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# Activity of Icacinol from *Icacina trichantha* on Seedling Growth of *Oryza sativa* and *Arabidopsis thaliana*

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### Abstract

Broadleaf weeds are very costly for crop growers. Additional herbicidal compounds need to be obtained, especially from natural sources. Extracts of *Icacina trichantha* were evaluated for responses in germinating seeds and seedlings of rice (*Oryza sativa*) and Arabidopsis (*Arabidopsis thaliana*). An ethyl acetate fraction of *I. trichantha* tuber and a diterpenoid constituent, icacinol (1), were found to have impacts on germination and growth of seedlings. The seed germination inhibitory activity on rice was minimal, but significant on Arabidopsis. While rice indicated some growth delay in leaf expansion in the presence of 1, the effects appeared temporary; chlorophyll and anthocyanins were not significantly altered compared to DMSO controls. Rice seedlings attained biomass similar to DMSO controls, and rice grains per panicle were not significantly different than the DMSO-controls. On the other hand, Arabidopsis exhibited damage to leaf expansion, reduced chlorophyll, and increased anthocyanins in aerial portions of the seedlings. Icacinol (1) may be a suitable chemical agent to investigate further for the treatment of eudicot weeds.

## **Graphical Abstract**



Weeds and invasive plant species negatively impact agriculture worldwide. They compete with crop plants for water, nutrients, and light, interfering with crop growth, and can ultimately reduce yields (for reviews, see Refs. $^{1-3}$ ). Weeds can be controlled by a variety of means including mechanical, chemical, and biological methods. The chemical methods employed utilize physiologically active compounds, both synthetic and naturally occurring, yet novel chemicals to combat weeds and invasive species require further exploration and validation. Novel chemicals can be identified by screening of chemical libraries, but functional herbicidal studies are inevitably required, as many experiments are done in plant cell cultures.<sup>4–6</sup> While seedlings are used in phenotype-based chemical screens to identify small molecules that disrupt processes,<sup>7</sup> screening pipelines require improvements in the detection of functional impact, in order to identify new types of molecules that impact plant growth, as well as candidates that do not pose environmental problems. Plants are rich sources of small-molecule organic compounds, with advantages of being renewable and biodegradable in the environment. Useful targeting would include finding compounds that elicit stress responses early in development in weeds, particularly in the seed to seedling transition, with low negative impact on crops of economic importance.

*Icacina trichantha* Oliv. (Icacinaceae), a drought-resistant shrub native to West and Central Africa, is a food and medicinal plant used by the indigenous tribes in Nigeria and neighboring countries.<sup>8</sup> Our group has reported a series of novel pimarane-type diterpene lactones in the tubers of the plant, including (9 $\beta$ H)-pimarane lactones, 17-*nor*-pimarane lactones, 17-*nor*-pimarane lactones, and di-*nor*-pimarane lactones.<sup>9–13</sup> Several of these secondary metabolites have displayed cytotoxic

activity against human cancer cell lines.<sup>11–14</sup> During fractionation and purification of a MeOH extract of the tubers of *I. trichantha*, it was observed that the EtOAc-soluble fraction induced growth dysregulation in the weed-like dicot plant *Arabidopsis thaliana*, but not in rice (*Oryza sativa*), leading to the isolation of icacinol (1) as an active principle. The activities of the EtOAc fraction and 1 are described herein, where the herbicidal characteristics may indicate a useful herbicidal component for monocot crops.

