

# Enhanced Solubility of Telmisartan Phthalic Acid Cocrystals within the pH Range of a Systemic Absorption Site

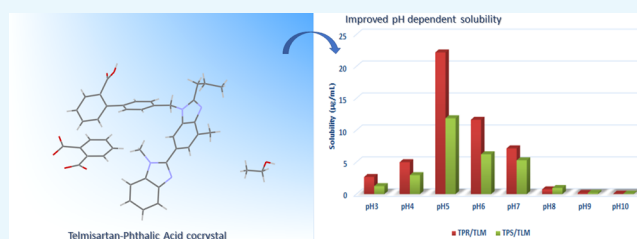
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## Supporting Information

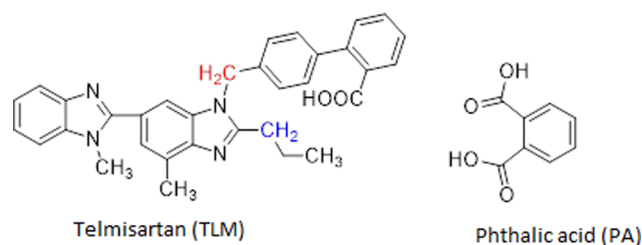
**ABSTRACT:** Telmisartan (TLM), a nonpeptide angiotensin II antagonist, is widely prescribed for treating arterial hypertension and marketed by the innovator with the trade name of Micardis and Micardis plus. Telmisartan exhibits low aqueous solubility in the pH range of 3–7, which is the physiological pH. For addressing the issue of poor solubility of TLM, its commercial form makes use of inorganic alkalinizers. The present work illustrates the attempt to improve the solubility of telmisartan via a crystal engineering approach. A novel solid form of telmisartan with phthalic acid was obtained through the solution crystallization method (TPS) and the reaction crystallization method (TPR). Both the forms (TPS and TPR) were thoroughly characterized by powder diffraction X-ray diffraction, differential scanning calorimetry, thermogravimetric analysis, Fourier transform infrared spectroscopy, and <sup>1</sup>H NMR and were identified to be two different crystalline forms. Solubility studies of TPS and TPR were conducted at varying pH of phosphate buffer, and they exhibited 11-fold and 22-fold increased solubility, respectively, when compared to that of the pure drug at pH 5, which is within the pH of small intestine at which telmisartan is best absorbed orally from the systemic circulation.



## INTRODUCTION

With the rise in the number of poorly soluble drugs in the pipeline and market place, manipulating aqueous solubility of these drugs has become a crucial step in preformulation research. Physicochemical properties of an active pharmaceutical ingredient (API), such as solubility, dissolution, bioavailability, stability, etc., are directed by the arrangement of molecules in its three-dimensional crystal lattice.<sup>1,2</sup> Thus, a variation in the structural arrangement of an API in its solid state through noncovalent interactions will aid in the development of an ideal crystal form with desired properties, and, at the same time, the intrinsic biological activity will be retained. Through exploitation of the crystal engineering technique, it has been possible to anticipate probable intermolecular interactions and design new solid forms with desired properties.<sup>3,4</sup> This in turn has spurred the growth and development of multicomponent solid forms such as salt,<sup>5,6</sup> cocrystals,<sup>7–9</sup> etc. and proved to be a promising approach to address issues such as poor solubility,<sup>10–12</sup> dissolution,<sup>13</sup> bioavailability,<sup>14</sup> stability,<sup>15,16</sup> mechanical properties,<sup>17,18</sup> etc.

In this regard, we aimed to investigate the applicability of the crystal engineering technique and explore a novel solid form of a poorly soluble drug, telmisartan (TLM). Telmisartan (TLM, Figure 1), chemically referred to as 2-[4-[[4-methyl-6-(1-methylbenzimidazol-2-yl)-2-propylbenzimidazol-1-yl]methyl]-phenyl]benzoic acid, is a nonpeptide angiotensin II antagonist,



