

Review

Nanomodulation of Macrophages in Multiple Sclerosis

Frances K. Nally, Chiara De Santi and Claire E. McCoy *

Molecular and Cellular Therapeutics, Royal College of Surgeons in Ireland, 123 St Stephen's Green, 2 D02 YN77 Dublin, Ireland; francesnally@rcsi.ie (F.K.N.); chiaradesanti@rcsi.ie (C.D.S.)

* Correspondence: clairemccoy@rcsi.ie; Tel.: +353-1-402-5017

Received: 15 May 2019; Accepted: 3 June 2019; Published: 5 June 2019



Abstract: Multiple Sclerosis (MS) is a chronic demyelinating autoimmune disease primarily affecting young adults. Despite an unclear causal factor, symptoms and pathology arise from the infiltration of peripheral immune cells across the blood brain barrier. Accounting for the largest fraction of this infiltrate, macrophages are functionally heterogeneous innate immune cells capable of adopting either a pro or an anti-inflammatory phenotype, a phenomenon dependent upon cytokine milieu in the CNS. This functional plasticity is of key relevance in MS, where the pro-inflammatory state dominates the early stage, instructing demyelination and axonal loss while the later anti-inflammatory state holds a key role in promoting tissue repair and regeneration in later remission. This review highlights a potential therapeutic benefit of modulating macrophage polarisation to harness the anti-inflammatory and reparative state in MS. Here, we outline the role of macrophages in MS and look at the role of current FDA approved therapeutics in macrophage polarisation. Moreover, we explore the potential of particulate carriers as a novel strategy to manipulate polarisation states in macrophages, whilst examining how optimising macrophage uptake via nanoparticle size and functionalisation could offer a novel therapeutic approach for MS.

Keywords: multiple sclerosis; experimental autoimmune encephalomyelitis; macrophage polarisation; monocytes; microglia; inflammation; nanoparticle; microparticle; drug delivery; CNS

1. Introduction

Multiple Sclerosis (MS) is a chronic inflammatory disease of the Central Nervous System (CNS), affecting an estimated 2.3 million people worldwide [1]. On average, disease onset and diagnosis occurs between the ages of 20 and 40, with occurrence 2–3 times higher in females than males, making MS the most common neurologically disabling disease in young adults [2]. Pathologically, the disease is characterised by the appearance of focal lesions in white and grey matter of the brain and spinal cord, indicative of extensive loss of oligodendrocytes and myelin sheath. Owing to the distribution of such lesions, clinical presentation is variable among patients and can include impairments in motor, sensory and cognitive functions, as well as pain and fatigue [3]. MS is a complex disease with incompletely understood aetiology with contribution from genetic predisposition [4], as well as environmental risk factors, including geographical location, Epstein Barr virus infection, human cytomegalovirus, lack of vitamin D and circadian rhythm disruption [5,6]. In 80–85% of newly diagnosed individuals, symptoms occur on a relapsing remitting basis (RRMS), with roughly two thirds of these go on to develop secondary progressive disease (SPMS) [7]. Primary progressive disease (PPMS) occurs in a smaller proportion of individuals [8].

Despite the fact the instigating factor in disease pathogenesis of MS remains elusive, plaque formation and disease symptoms are widely accepted as the result of immune cell infiltration, with the release of cytokines and inflammatory mediators leading to inflammation, myelin destruction,

oligodendrocyte loss and eventual axonal degeneration [9]. While there is a complex immune pathology at play with contributions from many immune cell types, MS has been traditionally viewed as a T-cell mediated disease [10–12]. While initial discussion centred on CD4+ T-cells subsets, the essential role of CD8+ T cells has been more recently discussed [11–13], as well as that of B lymphocytes [14,15]. However, emerging and accumulating evidence has highlighted a role for infiltrating monocytes and macrophages in human MS pathology, as comprehensively discussed in [16] and as further discussed in this review. These cells are the most predominant cell type in patient lesions [17–20], with their presence correlating with both demyelination [21,22] and axonal damage and degeneration [20,22–24].

Experimental autoimmune encephalomyelitis (EAE) is among the most frequently used models to study the demyelinating and immune pathology of MS [25]. EAE can be induced by direct immunization of myelin antigens PLP, MOG or MBP in Complete Freund's Adjuvant, or by the transfer of isolated activated CD4+ T cells [26], or less commonly by CD8+T cells to a naïve animal [27,28], which avoids the immunological consequences of adjuvant administration [29]. While disease course is variable, dependent upon both animal species and strain, and on the inoculating myelin antigen [26], overall pathology in this model is largely driven by CD4+ T helper (Th)1 and Th17 T cell subsets [25,30]. This, coupled with the fact that demyelination and lesion formation occur predominantly in the spinal cord rather than the CNS, indicates that EAE does not fully recapitulate human MS pathology [25,29,31]. Nonetheless, the utility of the EAE model is exemplified in the discovery and development of some of the current approved therapies, namely Glatiramer Acetate [32], Natalizumab [33], Fingolimod [34] and has been used to further elucidate the mechanism of action of others, including Alemtuzumab and Dimethyl Fumarate [35]. Moreover, the use of toxin induced models such as lysolecithin, cuprizone and ethidium bromide (EtBr) are seeing increased popularity, enabling different aspects of MS disease pathology to be addressed [35]. While there is no model available that can recapitulate the entirety of the molecular and cellular events involved in MS, the use of animal models has been invaluable to our current knowledge of mechanisms at play in MS, including understanding the role of macrophages in disease pathology, and are essential for use in preclinical models [36].

In this review, we examine this role played by macrophages in MS and MS animal models, and explore the potential for the use of nanotechnology in developing macrophage-centred therapeutics for preclinical efficacy in MS.

2. Macrophages in MS

2.1. Resident vs Infiltrating Macrophages

Macrophages are a highly plastic, highly diverse population of cells, with a multifaceted role in the normal immune response as well as in disease. Macrophages are professional phagocytes, and are the most numerous cells found in CNS lesions in both human MS and EAE models [17,21,37]. In the context of disease, these macrophages are a mixed population in terms of lineage, capable of arising from both CNS resident glial cells and infiltrating monocytes. Belonging to the former, microglia are the resident macrophages in the CNS parenchyma, with an essential role in neurological function and immunosurveillance under homeostatic conditions [38]. CNS resident macrophages are notable among myeloid populations; they arise from a distinct embryonic yolk sac population that enter the CNS prior to blood-brain barrier (BBB) closure and do not undergo replacement by hematopoietic precursors throughout life [39–41]. Non-parenchymal CNS resident macrophages, which include perivascular, meningeal and choroid plexus macrophages, also arise from embryonic populations and, with the exception of those in the choroid plexus, undergo little replacement by blood borne monocytes [42]. Monocytes and macrophages of peripheral origin are normally not present in the parenchyma of the healthy CNS, but are recruited in response to EAE induction and can be found in lesions in both EAE and MS pathology [17,21,37].

There is substantial evidence that the infiltrating monocytes, rather than resident microglia, play a more prominent role in driving disease pathogenesis. For example, microglial activation appears

to be a key early feature, distinct from monocyte entry, and does not ensure disease onset. In a model where reduced sensitivity to pharmacogenetic depletion resulted in microglial paralysis, EAE onset was delayed, with reduced clinical scoring, diminished monocyte infiltration and reduced myelin and axon destruction [43]. In other studies, microglial activation can be identified both prior to peripheral infiltration and appearance of symptoms, as well as in animals that fail to progress beyond the initial stage of EAE [44,45]. This suggests that although microglia play an important role in the initiation of disease in EAE, the continued progression of the disease is largely due to their role in monocyte recruitment. Monocyte depletion prior to symptom development delayed EAE onset and resulted in less severe clinical scoring [46–48] while depletion post-onset showed inhibited disease progression [46,49]. Furthermore, it has been demonstrated that in early disease, rising numbers of peripheral monocytes correlate with the severity of clinical scoring [45,50], with this infiltration correlating with progression to paralysis [45]. Moreover, observations from histological studies of human samples point to a role for microglia at similar early stages of lesion formation, noting their activation in normal appearing white matter prior to peripheral infiltration and myelin destruction [51–54]. The remainder of this review will focus on infiltrating monocytes and how they contribute to disease progression as well as to its resolution. We illustrate how this dual capacity of infiltrating monocytes could be manipulated as a novel therapeutic approach for MS.

2.2. A Dual Role for Macrophages in MS

Monocytes are mobilised by chemokine signalling and traverse the blood brain barrier in response to the induction of EAE, with the chemokine receptor CCR2 and ligand CCL2 particularly well explored in this context. A number of studies have demonstrated that the absence of CCR2, or one of its major ligands, CCL2, results in diminished or absent EAE development [55–58]. Crucially, this effect has been shown to be most prominently mediated through CCR2 engagement in the myeloid compartment [59]. CCR2 expression in monocytes occurs in a population primarily defined by high expression of Ly6c in mice that traffic to sites of inflammation [60]. Antibody mediated targeting of the CCR2+Ly6C^{hi} monocytes in the periphery during peak disease resulted in markedly reduced clinical scoring, while had no observable effect during the ‘priming’ phase of disease between EAE induction and onset [59]. In a relapsing model of EAE, this CCR2+Ly6C^{hi} population increase markedly in the blood prior to relapse in a GM-CSF dependent manner [61]. GM-CSF is a cytokine produced by Th subsets, and is essential to disease development [62,63]. Interestingly, a conditional receptor deletion strategy by Croxford and colleagues has shown the CCR2+ monocyte population to be the crucial facet of GM-CSF mediated pathogenesis in EAE [64].

While monocyte populations are somewhat differently defined in humans, consisting of CD14⁺⁺CD16[–] classical monocytes, CD14⁺CD16⁺⁺ non classical monocytes and CD14⁺⁺CD16⁺ intermediate subsets (outlined in [65]), changes to the monocyte populations in MS patients has been described. Increases in the non-classical populations in circulation have been shown in both RRMS and PPMS [66–68]. While this CD14⁺CD16⁺⁺ population is normally low in CCR2 expression [69], its expression has been shown to be significantly upregulated in this population of monocytes from the PBMC's of MS patients [68]. CCL2 expression has also been noted in both MS plaques and in the CSF of patients with ongoing disease [70–73]. There is ongoing debate as to whether this CCR2 mediated chemotaxis is as important in MS as it is in animal disease models [74–76], nonetheless, the potential for a therapeutic CCR2 targeting antibody is under investigation [77].

Following tissue entry into the spinal cord or CNS parenchyma, monocytes become activated, differentiate, becoming myeloid dendritic cells (DCs) or macrophages. With regard to macrophages, because this activation occurs in response to a variety of signals, there is considerable functional heterogeneity [78]. In EAE, T cell produced GM-CSF, IFN γ and TNF α cause polarisation to a pro-inflammatory phenotype or M1-like phenotype, with cells arising from CCR2+Ly6C^{hi} monocytes expressing high levels of MHCII, IL-6, IL-12p40, iNOS and inflammasome products IL-1 α and IL-1 β [59,61,64] (Figure 1). These macrophages facilitate damage to the CNS in a number of ways,

for example, MHCII, CD86 and CD40, as well as IL-12 are essential for antigen presentation and persistent T-cell activation in EAE [79,80]. The release of other pro-inflammatory cytokines, including $\text{TNF}\alpha$, $\text{IFN}\gamma$ and IL-6, as well as proteases, reactive oxygen species (ROS), reactive nitrogen species (RNS) establishes an inflammatory microenvironment that facilitates damage to the myelin sheath and the surrounding cells [81]. As well as their general role in the inflammatory cascade, there is strong evidence that macrophages directly mediate axon damage, an aspect of MS pathology that is attributed to permanent, progressive symptoms [82]. ROS and RNS of macrophage/microglial origin have been implicitly shown to cause axonal degeneration in EAE [83], with monocyte-derived macrophages in particular associated with direct axon contact [84]. Similarly, in human disease, axon loss in lesions is most prominently correlated with pro-inflammatory activity in macrophages [19,22–24], evident in both relapsing, primary progressive and secondary progressive disease [20]. Higher iNOS expression and activity is evidenced in both circulating patient monocytes as well as MS brain tissue versus healthy controls [85,86], and is significantly correlated with axon densities in lesions [19]. Crucially, iNOS expression in the CNS co-localises with CD64+ macrophages and is associated with the presence of nitrotyrosine and MBP fragments, highlighting the contribution of NO mediated damage at these sites [22].

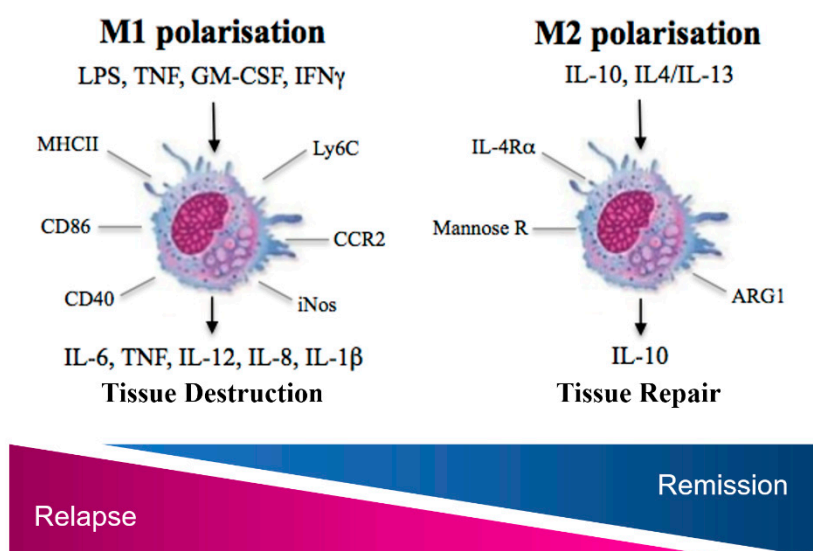


Figure 1. Schematic illustrating canonical M1 and M2 polarised macrophages that result in tissue destruction and tissue repair in the CNS in MS and experimental autoimmune encephalomyelitis (EAE). Agonists, cell surface markers, receptors and typical cytokines released are also highlighted.

While the pro-inflammatory population predominates in early disease, a gradual shift occurs through intermediate activation states [54,87,88]. By the remission phase of EAE, cells have a more alternatively activated or M2-like phenotype as clearly evidenced by an elegant fate tracing study labelling cells expressing iNOS and Arginase-1, canonical markers of M1-like and M2-like function, respectively [87]. Importantly, this functional shift can also be observed in MS patient lesions, with myelin-laden macrophages expressing high levels of M2 associated CD163 and CD206 [54], with a recent study highlighting the presence of CD206+ cells in inactive lesion centres, while iNOS expression was associated with areas of active pathology at lesion edges [88].

M2 polarisation can be driven by IL-4/IL-13 and IL-10 in vitro, with the latter driving a distinct transcriptional signature and overall, a more profound inhibition of pro-inflammatory processes [78,89]. IL-10 upregulation in the CNS in the recovery phase of the EAE model has been demonstrated [90,91], while the administration of IL-10 expressing mesenchymal stem cells is suppressive to disease development, with a capacity to suppress bone-marrow derived dendritic cell (BMDC) antigen presentation in vitro [92]. Macrophage IL-10 production is a general characteristic of alternative

activation, with autocrine signalling resulting in downregulation of MHCII and co-stimulatory molecules, suppressing antigen presentation and hampering CD4 T cell responses [89,93,94]. Suppression of the iNOS mediated respiratory burst also occurs, with the IL-4 driven upregulation of Arginase-1 [89], which depletes NO precursor arginine and drives the polyamine pathway, associated with collagen synthesis and tissue repair in the damaged CNS in animal models [95]. IL-4 is also associated with upregulation of scavenger receptors and enhanced endocytotic ability [89], facilitating clearance of myelin debris as evidenced in human MS lesions [96]. Moreover, IL-10 and IL-4 immunoreactivity has been shown in human MS brain tissue in active demyelinating lesions and at the rim in chronic active lesions, with receptors for these cytokines highly expressed by macrophages in parenchymal and perivascular areas [97].

This tissue reparative role, coupled with their inflammation limiting capacity, positions M2-like macrophages as key effectors in disease remission. Supporting this, the adoptive transfer of M2 polarised cells is beneficial in preventing EAE development [98,99]. There is evidence of a pro-repair action that can be mediated without CNS entry of adoptively transferred cells, suggesting that this effect may be mediated in the peripheral environment [100]. A more direct effect of the M2 phenotype on remyelination within CNS lesions has also been indicated. In toxin-induced demyelination, decreased oligodendrocyte precursors were linked to the loss of macrophage secreted growth factors as a result of depletion [101]. Additionally, Miron and colleagues have demonstrated a role for M2 macrophages in murine oligodendrocyte differentiation that is diminished by M2-specific depletion within lesions and highly dependent on M2-associated TGF- β family signalling molecule, Activin-A [102]. Interestingly, increased TGF- β is observed in blood cell cultures and in CSF of patients in remission compared to those with active disease [103,104]. The studies in animal models implicating macrophages in repair are additionally complemented by observations in human MS patients, where in resolving lesions with remyelination, macrophages persist, coexisting with this repair process and displaying different morphology and staining patterns than those observed in acute disease [105].

