expression profile.

Results

Differential miRs expression levels at Imatinib-resistant models. To check if the expression levels of miR-21, miR-26b and miR-451 may be related with Imatinib resistance, we evaluated the levels of each miR in three CML cell lines (one sensitive and two resistant to TKI, the K562-RC and the K562-RD cells respectively). The Imatinib-resistant cells presented different expression levels of miRs compared to the sensitive cell line (Fig. 1). The oncomiR miR-21 was up-regulated in both resistant models, with 4.3-fold higher expression in K562-RC cells ($p = 0.0038$; Fig. 1a). Similarly, miR-26b was increased in resistant cells, with 2.0-fold higher expression in K562-RD cells ($p = 0.0023$; Fig. 1b). By opposition, the tumour suppressor miR-451 was down-regulated in K562-RC and K562-RD cells, with 3.8-fold less expression in the discontinuation model (K562-RD) ($p = 0.0048$; Fig. 1c).

CML patients miRs expression levels at diagnosis. We subsequently assessed the expression of the same miRs in CML patients, samples at diagnosis (Fig. 2). Clinical and biological characteristics of our cohort of patients were summarised in Table 1. miR-26b and miR-451 were expressed in all patients, the tumour suppressor miR-451 exhibiting the highest expression at diagnosis [7.147 (95% confidence interval (CI): 5.201–8.912; Fig. 2c)], and the miR-26b the second one with a median of 0.087 (95% CI: 0.070–0.099; Fig. 2b). miR-21 was expressed in 93.3% of patients (28 out of 30 patients) with a median of 0.003 (95% CI: 0.00014–0.00033; Fig. 2a). The expression of this miR was inversely correlated with the expression of miR-451 ($R = -0.655$; $p = 0.0002$).

Characteristics	Diagnosis group (n = 30)		Follow-up group (n = 27)	
Demographic features				
Gender (%)				
Male	16	(53.3)	14	(51.9)
Female	14	(46.7)	13	(48.1)
Clinical features				
Age at diagnosis (years)				
Median	54	42		
Range	18–78	24–78		
Disease Phase				
Chronic Phase (%)	25	(83.3)		
Accelerate Phase or Blast Crisis (%)	5	(16.7)		
Treatment			<i>at analysis time point</i>	
TKI (%)	27	(90.0)	22	(81.5)
Imatinib (%)	26	(96.3)	14	(63.6)
2 nd TKI (%)	1	(3.7)	8	(36.4)
TKI plus INF- α (%)	3	(10.0)	5	(18.5)
Imatinib (%)	2	(66.7)	4	(80.0)
2 nd TKI (%)	1	(33.3)	1	(20.0)
Response			<i>at 6 Months</i>	
Optimal Response (%)	17	(56.7)	24	(88.9)
Without Optimal Response (%)	9	(30.0)	3	(11.1)
No determined (%)	4	(13.3)		
			<i>at 12 Months</i>	
Optimal Response (%)	15	(50.0)		
Without Optimal Response (%)	14	(46.7)		
No determined (%)	1	(3.3)		

Table 1. Biodemographic and clinical characteristics of CML patients. TKI: Tyrosine kinase inhibitors; INF- α : interferon-alpha.

miR-26b expression was positively correlated with miR-21 expression ($R = 0.611$; $p = 0.001$) and inversely associated with miR-451 ($R = -0.462$; $p = 0.010$).

The observed miR expression levels may be related with the disease phase at diagnosis, since miR-451 shown a tendency to be downregulated at accelerated phase or blast crisis [median of 4.28, Interquartile Range (IR) 1.82–8.92, $n = 5$] in comparison with patients in chronic phase (median of 7.31, IR 5.13–10.02, $n = 25$). However, the low number of patients in advanced phases did not allow statistically significant differences to be detected. For that it would be necessary at least 68 patients in each group (G power analysis with 80% of power).

TKI Response and miRs expression levels. To confirm if the miRs expression at diagnosis could be related with the degree of response achieved after TKI treatment, we evaluated the expression profile according to response levels after 6 and 12 months of treatment²⁰. Optimal response was considered when *BCR-ABL1* expression levels were lower than 1% at 6 months and $\leq 0.1\%$ at 12 months of treatment²⁰. Patients with higher levels of miR-451 at diagnosis had a higher rate of optimal response after 6 months ($p = 0.039$; Fig. 3a). Our results reveal that the lower expression of miR-21 ($p = 0.019$) and higher levels of miR-451 ($p = 0.012$) were associated with patients with optimal response at 12 months of treatment (Fig. 3b). At both time points, the levels of miR-26b were higher in patients without optimal response, but the differences were not statistically significant.

To explore the predictive power of miRs expression to predict response to TKI treatment, we performed ROC curve analysis (Fig. 4). miR-451 expression discriminated patients with optimal response from those without response after 6 months of treatment ($p = 0.038$; Fig. 4a). In addition, both miR-451 and miR-21 revealed significant potential as response biomarkers at 12 months of TKI (Fig. 4a). The expression of miR-451 was the most accurate parameter to predict response with an area under the curve (AUC) of 0.771 (95%CI: 0.594–0.949; $p = 0.013$). At diagnosis, miR-451 expression levels higher than 4.74 and 5.70 were the optimal cut-off values to discriminate optimal response after 6 and 12 months of treatment, respectively (Fig. 4b). These cut-off values present higher sensitivity, 94.1% at 6 months and 93.3% at 12 months, with a satisfactory specificity of 66.7% and 64.3% for both time points. Conversely, levels of oncomiR miR-21 below 1.16×10^{-4} at diagnosis was identified as the best cut-off value for optimal response at 12 months of treatment. This biomarker revealed the highest sensitivity (100%) and also a strong positive predictive value (PPV:100%) and a good negative predictive value (NPV: 70%) (Fig. 4b).

miRs profile and response after a year of treatment. Since we identified both miR-21 and miR-451 as good biomarkers of response after a year of treatment with TKI, we made a profile analysis and risk association.