The tubers of *I. trichantha* were extracted with 80% aqueous MeOH by percolation. The extract was dried, suspended in water, and successively partitioned with petroleum ether, EtOAc and *n*-BuOH into three fractions denoted S1 – S3, respectively, together with the water-soluble part (S4). All four fractions were tested for seed germination activity using rice (*Oryza sativa* L. "Nipponbare") and Arabidopsis (*Arabidopsis thaliana* Col) in initial screens. The EtOAc-soluble fraction (S2) displayed significant activity. Further plant response assays were conducted with S2 and subsequent chromatographic fractionation afforded an active compound, icacinol (1), which is a (9 $\beta$ H)-pimarane lactone previously found in *I. claessensii*,<sup>15</sup> *I. senegalensis*,<sup>16</sup> and *Casimirella* sp.<sup>14</sup> This compound has also shown cytotoxicity against the A2780 human ovarian cancer,<sup>14</sup> MDA-MB-435 human melanoma, MB-231 human breast cancer, and OVCAR3 human ovarian cancer<sup>12</sup> cell lines. No other biological activity of this compound has been reported.

Dilutions of fractions of *I. trichantha* were screened on 0.5X MS minimal medium, at 10 ng/mL and 100 ng/mL, as an initial test. Most fractions (S1, S3 and S4) did not impede or slow germination of either the monocot rice (*Oryza sativa*) or dicot Arabidopsis (*Arabidopsis thaliana*) (Figure S1, Supporting Information). However, the ethyl acetate fraction (S2) and icacinol (1) exhibited significant effects on seed germination (Figure 1). For rice seeds, the effect of S2 was significant at 100 ng/mL (p = 0.0267), with 87% of seeds germinating by 72 h compared to almost 100% of the untreated (DMSO control). Other dilutions were not significant (10 ng/mL shown). The icacinol (1) treatment was significant only at 265  $\mu$ M. Other dilutions shown did not affect germination. Arabidopsis seed responses, however, indicated much more sensitivity to S2 and 1. When seeds were grown in the presence of S2, the germination rates at 72 h were 62% (100 ng/mL, p = 0.037) and 88% (10 ng/mL, p = 0.0169), compared to the vehicle-only (DMSO) treatment. At 5.3  $\mu$ M, 1 reduced germination by 72 h, with about 80% of seeds germinating (p = 0.0039). Dosespecific responses were observed at 26.5  $\mu$ M (66%; p = 0.0459), 53  $\mu$ M (36%; p = 0.0084), and 265  $\mu$ M (<1%; p < 0.0001).

Icacinol (1) was found to be the most active among the tested components on plant germination responses. Importantly, there was a clear difference in seed germination between Arabidopsis (a eudicot) and rice (a monocot), where Arabidopsis germination was severely impaired under the conditions used. The chief difference of monocots and eudicots in germination is related to the fact that monocot seed leaf is thin and the endosperm to feed the seed leaf is outside of the leaf itself, whereas the eudicots like Arabidopsis have two seed leaves with an endosperm to feed the embryonic plant. In addition, Arabidopsis is a very fast-growing weed-like plant, with photosynthesizing cotyledons. It is possible that the metabolic interactions in the cotyledons of Arabidopsis are disrupted in the process of germination and the concomitant greening process, as Arabidopsis cotyledons develop and

photosynthesize within 2–3 days with summer (i.e., 16:8 light/dark cycle) conditions. Three pimarane-type diterpenoids, momilactones A-C, have been reported to inhibit germination of Arabidopsis.<sup>17–19</sup>