As outlined, we clearly see a dual role for macrophages in MS models and human disease owing to their polarisation potential, at the outset contributing substantially to disease pathogenesis, while studies in animal models highlight an essential reparative role, with evidence to support similar mechanisms in human MS. Some of the aforementioned studies illustrate that by depleting or increasing these populations experimentally, the outcome of EAE can be substantially altered. Thus, modulating these populations to minimise pro-inflammatory or M1-like and favour M2-like polarisation may hold potential for therapeutic translation in MS.

2.3. Current Therapeutics and Their Impact on Monocytes and Macrophages

While MS remains incurable, there has been considerable development over the last 25 years in terms of available disease modifying therapies (DMTs). First introduced in the mid-1990s, there are over 10 FDA approved treatments presently available (summarised in Table 1) which show efficacy in reducing disease relapses in RRMS patients [106]. Despite the advancement, Interferon Beta (IFN β) and Glatiramer Acetate (GA), the first two drugs to be introduced, remain the “first line” therapies for MS owing to their relative safety and proven efficacy [106,107]. Both drugs are administered by self-injection and function in an immunomodulatory capacity. Oral DMTs available for RRMS include immunosuppressives Fingolimod and Teriflunomide (TFM), as well as immunomodulatory Dimethyl fumarate (DMF). In addition, a number of monoclonal antibody-based therapeutics for RRMS have emerged, including natalizumab and alemtuzumab. While highly efficacious in preventing relapse and disability, these newer treatments carry high risk side effects including progressive multifocal leukoencephalopathy (PML) and development of secondary autoimmune diseases, respectively [108,109]. Moreover, Daclizumab, a monoclonal antibody against the alpha subunit of the IL-2 receptor has been recently removed from the market due to reports of encephalitis development [110].

Table 1. Approved Disease Modifying Treatments for Multiple Sclerosis (MS) and evidence for their effects on monocytes and/or macrophages.

	Type	FDA Approval	Format (Oral/Injectable)	Mechanism of Action	Studies in Monocytes/Macrophages	Adverse Effects
Interferon β	Cytokine	1993	Injection (SC or IM)	- Type 1 Interferon - Effect in B and T cells - Reduction in BBB disruption [111,112]	- IL-27 production by myeloid cells suppresses Th17 differentiation in EAE [113,114] - Increased response to IL10 in human monocytes [115]	Flu-like symptoms [116]
Glatiramer Acetate	Synthetic Copolymer [E,K,A,Y] _n	1995	Injection (SC)	- Shift from Th1 to Th2 responses - Increased foxp3+ Tregs [117,118]	- Shift treated patient monocytes to type II antigen presenting cells—Th2 T cell responses [119–121] - Increased phagocytosis in rat microglia and human monocytes [122,123]	Injection site reaction [111]
Natalizumab	Anti-alpha-4 integrin	2003	IV infusion	- Prevent α 4 integrin mediated T cell migration and CNS infiltration	- Reduced CNS accumulation of activated microglia and macrophages with early therapy in EAE [124] - Reduced pro-inflammatory mir-155 in patient monocytes [125]	PML risk, Allergic Reactions [108]
Fingolimod	Antagonist of sphingosine 1 phosphate receptor	2010	Oral	- Suppress lymphocyte migration from lymph nodes	- Microglial M2 polarisation in stroke model [126] - Alteration cytokine production in patient monocytes [127–129] - Reduced pro-inflammatory mir-155 in patient monocytes [125]	Cardiovascular complications [130]
Teriflunomide	dihydroorotate dehydrogenase inhibitor	2012	Oral	- Suppress rapid expansion of lymphocytes by inhibition of the pyrimidine <i>de novo</i> synthesis pathway	-	abnormal liver enzymes, gastrointestinal symptoms [131]

Table 1. Cont.

	Type	FDA Approval	Format (Oral/Injectable)	Mechanism of Action	Studies in Monocytes/Macrophages	Adverse Effects
Dimethyl Fumarate	Fumaric Acid Ester	2013	Oral	<ul style="list-style-type: none"> - reduction of Th1 responses - Nrf2 activator - NfkB inhibitor 	<ul style="list-style-type: none"> - Decreased monocyte infiltration in EAE [132,133] - Glycolysis inhibition in murine macrophages [134] - Decreased pro-inflammatory cytokines in EAE [135,136] - Decreased pro-inflammatory cytokines and mir-155 in patient monocytes [125] 	gastrointestinal symptoms, abnormal liver enzymes, flushing [137]
Alemtuzumab	Anti-CD52	2014	IV infusion	<ul style="list-style-type: none"> - Depletion of mainly mature T and B lymphocytes, to a lesser extent monocytes and dendritic cells 	<ul style="list-style-type: none"> - 	Development of other autoimmune disease, Intracerebral haemorrhage (rare) [109,138]
Mitoxantrone	Chemotherapeutic agent	2003	IV infusion	<ul style="list-style-type: none"> - DNA topoisomerase inhibitor - Suppressed cell proliferation - Impaired antigen presentation [139] 	<ul style="list-style-type: none"> - Reduced ex vivo migration capacity of patient monocytes [140] 	Leucopenia [141]
Ocrelizumab	Anti CD-20	2017	IV infusion	<ul style="list-style-type: none"> - Depletion of B cells - Note: the only FDA approved DMT for PPMS 	<ul style="list-style-type: none"> - 	Infusion related reaction, infections [142]

Broadly, these therapies act by either altering T-cell responses (IFN β , GA, DMF), inhibiting lymphocyte trafficking (Fingolimod, Natalizumab) or depleting lymphocyte populations (Alemtuzumab, Ocrelizumab, TFM, Mitoxantrone). How these therapies impact on monocyte and macrophages, however, has been less explored. Below and in Table 1 we consider evidence of any direct action of DMTs on monocyte and macrophage populations, which may contribute to their respective therapeutic efficacies.

2.3.1. Interferon- β

IFN β , a type 1 interferon, is an anti-inflammatory cytokine and was the first available DMT for the treatment of MS. In addition to affecting T and B lymphocyte function and reducing BBB transmigration [111,112], IFN β exerts effects on cells of the innate immune system in the context of MS. Of note, two studies demonstrate a key role of IL-27 production by DCs and macrophages in suppressing Th17 T cell mediated responses in EAE models [113,114]. An effect of IFN β treatment on human monocytes has also been documented, with monocytes from treated patients shifting towards a CD14⁺⁺CD16⁺ intermediate phenotype [66]. Notably, patient monocytes produce less IL-1 β in response to inflammatory stimuli [143], and show significantly reduced production of IL-8 and CCL2 after *ex vivo* T cell activation [144]. In terms of IFN β on macrophage and monocyte polarisation, a study by Liu and colleagues show enhanced sensitivity to IL-10, a driver of the M2 phenotype, through upregulation of the IL-10 receptor in both human monocytes and macrophages [115]. In conjunction with the increased serum IL-10 levels seen in IFN β treated MS patients [145,146], this indicates IL-10 modulation of macrophages and their monocyte precursors may occur in response to IFN β treatment.

2.3.2. Glatiramer Acetate

GA is a synthetic copolymer of lengths 50 to 90 residues of randomly arranged L-tyrosine (Y), L-glutamic acid (E), L-lysine (K), L-alanine (A), with its efficacy chiefly credited to its ability to modulate peripheral T cells towards a Th2 phenotype and increase the Treg population [117,118]. The effects of GA on the myeloid cell population are also believed to contribute to its therapeutic efficacy. This effect was initially demonstrated on human and animal cells *in vitro*, with GA treated monocytes showing decreased TNF α and cathepsin B levels in response to inflammatory stimuli, as well as increased production of anti-inflammatory IL-10 [147,148]. Similar findings were recapitulated in a number of studies utilising isolated monocytes from GA treated patients, showing decreased TNF α , IL-12 and IL-1 β in conjunction with increased IL-10, TGF- β and IL-1 receptor antagonist [119–121]. This cytokine shift in GA-treated monocytes is primarily explored in terms of the effects of antigen presentation by myeloid lineage cells on the T-cell response. The effects seen are consistent with type 2 antigen presenting cells, which induce development of Th2 responses. Interestingly, GA has been shown to increase phagocytosis in both rat microglia and MS patient monocytes [122,123] with debris clearance necessary for remyelination [149]. Monocyte modulation may be among the most long-lived responses to GA treatment, with a study showing increased anti-inflammatory monocytes as one of two significant changes in the leukocyte population that prevail following treatment periods of up to 16 years in MS patients [150].

2.3.3. Dimethyl Fumarate

DMF is an immunomodulatory drug originally used for psoriasis treatment, but is also therapeutically useful in the treatment of RRMS, resulting in diminished Th1 responses [151]. It is a known activator of the anti-inflammatory transcription factor Nrf2, as well as interfering in TLR signalling pathways upstream of NF κ B activation [152–155]. The immunomodulatory function of DMF extends to cells of the myeloid lineage, in EAE resulting in reduced monocyte infiltration and microglia with a more alternatively or 'M2' activated phenotype [132,133]. In murine monocytes, DMF treatment increases myeloid derived suppressor cells and results in an alternative activation profile in monocytes consistent with Type II APCs, while in macrophages decreases iNOS and TNF α expression,

with concomitant increases in Arg1 and IL-10 [135,136]. Similar effects on monocyte populations have been demonstrated in DMF treated MS patients, with peripheral monocytes showing reduced levels of mir-155, a micro-RNA associated with major pro-inflammatory effect [125], while in vitro treated human monocytes showed suppressed TNF α , IL-6 and IL-10 responses to a pro-inflammatory stimulus [125]. Furthermore, it has been shown the DMF treatment results in reduced expression of MHCII molecules and NF κ B in human myeloid DCs and a concomitant reduction in their capacity to activate T cells [156]. Interestingly, it has recently been shown that DMF has a profound effect on cell metabolism, blocking the glycolysis pathway by inhibition of glyceraldehyde 3-phosphate dehydrogenase (GAPDH) in murine macrophages [134]. This may contribute to the mechanism by which DMF promotes M2-like macrophages, as a preference for glycolysis is associated with M1 macrophages, while M2 macrophages see higher levels of oxidative phosphorylation [134,157].

2.3.4. Fingolimod

Fingolimod is an antagonist of sphingosine 1 phosphate receptor, which functions to inhibit leukocyte passage out of the lymph nodes. In monocytes isolated from Fingolimod treated patients, alteration in cytokine production has been observed with a reduction in pro-inflammatory cytokines such as TNF α , IL-1 β and IL-6 [127–129]. In addition, the capacity of fingolimod treatment to result in an M2 polarisation of microglia has been demonstrated in a murine model of stroke [126]. Similarly to MS patients treated with DMF and Natalizumab, Fingolimod also results in a reduction of pro-inflammatory mir-155 in circulating monocytes [125].

3. Nanoparticles and Microparticles in MS

The goal of most current MS therapies is dampening the immune response in the CNS, reducing the number and the severity of relapses and lesions in MS patients. Such treatment raises concerns due to the associated chronic, non-specific immunosuppression, which may pose a serious risk in the medium/long term [158]. Coupled with the fact that DMTs are not effective in all patients, this underlies an urgent need for the development of novel therapeutic strategies to overcome these issues. In recent years, nanotechnology has emerged as a promising method of drug delivery offering many advantages over conventional delivery mechanisms. Nanoparticle and microparticle carriers are colloidal particles, where sizes ranging from 10–1000 nm are generally designated nanoparticles (NPs), while those ranging from 1–250 μ m are designated microparticles (MPs) [159]. NPs and MPs can be prepared from a wide range of materials; among the most popular are lipid and polymer-based nanoparticles including poly-lactide co glycolic acid (PLGA) and chitosan [160]. Such carrier systems offer the ability to deliver otherwise challenging molecules, such as nucleic acid and low bioavailability drugs and facilitate controlled release. In addition, nanotechnology can mediate cell specific delivery and passage across biological barriers like the BBB through targeting by carrier type, size, surface charge, and conjugation of specific targeting ligands. This minimizes interaction with non-target cell types and can reduce the amount of drug required for sufficient accumulation in target cells. As outlined above, macrophage manipulation may offer novel opportunities in the treatment and management of MS. Macrophages are attractive targets for NP/MP-mediated delivery, usually seeing high uptake due to their phagocytic nature. In the following sections, we will give an insight on the role that NPs/MPs could play in either improving the efficacy of existing drugs or helping the delivery of newly developed therapeutics in preclinical models of MS, with particular emphasis on targeting macrophages. Although the majority of studies were performed in animal studies, it holds much promise for their translation in MS; as it was previously mentioned that many of the approved MS drugs have been tested for safety and efficacy in EAE models, as is recommended by preclinical guidelines [36]. A number of NP/MP based studies in MS models with impact in macrophages are explored in Table 2.

Table 2. NP/MP strategies in MS models with impact in macrophages.

Reference	NP/MP Chemistry	Size	Cargo	Functionalised	Route of Delivery	Model	Target Cells	Additional Points
[161]	PEG-PLL-PLLeu copolymers	not reported	c-Rel siRNA	-	IP	EAE	Macrophage	
[162]	inorganic-organic hybrid NP	60–80 nm	glucocorticoids	-	IP and IV(more effective)	EAE	Macrophage	
[163]	PEGylated liposome	<100 nm	Prednisolone	PEG	IV	EAE	<i>not specified</i>	liposomes were found mostly in macrophages, microglia and astrocytes
[164]	liposome	<100 nm	methylprednisolone	-	IV	EAE	<i>not specified</i>	Compared with free drug, only liposomal formulation resulted in significantly decreased CD68+ cells
[165]	liposome	<i>not reported</i>	methylprednisolone	short peptide fragments of ApoE or of β -amyloid	IV	EAE	<i>not specified</i>	
[166]	PEGylated liposome	95–120 nm	methylprednisolone	PEG + Glutathione	IV	EAE	<i>non specified</i>	Bigger reduction in disease score with the targeted vs non targeted liposome
[167]	PLGA	540 nm	(tNP) PLP (coated)	-	IV	EAE	APCs	Taken up by macrophages and DCs, most antigen presentation by DCs
[168]	PLGA	<i>not reported</i>	(tNP) PLP + rapamycin	-	SC prophylactic, IV peak disease	EAE	APCs	in vivo trafficking—IV-accumulation in liver and spleen most localisation to Macrophages and DCs in the spleen, but SC goes to the draining lymphnodes
[169]	PLGA	350–835 nm	(tNP) PLP	-	IV	EAE	APCs (Macrophage)	Immunofluorescence staining showing co localisation with F4/80 positive macrophages, lungs, spleen, lymph nodes
[170]	PLGA	80nm, 400 nm	(tNP) PLP	-	IV	EAE	APC's (DCs)	Larger particles show better uptake in BMDCs
[171]	PLGA	400–1500 nm	(tNP) MOG (coated)	-	IV or SC	EAE	APCs	SC admin not effective, non-significant trend to bring on disease more quickly

Table 2. Cont.

Reference	NP/MP Chemistry	Size	Cargo	Functionalised	Route of Delivery	Model	Target Cells	Additional Points
[172]	Au	60 nm	(tNP) MOG + small molecule (ITE)	PEG(to stabilize)	IV or IP	EAE	DC	ITE ligand activates the aryl hydrocarbon receptor (Ahr), which can induce tolerogenic DCs. Observed Ahr activation in Macrophages in vivo
[173]	poly(ϵ -caprolactone)	300–600 nm range	(tNP) Recombinant human MBP	-	SC	EAE	APCs	Histological observation of no macrophage or T cell infiltration in treated animals
[174]	PLGA	200 nm	(tNP) MOG and IL-10	-	SC	EAE	APCs	Authors suggest that observed T cell anergy and inhibited lymphocyte proliferation is due to induction of tolerance in macrophages
[175]	Acetalated Dextran	<i>not reported</i>	(tNP) MOG and Dexamethasone	-	SC		APC's (Macrophage)	Reduced macrophage GM-CSF and IL-17
[176]	PLGA	<i>not reported</i>	(tNP) MOG, Vitamin D3, TGF β , GM-CSF	-	SC	EAE	APCs	Macrophages have second highest MP uptake in axillary lymph after DC's, while these cells show equal uptake in inguinal lymph nodes. Treatment results in decreases numbers of activated macrophages in CNS
[177]	PLGA	400–500 nm	(tNP) PLP	-	IV	EAE		Localisation to spleen, liver, and lung at 3, 6, and 18 h post injection, cleared by 24 h
[178]	polystyrene, PLGA	500 nm	(tNP) PLP	-	IV	EAE	Macrophage	SC did not work as well as IV admin, NP show localisation to spleen marginal zone macrophages and uptake via MARCO receptor

3.1. Monocyte and Macrophage Depletion

The first studies to demonstrate NP/MP uptake by macrophages and efficiency in EAE were conducted as early as 1981, where intraperitoneal (IP) injections of silica quartz dust in rats was successful in lowering the clinical score in the EAE model by depletion of the peritoneal macrophage population [46]. Similar results were obtained by Huitinga and co-authors shortly after, where intravenous (IV) administration of mannosylated liposomes containing dichloromethylene diphosphonate (Cl₂MDP) in rats selectively depleted circulating monocytes and macrophages in the spleen and the liver [47]. This was accompanied by a lower infiltration rate of monocytes in the CNS and subsequent improvement of the clinical score in EAE model. Further research provided more details about the mechanisms of action of the Cl₂MDP-liposomes, showing that the myelin sheath of treated mice was not affected, with the expression of iNOS and TNF α by macrophages dramatically inhibited [48]. Importantly, the authors showed that the liposomes impede the CNS infiltration specifically of monocytes but not T-cells, suggesting again a key role of these innate immune cells in the onset and progression of the disease in the EAE model [48].