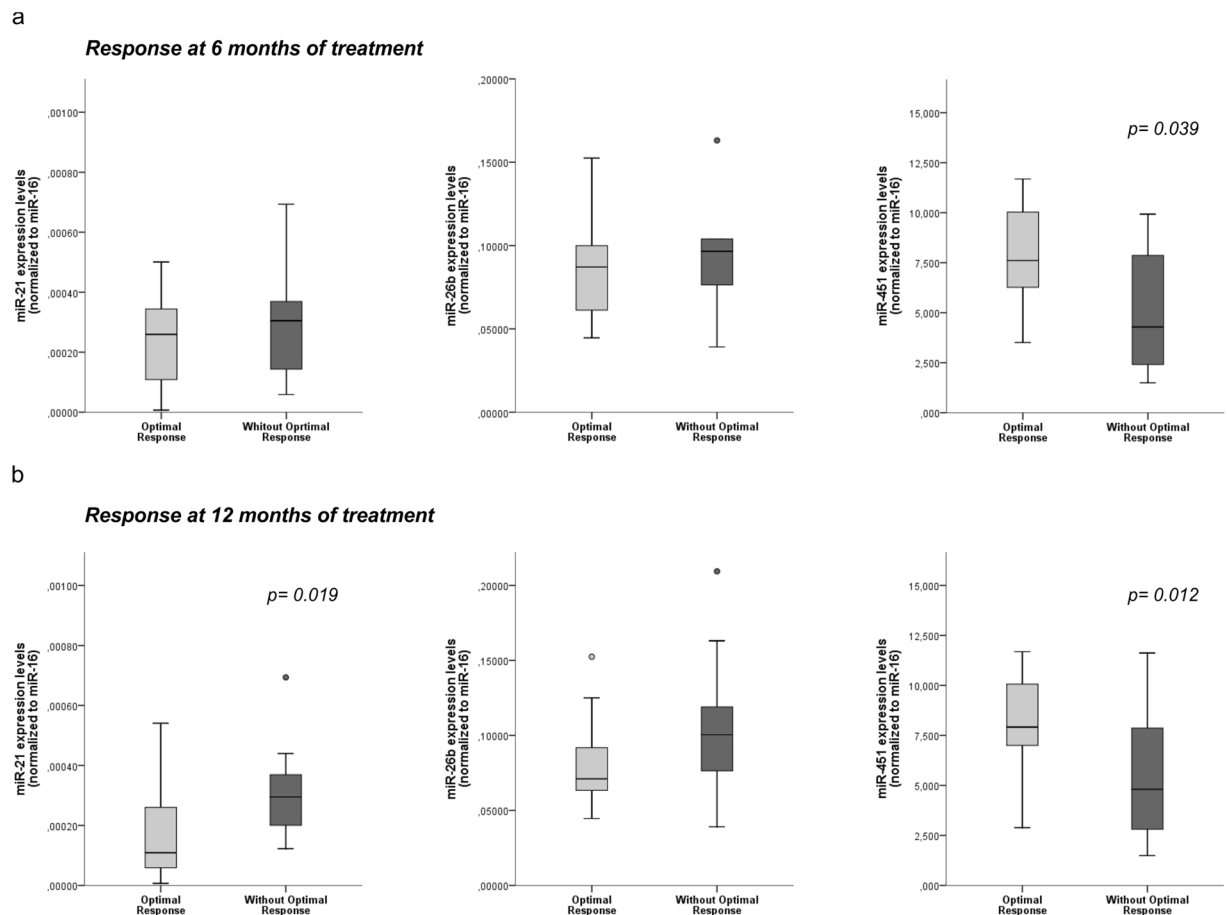


Figure 3. Correlation between TKI response and miRs expression levels at diagnosis. Patients were grouped according to TKI response in optimal response or without optimal response, reached after 6 and 12 months of treatment. The expression levels of miR-21, miR-26b and miR-451 at diagnosis were analysed according to the TKI response after 6 (a) and 12 (b) months of treatment. miR-451 was down-regulated in patients without optimal response after 6 and 12 months of TKI therapy, while miR-21 was up-regulated in patients without optimal response after 12 months of treatment. Statistical differences between groups are reported by the p-value.

Using the optimal cut-off values detected for each miR, we classified patients in four different profiles according to miR-21 and miR-451 expression levels: Profile 1: miR-451 > 5.69 and miR-21 < 1.16×10^{-4} ; Profile 2: miR-451 < 5.69 and miR-21 > 1.16×10^{-4} ; Profile 3: miR-451 > 5.69 and miR-21 > 1.16×10^{-4} ; Profile 4: miR-451 < 5.69 and miR-21 < 1.16×10^{-4} . After, we correlated these miRs profiles with the probability to achieve an optimal response after 12 months of treatment. All patients included in the first profile presented an optimal response (8 optimal responses out of 8), while most of those in profile 2 do not reach optimal response (1 optimal response out of 10). Risk analysis showed that profile 2 patients have 25 times lower probability to achieve optimal response after one year of treatment (OR: 0.040; 95%CI: 0.0039–0.398; $p = 0.001$). On the other hand, patients with profile 1 presented 42 times higher chances to accomplish an optimal response at the same time point (OR: 42.38; 95%CI: 2.129–843.9; $p = 0.0007$). Profile 3 was not statistically associated with response (4 optimal responses out of 9), and no patient presented profile 4.