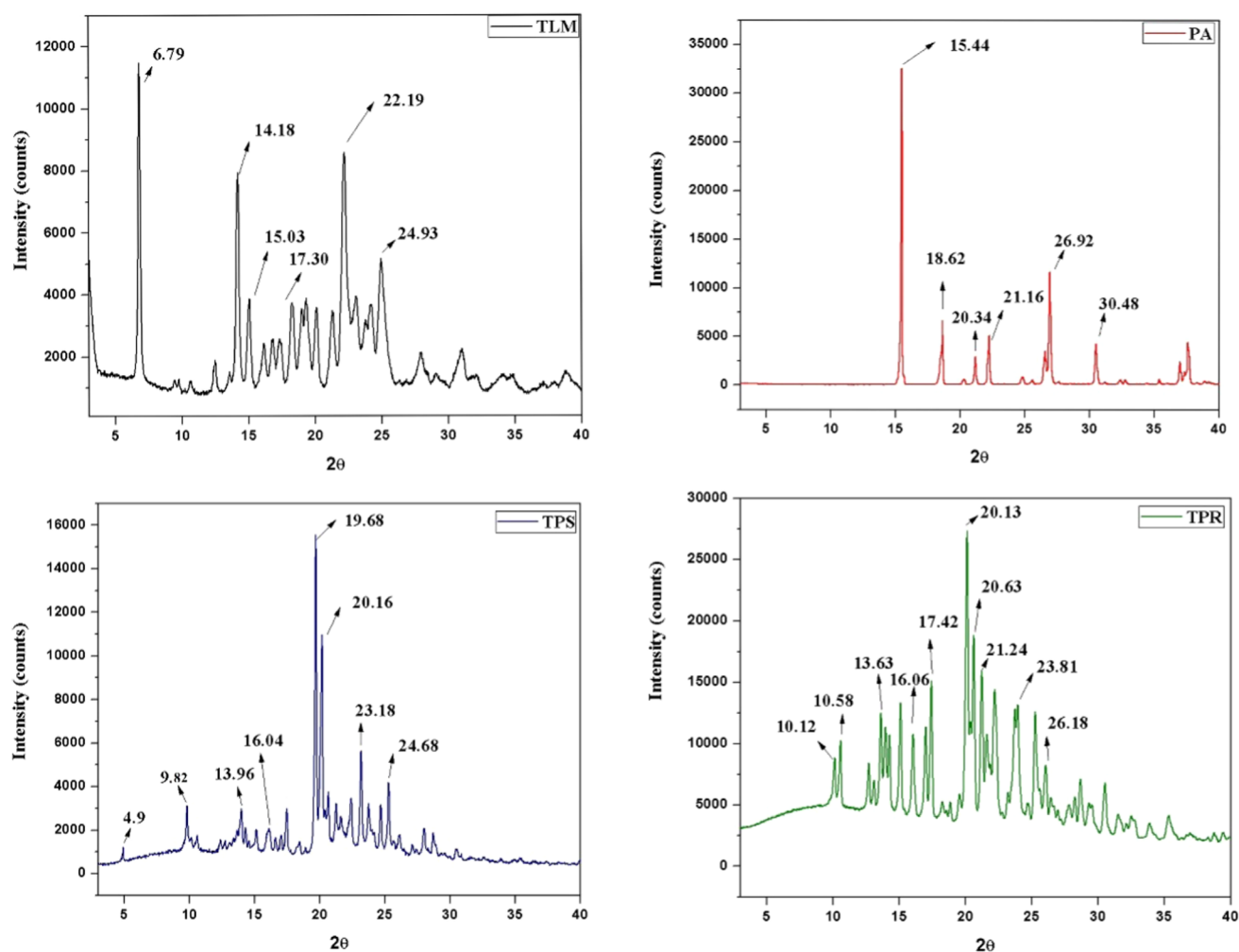
**Figure 1.** Chemical structure of telmisartan (TLM) highlighting distinguished CH<sub>2</sub> in color and phthalic acid (PA) used in this study.

prescribed for the treatment of arterial hypertension.<sup>19</sup> TLM exists in three crystalline forms, two polymorphs and one pseudopolymorph.<sup>20</sup> The aqueous solubility of TLM is strongly pH-dependent. The maximum solubility was observed at extremely low and high pH. TLM is practically insoluble at pH 3–7 (0.09 µg/mL in water), which is the physiological pH.<sup>19</sup> This poor solubility becomes a major hurdle in the bioavailability of TLM, which is only 42–58%. Nonetheless, TLM is commercially available and marketed by the innovator under the trade name of Micardis and Micardis plus. To

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**Figure 2.** Powder X-ray diffractograms of TPS and TPR compared with those of starting materials.