In line with these early findings, Getts and colleagues have shown that empty carboxylated PLGA and other negatively-charged MPs injected IV were taken up specifically by monocytes and monocyte-derived macrophages via the macrophage receptor with collagenous structure (MARCO) [179]. These cells were then less able to migrate into the brain and accumulated for a short period in the spleen, where they underwent apoptosis, accompanied by a reduction in the clinical score of the EAE. However, the authors did raise concerns about using this drastic approach in disorders where monocyte-derived macrophages are important also for disease resolution, as is indicated in MS, underpinning the limitation of total depletion as a viable therapeutic strategy.

3.2. NP/MP and Antigen Specific Tolerance Induction

Tolerogenic nanoparticles (tNPs) are typically packaged with the antigen that elicits the abnormal immune response in autoimmune disorders. They exert their immunomodulatory function by selectively targeting professional antigen-presenting cells (APCs), such as macrophages and DCs, due to their intrinsic ability to internalise tNPs via endocytosis. APCs loaded with tNPs are then able to elicit an effective antigen-specific immune response, as elegantly reviewed in Kishimoto & Maldonado [180]. In the context of MS, tNPs are prepared by encapsulating myelin antigens into different carriers, where they are taken up by APCs which then efficiently trigger tolerance induction in autoreactive T cells in in vivo models. The typical cargo is the myelin antigen in one of its different forms, including myelin oligodendrocyte glycoprotein (MOG) [171,172,174–176], myelin basic protein (MBP) [173] and proteolipid protein (PLP) [168,178,181]. The carriers vary from study to study, and they include PLGA [168,171,174,178,181], acetalated dextran [175], poly(ϵ -caprolactone) [173] and gold [172], among others. Interestingly, coupling the myelin antigen with immunomodulatory agents like dexamethasone [175], IL-10 [174], 2-(1'Hindole-3'-carbonyl)-thiazole-4-carboxylic acid methyl ester (ITE) [172], or rapamycin [168,181] has shown efficacy in the EAE model.

Considering APCs are essential for mediating the tNP effect, it is not surprising that some studies have shown a reduction in EAE clinical score after tNPs administration, due to the specific contribution of macrophages. For example, IV injection of PLGA and polystyrene PLP-tNPs in EAE mice models were taken up specifically by macrophages via interaction with the scavenger receptor MARCO and delayed the onset or improved the progression of the disease [178]. Similarly, PLGA PLP-tNPs administered with rapamycin were shown to co-localise with macrophages in the spleen, although the precise effects of tNPs on macrophages were not fully elucidated [168]. Functionally, treatment of PLGA MOG-tNPs in EAE mice dramatically reduced the number of activated macrophages and microglia in the CNS and this contributed to the overall decreased disease severity, while also reducing the number of CD86+MHCII+ DCs in lymph nodes [176]. Collectively, these works show that macrophages play a fundamental role in the uptake of tNPs, improving clinical progression in the EAE model and illustrating a promising NP-mediated, macrophage directed therapeutic approach.

3.3. Corticosteroid Delivery

Although not considered DMTs, steroids (including methylprednisolone, dexamethasone and prednisolone) are often used as first line response to treat acute relapses in MS patients due to their potent and quick ability to close the damaged blood brain barrier and reduce inflammation in the CNS. However, they are characterised by side-effects due to high doses and systemic administration. This issue can be addressed by administering them as liposomal drugs, which lead to higher tissue concentrations in the inflamed target organ compared to an equivalent dose of the free drug. Several research groups showed evidence of this advantage by using different carriers to enhance the efficacy of steroids in the EAE model, with macrophages frequently shown to be the primary effectors of this response [162–165,182].

IV administration of Prednisolone (PL) and methylprednisolone (MPL) encapsulated in PEGylated liposomes has been investigated in EAE, resulting in improved clinical score when compared to free drug [163,164]. Interestingly, liposomes in these studies were shown to be taken up mainly by macrophages in the spinal cord of injured animals, and by microglia and astrocytes. The therapeutic efficacy of the PL and MPL liposomes was coupled with decreased blood brain barrier disruption, decreased macrophage and T-cell infiltration in the CNS, and reduced demyelination and axonal loss [163,164]. Similarly, an inorganic-organic hybrid NP loaded with glucocorticoid betamethasone (BMP-NP) was shown *in vitro* to be preferentially taken up by macrophages rather than T cells or B cells [162]. *In vivo*, macrophages from BMP-NPs treated mice polarised towards an anti-inflammatory phenotype to the same extent as free glucocorticoid drug dexamethasone (DEX), as shown by a reduction in MHCII and CD86 positive cells and in TNF α secretion [162]. While the number of infiltrating T cells did not change between empty-NP and BMP-NPs treated mice, the number of macrophages in the spinal cord were significantly reduced, suggesting again that these cells are the primary target of this therapy. As a proof of concept for human translation, the authors treated peripheral blood monocytes from healthy individuals with BMP-NPs and observed an increased expression of anti-inflammatory genes and lower levels of pro-inflammatory genes in qRT-PCR [162]. This work strongly supports the notion that polarisation of macrophages towards an anti-inflammatory state could be of therapeutic benefit in MS.

3.4. NPs/MPs and Current MS Disease Modifying Therapies

In order to improve their bioavailability and lower their adverse effect, recent studies have investigated encapsulation of orally administered DMTs into NP/MPs. For example, Kumar and collaborators have shown that the formulation of DMF-loaded nanolipidic carriers (NLCs) coated with vitamin-based neuroprotective molecules like tocopherol acetate cholecalciferol and retinol acetate improved the clinical score in a cuprizone-induced demyelination mouse model when given once daily orally [183]. Treatment with DMF-NLCs, especially when coated with the vitamin-based compounds, showed improved locomotor activity, motor coordination and balance compared to free DMF treated mice. Myelination status was also measured in brain slices, again with the DMF-loaded vitamin NLCs showing higher remyelination compared to free DMF [183]. Further histopathological analyses showed none of the stomach tissue damage in DMF-NLC treated mice that was observed in those treated with free DMF.

TFM, an orally administered DMT for MS, is often associated with hepatotoxicity, possibly due to its delivery route which leads to higher exposure of the drug within the systemic circulation [184]. TFM was loaded into NLCs subsequently combined with mucoadhesive and gelling agents in order to overcome mucociliary clearance and achieve efficient delivery via the nose to brain route [185]. TFM-NCL and mucoadhesive TFM-NCL (TFM-MNLC) were given orally and intranasally to rats in a cuprizone-induced demyelination model and recovery was assessed by an exteroceptive behavioural model. Although very preliminary, the results showed a trend of improved neurological function in rats treated intranasally versus orally without hepatic or renal biomarker elevation [185].

These studies illustrate the potential for the use of nanocarriers to improve efficacy and specificity of existing DMTs for MS. Given that DMTs can produce effects in monocytes and macrophages, coupled with the high NP/MP uptake generally seen in these cells, macrophages could play an unexplored role in the reduced disease severity in these models, which may be worth further investigation.

3.5. NPs/MPs and Novel Drugs

Besides the use of NPs/MPs as carriers for existing drugs and tolerogenic molecules, many research groups have exploited the typical features of NPs/MPs (low cell toxicity, longer bioavailability, ability to cross the blood brain barrier among others) to design and test novel therapeutic strategies in *in vivo* models. This has been evidenced in targeting a number of cell populations in MS. For example, the pro-remyelination factor leukaemia inhibitory factor (LIF) was encapsulated in PLGA NPs, and specifically targeted oligodendrocyte precursor cells (OPCs) by conjugating the NPs with NG2 chondroitin sulphate proteoglycan antibodies [186]. NG2-targeted LIF-NPs were able to induce significantly higher percentage of remyelinated fibres and to increase the myelin thickness per axon compared to non-targeted LIF-NPs, suggesting that the conjugation of LIF-NPs with NG2 antibodies, and thus the OPC-specific targeting, was critical for the observed therapeutic effect [186]. Similarly, another group investigated delivery to OPCs, this time intranasally administering short-interfering RNA (siRNA) in chitosan NPs in ethidium bromide induced demyelination [187]. The target, LINGO-1 is a transmembrane protein that suppresses myelination and axonal regeneration and the antibody Opicinumab/BIIB033 directed towards this mediator has been tested in clinical trials for MS with limited effectiveness, potentially due to low CNS penetration [188,189]. Rats treated with LINGO-1 siRNAs NPs showed overall better motor activity and coordination and a more compact myelin sheath histologically, illustrating the effectiveness of this approach [187]. Drug repurposing has also been explored, with anti-inflammatory cancer drug lenalidomide delivered in combination with in anti-oxidant cerium oxide NPs, and antibiotic minocycline was encapsulated in PEG liposomes, both of which result in improved clinical scores in EAE, although the cellular target in these studies was not further investigated [190,191].

Moreover, the specific targeting of macrophages for siRNA delivery has been investigated in a recent study where the pathologic crosstalk between pro-inflammatory macrophages and auto-reactive Th1/Th17 T cells was tackled by silencing the transcription factor c-Rel [161]. C-Rel plays a key role in inducing pro-inflammatory cytokine secretion by macrophages and therefore in controlling the T cell response, with a siRNA encapsulated into PEG-PLL-PLLeu MPs and tested *in vitro* on macrophages [161]. Decreased levels of secreted IL-1 β among others were observed, suggesting that c-Rel silenced macrophages might be less able to induce Th1 and/or Th17 responses. Intraperitoneal (IP) injections of PEG-PLL-PLLeu-c-Rel MPs reduced the clinical score in the EAE model, and the fact that the MPs were preferentially localised in macrophages suggested that they played a crucial role in this phenotypic effect. Moreover, this was associated with a decrease of the number of infiltrating macrophages in the spinal cord and brain and lower serum levels of both IFN γ and IL-17A, suggesting that these MPs are able to dampen the Th1/Th17 response [161]. Together these studies illustrate the utility of nanotechnology for the delivery of novel therapeutics in MS models, with the latter showing the successful targeting of macrophages to induce a therapeutic response.

4. Optimising Delivery and NP/MP Uptake in Macrophages

As discussed, macrophages offer untapped potential in terms of therapeutic manipulation in MS, with nanotechnology as a promising means to realise this. While it is clear that macrophages are highly receptive to NP/MP mediated delivery, it is desirable to maximise the specificity of targeting to a therapeutically relevant population of cells, thus maximising the NP/MP payload and minimizing off target effects. In the functional modulation of macrophages in MS, specific targeting to M1-like macrophages in the CNS or their peripheral precursors could minimize the effects of this treatment on peripheral and other tissue macrophages, which are required in normal innate and adaptive

immune defence. Here we explore the enhancement of macrophage targeting through particle size and functionalisation, which could prove useful in the development of strategies for macrophage targeting in MS and associated models.

4.1. Size

Particle size is a key biophysical characteristic affecting cellular targeting and uptake. While endocytosis and pinocytosis facilitate the entry of NP around 200 nm or smaller into most cell types, phagocytosis can accommodate the uptake of particles up to 10 µm in diameter. This process is restricted to professional APCs, and thus larger particle sizes can be used to passively target macrophages, as reviewed extensively in [192]. In addition to restricting particle entry to phagocytosing cells, uptake studies show a trend for preferential uptake of larger NPs and MPs in macrophages compared with smaller NPs. In vivo studies with liposomes in both alveolar macrophages and in an atherosclerosis model show increased uptake with increasing size up to 2 µm and 500 nm respectively [193,194]. A similar trend is observed in vitro with PLGA and chitosan particles, with an optimal size at approximately 2 µm, and the latter study showing a second peak at 430 nm [195,196].

Despite the preference for larger NP/MP sizes, particles of smaller size are still frequently used for macrophage delivery [162,197–200], and may be more appropriate for a number of reasons. Administration route can factor in; nanomedicines delivered intravenously (IV) are restricted by small capillary size to avoid embolism [201], while small sizes are also preferred for intranasal administration for local CNS delivery (the nose to brain (NTB) route), which are restricted by axon diameter [202,203]. Additionally, for CNS entry from systemic circulation, smaller particles may have easier access across the BBB [201]. It is worth considering in designing strategies to target pro-inflammatory macrophages, that there is evidence that polarisation both contributes to size related uptake preferences of macrophages and can itself be influenced by NP/MP uptake. The investigation of silica NP (26 and 41 nm) and latex MP (1.75 µm) uptake by polarised cells indicates that while there is no significant difference in MP uptake, M1-like cells show significantly lower NP uptake than M2-like cells, proposed as a result of a higher endocytic capacity of M2-like cells [204]. Furthermore, there is evidence that smaller particles can increase the production of pro-inflammatory cytokines to a greater extent than larger particles [196].

4.2. Functionalisation

Enhancement of macrophage specific delivery can be achieved by the conjugation of ligands or antibodies to target highly expressed surface receptors on these cells. Mannose and galactose ligands have been exploited for their ability to bind the mannose receptor (CD206) and the galacto-type lectin, respectively. These receptors are highly expressed on macrophages and have been demonstrated for macrophage targeting in many disease contexts, among them infection [205,206], inflammatory bowel disease [207,208], cancerous tumours [209,210] tuberculosis [211] and atherosclerosis [212]. Notably, the use of mannose functionalised NPs has been demonstrated for the delivery of an antiretroviral drug targeted to macrophages in the CNS of rats, showing increased CNS drug concentrations following IV delivery compared with unmodified NPs or free drug [197].

The CD11b integrin is present on many leukocytes and is highly expressed in macrophages, their monocyte precursors, and microglia. A proof of concept study encapsulating leukaemia inhibitory factor (LIF) showed significantly increased therapeutic impact with anti-CD11b functionalisation in a myeloid cell line with M1-like characteristics, indicative of increased uptake [213]. In terms of microglial targeting, in a mixed glial culture model, anti-CD11b conjugation has been shown to maximise microglial uptake, while reducing the proportions of astrocytes and oligodendrocytes internalising NPs [214]. A separate study comparing anti-CD11b conjugation to that of non-specific IgG conjugation, showed that in vitro transfection efficiency of microglia with a microRNA cargo was increased from ~52% (IgG) to ~71% (CD11b) [200].

Additionally, CD64 may represent a promising candidate for mediating specific NP/MP delivery. CD64 or Fc γ receptor I (Fc γ RI) is constitutively expressed on monocytes and macrophages and inducible in neutrophils, but has shown to be substantially upregulated in macrophages with a pro-inflammatory or M1-like phenotype [215,216]. CD64 is also an MS relevant target, evidenced by its upregulation in macrophages of human disease samples [54]. As an NP/MP targeting ligand, in vitro macrophage uptake following conjugation of a CD64 antibody to solid lipid and PLGA NPs has been demonstrated, intended for future use in rheumatoid arthritis models [217,218]. Furthermore, Yong et al. demonstrate the utility of CD64 as a monocyte specific targeting ligand in a study delivering siRNA cargo to human peripheral blood mononuclear cells [219].

4.3. Macrophage Modulation: A Peripheral or CNS Centric Approach?

Worth considering in MS is the most pertinent location for targeted drug delivery to macrophages and how this relates to NP/MP design and administration. While macrophages that enter as monocytes in the hallmark immune infiltrate are capable of directly inflicting pathogenic damage as outlined above, peripheral monocytes also have a significant role in shaping the overarching immune landscape in which disease occurs, contributing to the cytokine environment and the shaping of T cell responses. An outstanding question thus remains as to whether targeting macrophages in the CNS or focusing delivery to peripheral monocyte precursors is the more appropriate therapeutic approach.

Peripheral monocyte uptake is readily achieved following systemic administration, which sees substantial NP uptake by circulating monocytes and DCs of the mononuclear phagocyte system (MPS) and potential accumulation in the spleen, liver, and kidneys owing to high numbers of those cells therein [220]. Manipulation of these cells may impact MS/EAE outcome by altering the immune response in the periphery, as evidenced in the aforementioned immune tolerance approaches [168,169,179] as well as in preventing CNS accumulation as a consequence of depletion [46–48]. Outside of immune tolerance approaches, however, peripheral modulation is not without concern owing to the potential for global immunosuppression. Directly targeting macrophages that have been mobilised to the CNS has the potential to avoid this complication, however, it represents a more complex target for drug delivery.

Systemically administered nanomedicines face two major hurdles before even considering their function in the brain; they typically must avoid uptake by peripheral macrophages in order to reach the CNS and also contend with the BBB, a highly selective barrier effected through the presence of tight junctions, degradative enzymes and selective transport proteins present where CNS microvessels interface with astrocytes [221]. With liposomes and hydrophobic carriers like PLGA, peripheral phagocyte evasion is most commonly achieved by polyethylene glycol (PEG) modification. PEG prolongs NP circulation by forming a hydrophilic layer around the NP and reducing opsonisation [220]. After overcoming peripheral uptake, most NP carriers, including PLGA, still cannot enter the CNS without the conjugation of ligands that exploit existing BBB transport mechanisms. Adsorptive-mediated transcytosis offers passage through the BBB by charge mediated binding of ligands to the negatively charged brain endothelial surface. PEG and also TAT, the HIV cell-penetrating peptide, have been used as ligands NP/MP targeted to macrophages within the CNS [222,223]. Receptor-mediated transcytosis offers a highly specific transport mechanism through the binding of conjugated ligands to specific receptors. With respect to CNS macrophage targeting, lactoferrin [222], transferrin receptor binding peptide [224], rabies virus glycoprotein [225], and mannose [197] are among those that have been explored.