In this context, miR-21 and miR-451 levels at diagnosis were used to create a predictive model for an optimal response after one year of treatment. From the different regression models tested, the main effect (ME) model that takes in consideration both miRs for optimal response, was the most predictive one accounting for a total of 66% of the variance (Table 2). Applying this model, we will be able to assess the individual probability of each patient to achieve an optimal response (P) Eq. (1).

$$P(\text{OptResp}) = \frac{1}{1 + e^{-(19,6 - 19,6 \times \text{miR}21 - 2,2 \times \text{miR}451)}} \quad (1)$$

In Eq. (1), miR21 and miR451 represent the miR expression in a dichotomized form (miR-21 $\leq 1.16 \times 10^{-4} = 0$ and miR-21 > $1.16 \times 10^{-4} = 1$; miR-451 > 5.7 = 0 and miR-451 $\leq 5.7 = 1$).

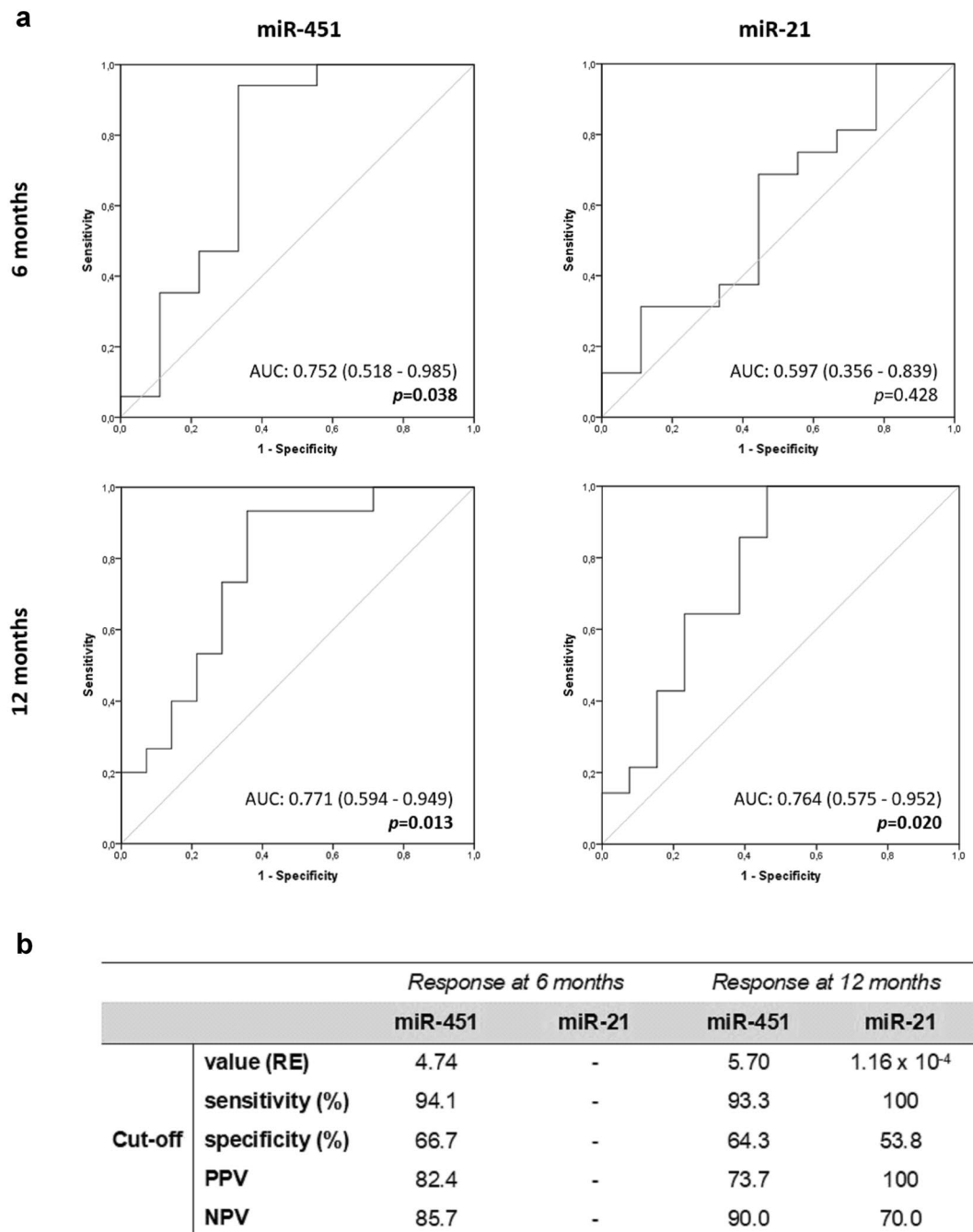


Figure 4. Receiver Operating Characteristic (ROC) curve analysis of miR-451 and miR-21. ROC curves (a) and the identification of optimal cut-off values to discriminate optimal response (b) point-out the ability of miR-451 and miR-21 expression at diagnosis to predict response to TKI treatment at 6 months and 12 months. For each curve was represent the Area Under the Curve (AUC) and the respective p-value. RE-Relative Expression; PPV- Positive Predictive Value; NPV- Negative Predictive Value.