Vegetative development in young seedlings exposed to S2 or 1 was also explored, with seedlings observed daily and specific characteristics evaluated weekly, post-sowing. It was noted over several dilutions of S2 and 1 that rice, while delayed in growth, grew without mortality in vegetative stage, then switched to reproductive mode to seed. Growth (Figure 2) observed at two weeks post-sowing was delayed for rice, where by six weeks the biomass of whole plant presented comparable means (Figure 3; DMSO-control mean 99.6% vs. 94.2% for S2, 90.4% for 1 at 53 µM), yet, based on comparison of the means, data were statistically significant. In addition, rice grains per panicle (when plants were permitted to go to seed) were highly similar (DMSO control compared to 1 at 26.5 µM; Figure S2, Supporting Information). However, Arabidopsis treated with 1 showed greater mean biomass differences (Figure 3; DMSO-control mean 99.8% vs. 92.8% for S2, 64.4% for 1 at 53  $\mu$ M). Arabidopsis did not achieve transition to reproductive stage within typical time when treated with 1 (compared to untreated). Rice treated with 1 also showed no difference in chlorophylls (no reductions) and no increase in anthocyanins by two weeks post-germination (Figure S3). However, Arabidopsis treated with 1 showed indications of stress, where anthocyanins increased and chlorophyll was reduced (Figure S3). The tissue size at this same age (shoot 1.79 vs. 1.59 n.s.; root 5.09 vs. 4.075, p < 0.01) was significant for the roots of Arabidopsis. Arabidopsis signs of damage can be seen in cotyledons (less chlorophyll) and anthocyanin accumulation manifested at the apical hook (Figure 4). Anthocyanin accumulation can occur in response to many chemicals including herbicides or growth regulators.<sup>20</sup> Damage to the Arabidopsis leaves by 1 (53 µM) treatment also included chlorosis and lack of cotyledon expansion, with an appearance similar to auxin-related herbicides that in dicots can cause oxidative stress and destruction of the chloroplast (leaving rice and other crops lesser affected) by still poorly understood mechanisms.<sup>21</sup> The present experimentation was not on soil for the majority of tests (60 days or less) and this may affect the plant responses. The growth to seed set occurred on soil (transplants, see methods in Experimental Section).

In the literature, two (9 $\beta$ H)-pimarane diterpenes, momilactones A and B, found in rice and the moss *Hypnum plumaeforme*, have shown allelopathic or herbicidal activities.<sup>22–25</sup> Momilactone B has exhibited growth inhibition on barnyard grass (*Echinochloa crus-galli*), a major noxious weed associated with rice and other monocot and dicot plants.<sup>26</sup> Further, momilactone B exhibited significant antifungal activity against the rice blast pathogen *Magnaporthe oryzae*, which is responsible for the devastating disease that has been estimated to cause the loss of 10–30% of the total rice harvest and also infects wheat, barley, and millet crops.<sup>27</sup> Momilactones A and B were reported to be allelochemicals in the defense responses of moss (*Hypnum plumaeforme* and *Pseudoleskeella papillosa*) as well.<sup>17,28</sup> Individual momilactones A and B have already been patented as herbicides and potential anticancer agents.

The discovery of the potential herbicidal activity of icacinol (1) now adds to our knowledge of the (9 $\beta$ *H*)-pimarane diterpenes. These versatile molecules seem to be the natural choice

for plant defense during evolution, and they can serve as prototypes for other lead compounds in the search for crop-friendly herbicides, antifungal agents, and related agricultural substances. Further studies on natural sources to enrich structural diversity of momilactone-like molecules and discovery of more promising lead compounds and adjuvants for treatment are desirable goals for weed management.

#### EXPERIMENTAL SECTION

#### **General Experimental Procedures**

Chemicals for plant experiments, and sterile plastics (square petri dishes, phytatrays<sup>TM</sup> and phytatray II<sup>TM</sup>) unless otherwise noted, were obtained from Sigma-Aldrich (St. Louis, MO USA). Seeds were sterilized in bleach solution (49% sodium hypochlorite [6.05%], 49% sterile water, 2% 0.1% Triton X-100) with vigorous shaking for 5 min then 4 washes with sterile water. All materials in contact with seeds or plants were sterilized by autoclaving or purchased sterile (gamma irradiated) or sterile (0.22µm) filtered (molecular biology grade water). In order to test specific chemical concentrations per area, 0.5XMS minimal medium trays were made (50mL phytatray<sup>TM</sup>, 50 mL vertical tray) for seed germination and plant growth.<sup>31</sup> Rice when transplanted went into autoclaved soil/sand mix.

#### **Plant Material**

Tubers of *Icacina trichantha* were collected in June 2011 from Orba village, Nsukka, Enugu State, Nigeria, and authenticated by Prof. B. O. Olorede of the Botany Department, University of Abuja, Nigeria, and Mr. A. Ozioko, botanist at BDCP Laboratories, Nsukka, Nigeria. A voucher specimen (UNN/FVM 456) was deposited at the pharmacology laboratory at the University of Nigeria, Nsukka, Nigeria.