Alternatively, localised CNS delivery can be achieved through the more recently investigated NTB route (fully reviewed in [226–228]), which notably bypasses the BBB. This may avoid the seeming contradiction in systemic delivery associated with trying to avoid phagocytosis in the periphery, while trying to target this property in CNS. NTB is not without challenges, however, and small administration volumes, restricted particle size and NP mucoadhesive properties must all be considered. Promisingly, respective targeting of microglia and macrophages by NTB administered NP/MPS has been demonstrated in both an LPS induced neuroinflammation model of Parkinson's

Disease and HIV infection [198,229]. Regarding MS and associated animal models, the NTB route is under investigation in terms of both naked [230,231] and NP encapsulated therapeutics for both existing and novel therapeutics, as explored above [185,187,232], and could offer a novel avenue for MS centric therapeutics.

5. Concluding Remarks

In conclusion, peripherally derived macrophages play a dominant role in MS onset, progression and repair. The aforementioned studies suggest that modulating the polarisation status of macrophages with NP/MP-loaded drugs to enhance an M2-“switched” state could represent a valid and partially unexplored area of research. It is worth noting that future studies embarking in NP/MP administration in MS should seriously consider assessing the macrophage uptake and polarisation states as it is highly likely that this phenomenon contributes to overall efficacy in disease progression.

Funding: This research was funded by Science Foundation Ireland, grant number 16/FRL/3855, and The Irish Research Council, grant number GOIPD/2018/875.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. MS Prevalence: National Multiple Sclerosis Society. Available online: <https://www.nationalmssociety.org/About-the-Society/MS-Prevalence> (accessed on 23 October 2018).
2. Pugliatti, M.; Rosati, G.; Carton, H.; Riise, T.; Drulovic, J.; Vecsei, L.; Milanov, I. The Epidemiology of Multiple Sclerosis in Europe. *Eur. J. Neurol.* **2006**, *13*, 700–722. [[CrossRef](#)] [[PubMed](#)]
3. Compston, A.; Coles, A. Multiple Sclerosis. *Lancet* **2008**, *372*, 1502–1517. [[CrossRef](#)]
4. Beecham, A.H.; Patsopoulos, N.A.; Xifara, D.K.; Davis, M.F.; Kempainen, A.; Cotsapas, C.; Shah, T.S.; Spencer, C.; Booth, D.; Goris, A.; et al. Analysis of Immune-Related Loci Identifies 48 New Susceptibility Variants for Multiple Sclerosis. *Nat. Genet.* **2013**, *45*, 1353–1360. [[CrossRef](#)] [[PubMed](#)]
5. Belbasis, L.; Bellou, V.; Evangelou, E.; Ioannidis, J.P.A.; Tzoulaki, I. Environmental Risk Factors and Multiple Sclerosis: An Umbrella Review of Systematic Reviews and Meta-Analyses. *Lancet Neurol.* **2015**, *14*, 263–273. [[CrossRef](#)]
6. Hedström, A.K.; Åkerstedt, T.; Hillert, J.; Olsson, T.; Alfredsson, L. Shift Work at Young Age Is Associated with Increased Risk for Multiple Sclerosis. *Ann. Neurol.* **2011**, *70*, 733–741. [[CrossRef](#)] [[PubMed](#)]
7. Balk, L.; Tewarie, P.; Killestein, J.; Polman, C.; Uitdehaag, B.; Petzold, A. Disease Course Heterogeneity and OCT in Multiple Sclerosis. *Mult. Scler. J.* **2014**, *20*, 1198–1206. [[CrossRef](#)]
8. Lund, C.; Nakken, K.O.; Edland, A.; Celius, E.G. Multiple Sclerosis and Seizures: Incidence and Prevalence over 40 Years. *Acta Neurol. Scand.* **2014**, *130*, 368–373. [[CrossRef](#)]
9. Dendrou, C.A.; Fugger, L.; Friese, M.A. Immunopathology of Multiple Sclerosis. *Nat. Rev. Immunol.* **2015**, *15*, 545–558. [[CrossRef](#)]
10. McFarland, H.F.; Martin, R. Multiple Sclerosis: A Complicated Picture of Autoimmunity. *Nat. Immunol.* **2007**, *8*, 913–919. [[CrossRef](#)]
11. Fletcher, J.M.; Lalor, S.J.; Sweeney, C.M.; Tubridy, N.; Mills, K.H.G. T Cells in Multiple Sclerosis and Experimental Autoimmune Encephalomyelitis. *Clin. Exp. Immunol.* **2010**, *162*, 1–11. [[CrossRef](#)]
12. Kaskow, B.J.; Baecher-Allan, C. Effector T Cells in Multiple Sclerosis. *Cold Spring Harb. Perspect. Med.* **2018**, *8*, a029025. [[CrossRef](#)] [[PubMed](#)]
13. Friese, M.A.; Fugger, L. Pathogenic CD8+ T Cells in Multiple Sclerosis. *Ann. Neurol.* **2009**, *66*, 132–141. [[CrossRef](#)] [[PubMed](#)]
14. Li, R.; Patterson, K.R.; Bar-Or, A. Reassessing B Cell Contributions in Multiple Sclerosis. *Nat. Immunol.* **2018**, *19*, 696–707. [[CrossRef](#)] [[PubMed](#)]
15. Pröbstel, A.-K.; Sanderson, N.; Derfuss, T.; Pröbstel, A.-K.; Sanderson, N.S.R.; Derfuss, T. B Cells and Autoantibodies in Multiple Sclerosis. *Int. J. Mol. Sci.* **2015**, *16*, 16576–16592. [[CrossRef](#)] [[PubMed](#)]
16. Mishra, M.K.; Yong, V.W. Myeloid Cells—Targets of Medication in Multiple Sclerosis. *Nat. Rev. Neurol.* **2016**, *12*, 539–551. [[CrossRef](#)] [[PubMed](#)]

17. Henderson, A.P.D.; Barnett, M.H.; Parratt, J.D.E.; Prineas, J.W. Multiple Sclerosis: Distribution of Inflammatory Cells in Newly Forming Lesions. *Ann. Neurol.* **2009**, *66*, 739–753. [[CrossRef](#)] [[PubMed](#)]
18. Lucchinetti, C.; Bruck, W.; Parisi, J.; Scheithauer, B.; Rodriguez, M.; Lassmann, H. Heterogeneity of Multiple Sclerosis Lesions. *Ann. Neurol.* **2000**, *47*, 707–717. [[CrossRef](#)]
19. Bitsch, A.; Schuchardt, J.; Bunkowski, S.; Kuhlmann, T.; Bruck, W. Acute Axonal Injury in Multiple Sclerosis Correlation with Demyelination and Inflammation. *Brain* **2000**, *123*, 1174–1183. [[CrossRef](#)] [[PubMed](#)]
20. Frischer, J.M.; Bramow, S.; Dal-Bianco, A.; Lucchinetti, C.F.; Rauschka, H.; Schmidbauer, M.; Laursen, H.; Sorensen, P.S.; Lassmann, H. The Relation between Inflammation and Neurodegeneration in Multiple Sclerosis Brains. *Brain* **2009**, *132*, 1175–1189. [[CrossRef](#)]
21. Brück, W.; Sommermeier, N.; Bergmann, M.; Zettl, U.; Goebel, H.H.; Kretzschmar, H.A.; Lassmann, H. Macrophages in Multiple Sclerosis. *Immunobiology* **1996**, *195*, 588–600. [[CrossRef](#)]
22. Hill, K.; Zollinger, L.; Watt, H.; Carlson, N.; Rose, J. Inducible Nitric Oxide Synthase in Chronic Active Multiple Sclerosis Plaques: Distribution, Cellular Expression and Association with Myelin Damage. *Neuroimmunology* **2004**, *151*, 171–179. [[CrossRef](#)] [[PubMed](#)]
23. Kuhlmann, T.; Lingfeld, G.; Bitsch, A.; Schuchardt, J.; Brück, W. Acute Axonal Damage in Multiple Sclerosis Is Most Extensive in Early Disease Stages and Decreases over Time. *Brain* **2002**, *125*, 2202–2212. [[CrossRef](#)] [[PubMed](#)]
24. Ferguson, B.; Matyszak, M.K.; Esiri, M.M.; Perry, V.H. Axonal Damage in Acute Multiple Sclerosis Lesions. *Brain* **1997**, *120*, 393–399. [[CrossRef](#)] [[PubMed](#)]
25. Lassmann, H.; Bradl, M. Multiple Sclerosis: Experimental Models and Reality. *Acta Neuropathol.* **2017**, *133*, 223–244. [[CrossRef](#)] [[PubMed](#)]
26. Terry, R.L.; Ifergan, I.; Miller, S.D. Experimental Autoimmune Encephalomyelitis in Mice. *Methods Mol. Biol.* **2016**, *1304*, 145–160. [[CrossRef](#)] [[PubMed](#)]
27. Huseby, E.S.; Liggitt, D.; Brabb, T.; Schnabel, B.; Öhlén, C.; Goverman, J. A Pathogenic Role for Myelin-Specific CD8(+) T Cells in a Model for Multiple Sclerosis. *J. Exp. Med.* **2001**, *194*, 669–676. [[CrossRef](#)] [[PubMed](#)]
28. Sun, D.; Whitaker, J.N.; Huang, Z.; Liu, D.; Coleclough, C.; Wekerle, H.; Raine, C.S. Myelin Antigen-Specific CD8+ T Cells Are Encephalitogenic and Produce Severe Disease in C57BL/6 Mice. *J. Immunol.* **2001**, *166*, 7579–7587. [[CrossRef](#)] [[PubMed](#)]
29. Bjelobaba, I.; Begovic-Kupresanin, V.; Pekovic, S.; Lavrnja, I. Animal Models of Multiple Sclerosis: Focus on Experimental Autoimmune Encephalomyelitis. *J. Neurosci. Res.* **2018**, *96*, 1021–1042. [[CrossRef](#)] [[PubMed](#)]
30. Kroenke, M.A.; Carlson, T.J.; Andjelic, A.V.; Segal, B.M. IL-12- and IL-23-Modulated T Cells Induce Distinct Types of EAE Based on Histology, CNS Chemokine Profile, and Response to Cytokine Inhibition. *J. Exp. Med.* **2008**, *205*, 1535–1541. [[CrossRef](#)] [[PubMed](#)]
31. Robinson, A.P.; Harp, C.T.; Noronha, A.; Miller, S.D. The Experimental Autoimmune Encephalomyelitis (EAE) Model of MS. In *Handbook of Clinical Neurology*; Elsevier: Amsterdam, The Netherlands, 2014; Volume 122, pp. 173–189. [[CrossRef](#)]
32. Teitelbaum, D.; Meshorer, A.; Hirshfeld, T.; Arnon, R.; Sela, M. Suppression of Experimental Allergic Encephalomyelitis by a Synthetic Polypeptide. *Eur. J. Immunol.* **1971**, *1*, 242–248. [[CrossRef](#)] [[PubMed](#)]
33. Yednock, T.A.; Cannon, C.; Fritz, L.C.; Sanchez-Madrid, F.; Steinman, L.; Karin, N. Prevention of Experimental Autoimmune Encephalomyelitis by Antibodies against A4 β 1 Integrin. *Nature* **1992**, *356*, 63–66. [[CrossRef](#)] [[PubMed](#)]
34. Fujino, M. Amelioration of Experimental Autoimmune Encephalomyelitis in Lewis Rats by FTY720 Treatment. *J. Pharmacol. Exp. Ther.* **2003**, *305*, 70–77. [[CrossRef](#)] [[PubMed](#)]
35. Kipp, M.; Nyamoya, S.; Hochstrasser, T.; Amor, S. Multiple Sclerosis Animal Models: A Clinical and Histopathological Perspective. *Brain Pathol.* **2017**, *27*, 123–137. [[CrossRef](#)] [[PubMed](#)]
36. Committee for Medicinal Products for Human Use (CHMP). *Guideline on Clinical Investigation of Medicinal Products for the Treatment of Multiple Sclerosis*; CHMP/771815/2011 Rev. 2; European Medicines Agency: London, UK, 2015; p. 14.
37. Polman, C.H.; Dijkstra, C.D.; Sminia, T.; Koetsier, J.C. Immunohistological Analysis of Macrophages in the Central Nervous System of Lewis Rats with Acute Experimental Allergic Encephalomyelitis. *J. Neuroimmunol.* **1986**, *11*, 215–222. [[CrossRef](#)]
38. Nimmerjahn, A.; Kirchhoff, F.; Helmchen, F. Resting Microglial Cells Are Highly Dynamic Surveillants of Brain Parenchyma in Vivo. *Science* **2005**, *308*, 1314–1318. [[CrossRef](#)] [[PubMed](#)]

39. Ginhoux, F.; Greter, M.; Leboeuf, M.; Nandi, S.; See, P.; Gokhan, S.; Mehler, M.F.; Conway, S.J.; Ng, L.G.; Stanley, E.R.; et al. Microglia Derive from Primitive Macrophages. *Science* **2010**, *701*, 841–845. [[CrossRef](#)] [[PubMed](#)]
40. Schulz, C.; Perdiguero, E.G.; Chorro, L.; Szabo-Rogers, H.; Cagnard, N.; Kierdorf, K.; Prinz, M.; Wu, B.; Jacobsen, S.E.W.; Pollard, J.W.; et al. A Lineage of Myeloid Cells Independent of Myb and Hematopoietic Stem Cells. *Science* **2012**, *336*, 86–90. [[CrossRef](#)]
41. Ajami, B.; Bennett, J.L.; Kriegler, C.; Tetzlaff, W.; Rossi, F.M.V. Local Self-Renewal Can Sustain CNS Microglia Maintenance and Function throughout Adult Life. *Nat. Neurosci.* **2007**, *10*, 1538–1543. [[CrossRef](#)]
42. Goldmann, T.; Wieghofer, P.; Jordão, M.J.C.; Prutek, F.; Hagemeyer, N.; Frenzel, K.; Amann, L.; Staszewski, O.; Kierdorf, K.; Krueger, M.; et al. Origin, Fate and Dynamics of Macrophages at Central Nervous System Interfaces. *Nat. Immunol.* **2016**, *17*, 797–805. [[CrossRef](#)]
43. Heppner, F.L.; Greter, M.; Marino, D.; Falsig, J.; Raivich, G.; Hövelmeyer, N.; Waisman, A.; Rüllicke, T.; Prinz, M.; Priller, J.; et al. Experimental Autoimmune Encephalomyelitis Repressed by Microglial Paralysis. *Nat. Med.* **2005**, *11*, 146–152. [[CrossRef](#)]
44. Ponomarev, E.D.; Shriver, L.P.; Maresz, K.; Dittel, B.N. Microglial Cell Activation and Proliferation Precedes the Onset of CNS Autoimmunity. *J. Neurosci. Res.* **2005**, *81*, 374–389. [[CrossRef](#)] [[PubMed](#)]
45. Ajami, B.; Bennett, J.L.; Kriegler, C.; McNagny, K.M.; Rossi, F.M.V. Infiltrating Monocytes Trigger EAE Progression, but Do Not Contribute to the Resident Microglia Pool. *Nat. Neurosci.* **2011**, *14*, 1142–1150. [[CrossRef](#)] [[PubMed](#)]
46. Brosnan, C.F.; Bornstein, M.B.; Bloom, B.R. The Effects of Macrophage Depletion on the Clinical and Pathologic Expression of Experimental Allergic Encephalomyelitis. *J. Immunol.* **1981**, *126*, 614–620. [[PubMed](#)]
47. Huitinga, I.; van Rooijen, N.; de Groot, C.J.; Uitdehaag, B.M.; Dijkstra, C.D. Suppression of Experimental Allergic Encephalomyelitis in Lewis Rats after Elimination of Macrophages. *J. Exp. Med.* **1990**, *172*, 1025–1033. [[CrossRef](#)] [[PubMed](#)]
48. Tran, E.H.; Hoekstra, K.; Van Rooijen, N.; Dijkstra, D.; Owens, T. Immune Invasion of the Central Nervous System Parenchyma and Experimental Allergic Encephalomyelitis, But Not Leukocyte Extravasation from Blood, Are Prevented in Macrophage-Depleted Mice. *J. Immunol.* **1998**, *161*, 3767–3775. [[PubMed](#)]
49. Moreno, M.A.; Burns, T.; Yao, P.; Miers, L.; Pleasure, D.; Soulika, A.M. Therapeutic Depletion of Monocyte-Derived Cells Protects from Long-Term Axonal Loss in Experimental Autoimmune Encephalomyelitis. *J. Neuroimmunol.* **2016**, *290*, 36–46. [[CrossRef](#)] [[PubMed](#)]
50. Lewis, N.D.; Hill, J.D.; Juchem, K.W.; Stefanopoulos, D.E.; Modis, L.K. RNA Sequencing of Microglia and Monocyte-Derived Macrophages from Mice with Experimental Autoimmune Encephalomyelitis Illustrates a Changing Phenotype with Disease Course. *J. Neuroimmunol.* **2014**, *277*, 26–38. [[CrossRef](#)] [[PubMed](#)]
51. Barnett, M.H.; Prineas, J.W. Relapsing and Remitting Multiple Sclerosis: Pathology of the Newly Forming Lesion. *Ann. Neurol.* **2004**, *55*, 458–468. [[CrossRef](#)] [[PubMed](#)]
52. Howell, O.W.; Rundle, J.L.; Garg, A.; Komada, M.; Brophy, P.J.; Reynolds, R. Activated Microglia Mediate Axoglial Disruption That Contributes to Axonal Injury in Multiple Sclerosis. *J. Neuropathol. Exp. Neurol.* **2010**, *69*, 1017–1033. [[CrossRef](#)] [[PubMed](#)]
53. Van Noort, J.M.; van den Elsen, P.J.; van Horssen, J.; Geurts, J.J.G.; van der Valk, P.; Amor, S. Preactive Multiple Sclerosis Lesions Offer Novel Clues for Neuroprotective Therapeutic Strategies. *CNS Neurol. Disord. Drug Targets* **2011**, *10*, 68–81. [[CrossRef](#)]
54. Vogel, D.Y.S.; Vereyken, E.J.F.; Glim, J.E.; Heijnen, P.D.A.M.; Moeton, M.; van der Valk, P.; Amor, S.; Teunissen, C.E.; van Horssen, J.; Dijkstra, C.D. Macrophages in Inflammatory Multiple Sclerosis Lesions Have an Intermediate Activation Status. *J. Neuroinflammation* **2013**, *10*, 1. [[CrossRef](#)] [[PubMed](#)]
55. Izikson, L.; Klein, R.S.; Charo, I.F.; Weiner, H.L.; Luster, A.D. Resistance to Experimental Autoimmune Encephalomyelitis in Mice Lacking the CC Chemokine Receptor (CCR)2. *J. Exp. Med.* **2000**, *192*, 1075–1080. [[CrossRef](#)] [[PubMed](#)]
56. Fife, B.T.; Huffnagle, G.B.; Kuziel, W.A.; Karpus, W.J. CC Chemokine Receptor 2 Is Critical for Induction of Experimental Autoimmune Encephalomyelitis. *J. Exp. Med.* **2000**, *192*, 899–905. [[CrossRef](#)] [[PubMed](#)]
57. Gaupp, S.; Pitt, D.; Kuziel, W.A.; Cannella, B.; Raine, C.S. Experimental Autoimmune Encephalomyelitis (EAE) in CCR2^{-/-} Mice: Susceptibility in Multiple Strains. *Am. J. Pathol.* **2003**, *162*, 139–150. [[CrossRef](#)]