resolve the issue of pH-dependent solubility of TLM, the marketed formulations make use of strong alkalinizers such as sodium hydroxide, potassium hydroxide, meglumine, or combination thereof as pH modulators.<sup>21</sup> The inorganic alkalinizers increase the microenvironmental pH for achieving the optimum solubility required for absorption. However, this leads to disruption and degeneration of duodenal and jejunal mucosal tissues.<sup>22</sup> Apart from the solubility issues, telmisartan is rapidly absorbed throughout the gastrointestinal tract.<sup>23</sup> However, the maximum absorption is found to be in the small intestine where the intraluminal pH is within the range of 5–7.<sup>23,24</sup> Therefore, the aim of our study is to improve the aqueous solubility of telmisartan within the pH range of small intestine (pH 5–7) to achieve the best possible oral absorption without using any alkalinizers. In comparison with formulation approaches for improving the solubility of telmisartan,<sup>25–28</sup> the investigations through the crystal engineering approach are less and limited to only four reports in the literature.<sup>29–32</sup> Hence, there is a need to limit the use of inorganic alkalinizers and find a suitable crystal form of TLM that can address its poor aqueous solubility and may be an alternative to the existing dosage form.

In this article, we describe the screening studies for a new solid form of telmisartan, isolation and preparation of novel multicomponent solid forms (TPS and TPR) of telmisartan with phthalic acid (PA, Figure 1), and their comprehensive

characterization by powder X-ray diffraction (PXRD), <sup>1</sup>H NMR, differential scanning calorimetry (DSC), thermogravimetric analysis (TGA), and Fourier transform infrared spectroscopy (FTIR) followed by solubility measurements at various pH.

## RESULTS AND DISCUSSION

The structure of TLM possesses one acidic and two basic functionalities, or, in other words, it offers one hydrogen bond donor and two acceptor sites. Thus, it is expected to form hydrogen bonds with certain cofomers that are having acid or amide functional groups. For modulating pH-dependent solubility, pK<sub>a</sub> of the cofomers was also taken into consideration. Therefore, in this study, a series of cofomers containing carboxylic group and amide group with varying pK<sub>a</sub> values were selected for the screening purpose (Table S1, Supporting Information) by employing the solution crystallization method. A novel multicomponent solid form of telmisartan with phthalic acid was obtained by the solution crystallization method (TPS), which was further prepared by the reaction crystallization method (TPR). The obtained forms (TPS and TPR) were characterized through various analytical techniques.

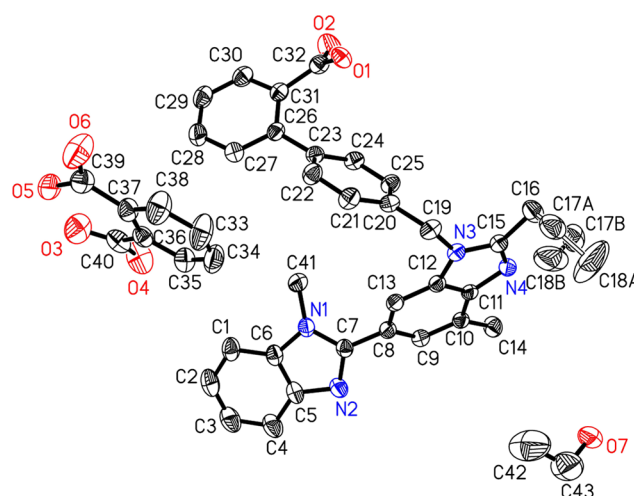
**Powder X-ray Diffraction Analysis.** The PXRD pattern with new diffraction peaks and absence of characteristic peaks of the individual components confirms that a new crystalline

form is produced as the peaks are representatives of reflections of each atomic plane.<sup>33</sup> The characteristic peak of TLM appeared at  $2\theta$  values of 6.8, 14.18, 20.10, and 22.19, and phthalic acid showed the characteristic reflection at 15.44, 18.62, and 26.92. The PXRD pattern of TPS and TPR compared with the starting material, telmisartan (TLM) and phthalic acid (PA), is presented in Figure 2. The PXRD pattern revealed that the characteristic peaks of the drug and coformer were absent in it. New diffraction peaks at  $2\theta$  values of 4.9, 12.76, 17.04, 22.62, 25.3, 28.02, and 28.72 were observed in TPS. In TPR, unique diffraction peaks were present at  $2\theta$  values of 13.12, 13.98, 17.04, 20.66, 23.94, 25.32, 28.32, and 28.72. The characteristic peaks of telmisartan and phthalic acid shifted, and the appearance of new peaks in TPS and TPR confirmed the formation of a new crystalline phase. The literature suggests that various methods used for the synthesis of cocrystals do not lead to the production of the same cocrystal form or composition.<sup>34</sup> Chow et al. suggested that accelerated crystallization by rapid solvent removal often yields a metastable solid, as speculated by Ostwald's rule of successive stages.<sup>34</sup> In this case too, the obtained TPS and TPR possess different cocrystalline forms owing to their different synthesis methods. The difference in the PXRD pattern, thermal behavior, and solubility studies of TPR and TPS proved the same.