#### **Extraction and Isolation**

The powdered tubers of *I. trichantha* (1.5 kg) were extracted with 80% aq. MeOH by percolation to yield 166 g of dry crude extract, which was partitioned into petroleum ethersoluble (S1, 11 g), EtOAc-soluble (S2, 17 g), BuOH-soluble (S3, 15 g), and H<sub>2</sub>O-soluble (S4, 128 g) fractions. The EtOAc-soluble fraction (S2) was further fractionated. A precipitate from sub-fraction 47–53 was purified by further chromatography using a mixture of CH<sub>2</sub>Cl<sub>2</sub> and MeOH to yield icacinol (1), a crop of 50 mg crystals of which was finally obtained. This compound exhibited physical and spectroscopic parameters consistent with literature data,<sup>12</sup> NMR data is shown (Figure S4, Supporting Information).

#### Plant Growth Response Screening Materials

Stock solutions were made of all final extract fractions at 10 µg/mL, with icacinol (1) resuspended to a 265 µM solution in sterile DMSO, then were diluted for use in  $0.5 \times MS^{27}$  for experiments described herein. *Arabidopsis thaliana* (Arabidopsis) Columbia WT seed was originally obtained from ABRC/TAIR.<sup>29</sup> Bulk seed stocks were grown as previously reported.<sup>30</sup> Seeds of *Oryza sativa* (rice) Nipponbare (obtained from KW) were sterilized and sown in the same manner as Arabidopsis seed, as described,<sup>31</sup> but incubated at 28 °C for germination.

#### Assays for Growth Responses

Seeds were surface sterilized in bleach solution, then rinsed in sterile water, and plated on 0.5X MS (pH 5.8) in Phytatrays (Sigma, St. Louis, USA) with no sucrose and no added vitamins as described,<sup>31</sup> but instead of a cold treatment, were given a 48 h darkness treatment at 20 °C. At the time of planting, a plant extract of S2 or **1** or DMSO (empty vehicle control) was diluted in 0.5X MS, then was added to 0.8% low melt agarose (0.5X MS; top agarose) when the autoclaved media fell below 50 °C, mixed well, then poured and allowed to solidify with sterilized and washed seeds in the top agarose in a thin layer. Dilutions of chemicals derived from plant were initially made at 1:20 to 1:5000 (depending on the experiment) from extract stock in DMSO (described above), diluted with 0.5X MS right before plating. After sowing seeds on plates, plates were placed into black boxes and moved to 20 °C complete darkness room for 48 h, then subsequently moved to white light at 120 µmol m<sup>-2</sup>s<sup>-1</sup>, at 20 °C for Arabidopsis and 28 °C for rice.

#### Vertical Growth Assays of Arabidopsis

On sterile square petri plates seeds were lined up in a row, 2.5 cm from the designated top edge, and 2.0 cm between each row. A 10- $\mu$ L aliquot of seed in top agarose spotted onto a 0.5X MS phytatray base plate of different dilutions of S2 or **1**. Plated, sterilized seed plates were turned vertical, secured into light-tight black Plexiglas<sup>TM</sup> boxes, sealed with aluminum foil then were cold placed in a dark room at 20 °C for 48 h. After 48 h the plates were moved to one of several locations: to a 24 h 20 °C dark environmental room (continuous darkness, i.e., Dc), or white light environmental room where lights were on for 24 h continuous (Wc) or lighting regime was 16:8 (16 h light, 8 h darkness),<sup>32</sup> for varying numbers of days, ranging from 0 – 60 days, depending on the experiment. Lights sources, including dim green light for handling dark-grown seedlings, have been described.<sup>30, 32</sup> The fluence rate received by the vertical plates and phytatrays determined by a LiCor meter was 120  $\mu$ mol m<sup>-2</sup>s<sup>-1</sup>.