58. Huang, D.; Wang, J.; Kivisakk, P.; Rollins, B.J.; Ransohoff, R.M. Absence of Monocyte Chemoattractant Protein 1 in Mice Leads to Decreased Local Macrophage Recruitment and Antigen-Specific T Helper Cell Type 1 Immune Response in Experimental Autoimmune Encephalomyelitis. *J. Exp. Med.* **2001**, *193*, 713–726. [[CrossRef](#)] [[PubMed](#)]
59. Mildner, A.; Mack, M.; Schmidt, H.; Bruck, W.; Djukic, M.; Zabel, M.; Hille, A.; Priller, J.; Prinz, M. CCR2+Ly-6Chi Monocytes Are Crucial for the Effector Phase of Autoimmunity in the Central Nervous System. *Brain* **2009**, *132*, 2487–2500. [[CrossRef](#)] [[PubMed](#)]
60. Gordon, S.; Taylor, P.R. Monocyte and Macrophage Heterogeneity. *Nat. Rev. Immunol.* **2005**, *5*, 953–964. [[CrossRef](#)] [[PubMed](#)]
61. King, I.L.; Dickendesher, T.L.; Segal, B.M. Circulating Ly-6C+ Myeloid Precursors Migrate to the CNS and Play a Pathogenic Role during Autoimmune Demyelinating Disease. *Blood* **2009**, *113*, 3190–3197. [[CrossRef](#)] [[PubMed](#)]
62. Codarri, L.; Gyölvéshi, G.; Tosevski, V.; Hesske, L.; Fontana, A.; Magnenat, L.; Suter, T.; Becher, B. ROR γ t Drives Production of the Cytokine GM-CSF in Helper T Cells, Which Is Essential for the Effector Phase of Autoimmune Neuroinflammation. *Nat. Immunol.* **2011**, *12*, 560–567. [[CrossRef](#)] [[PubMed](#)]
63. McQualter, J.L.; Darwiche, R.; Ewing, C.; Onuki, M.; Kay, T.W.; Hamilton, J.A.; Reid, H.H.; Bernard, C.C.A. Granulocyte Macrophage Colony-Stimulating Factor: A New Putative Therapeutic Target in Multiple Sclerosis. *J. Exp. Med.* **2001**, *194*, 873–882. [[CrossRef](#)]
64. Croxford, A.L.; Lanzinger, M.; Hartmann, F.J.; Schreiner, B.; Mair, F.; Pelczar, P.; Clausen, B.E.; Jung, S.; Greter, M.; Becher, B. The Cytokine GM-CSF Drives the Inflammatory Signature of CCR2+ Monocytes and Licenses Autoimmunity. *Immunity* **2015**, *43*, 502–514. [[CrossRef](#)] [[PubMed](#)]
65. Ziegler-Heitbrock, L.; Ancuta, P.; Crowe, S.; Dalod, M.; Grau, V.; Hart, D.N.; Leenen, P.J.M.; Liu, Y.-J.; MacPherson, G.; Randolph, G.J.; et al. Nomenclature of Monocytes and Dendritic Cells in Blood. *Blood* **2010**, *116*, e74–e80. [[CrossRef](#)] [[PubMed](#)]
66. Bergh, F.T.; Dayyani, F.; Ziegler-Heitbrock, L. Impact of Type-I-Interferon on Monocyte Subsets and Their Differentiation to Dendritic Cells: An in Vivo and Ex Vivo Study in Multiple Sclerosis Patients Treated with Interferon-Beta. *J. Neuroimmunol.* **2004**, *146*, 176–188. [[CrossRef](#)]
67. Chuluundorj, D.; Harding, S.A.; Abernethy, D.; La Flamme, A.C. Glatiramer Acetate Treatment Normalized the Monocyte Activation Profile in MS Patients to That of Healthy Controls. *Immunol. Cell Biol.* **2017**, *95*, 297–305. [[CrossRef](#)] [[PubMed](#)]
68. Chuluundorj, D.; Harding, S.A.; Abernethy, D.; La Flamme, A.C. Expansion and Preferential Activation of the CD14+ CD16+ Monocyte Subset during Multiple Sclerosis. *Immunol. Cell Biol.* **2014**, *92*, 509–517. [[CrossRef](#)] [[PubMed](#)]
69. Ingersoll, M.; Spanbroek, R.; Lottaz, C.; Gautier, E.; Frankenberger, M.; Hoffman, R.; Lang, R.; Hannifa, M.; Collin, M.; Tacke, F.; et al. Comparison of Gene Expression Profiles between Human and Mouse Monocyte Subsets. *Blood* **2010**, *115*, e10–e19. [[CrossRef](#)]
70. Simpson, J.; Newcombe, J.; Cuzner, M.; Woodroffe, M. Expression of Monocyte Chemoattractant Protein-1 and Other β -Chemokines by Resident Glia and Inflammatory Cells in Multiple Sclerosis Lesions. *J. Neuroimmunol.* **1998**, *84*, 238–249. [[CrossRef](#)]
71. McManus, C.; Berman, J.W.; Brett, F.M.; Staunton, H.; Farrell, M.; Brosnan, C.F. MCP-1, MCP-2 and MCP-3 Expression in Multiple Sclerosis Lesions: An Immunohistochemical and in Situ Hybridization Study. *J. Neuroimmunol.* **1998**, *86*, 20–29. [[CrossRef](#)]
72. Prins, M.; Dutta, R.; Baselmans, B.; Brevé, J.J.P.; Bol, J.G.J.M.; Deckard, S.A.; van der Valk, P.; Amor, S.; Trapp, B.D.; de Vries, H.E.; et al. Discrepancy in CCL2 and CCR2 Expression in White versus Grey Matter Hippocampal Lesions of Multiple Sclerosis Patients. *Acta Neuropathol. Commun.* **2014**, *2*, 98. [[CrossRef](#)]
73. Van Der Voorn, P.; Tekstra, J.; Beelen, R.H.J.; Tensen, C.P.; Van Der Valk, P.; De Groot, C.J.A. Expression of MCP-1 by Reactive Astrocytes in Demyelinating Multiple Sclerosis Lesions. *Am. J. Pathol.* **1999**, *154*, 45–51. [[CrossRef](#)]
74. Sorensen, T.L.; Ransohoff, R.M.; Strieter, R.M.; Sellebjerg, F. Chemokine CCL2 and Chemokine Receptor CCR2 in Early Active Multiple Sclerosis. *Eur. J. Neurol.* **2004**, *11*, 445–449. [[CrossRef](#)] [[PubMed](#)]
75. Mahad, D.J.; Ransohoff, R.M. The Role of MCP-1 (CCL2) and CCR2 in Multiple Sclerosis and Experimental Autoimmune Encephalomyelitis (EAE). *Semin. Immunol.* **2003**, *15*, 23–32. [[CrossRef](#)]

76. Bose, S.; Cho, J. Role of Chemokine CCL2 and Its Receptor CCR2 in Neurodegenerative Diseases. *Arch. Pharm. Res.* **2013**, *36*, 1039–1050. [[CrossRef](#)] [[PubMed](#)]
77. Lagumersindez-Denis, N.; Wrzos, C.; Mack, M.; Winkler, A.; van der Meer, F.; Reinert, M.C.; Hollasch, H.; Flach, A.; Brühl, H.; Cullen, E.; et al. Differential Contribution of Immune Effector Mechanisms to Cortical Demyelination in Multiple Sclerosis. *Acta Neuropathol.* **2017**, *134*, 15–34. [[CrossRef](#)] [[PubMed](#)]
78. Mantovani, A.; Sica, A.; Sozzani, S.; Allavena, P.; Vecchi, A.; Locati, M. The Chemokine System in Diverse Forms of Macrophage Activation and Polarization. *Trends Immunol.* **2004**, *25*, 677–686. [[CrossRef](#)] [[PubMed](#)]
79. Girvin, A.M.; Dal Canto, M.C.; Miller, S.D. CD40/CD40L Interaction Is Essential for the Induction of EAE in the Absence of CD28-Mediated Co-Stimulation. *J. Autoimmun.* **2002**, *18*, 83–94. [[CrossRef](#)] [[PubMed](#)]
80. Bartholomäus, I.; Kawakami, N.; Odoardi, F.; Schläger, C.; Miljkovic, D.; Ellwart, J.W.; Klinkert, W.E.F.; Flügel-Koch, C.; Issekutz, T.B.; Wekerle, H.; et al. Effector T Cell Interactions with Meningeal Vascular Structures in Nascent Autoimmune CNS Lesions. *Nature* **2009**, *462*, 94–98. [[CrossRef](#)]
81. Jiang, Z.; Jiang, J.X.; Zhang, G.X. Macrophages: A Double-Edged Sword in Experimental Autoimmune Encephalomyelitis. *Immunol. Lett.* **2014**, *160*, 1722. [[CrossRef](#)]
82. Kornek, B.; Storch, M.K.; Weissert, R.; Wallstroem, E.; Stefferl, A.; Olsson, T.; Linington, C.; Schmidbauer, M.; Lassmann, H. Multiple Sclerosis and Chronic Autoimmune Encephalomyelitis: A Comparative Quantitative Study of Axonal Injury in Active, Inactive, and Remyelinated Lesions. *Am. J. Pathol.* **2000**, *157*, 267–276. [[CrossRef](#)]
83. Nikić, I.; Merkler, D.; Sorbara, C.; Brinkoetter, M.; Kreutzfeldt, M.; Bareyre, F.M.; Brück, W.; Bishop, D.; Misgeld, T.; Kerschensteiner, M. A Reversible Form of Axon Damage in Experimental Autoimmune Encephalomyelitis and Multiple Sclerosis. *Nat. Med.* **2011**, *17*, 495–499. [[CrossRef](#)]
84. Yamasaki, R.; Lu, H.; Butovsky, O.; Ohno, N.; Rietsch, A.M.; Cialic, R.; Wu, P.; Doykan, C.; Lin, J.; Cotleur, A.C.; et al. Differential Roles of Microglia and Monocytes in the Inflamed Central Nervous System. *J. Exp. Med.* **2014**, *211*, 1533–1549. [[CrossRef](#)] [[PubMed](#)]
85. López-Moratalla, N.; González, Á.; Aymerich, M.; Lopez-Zabalza, M.; Pio, R.; de Castro, P.; Santiago, E. Monocyte Inducible Nitric Oxide Synthase in Multiple Sclerosis: Regulatory Role of Nitric Oxide. *Nitric Oxide* **1997**, *1*, 95–104. [[CrossRef](#)] [[PubMed](#)]
86. Bö, L.; Dawson, T.M.; Wesselingh, S.; Möurk, S.; Choi, S.; Kong, P.A.; Hanley, D.; Trapp, B.D. Induction of Nitric Oxide Synthase in Demyelinating Regions of Multiple Sclerosis Brains. *Ann. Neurol.* **1994**, *36*, 778–786. [[CrossRef](#)] [[PubMed](#)]
87. Locatelli, G.; Theodorou, D.; Kendirli, A.; Joana, M.; Jordão, C.; Staszewski, O.; Phulphagar, K.; Cantuti-castelvetri, L.; Dagkalis, A.; Bessis, A.; et al. Mononuclear Phagocytes Locally Specify and Adapt Their Phenotype in a Multiple Sclerosis Model. *Nat. Neurosci.* **2018**, *21*, 1196–1208. [[CrossRef](#)] [[PubMed](#)]
88. Giles, D.A.; Washnock-Schmid, J.M.; Duncker, P.C.; Dahlawi, S.; Ponath, G.; Pitt, D.; Segal, B.M. Myeloid Cell Plasticity in the Evolution of Central Nervous System Autoimmunity. *Ann. Neurol.* **2018**, *83*, 131–141. [[CrossRef](#)] [[PubMed](#)]
89. Gordon, S. Alternative Activation of Macrophages. *Nat. Rev. Immunol.* **2003**, *3*, 23–35. [[CrossRef](#)] [[PubMed](#)]
90. Kennedy, M.; Torrance, D.; Picha, K.; Mohler, K. Analysis of Cytokine mRNA Expression in the Central Nervous System of Mice with Experimental Autoimmune Encephalomyelitis Reveals That IL-10 mRNA Expression Correlates with Recovery. *J. Immunol.* **1992**, *149*, 2496–2505. [[PubMed](#)]
91. Jander, S.; Pohl, J.; D’Urso, D.; Gillen, C.; Stoll, G. Time Course and Cellular Localization of Interleukin-10 mRNA and Protein Expression in Autoimmune Inflammation of the Rat Central Nervous System. *Am. J. Pathol.* **1998**, *152*, 975–982. [[PubMed](#)]
92. Payne, N.L.; Sun, G.; McDonald, C.; Moussa, L.; Emerson-Webber, A.; Loisel-Meyer, S.; Medin, J.A.; Siatskas, C.; Bernard, C.C. Human Adipose-Derived Mesenchymal Stem Cells Engineered to Secrete IL-10 Inhibit APC Function and Limit CNS Autoimmunity. *Brain. Behav. Immun.* **2013**, *30*, 103–114. [[CrossRef](#)] [[PubMed](#)]
93. Fiorentino, D.F.; Zlotnik, A.; Vieira, P.; Mosmann, T.R.; Howard, M.; Moore, K.W.; O’Garra, A. IL-10 Acts on the Antigen-Presenting Cell to Inhibit Cytokine Production by Th1 Cells. *J. Immunol.* **1991**, *146*, 3444–3451. [[PubMed](#)]
94. O’Garra, A.; Vieira, P.L.; Vieira, P.; Goldfeld, A.E. IL-10-producing and Naturally Occurring CD4+ Tregs: Limiting Collateral Damage. *J. Clin. Invest.* **2004**, *114*, 1372–1378. [[CrossRef](#)] [[PubMed](#)]