**Calculation of Stoichiometric Proportion in TPS and TPR.** Solution  $^1\text{H}$  NMR was used to identify and confirm the chemical components and the stoichiometry in the new crystalline phase.<sup>35,36</sup> The resulting  $^1\text{H}$  NMR spectrum of the TPS and TPR cocrystal systems comprises the peaks from TLM, PA, and ethanol. It is evident from NMR that the acidic protons from the carboxylic group of TPS and TPR are in downfield at 13.35 ppm (Figure S1) and 13.26 ppm (Figure S4), respectively, which confirms H-bonded acidic protons and voids any proton transfer. This study further confirms the existence of a solvated cocrystal for both TPS and TPR. Both the  $\text{D}_2\text{O}$  exchange spectra of TPS (Figure S1) and TPR (Figure S5) show the disappearance of the peaks corresponding to acidic protons. The peak assignment from  $^1\text{H}$  NMR data confirms the distinguished peak of two  $\text{CH}_2$ , as shown in blue and red (Figure 1) at  $\delta$  2.90 (t, 2H) and  $\delta$  5.54 (s, 2H), respectively, for TLM in the TPS cocrystal system. A total of 14 aromatic protons (ArHs) appear as multiples in the region of  $\delta$  7.14–7.57. Furthermore, phthalic acid has 4 ArHs as multiplets in the region of  $\delta$  7.64–7.69, which merge with aromatic protons of TLM. The relative integration of distinguished  $\text{CH}_2$  (red as in Figure 1) of TLM at  $\delta$  5.54 shows integration of 2H and 18H found in the aromatic region. Thus, quantitative integration (Figure S2) confirms the 1:1 ratio of TLM (14 ArHs) and PA (4 ArHs) in the TPS cocrystal system. Similar results were obtained for the TPR cocrystal system, which confirmed the 1:1 ratio of TLM (14 ArHs) and PA (4 ArHs) in this system. In another quantitative integration study from  $^1\text{H}$  NMR spectrum of TPS cocrystal system showing distinguished  $\text{CH}_2$  peak of TLM (blue  $\delta$ : 2.90, t, 2H) with 1 integration in comparison to 0.5 integration for  $\text{CH}_2$  proton of ethanol ( $\delta$ : 3.35–3.40). Thus, the presence of half ratio of ethanol in comparison to unit ratio of TLM confirms the presence of TLM/ethanol in 1:0.5 ratio (Figure S3) in TPS cocrystal. Thus, overall, the ratio of TLM, PA, and ethanol was stoichiometrically confirmed as 1:1:0.5 in the TPS cocrystal system. Similar quantitative integration studies were conducted to identify the stoichiometric proportion of TLM, PA, and

ethanol in the TPR cocrystal system, and it was found to be 1:1:0.3, as shown in Figure S5.

It is interesting to note that the presence of TLM, PA, and ethanol was further confirmed through single-crystal X-ray diffraction (SCXRD) analysis. The asymmetric unit comprises TLM, PA, and ethanol in the TPS cocrystal, as shown in the Oak Ridge Thermal Ellipsoid Plot (ORTEP) diagram with 30% probability displacement ellipsoids (Figure 3).<sup>37</sup> The single-crystal parameters of TPS are mentioned in the foot note of Figure 3.