miRs expression at follow-up. Due to lack of samples, it was not possible to determine the miRs expression at diagnosis in the follow-up group. To determine the fold-change of this group, we used the median of each miR observed in the diagnosis group as a reference, since the values did not present a normal distribution. miR-21 was up-regulated 15.0 times during the treatment, while miR-26b and miR-451 were down-regulated 2.4 and 9.2 times, respectively. Correlating the expression levels with clinical and laboratory data, we observed higher levels of miR-21 ($p = 0.008$; Fig. 5a) and miR-26b ($p = 0.008$; Figure 5b) in patients that fail 1st line of treatment and switch for other TKI. Additionally, miR-26b levels were significantly downregulated in patients with lower *BCR-ABL1* quantification ($p = 0.041$; Fig. 5c). Furthermore, we observed that patients with a deeper molecular response (MR) presented lower levels of miR-26b than those with MR of 3.0 (MR 4.0 $p = 0.032$. and MR 4.5 $p = 0.041$) (Fig. 5d).

Model	Variables dichotomize			
	miR-21	miR-451	Interaction	Main Effect
Raw Agreement (%)	79.31	79.31	79.31	79.31
kappa (Cohen)	0.59	0.58	0.58	0.58
Sensitivity (%)	100.00	64.29	64.29	64.29
Specificity (%)	60.00	93.33	93.33	93.33
PPV (%)	70.00	90.00	90.00	90.00
NPV (%)	100.00	73.68	73.68	73.68
accuracy (Youden)	60.00	57.62	57.62	57.62
R ² (Nagelkerke)	0.56	0.45	0.45	0.66
AIC	28.44	32.40	32.40	26.36

Table 2. Statistical measures for predicting models for optimal response. PPV: Positive predictive value; NPV: Negative predictive value; AIC: Akaike information criterion.

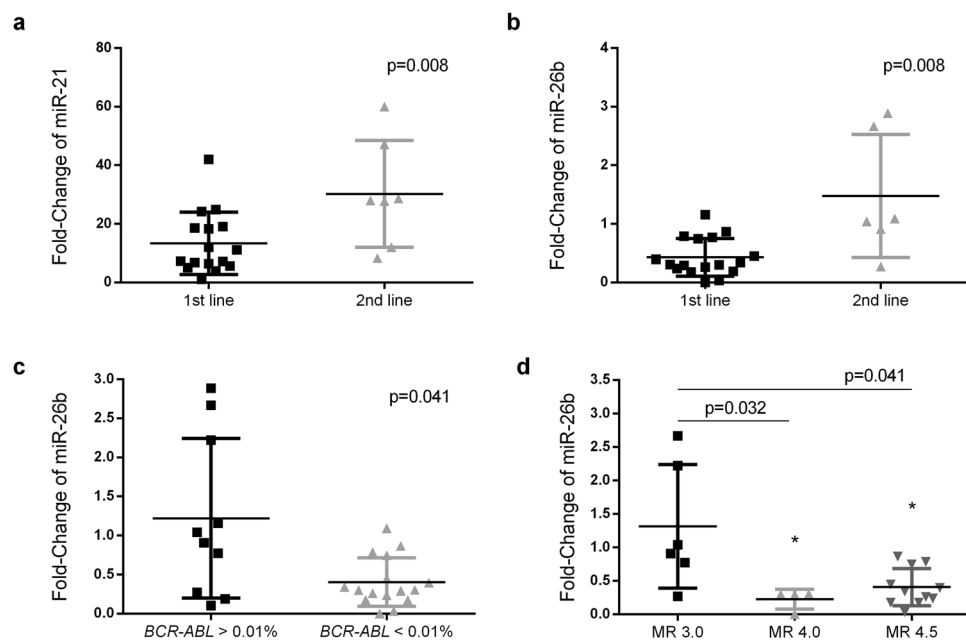


Figure 5. Analysis of miR-21 and miR-26b fold-change in patients under TKI treatment. miR-21 (a) and miR-26b (b) fold-change were analysed according to the number of TKI that patients were exposed. The miR-26 expression levels were also studied in patients stratified according to *BCR-ABL1* values (c) and to molecular response rates (d). Higher levels of miR-21 and miR-26b were detected in patients that fail 1st line of treatment, whereas miR-26b were down-regulated in patients with lower *BCR-ABL1* levels and also in patients with deeper responses to treatment (MR 4.0 and MR 4.5). Statistical differences between groups are reported by the p-value.

Discussion

Most deregulated miRs act on specific signalling pathways that seem to be essential to promote and maintain cancer cell survival and growth⁴. This critical regulatory role, led to associations between miRs expression and response to anticancer drugs proposed by several authors¹⁰. Our results from *in vitro* models highlighted the relation of miR-21 and miR-26b overexpression and miR-451 downregulation with imatinib-resistant phenotype. As expected, an unbalance between oncomiRs and tumour suppressor miRs were observed in the resistant models. In line with *in vitro* models results, we found a correlation between miR-21 and miR-451 expression at diagnosis with the response to TKI treatment. In our cohort, we observed that the optimal response to TKI after 6 months was related to high levels of miR-451. After 12 months of treatment, patients with an optimal response presented not only high levels of miR-451 but also low levels of miR-21, compared to those without optimal response.