95. Greenhalgh, A.D.; Passos dos Santos, R.; Zarruk, J.G.; Salmon, C.K.; Kroner, A.; David, S. Arginase-1 Is Expressed Exclusively by Infiltrating Myeloid Cells in CNS Injury and Disease. *Brain. Behav. Immun.* **2016**, *56*, 61–67. [[CrossRef](#)] [[PubMed](#)]
96. Boven, L.A.; Van Meurs, M.; Van Zwam, M.; Wierenga-Wolf, A.; Hintzen, R.Q.; Boot, R.G.; Aerts, J.M.; Amor, S.; Nieuwenhuis, E.E.; Laman, J.D. Myelin-Laden Macrophages Are Anti-Inflammatory, Consistent with Foam Cells in Multiple Sclerosis. *Brain* **2006**, *129*, 517–526. [[CrossRef](#)] [[PubMed](#)]
97. Hulshof, S.; Montagne, L.; De Groot, C.J.A.; Van Der Valk, P. Cellular Localization and Expression Patterns of Interleukin-10, Interleukin-4, and Their Receptors in Multiple Sclerosis Lesions. *Glia* **2002**, *38*, 24–35. [[CrossRef](#)] [[PubMed](#)]
98. Tierney, J.B.; Kharrang, M.; La Flamme, A.C. Type II-Activated Macrophages Suppress the Development of Experimental Autoimmune Encephalomyelitis. *Immunol. Cell Biol.* **2009**, *87*, 235–240. [[CrossRef](#)] [[PubMed](#)]
99. Jiang, H.-R.; Milovanović, M.; Allan, D.; Niedbala, W.; Besnard, A.-G.; Fukada, S.Y.; Alves-Filho, J.C.; Togbe, D.; Goodyear, C.S.; Linington, C.; et al. IL-33 Attenuates EAE by Suppressing IL-17 and IFN- γ Production and Inducing Alternatively Activated Macrophages. *Eur. J. Immunol.* **2012**, *42*, 1804–1814. [[CrossRef](#)] [[PubMed](#)]
100. Mikita, J.; Dubourdieu-Cassagno, N.; Deloire, M.S.; Vekris, A.; Biran, M.; Raffard, G.; Brochet, B.; Canron, M.H.; Franconi, J.M.; Boiziau, C.; et al. Altered M1/M2 Activation Patterns of Monocytes in Severe Relapsing Experimental Rat Model of Multiple Sclerosis. Amelioration of Clinical Status by M2 Activated Monocyte Administration. *Mult. Scler. J.* **2011**, *17*, 2–15. [[CrossRef](#)] [[PubMed](#)]
101. Kotter, M.R.; Zhao, C.; Van Rooijen, N.; Franklin, R.J.M. Macrophage-Depletion Induced Impairment of Experimental CNS Remyelination Is Associated with a Reduced Oligodendrocyte Progenitor Cell Response and Altered Growth Factor Expression. *Neurobiol. Dis.* **2005**, *18*, 166–175. [[CrossRef](#)]
102. Miron, V.E.; Boyd, A.; Zhao, J.W.; Yuen, T.J.; Ruckh, J.M.; Shadrach, J.L.; Van Wijngaarden, P.; Wagers, A.J.; Williams, A.; Franklin, R.J.M.; et al. M2 Microglia and Macrophages Drive Oligodendrocyte Differentiation during CNS Remyelination. *Nat. Neurosci.* **2013**, *16*, 1211–1218. [[CrossRef](#)]
103. Mokhtarian, F.; Shi, Y.; Shirazian, D.; Morgante, L.; Miller, A.; Grob, D. Defective Production of Anti-Inflammatory Cytokine, TGF-Beta by T Cell Lines of Patients with Active Multiple Sclerosis. *J. Immunol.* **1994**, *152*, 6003–6010.
104. Carrieri, P.B.; Provitera, V.; Bruno, R.; Perrella, M.; Tartaglia, G.; Busto, A.; Perrella, O. Possible Role of Transforming Growth Factor-Beta in Relapsing-Remitting Multiple Sclerosis. *Neurol. Res.* **1997**, *19*, 599–600. [[CrossRef](#)] [[PubMed](#)]
105. Raine, C.S. Multiple Sclerosis: The Resolving Lesion Revealed. *J. Neuroimmunol.* **2017**, *304*, 2–6. [[CrossRef](#)] [[PubMed](#)]
106. Wingerchuk, D.M.; Carter, J.L. Multiple Sclerosis: Current and Emerging Disease-Modifying Therapies and Treatment Strategies. *Mayo Clin. Proc.* **2014**, *89*, 225–240. [[CrossRef](#)] [[PubMed](#)]
107. Cross, A.H.; Naismith, R.T. Established and Novel Disease-Modifying Treatments in Multiple Sclerosis. *J. Intern. Med.* **2014**, *275*, 350–363. [[CrossRef](#)] [[PubMed](#)]
108. Polman, C.H.; O'Connor, P.W.; Havrdova, E.; Hutchinson, M.; Kappos, L.; Miller, D.H.; Phillips, J.T.; Lublin, F.D.; Giovannoni, G.; Wajgt, A.; et al. A Randomized, Placebo-Controlled Trial of Natalizumab for Relapsing Multiple Sclerosis. *N. Engl. J. Med.* **2006**, *354*, 899–910. [[CrossRef](#)] [[PubMed](#)]
109. Coles, A.J.; Twyman, C.L.; Arnold, D.L.; Cohen, J.A.; Confavreux, C.; Fox, E.J.; Hartung, H.-P.; Havrdova, E.; Selmaj, K.W.; Weiner, H.L.; et al. Alemtuzumab for Patients with Relapsing Multiple Sclerosis after Disease-Modifying Therapy: A Randomised Controlled Phase 3 Trial. *Lancet (London, England)* **2012**, *380*, 1829–1839. [[CrossRef](#)]
110. US Food and Drug Administration. Drug Safety and Availability—FDA Working with Manufacturers to Withdraw Zinbryta from the Market in the United States. US Department of Health and Human Services. Available online: <https://www.fda.gov/drugs/drug-safety-and-availability/fda-working-manufacturers-withdraw-zinbryta-market-united-states> (accessed on 10 April 2019).
111. O'Connor, P.; Filippi, M.; Arnason, B.; Comi, G.; Cook, S.; Goodin, D.; Hartung, H.-P.; Jeffery, D.; Kappos, L.; Boateng, F.; et al. 250 Mg or 500 Mg Interferon Beta-1b versus 20 Mg Glatiramer Acetate in Relapsing-Remitting Multiple Sclerosis: A Prospective, Randomised, Multicentre Study. *Lancet Neurol.* **2009**, *8*, 889–897. [[CrossRef](#)]
112. Dhib-Jalbut, S.; Marks, S. Interferon-Beta Mechanisms of Action in Multiple Sclerosis. *Neurology* **2010**, *74* (Suppl. 1), S17–S24. [[CrossRef](#)]

113. Guo, B.; Chang, E.Y.; Cheng, G. The Type I IFN Induction Pathway Constrains Th17-Mediated Autoimmune Inflammation in Mice. *J. Clin. Invest.* **2008**, *118*, 1680–1690. [[CrossRef](#)] [[PubMed](#)]
114. Shinohara, M.L.; Kim, J.-H.; Garcia, V.A.; Cantor, H. Engagement of the Type I Interferon Receptor on Dendritic Cells Inhibits T Helper 17 Cell Development: Role of Intracellular Osteopontin. *Immunity* **2008**, *29*, 68–78. [[CrossRef](#)]
115. Liu, B.-S.; Janssen, H.L.A.; Boonstra, A. Type I and III Interferons Enhance IL-10R Expression on Human Monocytes and Macrophages, Resulting in IL-10-Mediated Suppression of TLR-Induced IL-12. *Eur. J. Immunol.* **2012**, *42*, 2431–2440. [[CrossRef](#)] [[PubMed](#)]
116. PRISMS (Prevention of Relapses and Disability by Interferon Beta-1a Subcutaneously in Multiple Sclerosis) Study Group. Randomised Double-Blind Placebo-Controlled Study of Interferon Beta-1a in Relapsing/Remitting Multiple Sclerosis. *Lancet (London, England)* **1998**, *352*, 1498–1504.
117. Duda, P.W.; Schmied, M.C.; Cook, S.L.; Krieger, J.I.; Hafler, D.A. Glatiramer Acetate (Copaxone®) Induces Degenerate, Th2-Polarized Immune Responses in Patients with Multiple Sclerosis. *J. Clin. Investig.* **2000**, *105*, 967–976. [[CrossRef](#)] [[PubMed](#)]
118. Gran, B.; Tranquill, L.R.; Chen, M.; Bielekova, B.; Zhou, W.; Dhib-Jalbut, S.; Martin, R. Mechanisms of Immunomodulation by Glatiramer Acetate. *Neurology* **2000**, *55*, 1704–1714. [[CrossRef](#)] [[PubMed](#)]
119. Kim, H.J.; Ifergan, I.; Antel, J.P.; Seguin, R.; Duddy, M.; Lapierre, Y.; Jalili, F.; Bar-Or, A. Type 2 Monocyte and Microglia Differentiation Mediated by Glatiramer Acetate Therapy in Patients with Multiple Sclerosis. *J. Immunol.* **2004**, *172*, 7144–7153. [[CrossRef](#)] [[PubMed](#)]
120. Burger, D.; Molnarfi, N.; Weber, M.S.; Brandt, K.J.; Benkhoucha, M.; Gruaz, L.; Chofflon, M.; Zamvil, S.S.; Lalive, P.H. Glatiramer Acetate Increases IL-1 Receptor Antagonist but Decreases T Cell-Induced IL-1 in Human Monocytes and Multiple Sclerosis. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 4355–4359. [[CrossRef](#)] [[PubMed](#)]
121. Weber, M.S.; Starck, M.; Wagenpfeil, S.; Meinl, E.; Hohlfeld, R.; Farina, C. Multiple Sclerosis: Glatiramer Acetate Inhibits Monocyte Reactivity in Vitro and in Vivo. *Brain* **2004**, *127*, 1370–1378. [[CrossRef](#)] [[PubMed](#)]
122. Pul, R.; Moharreggh-Khiabani, D.; Škuljec, J.; Skripuletz, T.; Garde, N.; Voß, E.V.; Stangel, M. Glatiramer Acetate Modulates TNF- α and IL-10 Secretion in Microglia and Promotes Their Phagocytic Activity. *J. Neuroimmune Pharm.* **2011**, *6*, 381–388. [[CrossRef](#)]
123. Pul, R.; Morbiducci, F.; Škuljec, J.; Skripuletz, T.; Singh, V.; Diederichs, U.; Garde, N.; Voss, E.V.; Trebst, C.; Stangel, M. Glatiramer Acetate Increases Phagocytic Activity of Human Monocytes In Vitro and in Multiple Sclerosis Patients. *PLoS ONE* **2012**, *7*. [[CrossRef](#)]
124. Mindur, J.E.; Ito, N.; Dhib-Jalbut, S.; Ito, K. Early Treatment with Anti-VLA-4 MAb Can Prevent the Infiltration and/or Development of Pathogenic CD11b+CD4+ T Cells in the CNS during Progressive EAE. *PLoS ONE* **2014**, *9*, e99068. [[CrossRef](#)]
125. Michell-Robinson, M.A.; Moore, C.S.; Healy, L.M.; Osso, L.A.; Zorko, N.; Grouza, V.; Touil, H.; Poliquin-Lasnier, L.; Trudelle, A.M.; Giacomini, P.S.; et al. Effects of Fumarates on Circulating and CNS Myeloid Cells in Multiple Sclerosis. *Ann. Clin. Transl. Neurol.* **2016**, *3*, 27–41. [[CrossRef](#)] [[PubMed](#)]
126. Qin, C.; Fan, W.-H.; Liu, Q.; Shang, K.; Murugan, M.; Wu, L.-J.; Wang, W.; Tian, D.-S. Fingolimod Protects Against Ischemic White Matter Damage by Modulating Microglia Toward M2 Polarization via STAT3 Pathway. *Stroke* **2017**, *48*, 3336–3346. [[CrossRef](#)] [[PubMed](#)]
127. Luessi, F.; Kraus, S.; Trinschek, B.; Lerch, S.; Ploen, R.; Paterka, M.; Roberg, T.; Poisa-Beiro, L.; Klotz, L.; Wiendl, H.; et al. FTY720 (Fingolimod) Treatment Tips the Balance towards Less Immunogenic Antigen-Presenting Cells in Patients with Multiple Sclerosis. *Mult. Scler. J.* **2015**, *21*, 1811–1822. [[CrossRef](#)] [[PubMed](#)]
128. Thomas, K.; Sehr, T.; Proschmann, U.; Rodriguez-Leal, F.A.; Haase, R.; Ziemssen, T. Fingolimod Additionally Acts as Immunomodulator Focused on the Innate Immune System beyond Its Prominent Effects on Lymphocyte Recirculation. *J. Neuroinflamm.* **2017**, *14*, 41. [[CrossRef](#)] [[PubMed](#)]
129. Di Dario, M.; Colombo, E.; Govi, C.; De Feo, D.; Messina, M.J.; Romeo, M.; Sangalli, F.; Muiola, L.; Rodegher, M.; Martino, G.; et al. Myeloid Cells as Target of Fingolimod Action in Multiple Sclerosis. *Neurol.-Neuroimmunol. Neuroinflamm.* **2015**, *2*, e157. [[CrossRef](#)]
130. Cohen, J.A.; Barkhof, F.; Comi, G.; Hartung, H.-P.; Khatir, B.O.; Montalban, X.; Pelletier, J.; Capra, R.; Gallo, P.; Izquierdo, G.; et al. Oral Fingolimod or Intramuscular Interferon for Relapsing Multiple Sclerosis. *N. Engl. J. Med.* **2010**, *362*, 402–415. [[CrossRef](#)]

131. O'Connor, P.; Wolinsky, J.S.; Confavreux, C.; Comi, G.; Kappos, L.; Olsson, T.P.; Benzerdjeb, H.; Truffinet, P.; Wang, L.; Miller, A.; et al. Randomized Trial of Oral Teriflunomide for Relapsing Multiple Sclerosis. *N. Engl. J. Med.* **2011**, *365*, 1293–1303. [[CrossRef](#)]
132. Schilling, S.; Goelz, S.; Linker, R.; Luehder, F.; Gold, R. Fumaric Acid Esters Are Effective in Chronic Experimental Autoimmune Encephalomyelitis and Suppress Macrophage Infiltration. *Clin. Exp. Immunol.* **2006**, *145*, 101–107. [[CrossRef](#)]
133. Parodi, B.; Rossi, S.; Morando, S.; Cordano, C.; Bragoni, A.; Motta, C.; Usai, C.; Wipke, B.T.; Scannevin, R.H.; Mancardi, G.L.; et al. Fumarates Modulate Microglia Activation through a Novel HCAR2 Signaling Pathway and Rescue Synaptic Dysregulation in Inflamed CNS. *Acta Neuropathol.* **2015**, *130*, 279–295. [[CrossRef](#)]
134. Kornberg, M.D.; Bhargava, P.; Kim, P.M.; Putluri, V.; Snowman, A.M.; Putluri, N.; Calabresi, P.A.; Snyder, S.H. Dimethyl Fumarate Targets GAPDH and Aerobic Glycolysis to Modulate Immunity. *Science* **2018**, *360*, 449–453. [[CrossRef](#)]
135. Schulze-Topphoff, U.; Varrin-Doyer, M.; Pekarek, K.; Spencer, C.M.; Shetty, A.; Sagan, S.A.; Cree, B.A.C.; Sobel, R.A.; Wipke, B.T.; Steinman, L.; et al. Dimethyl Fumarate Treatment Induces Adaptive and Innate Immune Modulation Independent of Nrf2. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, 4777–4782. [[CrossRef](#)] [[PubMed](#)]
136. Han, R.; Xiao, J.; Zhai, H.; Hao, J. Dimethyl Fumarate Attenuates Experimental Autoimmune Neuritis through the Nuclear Factor Erythroid-Derived 2-Related Factor 2/Hemoxygenase-1 Pathway by Altering the Balance of M1/M2 Macrophages. *J. Neuroinflamm.* **2016**, *13*, 97. [[CrossRef](#)] [[PubMed](#)]
137. Gold, R.; Kappos, L.; Arnold, D.L.; Bar-Or, A.; Giovannoni, G.; Selmaj, K.; Tornatore, C.; Sweetser, M.T.; Yang, M.; Sheikh, S.I.; et al. Placebo-Controlled Phase 3 Study of Oral BG-12 for Relapsing Multiple Sclerosis. *N. Engl. J. Med.* **2012**, *367*, 1098–1107. [[CrossRef](#)] [[PubMed](#)]
138. Azevedo, C.J.; Kutz, C.; Dix, A.; Boster, A.; Sanossian, N.; Kaplan, J. Intracerebral Haemorrhage during Alemtuzumab Administration. *Lancet. Neurol.* **2019**, *18*, 329–331. [[CrossRef](#)]
139. Fox, E.J. Mechanism of Action of Mitoxantrone. *Neurology* **2004**, *63* (12 Suppl. 6), S15–S18. [[CrossRef](#)] [[PubMed](#)]
140. Kopadze, T.; Dehmel, T.; Hartung, H.-P.; Stüve, O.; Kieseier, B.C. Inhibition by Mitoxantrone of In Vitro Migration of Immunocompetent Cells. *Arch. Neurol.* **2006**, *63*, 1572. [[CrossRef](#)] [[PubMed](#)]
141. Mitoxantrone in Progressive Multiple Sclerosis: A Placebo-Controlled, Double-Blind, Randomised, Multicentre Trial. *Lancet* **2002**, *360*, 2018–2025. [[CrossRef](#)]
142. Montalban, X.; Hauser, S.L.; Kappos, L.; Arnold, D.L.; Bar-Or, A.; Comi, G.; de Seze, J.; Giovannoni, G.; Hartung, H.-P.; Hemmer, B.; et al. Ocrelizumab versus Placebo in Primary Progressive Multiple Sclerosis. *N. Engl. J. Med.* **2017**, *376*, 209–220. [[CrossRef](#)]
143. Guarda, G.; Braun, M.; Staehli, F.; Tardivel, A.; Mattmann, C.; Förster, I.; Farlik, M.; Decker, T.; Du Pasquier, R.A.; Romero, P.; et al. Type I Interferon Inhibits Interleukin-1 Production and Inflammasome Activation. *Immunity* **2011**, *34*, 213–223. [[CrossRef](#)]
144. Comabella, M.; Imitola, J.; Weiner, H.L.; Houry, S.J. Interferon-Beta Treatment Alters Peripheral Blood Monocytes Chemokine Production in MS Patients. *J. Neuroimmunol.* **2002**, *126*, 205–212. [[CrossRef](#)]
145. Rep, M.H.; Schrijver, H.M.; van Lopik, T.; Hintzen, R.Q.; Roos, M.T.; Adèr, H.J.; Polman, C.H.; van Lier, R.A. Interferon (IFN)-Beta Treatment Enhances CD95 and Interleukin 10 Expression but Reduces Interferon-Gamma Producing T Cells in MS Patients. *J. Neuroimmunol.* **1999**, *96*, 92–100. [[CrossRef](#)]
146. Ersoy, E.; Kus, C.N.S.; Sener, U.; Coker, I.; Zorlu, Y. The Effects of Interferon-Beta on Interleukin-10 in Multiple Sclerosis Patients. *Eur. J. Neurol.* **2005**, *12*, 208–211. [[CrossRef](#)] [[PubMed](#)]
147. Li, Q.; Milo, R.; Panitch, H.; Swoveland, P.; Bever, C.T. Glatiramer Acetate Blocks the Activation of THP-1 Cells by Interferon- γ . *Eur. J. Pharmacol.* **1998**, *342*, 303–310. [[CrossRef](#)]
148. Jung, S.; Siglienti, I.; Grauer, O.; Magnus, T.; Scarlato, G.; Toyka, K. Induction of IL-10 in Rat Peritoneal Macrophages and Dendritic Cells by Glatiramer Acetate. *J. Neuroimmunol.* **2004**, *148*, 63–73. [[CrossRef](#)] [[PubMed](#)]
149. Robinson, S.; Miller, R.H. Contact with Central Nervous System Myelin Inhibits Oligodendrocyte Progenitor Maturation. *Dev. Biol.* **1999**, *216*, 359–368. [[CrossRef](#)] [[PubMed](#)]
150. Spadaro, M.; Montarolo, F.; Perga, S.; Martire, S.; Brescia, F.; Malucchi, S.; Bertolotto, A. Biological Activity of Glatiramer Acetate on Treg and Anti-Inflammatory Monocytes Persists for More than 10 Years in Responder Multiple Sclerosis Patients. *Clin. Immunol.* **2017**, *181*, 83–88. [[CrossRef](#)] [[PubMed](#)]