**Figure 3.** X-ray crystal structure of the TPS cocrystal showing the ORTEP diagram using a 30% ellipsoidal plot. Telmisartan (TLM), phthalic acid (PA), and ethanol were found in the asymmetric unit, and hydrogen atoms are not shown for clarity.  $\text{C}_{43}\text{H}_{42}\text{N}_4\text{O}_7$ ,  $M = 726.81$ , monoclinic, space group:  $I 2/a$ ,  $a = 15.9027(10)$ ,  $b = 13.2297(9)$ ,  $c = 36.609(2)$  Å,  $\alpha = 90.000(0)$ ,  $\beta = 99.754(6)$ ,  $\gamma = 90.000(0)$ ,  $V = 7590.7(8)$  Å<sup>3</sup>,  $T = 293(2)$  K,  $Z = 8$ ,  $\mu = 0.087$  mm<sup>-1</sup>,  $F(000) = 3072.0$ ,  $D_c = 1.272$  Mg/m, crystal size:  $0.15 \times 0.12 \times 0.10$  mm, 140 72 reflections measured, 6687 unique,  $R1 = 0.0741$  for 4357  $F_o > 4\sigma(F_o)$  and 0.1075 for all 6687 data and 528 parameters. Unit cell determination and intensity data collection were performed with 99% completeness at 293(2) K. Structure solutions by direct methods and refinements by full-matrix least-squares methods on  $F_2$  [CCDC No: 1520132].

**Thermal Analysis.** The distinct melting points of TPS and TPR from those of TLM (270.24 °C) and PA (209.94 °C) suggested the formation of a new crystalline phase. However, TPS and TPR exhibited an unfamiliar thermal behavior (Figures 4A and 5A). Desolvation of the ethanol molecule occurred in TPS and TPR at 124 and 156 °C, respectively, indicating the different binding energy of ethanol in TPS and TPR. Melting of desolvated TPS and TPR was observed at 199.8 °C, heat of fusion ( $\Delta H_f$ ) = 136.40 J/g, and at 200.53 °C,  $\Delta H_f = 157.66$  J/g, respectively. Upon desolvation, TPS and TPR converted into the same form, similar to the phenomenon observed in the telmisartan–saccharine cocrystal.<sup>29</sup> The melting of desolvated TPS and TPR was found to be accompanied by decomposition of PA (endothermic event) and subsequent recrystallization (exothermic event) of TLM. The recrystallization peak of TLM does not appear in any of the DSC thermogram due to partial cancellation of the exothermic and endothermic events occurring simultaneously.<sup>38,39</sup> Surprisingly, the endothermic melting peak of recrystallized TLM did not appear in TPR. A DSC experiment

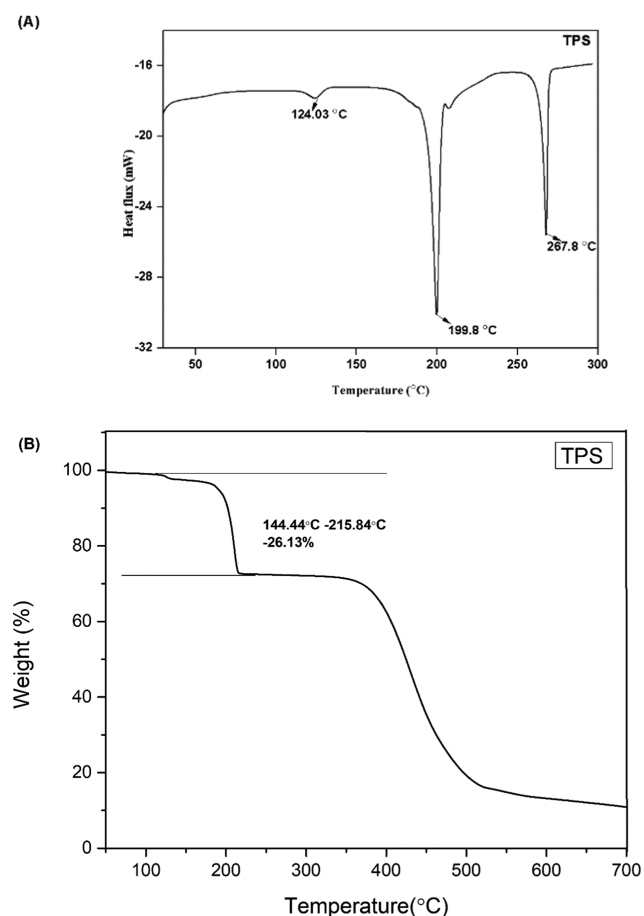


Figure 4. (A) DSC thermogram of TPS and (B) TGA curve of TPS.

was conducted to unveil the reason behind the phenomenon. After melting of the desolvated TPR form, it was isothermally held at 200 °C for 1 h and then cooled to 30 °C to ensure complete recrystallization of TLM from the melt. The endothermic peak of recrystallized TLM appeared in the second heating cycle and is presented in Figure 6.