#### **Germination and Phenotyping**

Successful germination was determined under a dissecting scope by viewing rice and Arabidopsis seeds every 12 h post-sowing, scoring complete emergence of the radical as previously described.<sup>30</sup> Phenotypic responses on untreated (control) and experimental extract plates were determined by comparison of chemically treated seeds to untreated plants (DMSO vehicle control), and counted on a Zeiss Stereo Discovery V.8 microscope at 1X using Axiovision (Zeiss). Plates were photographed at different times after stratification using Nikon Coolpix on a light box or black background. At least three sets of 30 seeds were scored for each germination assay on phytatrays at 24 h, 48 h, and 72 h post-stratification.

#### Vegetative Assessment

Rice and Arabidopsis seeds were sown on phytatray II to assess how they would grow over 7–60 days, depending on experiments. At four weeks, rice seedlings were transferred to soil/ sand mix submerged in water in white catering 1-gallon buckets, where they were moved to greenhouse of similar fluence levels, and where daytime temperature was above 25 °C, in order to grow long-term, including for seed set (rice grain per panicle). Sets of plants were

washed at dried to determine root and shoot dried weight at six weeks. Images and samples (100 seedlings  $\times$  3 replicates) for chlorophylls extraction<sup>33</sup> and anthocyanins extraction<sup>34</sup> were taken at two weeks. Seedlings were imaged on dissecting scope at various times to capture phenotypes as indicated in Figures.

#### Statistical Analysis

Phenotypes and plant responses were quantitated and the values entered into GraphPad/ Prism V.5 for Mac, similar in the manner described.<sup>31</sup> Equal variances were not assumed in plant materials (seeds and seedlings) and differences of means compared (typically to the DMSO-control data) were assessed by unpaired t-test (two-tailed) with Welch's correction.

#### Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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#### Figure 1.

Germination at 72 h

*Icacina tricantha* fractions and icacinol (1) effects on seed germination. Extractions were conducted as described in Experimental Section. S2 (EtOAc-soluble fraction) and **1** were mixed in top agarose with seeds at concentrations indicated in the figure; 100 seeds per replicate were sown. Seed germination was scored 72 h postplating at 20°C for *Arabidopsis* and at 28°C for rice.





#### Figure 2.

*Icacina tricantha* extracts and icacinol (1) impacts on vegetative growth. Extractions were conducted as described in the Experimental Section. S2 (EtOAc-soluble fraction) and 1 were mixed in top agarose with seeds at concentration indicated in the figure. Plants were grown on that medium for 2 weeks at 20°C (28°C for rice) in 16:8, then photographed as shown. Images are representative. Top panel rice (size bar 2 cm), lower panel *Arabidopsis* (size bar 250 µm).



# Mass of seedlings at 6 weeks

#### Figure 3.

*Icacina tricantha* extract and icacinol (1) impacts on longer term vegetative growth. Rice and *Arabidopsis* seeds were sown; then seedlings were grown for 6 weeks in 16:8 (rice at 28°C; *Arabidopsis* at 20°C), with S2 or **1** in the medium as described in Figure 2. Ten random representative seedlings were selected from each set. Tissue was harvested, dried, then weighed for comparison of the mass to the untreated control (DMSO control; % of control shown). DMSO controls were compared to completely untreated controls to determine their percent. \* P<0.05; \*\* P<0.01; \*\*\*\*P<0.001



#### Figure 4.

Icacinol (1) impact on *Arabidopsis* seedling growth. **1** was mixed in top agarose with seeds at the concentration indicated, sown on vertical plates. Plants were grown for 14 days at 20°C in 16:8, sown on vertical plates (there is some limited contact with gel by stem and leaf edge at emergence), and then photographed on adissecting microscope as shown. Images are representative. The manifestation of anthocyanin response at the apical hook and leaf base is indicated by circled pigmented area and enlargement. Upper 2 panels images grown in

continuous white light (Wc); lower image shows seedlings grown in continuous darkness (Dc) on Phytatrays.