151. Schimrigk, S.; Brune, N.; Hellwig, K.; Lukas, C.; Bellenberg, B.; Rieks, M.; Hoffmann, V.; Pohlau, D.; Przuntek, H. Oral Fumaric Acid Esters for the Treatment of Active Multiple Sclerosis: An Open-Label, Baseline-Controlled Pilot Study. *Eur. J. Neurol.* **2006**, *13*, 604–610. [[CrossRef](#)] [[PubMed](#)]
152. Linker, R.A.; Lee, D.; Ryan, S.; van Dam, A.; Conrad, R.; Bista, P.; Zeng, W.; Hronowsky, X.; Buko, A.; Chollate, S.; et al. Fumaric Acid Esters Exert Neuroprotective Effects in Neuroinflammation via Activation of the Nrf2 Antioxidant Pathway. *Brain* **2011**, *134*, 678–692. [[CrossRef](#)]
153. McGuire, V.A.; Ruiz-Zorrilla Diez, T.; Emmerich, C.H.; Strickson, S.; Ritorto, M.S.; Sutavani, R.V.; Weiβ, A.; Houslay, K.F.; Knebel, A.; Meakin, P.J.; et al. Dimethyl Fumarate Blocks Pro-Inflammatory Cytokine Production via Inhibition of TLR Induced M1 and K63 Ubiquitin Chain Formation. *Sci. Rep.* **2016**, *6*, 31159. [[CrossRef](#)]
154. Diebold, M.; Sievers, C.; Bantug, G.; Sanderson, N.; Kappos, L.; Kuhle, J.; Lindberg, R.L.P.; Derfuss, T. Dimethyl Fumarate Influences Innate and Adaptive Immunity in Multiple Sclerosis. *J. Autoimmun.* **2018**, *86*, 39–50. [[CrossRef](#)]
155. Lim, J.L.; van der Pol, S.M.A.; Di Dio, F.; van het Hof, B.; Kooij, G.; de Vries, H.E.; van Horsen, J. Protective Effects of Monomethyl Fumarate at the Inflamed Blood–brain Barrier. *Microvasc. Res.* **2016**, *105*, 61–69. [[CrossRef](#)] [[PubMed](#)]
156. Mazzola, M.A.; Raheja, R.; Regev, K.; Beynon, V.; von Glehn, F.; Paul, A.; Pierre, I.; Kivisakk, P.; Weiner, H.L.; Gandhi, R. Monomethyl Fumarate Treatment Impairs Maturation of Human Myeloid Dendritic Cells and Their Ability to Activate T Cells. *Mult. Scler. J.* **2019**, *25*, 63–71. [[CrossRef](#)] [[PubMed](#)]
157. Angiari, S.; O’Neill, L.A. Dimethyl Fumarate: Targeting Glycolysis to Treat MS. *Cell Res.* **2018**, *28*, 613–615. [[CrossRef](#)] [[PubMed](#)]
158. Klotz, L.; Havla, J.; Schwab, N.; Hohlfeld, R.; Barnett, M.; Reddel, S.; Wiendl, H. Risks and Risk Management in Modern Multiple Sclerosis Immunotherapeutic Treatment. *Ther. Adv. Neurol. Disord.* **2019**, *12*, 1756286419836571. [[CrossRef](#)] [[PubMed](#)]
159. Mundargi, R.C.; Babu, V.R.; Rangaswamy, V.; Patel, P.; Aminabhavi, T.M. Nano/Micro Technologies for Delivering Macromolecular Therapeutics Using Poly(d,l-Lactide-Co-Glycolide) and Its Derivatives. *J. Control. Release* **2008**, *125*, 193–209. [[CrossRef](#)] [[PubMed](#)]
160. Singh, A.; Talekar, M.; Raikar, A.; Amiji, M. Macrophage-Targeted Delivery Systems for Nucleic Acid Therapy of Inflammatory Diseases. *J. Control. Release* **2014**, *190*, 515–530. [[CrossRef](#)] [[PubMed](#)]
161. Zhang, H.; Bi, J.; Yi, H.; Fan, T.; Ruan, Q.; Cai, L.; Chen, Y.H.; Wan, X. Silencing C-Rel in Macrophages Dampens Th1 and Th17 Immune Responses and Alleviates Experimental Autoimmune Encephalomyelitis in Mice. *Immunol. Cell Biol.* **2017**, *95*, 593–600. [[CrossRef](#)] [[PubMed](#)]
162. Montes-Cobos, E.; Ring, S.; Fischer, H.J.; Heck, J.; Strauß, J.; Schwaninger, M.; Reichardt, S.D.; Feldmann, C.; Lühder, F.; Reichardt, H.M. Targeted Delivery of Glucocorticoids to Macrophages in a Mouse Model of Multiple Sclerosis Using Inorganic-Organic Hybrid Nanoparticles. *J. Control. Release* **2017**, *245*, 157–169. [[CrossRef](#)]
163. Schmidt, J.; Metselaar, J.M.; Wauben, M.H.M.; Toyka, K.V.; Storm, G.; Gold, R. Drug Targeting by Long-Circulating Liposomal Glucocorticosteroids Increases Therapeutic Efficacy in a Model of Multiple Sclerosis. *Brain* **2003**, *126*, 1895–1904. [[CrossRef](#)]
164. Linker, R.A.; Weller, C.; Lühder, F.; Mohr, A.; Schmidt, J.; Knauth, M.; Metselaar, J.M.; Gold, R. Liposomal Glucocorticosteroids in Treatment of Chronic Autoimmune Demyelination: Long-Term Protective Effects and Enhanced Efficacy of Methylprednisolone Formulations. *Exp. Neurol.* **2008**, *211*, 397–406. [[CrossRef](#)]
165. Turjeman, K.; Bavli, Y.; Kizelsztejn, P.; Schilt, Y.; Allon, N.; Katzir, T.B.; Sasson, E.; Raviv, U.; Ovadia, H.; Barenholz, Y. Nano-Drugs Based on Nano Sterically Stabilized Liposomes for the Treatment of Inflammatory Neurodegenerative Diseases. *PLoS ONE* **2015**, *10*, e0130442. [[CrossRef](#)] [[PubMed](#)]
166. Gaillard, P.J.; Appeldoorn, C.C.M.; Rip, J.; Dorland, R.; van der Pol, S.M.A.; Kooij, G.; de Vries, H.E.; Reijerkerk, A. Enhanced Brain Delivery of Liposomal Methylprednisolone Improved Therapeutic Efficacy in a Model of Neuroinflammation. *J. Control. Release* **2012**, *164*, 364–369. [[CrossRef](#)] [[PubMed](#)]
167. Kuo, R.; Saito, E.; Miller, S.D.; Shea, L.D. Peptide-Conjugated Nanoparticles Reduce Positive Co-Stimulatory Expression and T Cell Activity to Induce Tolerance. *Mol. Ther.* **2017**, *25*, 1676–1685. [[CrossRef](#)] [[PubMed](#)]
168. Maldonado, R.A.; LaMothe, R.A.; Ferrari, J.D.; Zhang, A.-H.; Rossi, R.J.; Kolte, P.N.; Griset, A.P.; O’Neil, C.; Altretter, D.H.; Browning, E.; et al. Polymeric Synthetic Nanoparticles for the Induction of Antigen-Specific Immunological Tolerance. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, E156–E165. [[CrossRef](#)] [[PubMed](#)]

169. McCarthy, D.P.; Yap, J.W.T.; Harp, C.T.; Song, W.K.; Chen, J.; Pearson, R.M.; Miller, S.D.; Shea, L.D. An Antigen-Encapsulating Nanoparticle Platform for TH1/17 Immune Tolerance Therapy. *Nanomed. Nanotechnol. Biol. Med.* **2017**, *13*, 191–200. [[CrossRef](#)] [[PubMed](#)]
170. Pearson, R.M.; Casey, L.M.; Hughes, K.R.; Wang, L.Z.; North, M.G.; Getts, D.R.; Miller, S.D.; Shea, L.D. Controlled Delivery of Single or Multiple Antigens in Tolerogenic Nanoparticles Using Peptide-Polymer Bioconjugates. *Mol. Ther.* **2017**, *25*, 1655–1664. [[CrossRef](#)]
171. Gholamzad, M.; Ebtekar, M.; Ardestani, M. Intravenous Injection of Myelin Oligodendrocyte Glycoprotein-Coated PLGA Microparticles Have Tolerogenic Effects in Experimental Autoimmune Encephalomyelitis. *Iran. J. Allergy Asthma Immunol.* **2017**, *16*, 271–281.
172. Yeste, A.; Nadeau, M.; Burns, E.J.; Weiner, H.L.; Quintana, F.J. Nanoparticle-Mediated Codelivery of Myelin Antigen and a Tolerogenic Small Molecule Suppresses Experimental Autoimmune Encephalomyelitis. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 11270–11275. [[CrossRef](#)]
173. Al-Ghobashy, M.A.; Elmehad, A.N.; Abdelsalam, R.M.; Nooh, M.M.; Al-Shorbagy, M.; Laible, G. Development and Pre-Clinical Evaluation of Recombinant Human Myelin Basic Protein Nano Therapeutic Vaccine in Experimental Autoimmune Encephalomyelitis Mice Animal Model. *Sci. Rep.* **2017**, *7*, 1–16. [[CrossRef](#)]
174. Cappellano, G.; Woldetsadik, A.D.; Orilieri, E.; Shivakumar, Y.; Rizzi, M.; Carniato, F.; Gigliotti, C.L.; Boggio, E.; Clemente, N.; Comi, C.; et al. Subcutaneous Inverse Vaccination with PLGA Particles Loaded with a MOG Peptide and IL-10 Decreases the Severity of Experimental Autoimmune Encephalomyelitis. *Vaccine* **2014**, *32*, 5681–5689. [[CrossRef](#)]
175. Peine, K.J.; Guerau-De-Arellano, M.; Lee, P.; Kanthamneni, N.; Severin, M.; Probst, G.D.; Peng, H.; Yang, Y.; Vangundy, Z.; Papenfuss, T.L.; et al. Treatment of Experimental Autoimmune Encephalomyelitis by Codelivery of Disease Associated Peptide and Dexamethasone in Acetalated Dextran Microparticles. *Mol. Pharm.* **2014**, *11*, 828–835. [[CrossRef](#)] [[PubMed](#)]
176. Cho, J.J.; Stewart, J.M.; Drashansky, T.T.; Brusko, M.A.; Zuniga, A.N.; Lorentsen, K.J.; Keselowsky, B.G.; Avram, D. An Antigen-Specific Semi-Therapeutic Treatment with Local Delivery of Tolerogenic Factors through a Dual-Sized Microparticle System Blocks Experimental Autoimmune Encephalomyelitis. *Biomaterials* **2017**, *143*, 79–92. [[CrossRef](#)] [[PubMed](#)]
177. Hunter, Z.; McCarthy, D.P.; Yap, W.T.; Harp, C.T.; Getts, D.R.; Shea, L.D.; Miller, S.D. A Biodegradable Nanoparticle Platform for the Induction of Antigen-Specific Immune Tolerance for Treatment of Autoimmune Disease. *ACS Nano* **2014**, *8*, 2148–2160. [[CrossRef](#)] [[PubMed](#)]
178. Getts, D.R.; Martin, A.J.; McCarthy, D.P.; Terry, R.L.; Hunter, Z.N.; Yap, W.T.; Getts, M.T.; Pleiss, M.; Luo, X.; King, N.J.C.; et al. Microparticles Bearing Encephalitogenic Peptides Induce T-Cell Tolerance and Ameliorate Experimental Autoimmune Encephalomyelitis. *Nat. Biotechnol.* **2012**, *30*, 1217–1224. [[CrossRef](#)] [[PubMed](#)]
179. Getts, D.R.; Terry, R.L.; Getts, M.T.; Deffrasnes, C.; Müller, M.; Vreden, C.; Ashhurst, T.M.; Chami, B.; McCarthy, D.; Wu, H.; et al. Therapeutic Inflammatory Monocyte Modulation Using Immune-Modifying Microparticles. *Sci. Transl. Med.* **2014**, *6*. [[CrossRef](#)] [[PubMed](#)]
180. Kishimoto, T.K.; Maldonado, R.A. Nanoparticles for the Induction of Antigen-Specific Immunological Tolerance. *Front. Immunol.* **2018**, *9*, 230. [[CrossRef](#)] [[PubMed](#)]
181. LaMothe, R.A.; Kolte, P.N.; Vo, T.; Ferrari, J.D.; Gelsing, T.C.; Wong, J.; Chan, V.T.; Ahmed, S.; Srinivasan, A.; Deitemeyer, P.; et al. Tolerogenic Nanoparticles Induce Antigen-Specific Regulatory T Cells and Provide Therapeutic Efficacy and Transferrable Tolerance against Experimental Autoimmune Encephalomyelitis. *Front. Immunol.* **2018**, *9*, 281. [[CrossRef](#)]
182. Avnir, Y.; Turjeman, K.; Tulchinsky, D.; Sigal, A.; Kizelsztejn, P.; Tzemach, D.; Gabizon, A.; Barenholz, Y. Fabrication Principles and Their Contribution to the Superior in Vivo Therapeutic Efficacy of Nano-Liposomes Remote Loaded with Glucocorticoids. *PLoS ONE* **2011**, *6*, e25721. [[CrossRef](#)]
183. Kumar, P.; Sharma, G.; Gupta, V.; Kaur, R.; Thakur, K.; Malik, R.; Kumar, A.; Kaushal, N.; Raza, K. Preclinical Explorative Assessment of Dimethyl Fumarate-Based Biocompatible Nanolipoidal Carriers for the Management of Multiple Sclerosis. *ACS Chem. Neurosci.* **2018**, *9*, 1152–1158. [[CrossRef](#)]
184. Antonazzo, I.C.; Poluzzi, E.; Forcesi, E.; Riise, T.; Bjornevik, K.; Baldin, E.; Muratori, L.; De Ponti, F.; Raschi, E. Liver Injury with Drugs Used for Multiple Sclerosis: A Contemporary Analysis of the FDA Adverse Event Reporting System. *Mult. Scler.* **2018**, 1352458518799598. [[CrossRef](#)]