miR-451 acts as a tumour suppressor and has been reported to be downregulated in some neoplasias, such as gastric cancer, glioblastoma and leukaemia^{21–23}. Particularly in CML, the expression of miR-451 was associated with a favourable prognosis, since this miR targets *ABL* and *BCR-ABL1* directly¹². According to Lopotová *et al.*²⁴, the miR-451 expression is downregulated in CML at diagnosis compared with healthy controls, and this could be due to *BCR-ABL* activity²⁵. The same authors suggested a reciprocal regulatory loop between this miR and *BCR-ABL1* levels that could be interrupted by imatinib treatment²⁴. This interaction could have a critical impact on response level, and levels of miR-451 constitute an excellent marker to predict response to TKI. In our results,

miR-451 levels were able to predict patients that will achieve response at an early point (6 months) and after one year of treatment, with very good sensitivity and specificity. In the same line, Su *et al.* (2015) suggested miR-451 as a potential prognostic biomarker in gastric cancer²³. The introduction of new biomarkers, as could be the case of miR-451, will improve patient monitoring and could have an impact on therapeutic selection.

With opposite functions, miR-21 is one of the most overexpressed miRNAs in solid and haematological tumours¹³. We observed that low levels of this miRNA at CML diagnosis were related with the achievement of optimal response to TKI therapy. miR-21 presents multiple targets as PTEN, PDCD4 and TIMPs that contribute to the promotion of proliferation, survival and invasion pathways, however according to cell type, the principal mechanism of action may be different^{13,26}. In some cells, it could be related to the up-regulation of BCL-2 to prevent apoptosis, while in others the most significant effect may be the inhibition of PTEN and consequently the activation of PI3K/AKT pathway^{13,27}. In CML, BCR-ABL, with its constitutive tyrosine kinase activity, stimulates multiple signalling pathways such as PI3K/AKT and MAPK/ERK. The downregulation of PTEN and inhibition of PDCD4 has been described as the most crucial action of miR-21 in CML cells^{26,28}. The correlation between miR-21 expression and resistance to anticancer agents, described by us and other groups, highlights the possible clinical application of this miRNA as a target for therapy²⁹. Several groups showed an increase in cell death and a synergistic effect with other drugs when miR-21 was inhibited by antagomiRNAs or some molecules that modulate miR-21, as curcumin^{30–32}. In this work, we did not test miR-21 as a therapeutic target, but we assess the power as a tool for predicting a good response to TKI treatment. Levels of miR-21 measured at diagnosis work as a biomarker of response after one year of TKI therapy. In childhood B cell acute lymphoblastic leukaemia, the same miRNA is an independent prognostic marker, where high levels were associated with poor response and shorter overall survival³³.

Using a single measure to predict prognosis, progression, response and many other aspects seem very simple and sometimes difficult to explain the cell complexity. Based on this, we look for the combinatorial miRNA expression profile since two miRNAs showed biomarker characteristics. Based on cut-off values identified for miR-21 and miR-451, we created 4 different profiles to assess the risk/probability to achieve optimal response after 12 months of TKI treatment. Patients with a high level of miR-451 and low levels of the oncomiRNA miR-21 at diagnosis (profile 1) presented 42 times more probability of achieving an optimal response. According to miRNA functions, profile 1 appears as the best combination possible supporting a more controlled environment against cancer cells growth. In opposition, profile 2, which comprehends higher oncomiRNA expression, leads to a very low probability of these patients reaching optimal response.

By identifying patients at high risk of relapse or progression under treatment, we can prevent several complications and disease progression. Improving the disease monitoring or introducing changes in therapeutic options for these subgroups of patients could be the best approach to achieve better results. These predictive models might play an important role in identifying subgroups of patients based on clinical and biological features. Sokal, EURO, EUTOS and ELTS are scores used in CML for discriminating overall survival (Sokal and EURO), predicting complete cytogenetic remission (CCgR) 18 months after the start of therapy, and predicting the probability of CML-related death, respectively^{34–37}. These scores are based on age, spleen size and haematological parameters at diagnosis, such as basophils, blast cells and platelets. However, none of them were designed to predict optimal response to TKI treatment. Using our miRNA results, we create a predictive model (Eq. 1) to assess the probability of each patient achieving an optimal response to TKI treatment after one year of therapy. The identification of new biomarkers of response/resistance to TKI is in constant progress, and several molecules present good discriminating power, namely specific gene signatures. Nevertheless, independent validation of all new biomarkers and scores would be very helpful to implement in the clinic the most accurate one to predict the optimal response.

In order to increase the number of patients studied and due to the impossibility of obtaining samples at diagnosis from all patients, we use the medians of miRNA expression at diagnosis to estimate the fold-change in miRNA expression induced by TKI treatment. Although not statistically significant, the expression levels of miR-26b at diagnosis were higher in patients that did not reach optimal response with TKI. However, this miRNA presented some relevance at follow-up measures in a group of treated patients. The majority of studies describe miR-26b as a tumour suppressor miRNA that can inhibit cell proliferation in breast cancer, hepatocellular carcinoma and cervical cancers^{16,38,39}. However, some contradictory results have suggested that this miRNA could act as an oncomiRNA. Palumbo *et al.*¹⁵ revealed that the regulation of PTEN in pituitary tumour cells is mediated by miR-26b action¹⁵. miR-26b by targeting PTEN, as also described for miR-21, leaves the PI3K/AKT signalling pathway without regulation, promoting tumorigenesis. In agreement with these authors, our results suggest that lower levels of miR-26b were associated with a good response to the first line of treatment and with more profound responses. These results suggest that miR-26b might be an indicator of good response levels.