185. Gadhave, D.G.; Kokare, C.R. Nanostructured Lipid Carriers Engineered for Intranasal Delivery of Teriflunomide in Multiple Sclerosis: Optimization and in Vivo Studies. *Drug Dev. Ind. Pharm.* **2019**, *45*, 839–851. [[CrossRef](#)] [[PubMed](#)]
186. Rittchen, S.; Boyd, A.; Burns, A.; Park, J.; Fahmy, T.M.; Metcalfe, S.; Williams, A. Myelin Repair in Vivo Is Increased by Targeting Oligodendrocyte Precursor Cells with Nanoparticles Encapsulating Leukaemia Inhibitory Factor (LIF). *Biomaterials* **2015**, *56*, 78–85. [[CrossRef](#)] [[PubMed](#)]
187. Youssef, A.E.H.; Dief, A.E.; El Azhary, N.M.; Abdelmonsif, D.A.; El-fetiany, O.S. LINGO-1 SiRNA Nanoparticles Promote Central Remyelination in Ethidium Bromide-Induced Demyelination in Rats. *J. Physiol. Biochem.* **2019**, *75*, 89–99. [[CrossRef](#)] [[PubMed](#)]
188. Tran, J.Q.; Rana, J.; Barkhof, F.; Melamed, I.; Gevorkyan, H.; Wattjes, M.P.; de Jong, R.; Brososky, K.; Ray, S.; Xu, L.; et al. Randomized Phase I Trials of the Safety/Tolerability of Anti-LINGO-1 Monoclonal Antibody BIIB033. *Neurol. Neuroimmunol. Neuroinflamm.* **2014**, *1*, e18. [[CrossRef](#)] [[PubMed](#)]
189. Mellion, M.; Edwards, K.R.; Hupperts, R.; Drulović, J.; Montalban, X.; Hartung, H.-P.; Brochet, B.; Calabresi, P.A.; Rudick, R.; Ibrahim, A.; et al. Efficacy Results from the Phase 2b SYNERGY Study: Treatment of Disabling Multiple Sclerosis with the Anti-LINGO-1 Monoclonal Antibody Opicinumab (S33.004). *Neurology* **2017**, *88* (16 Supplement), S33.004.
190. Eitan, E.; Hutchison, E.R.; Greig, N.H.; Tweedie, D.; Celik, H.; Ghosh, S.; Fishbein, K.W.; Spencer, R.G.; Sasaki, C.Y.; Ghosh, P.; et al. Combination Therapy with Lenalidomide and Nanoceria Ameliorates CNS Autoimmunity. *Exp. Neurol.* **2015**, *273*, 151–160. [[CrossRef](#)] [[PubMed](#)]
191. Hu, W.; Metselaar, J.; Ben, L.-H.; Cravens, P.D.; Singh, M.P.; Frohman, E.M.; Eagar, T.N.; Racke, M.K.; Kieseier, B.C.; Stüve, O. PEG Minocycline-Liposomes Ameliorate CNS Autoimmune Disease. *PLoS ONE* **2009**, *4*, e4151. [[CrossRef](#)]
192. Kohane, D.S. Microparticles and Nanoparticles for Drug Delivery. *Biotechnol. Bioeng.* **2007**, *96*, 203–209. [[CrossRef](#)]
193. Chono, S.; Tauchi, Y.; Morimoto, K. Influence of Particle Size on the Distributions of Liposomes to Atherosclerotic Lesions in Mice. *Drug Dev. Ind. Pharm.* **2006**, *32*, 125–135. [[CrossRef](#)]
194. Chono, S.; Tanino, T.; Seki, T.; Morimoto, K. Uptake Characteristics of Liposomes by Rat Alveolar Macrophages: Influence of Particle Size and Surface Mannose Modification. *J. Pharm. Pharmacol.* **2007**, *59*, 75–80. [[CrossRef](#)]
195. Lawlor, C.; O'sullivan, M.P.; Sivadas, N.; O'leary, S.; Gallagher, P.J.; Keane, J.; Cryan, S.-A. The Application of High-Content Analysis in the Study of Targeted Particulate Delivery Systems for Intracellular Drug Delivery to Alveolar Macrophages. *Mol. Pharm.* **2011**, *8*, 1101–1112. [[CrossRef](#)]
196. Yue, H.; Wei, W.; Yue, Z.; Lv, P.; Wang, L.; Ma, G.; Su, Z. Particle Size Affects the Cellular Response in Macrophages. *Eur. J. Pharm. Sci.* **2010**, *41*, 650–657. [[CrossRef](#)]
197. Patel, B.K.; Parikh, R.H.; Patel, N. Targeted Delivery of Mannosylated-PLGA Nanoparticles of Antiretroviral Drug to Brain. *Int. J. Nanomed.* **2018**, *13*, 97–100. [[CrossRef](#)]
198. Yadav, S.; Gandham, S.K.; Panicucci, R.; Amiji, M.M. Intranasal Brain Delivery of Cationic Nanoemulsion-Encapsulated TNF α SiRNA in Prevention of Experimental Neuroinflammation. *Nanomed. Nanotechnol. Biol. Med.* **2016**, *12*, 987–1002. [[CrossRef](#)]
199. Gao, W.; Li, J. Targeted SiRNA Delivery Reduces Nitric Oxide Mediated Cell Death after Spinal Cord Injury. *J. Nanobiotechnol.* **2017**, *15*, 1–11. [[CrossRef](#)]
200. Zhou, Y.L.; Zhang, L.; Zhou, Z.; Liu, W.; Lu, Y.; He, S.; Cui, Y.; Qin, Y.; Hua, M. Antibody Modified Nanoparticle-Mediated Delivery of MiR-124 Regulates Apoptosis via Repression the Stat3 Signal in Mycobacterial-Infected Microglia. *J. Biomed. Nanotechnol.* **2018**, *14*, 2185–2197. [[CrossRef](#)] [[PubMed](#)]
201. Singh, R.; Lillard, J.W., Jr. Nanoparticle-Based Targeted Drug Delivery. *Exp. Mol. Pathol.* **2009**, *86*, 215–223. [[CrossRef](#)] [[PubMed](#)]
202. Mistry, A.; Glud, S.Z.; Kjemis, J.; Randel, J.; Howard, K.A.; Stolnik, S.; Illum, L. Effect of Physicochemical Properties on Intranasal Nanoparticle Transit into Murine Olfactory Epithelium. *J. Drug Target.* **2009**, *17*, 543–552. [[CrossRef](#)] [[PubMed](#)]
203. Ahmad, E.; Feng, Y.; Qi, J.; Fan, W.; Ma, Y.; He, H.; Xia, F.; Dong, X.; Zhao, W.; Lu, Y.; et al. Evidence of Nose-to-Brain Delivery of Nanoemulsions: Cargoes but Not Vehicles. *Nanoscale* **2017**, *9*, 1174–1183. [[CrossRef](#)]
204. Hoppstädter, J.; Seif, M.; Dembek, A.; Cavellius, C.; Huwer, H.; Kraegeloh, A.; Kiemer, A.K. M2 Polarization Enhances Silica Nanoparticle Uptake by Macrophages. *Front. Pharm.* **2015**, *6*, 1–12. [[CrossRef](#)]

205. Rathore, A.; Jain, A.; Gulbake, A.; Shilpi, S.; Khare, P.; Jain, A.; Jain, S.K. Mannosylated Liposomes Bearing Amphotericin B for Effective Management of Visceral Leishmaniasis. *J. Liposome Res.* **2011**, *21*, 333–340. [[CrossRef](#)] [[PubMed](#)]
206. Nahar, M.; Jain, N.K. Preparation, Characterization and Evaluation of Targeting Potential of Amphotericin B-Loaded Engineered PLGA Nanoparticles. *Pharm. Res.* **2009**, *26*, 2588–2598. [[CrossRef](#)] [[PubMed](#)]
207. Xiao, B.; Laroui, H.; Ayyadurai, S.; Viennois, E.; Charania, M.A.; Zhang, Y.; Merlin, D. Mannosylated Bioreducible Nanoparticle-Mediated Macrophage-Specific TNF- α RNA Interference for IBD Therapy. *Biomaterials* **2013**, *34*, 7471–7482. [[CrossRef](#)] [[PubMed](#)]
208. Huang, Y.; Guo, J.; Gui, S. Orally Targeted Galactosylated Chitosan Poly(Lactic-Co-Glycolic Acid) Nanoparticles Loaded with TNF- α siRNA Provide a Novel Strategy for the Experimental Treatment of Ulcerative Colitis. *Eur. J. Pharm. Sci.* **2018**, *125*, 232–243. [[CrossRef](#)] [[PubMed](#)]
209. Liu, L.; Yi, H.; He, H.; Pan, H.; Cai, L.; Ma, Y. Tumor Associated Macrophage-Targeted MicroRNA Delivery with Dual-Responsive Polypeptide Nanovectors for Anti-Cancer Therapy. *Biomaterials* **2017**, *134*, 166–179. [[CrossRef](#)] [[PubMed](#)]
210. Niu, M.; Valdes, S.; Naguib, Y.W.; Hursting, S.D.; Cui, Z. Tumor-Associated Macrophage-Mediated Targeted Therapy of Triple-Negative Breast Cancer. *Mol. Pharm.* **2016**, *13*, 1833–1842. [[CrossRef](#)] [[PubMed](#)]
211. Vieira, A.C.; Magalhães, J.; Rocha, S.; Cardoso, M.S.; Santos, S.G.; Borges, M.; Pinheiro, M.; Reis, S. Targeted Macrophages Delivery of Rifampicin-Loaded Lipid Nanoparticles to Improve Tuberculosis Treatment. *Nanomedicine* **2017**, *12*, 2721–2736. [[CrossRef](#)] [[PubMed](#)]
212. He, H.; Yuan, Q.; Bie, J.; Wallace, R.L.; Yannie, P.J.; Wang, J.; Lancina, M.G.; Zolotarskaya, O.Y.; Korzun, W.; Yang, H.; et al. Development of Mannose Functionalized Dendrimeric Nanoparticles for Targeted Delivery to Macrophages: Use of This Platform to Modulate Atherosclerosis. *Transl. Res.* **2018**, *193*, 13–30. [[CrossRef](#)] [[PubMed](#)]
213. Davis, S.M.; Reichel, D.; Bae, Y.; Pennypacker, K.R. Leukemia Inhibitory Factor-Loaded Nanoparticles with Enhanced Cytokine Metabolic Stability and Anti-Inflammatory Activity. *Pharm. Res.* **2018**, *35*. [[CrossRef](#)]
214. Cerqueira, S.R.; Silva, B.L.; Oliveira, J.M.; Mano, J.F.; Sousa, N.; Salgado, A.J.; Reis, R.L. Multifunctionalized CMCh/PAMAM Dendrimer Nanoparticles Modulate the Cellular Uptake by Astrocytes and Oligodendrocytes in Primary Cultures of Glial Cells. *Macromol. Biosci.* **2012**, *12*, 591–597. [[CrossRef](#)]
215. Akinrinmade, O.A.; Chetty, S.; Daramola, A.K.; Islam, M.-U.; Thepen, T.; Barth, S. CD64: An Attractive Immunotherapeutic Target for M1-Type Macrophage Mediated Chronic Inflammatory Diseases. *Biomedicines* **2017**, *5*, 56. [[CrossRef](#)] [[PubMed](#)]
216. Hristodorov, D.; Mladenov, R.; von Felbert, V.; Huhn, M.; Fischer, R.; Barth, S.; Thepen, T. Targeting CD64 Mediates Elimination of M1 but Not M2 Macrophages in Vitro and in Cutaneous Inflammation in Mice and Patient Biopsies. *MABS* **2015**, *7*, 853–862. [[CrossRef](#)] [[PubMed](#)]
217. Moura, C.C.; Segundo, M.A.; das Neves, J.; Reis, S.; Sarmiento, B. Co-Association of Methotrexate and SPIONs into Anti-CD64 Antibody-Conjugated PLGA Nanoparticles for Theranostic Application. *Int. J. Nanomed.* **2014**, *9*, 4911–4922. [[CrossRef](#)]
218. Albuquerque, J.; Moura, C.C.; Sarmiento, B.; Reis, S. Solid Lipid Nanoparticles: A Potential Multifunctional Approach towards Rheumatoid Arthritis Theranostics. *Molecules* **2015**, *20*, 11103–11118. [[CrossRef](#)] [[PubMed](#)]
219. Yong, S.-B.; Kim, H.J.; Kim, J.K.; Chung, J.Y.; Kim, Y.-H. Human CD64-Targeted Non-Viral siRNA Delivery System for Blood Monocyte Gene Modulation. *Sci. Rep.* **2017**, *7*, 42171. [[CrossRef](#)] [[PubMed](#)]
220. Blanco, E.; Shen, H.; Ferrari, M. Principles of Nanoparticle Design for Overcoming Biological Barriers to Drug Delivery. *Nat. Biotechnol.* **2015**, *33*, 941–951. [[CrossRef](#)] [[PubMed](#)]
221. Engelhardt, B.; Sorokin, L. The Blood–brain and the Blood–cerebrospinal Fluid Barriers: Function and Dysfunction. *Semin. Immunopathol.* **2009**, *31*, 497–511. [[CrossRef](#)] [[PubMed](#)]
222. Mo, X.; Zheng, Z.; He, Y.; Zhong, H.; Kang, X.; Shi, M.; Liu, T.; Jiao, Z.; Huang, Y. Antiglioma via Regulating Oxidative Stress and Remodeling Tumor-Associated Macrophage Using Lactoferrin-Mediated Biomimetic Codelivery of Simvastatin/Fenretinide. *J. Control. Release* **2018**, *287*, 12–23. [[CrossRef](#)]
223. Calvo, P.; Gouritin, B.; Villarroya, H.; Eclancher, F.; Giannavola, C.; Klein, C.; Andreux, J.P.; Couvreur, P. Quantification and Localization of PEGylated Polycyanoacrylate Nanoparticles in Brain and Spinal Cord during Experimental Allergic Encephalomyelitis in the Rat. *Eur. J. Neurosci.* **2002**, *15*, 1317–1326. [[CrossRef](#)]

224. Zhao, P.; Wang, Y.; Kang, X.; Wu, A.; Yin, W.; Tang, Y.; Wang, J.; Zhang, M.; Duan, Y.; Huang, Y. Dual-Targeting Biomimetic Delivery for Anti-Glioma Activity: Via Remodeling the Tumor Microenvironment and Directing Macrophage-Mediated Immunotherapy. *Chem. Sci.* **2018**, *9*, 2674–2689. [[CrossRef](#)]
225. Zou, L.; Tao, Y.; Payne, G.; Do, L.; Thomas, T.; Rodriguez, J.; Dou, H. Targeted Delivery of Nano-PTX to the Brain Tumor-Associated Macrophages. *Oncotarget* **2017**, *8*, 6564–6578. [[CrossRef](#)] [[PubMed](#)]
226. Bourganis, V.; Kammona, O.; Alexopoulos, A.; Kiparissides, C. Recent Advances in Carrier Mediated Nose-to-Brain Delivery of Pharmaceuticals. *Eur. J. Pharm. Biopharm.* **2018**, *128*, 337–362. [[CrossRef](#)] [[PubMed](#)]
227. Samaridou, E.; Alonso, M.J. Nose-to-Brain Peptide Delivery—The Potential of Nanotechnology. *Bioorg. Med. Chem.* **2018**, *26*, 2888–2905. [[CrossRef](#)] [[PubMed](#)]
228. Warnken, Z.N.; Smyth, H.D.C.; Watts, A.B.; Weitman, S.; Kuhn, J.G.; Williams, R.O. Formulation and Device Design to Increase Nose to Brain Drug Delivery. *J. Drug Deliv. Sci. Technol.* **2016**, *35*, 213–222. [[CrossRef](#)]
229. Dalpiaz, A.; Fogagnolo, M.; Ferraro, L.; Capuzzo, A.; Pavan, B.; Rassu, G.; Salis, A.; Giunchedi, P.; Gavini, E. Nasal Chitosan Microparticles Target a Zidovudine Prodrug to Brain HIV Sanctuaries. *Antivir. Res.* **2015**, *123*, 146–157. [[CrossRef](#)] [[PubMed](#)]
230. Fransson, M.; Piras, E.; Wang, H.; Burman, J.; Duprez, I.; Harris, R.A.; LeBlanc, K.; Magnusson, P.U.; Brittebo, E.; Loskog, A.S.I. Intranasal Delivery of Central Nervous System-Retargeted Human Mesenchymal Stromal Cells Prolongs Treatment Efficacy of Experimental Autoimmune Encephalomyelitis. *Immunology* **2014**, *142*, 431–441. [[CrossRef](#)] [[PubMed](#)]
231. Mayo, L.; Da Cunha, A.P.; Madi, A.; Beynon, V.; Yang, Z.; Alvarez, J.I.; Prat, A.; Sobel, R.A.; Kobzik, L.; Lassmann, H.; et al. IL-10-Dependent Tr1 Cells Attenuate Astrocyte Activation and Ameliorate Chronic Central Nervous System Inflammation. *Brain* **2016**, *139*, 1939–1957. [[CrossRef](#)]
232. Esposito, E.; Cortesi, R.; Drechsler, M.; Fan, J.; Fu, B.M.; Calderan, L.; Mannucci, S.; Boschi, F.; Nastruzzi, C. Nanoformulations for Dimethyl Fumarate: Physicochemical Characterization and in Vitro / in Vivo Behavior. *Eur. J. Pharm. Biopharm.* **2017**, *115*, 285–296. [[CrossRef](#)]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).