The link between miR-21, miR-26b and miR-451 and TKI response levels need to be validated in a bigger cohort of patients before moving into clinical practice. On the other hand, understanding the crucial role of these miRNAs in disease progression and treatment response might be very useful for exploring these miRNAs as a new therapeutic target in CML. In conclusion, expression levels of miR-21 and miR-451 at diagnosis play a crucial role in CML response to TKI treatment and may constitute a new potential biomarker of response and for the guidance of therapeutic options.

Material and Methods

Cell lines. K562 cells sensitive to Imatinib and two Imatinib-resistant models (K562-RC and K562-RD cells) were used as *in vitro* models of CML. The sensitive cell line was obtained from ATCC, and the Imatinib-resistant cell lines were developed in our laboratory based on two strategies: a continuous exposure (K562-RC) and a discontinuous exposure to Imatinib (K562-RD), as described in Alves *et al.*⁴⁰.

Study population. Fifty-seven CML patients were enrolled in the present study and recruited at Clinical Hematology Department of Centro Hospitalar Universitário de Coimbra (CHUC), Portugal. Patients were

grouped according to different time points: at diagnosis (diagnosis group, $n = 30$) and follow-up (follow-up group, $n = 27$). Clinical and biological characteristics were summarised in Table 1. Treatment response criteria were defined according to the European Leukemia Net (ELN) guidelines²⁰, where optimal response corresponds to *BCR-ABL* levels lower than 1% at 6 months and $\leq 0.1\%$ at 12 months. Patients that presented levels higher than the established cut-off were incorporated in “without optimal response” group. The study was conducted according to the Helsinki Declaration, and all participants provided informed consent for participation before enrolment. The Ethics Committee of the Faculty of Medicine (University of Coimbra, Portugal) approved all research procedures (ref. CE-014/2014).

RNA extraction, cDNA synthesis, and real-time PCR. Total RNA was extracted from CML patients' samples and cell lines using the mirVana miRNA Isolation Kit (Ambion), according to the manufacturer's protocol. RNA quantity and quality were measured using a NanoDrop ND-1000 spectrophotometer. TaqMan[®] Advanced miRNA cDNA Synthesis Kit (Applied Biosystems) was used to convert 10 ng of total RNA, according to the manufacturer's protocol. Briefly, this kit included several steps: the Poly(A)-tailing reaction, the ligation reaction, reverse transcriptase reaction followed by a miR-Amp reaction. cDNA was diluted to 1:10 with DNase/RNase free water before use. The levels of miR-21, miR-451, miR-26b and miR-16 were determined in each sample. The quantitative real-time polymerase chain reaction (qPCR) was performed using TaqMan Advanced miRNA Assays (Applied Biosystems) in a QuantStudio 3 system (Applied Biosystems). For each miR, we used one TaqMan Advanced miRNA Assays with specific probes and primers. miR-16 exhibited the most constant expression levels between the samples and was identified as endogenous control by relative quantification software (Thermo Fisher Scientific). Based on this and supported with statistical analysis (Supplementary Figure S1) and miR-16 was used as endogenous control. Relative expression levels of the studied miRNAs were calculated using the relative quantification $2^{-\Delta CT}$ method and normalised with miR-16 ($\Delta CT = CT$ of target $- CT$ of miR-16). The fold change was calculated by $2^{-\Delta\Delta CT}$ method using K562 as a reference for cell lines and the expression at diagnosis for patients follow-up.

Statistical analysis. The statistical analyses were performed using IBM.SPSS[®], version 22, except when otherwise indicated. Normal distribution was tested and the following analyses were performed according to these results. Differences between two groups were assessed by nonparametric Mann-Whitney U test and for three or more groups by nonparametric Kruskal-Wallis test followed by Dunn's multiple comparison tests. The receiver operating characteristic (ROC) curves analysis was performed to evaluate the variables accuracy as biomarkers. For each ROC curve, an optimal cut-off point was determined by the maximum Youden's J Index, which corresponds to the value of the parameter that maximised the sum of specificity and sensitivity. The positive predictive value (PPV) and negative predictive value (NPV) were calculated for each identified biomarker. The association between profiles and the optimal response was analysed by calculating the odds ratio (OR) and its 95% confidence interval (CI), using Fisher's exact test with GraphPad Prism version 6.0. G power analysis was performed Gpower software (version 3.1.9.2). The formulas for predicting an optimal response using both miR-21 and miR-451 expression levels were achieved using R software and its general linear model (glm) function with binomial family and logit link, either using continuous or binary miR profile, considering the single effect, main effect, interaction or full factorial models. The Akaike information criterion (AIC's) minimal value identified the best model. All the statistical tests were considered significant when $p < 0.05$.

Data Availability

All data generated or analysed during this study are included in this published article.

References

- Esquela-Kerscher, A. & Slack, F. J. Oncomirs — microRNAs with a role in cancer. *Nature Reviews Cancer* **6**, 259, <https://doi.org/10.1038/nrc1840> (2006).
- Alizadeh, S., Azizi, S. G., Soleimani, M., Farshi, Y. & Kashani Khatib, Z. The Role of MicroRNAs in Myeloproliferative Neoplasia. *International Journal of Hematology-Oncology and Stem Cell Research* **10**, 172–185 (2016).
- Negrini, M., Nicoloso, M. S. & Calin, G. A. MicroRNAs and cancer—new paradigms in molecular oncology. *Current Opinion in Cell Biology* **21**, 470–479, <https://doi.org/10.1016/j.ceb.2009.03.002> (2009).
- Ichimura, A., Ruike, Y., Terasawa, K. & Tsujimoto, G. miRNAs and regulation of cell signaling. *The FEBS Journal* **278**, 1610–1618, <https://doi.org/10.1111/j.1742-4658.2011.08087.x> (2011).
- Berindan-Neagoe, I., Monroig, P. D. C., Pasculli, B. & Calin, G. A. MicroRNAome genome: A treasure for cancer diagnosis and therapy. *CA: A Cancer Journal for Clinicians* **64**, 311–336, <https://doi.org/10.3322/caac.21244> (2014).
- Litwińska, Z. & Machaliński, B. miRNAs in chronic myeloid leukemia: small molecules, essential function. *Leukemia & Lymphoma* **58**, 1297–1305, <https://doi.org/10.1080/10428194.2016.1243676> (2017).
- Fernandes, Q. MicroRNA: Defining a new niche in Leukemia. *Blood Reviews* **31**, 129–138, <https://doi.org/10.1016/j.blre.2016.11.003> (2017).
- Apperley, J. F. Chronic myeloid leukaemia. *The Lancet* **385**, 1447–1459, [https://doi.org/10.1016/S0140-6736\(13\)62120-0](https://doi.org/10.1016/S0140-6736(13)62120-0) (2015).
- Volpe, G., Panuzzo, C., Uliciani, S. & Cilloni, D. Imatinib resistance in CML. *Cancer Letters* **274**, 1–9, <https://doi.org/10.1016/j.canlet.2008.06.003> (2009).
- An, X., Sarmiento, C., Tan, T. & Zhu, H. Regulation of multidrug resistance by microRNAs in anti-cancer therapy. *Acta Pharmaceutica Sinica B* **7**, 38–51, <https://doi.org/10.1016/j.apsb.2016.09.002> (2017).
- Gordon, J. E. A., Wong, J. J. L. & Rasko, J. E. J. MicroRNAs in myeloid malignancies. *British Journal of Haematology* **162**, 162–176, <https://doi.org/10.1111/bjh.12364> (2013).
- Yeh, C.-H., Moles, R. & Nicot, C. Clinical significance of microRNAs in chronic and acute human leukemia. *Molecular Cancer* **15**, 37, <https://doi.org/10.1186/s12943-016-0518-2> (2016).
- Feng, Y.-H. & Tsao, C.-J. Emerging role of microRNA-21 in cancer. *Biomedical Reports* **5**, 395–402, <https://doi.org/10.3892/br.2016.747> (2016).
- Bai, H., Xu, R., Cao, Z., Wei, D. & Wang, C. Involvement of miR-21 in resistance to daunorubicin by regulating PTEN expression in the leukaemia K562 cell line. *FEBS Letters* **585**, 402–408, <https://doi.org/10.1016/j.febslet.2010.12.027> (2010).

15. Palumbo, T. *et al.* Functional screen analysis reveals miR-26b and miR-128 as central regulators of pituitary somatotrophic tumor growth through activation of the PTEN–AKT pathway. *Oncogene* **32**, 1651, <https://doi.org/10.1038/onc.2012.190> (2012).
16. Luo, M., Shen, D., Wang, W. & Xian, J. Aberrant expression of microRNA-26b and its prognostic potential in human cervical cancer. *International Journal of Clinical and Experimental Pathology* **8**, 5542–5548 (2015).
17. Xishan, Z., Ziying, L., Jing, D. & Gang, L. MicroRNA-320a acts as a tumor suppressor by targeting BCR/ABL oncogene in chronic myeloid leukemia. *Scientific Reports* **5**, 12460, <https://doi.org/10.1038/srep12460> (2015).
18. Kotagama, K., Chang, Y. & Mangone, M. miRNAs as Biomarkers in Chronic Myelogenous Leukemia. *Drug Development Research* **76**, 278–285, <https://doi.org/10.1002/ddr.21266> (2015).
19. Avigad, S. *et al.* miR expression profiling at diagnosis predicts relapse in pediatric precursor B-cell acute lymphoblastic leukemia. *Genes, Chromosomes and Cancer* **55**, 328–339, <https://doi.org/10.1002/gcc.22334> (2015).
20. Baccarani, M. *et al.* European LeukemiaNet recommendations for the management of chronic myeloid leukemia: 2013. *Blood* **122**, 872–884, <https://doi.org/10.1182/blood-2013-05-501569> (2013).
21. Nan, Y. *et al.* MiRNA-451 plays a role as tumor suppressor in human glioma cells. *Brain Research* **1359**, 14–21, <https://doi.org/10.1016/j.brainres.2010.08.074> (2010).
22. Soltani, I. *et al.* Downregulation of miR-451 in Tunisian chronic myeloid leukemia patients: potential implication in imatinib resistance. *Hematology* **22**, 201–207, <https://doi.org/10.1080/10245332.2016.1252020> (2017).
23. Su, Z., Zhao, J., Rong, Z., Geng, W. & Wang, Z. MiR-451, a potential prognostic biomarker and tumor suppressor for gastric cancer. *International Journal of Clinical and Experimental Pathology* **8**, 9154–9160 (2015).
24. Lopotová, T., Žáčková, M., Klamová, H. & Moravcová, J. MicroRNA-451 in chronic myeloid leukemia: miR-451–BCR–ABL regulatory loop? *Leukemia Research* **35**, 974–977, <https://doi.org/10.1016/j.leukres.2011.03.029> (2011).
25. Scholl, V., Hassan, R. & Zalcberg, I. R. miRNA-451: A putative predictor marker of Imatinib therapy response in chronic myeloid leukemia. *Leukemia Research* **36**, 119–121, <https://doi.org/10.1016/j.leukres.2011.08.023> (2012).
26. Panagal, M. *et al.* MicroRNA21 and the various types of myeloid leukemia. *Cancer Gene Therapy*. <https://doi.org/10.1038/s41417-018-0025-2> (2018).
27. Ferreira, A. F. *et al.* ApoptomiRs expression modulated by BCR–ABL is linked to CML progression and imatinib resistance. *Blood Cells, Molecules, and Diseases* **53**, 47–55, <https://doi.org/10.1016/j.bcmd.2014.02.008> (2014).
28. Chen, J., Xu, T. & Chen, C. The critical roles of miR-21 in anti-cancer effects of curcumin. *Annals of Translational Medicine* **3**, 330, <https://doi.org/10.3978/j.issn.2305-5839.2015.09.20> (2015).
29. Abba, M. L. *et al.* MicroRNAs as novel targets and tools in cancer therapy. *Cancer Letters* **387**, 84–94, <https://doi.org/10.1016/j.canlet.2016.03.043> (2017).
30. Taverna, S. *et al.* Curcumin inhibits *in vitro* and *in vivo* chronic myelogenous leukemia cells growth: a possible role for exosomal disposal of miR-21. *Oncotarget* **6**, 21918–21933, <https://doi.org/10.18632/oncotarget.4204> (2015).
31. Li, Y. *et al.* Anti-miR-21 oligonucleotide sensitizes leukemic K562 cells to arsenic trioxide by inducing apoptosis. *Cancer Science* **101**, 948–954, <https://doi.org/10.1111/j.1349-7006.2010.01489.x> (2010).
32. Wang, W.-Z. *et al.* Targeting miR-21 sensitizes Ph+ ALL Sup-b15 cells to imatinib-induced apoptosis through upregulation of PTEN. *Biochemical and Biophysical Research Communications* **454**, 423–428, <https://doi.org/10.1016/j.bbrc.2014.10.107> (2014).
33. Labib, H. A., Elantouny, N. G., Ibrahim, N. F. & Alnagar, A. A. Upregulation of microRNA-21 is a poor prognostic marker in patients with childhood B cell acute lymphoblastic leukemia. *Hematology* **22**, 392–397, <https://doi.org/10.1080/10245332.2017.1292204> (2017).
34. Sokal, J. E. *et al.* Prognostic discrimination in “good-risk” chronic granulocytic leukemia. *Blood* **63**, 789 (1984).
35. Hasford, J. *et al.* A New Prognostic Score for Survival of Patients With Chronic Myeloid Leukemia Treated With Interferon Alfa Writing Committee for the Collaborative CML Prognostic Factors Project Group. *JNCI: Journal of the National Cancer Institute* **90**, 850–859, <https://doi.org/10.1093/jnci/90.11.850> (1998).
36. Hasford, J. *et al.* Predicting complete cytogenetic response and subsequent progression-free survival in 2060 patients with CML on imatinib treatment: the EUTOS score. *Blood* **118**, 686, <https://doi.org/10.1182/blood-2010-12-319038> (2011).
37. Pfirrmann, M. *et al.* Prognosis of long-term survival considering disease-specific death in patients with chronic myeloid leukemia. *Leukemia* **30**, 48–56, <https://doi.org/10.1038/leu.2015.261> (2016).
38. Jin, F. *et al.* MiR-26 enhances chemosensitivity and promotes apoptosis of hepatocellular carcinoma cells through inhibiting autophagy. *Cell Death & Disease* **8**, e2540, <https://doi.org/10.1038/cddis.2016.461> (2017).
39. Li, J. *et al.* MiRNA-26b inhibits proliferation by targeting PTGS2 in breast cancer. *Cancer Cell International* **13**, 7, <https://doi.org/10.1186/1475-2867-13-7> (2013).
40. Alves, R. *et al.* Drug transporters play a key role in the complex process of Imatinib resistance *in vitro*. *Leukemia Research* **39**, 355–360, <https://doi.org/10.1016/j.leukres.2014.12.008> (2015).

Acknowledgements

R.A. was supported by the Portuguese Foundation for Science and Technology (FCT) with a PhD grant (SFRH/BD/51994/2012). The work was supported by CIMAGO (Project 10/14), by Faculty of Medicine of the University of Coimbra and Santander Totta Bank with the grant (FMUC-BST-2016-214), by funds from FEDER through the Operational Program Competitiveness Factors (COMPETE) and by FCT under the strategic projects from FCT/MCTES/PIDDAC (CNC.IBILI. Centre Reference: UID/NEU/04539/2013).

Author Contributions

R.A., A.M.A. and A.B.S.R. conceived of the study. P.F.T., A.B.R., D.L. and G.M. recruited and provided the clinical information of the participants. R.A., A.C.G. and J.J. conducted the experiments. R.A., A.C.G. and B.O. performed the statistical analysis. R.A., A.C.G. and J.J. wrote the paper. A.M.A. and A.B.S.R. revised the manuscript. All authors read the final version and approved of its submission.

Additional Information

Supplementary information accompanies this paper at <https://doi.org/10.1038/s41598-019-46132-9>.

Competing Interests: The authors declare no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2019