On analyzing the TGA profile of TPS and TPR (Figures 4B and 5B), there was a weight loss of 26.13 and 26.38% in the temperature range of 144.44–215.84 and 146–238 °C, respectively. The weight loss in TPS corroborates with the theoretical weight loss of 27.16% for the loss of 0.5 mol of ethanol and 1 mol of phthalic acid from the crystal lattice. In the same manner, the weight loss in TPR corroborates with the theoretical weight loss of 25.90% for the loss of 0.3 mol of ethanol and 1 mol of phthalic acid from the crystal lattice. This suggests that stoichiometry of TLM/PA/EtOH in TPS and TPR is 1:1:0.5 and 1:1:0.3, respectively, and this also corroborates with the stoichiometry established from  $^1\text{H}$  NMR analysis.

#### Fourier Transform Infrared (FTIR) Spectroscopy.

Fourier transform infrared (FTIR) spectroscopy can be used as a reliable technique to detect the formation of the multicomponent crystal. The FTIR spectra of telmisartan, phthalic acid, TPS, and TPR are shown in Figure 7, and relevant bands are presented in Table 1. Carboxylic acid (COOH) absorbs strongly at 1700  $\text{cm}^{-1}$  for the C=O stretching band and a weaker C–O stretch around 1200  $\text{cm}^{-1}$ , whereas the carboxylate group (COO $^-$ ) exhibits two characteristic coupled carbonyl absorption band at 1550–

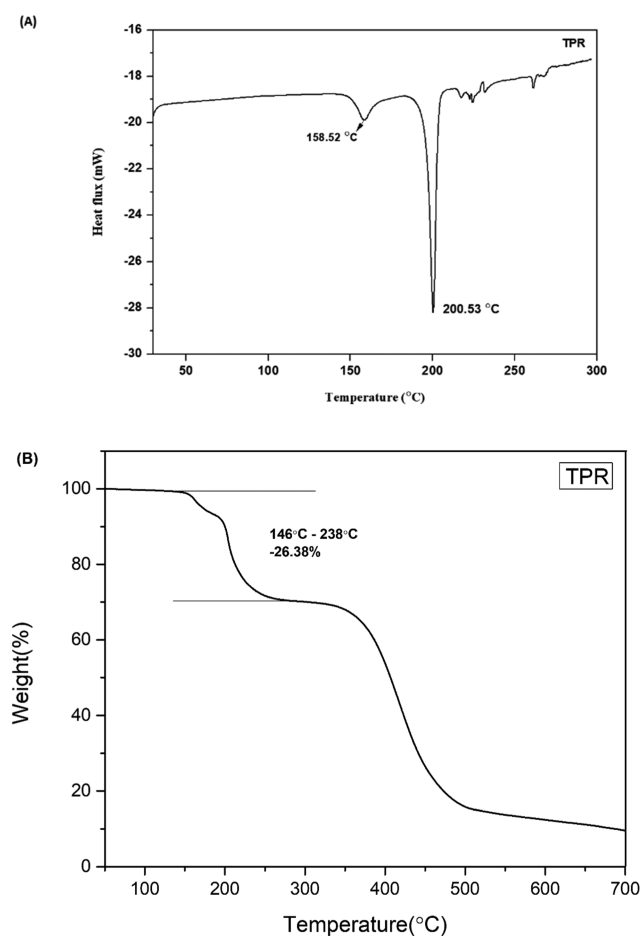


Figure 5. (A) DSC thermogram of TPR and (B) TGA curve of TPR.

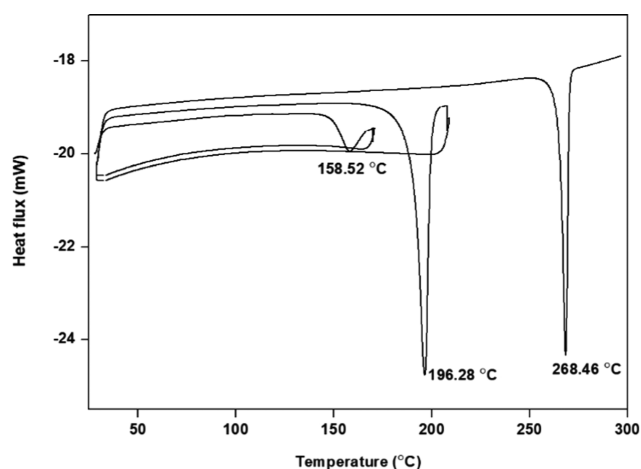
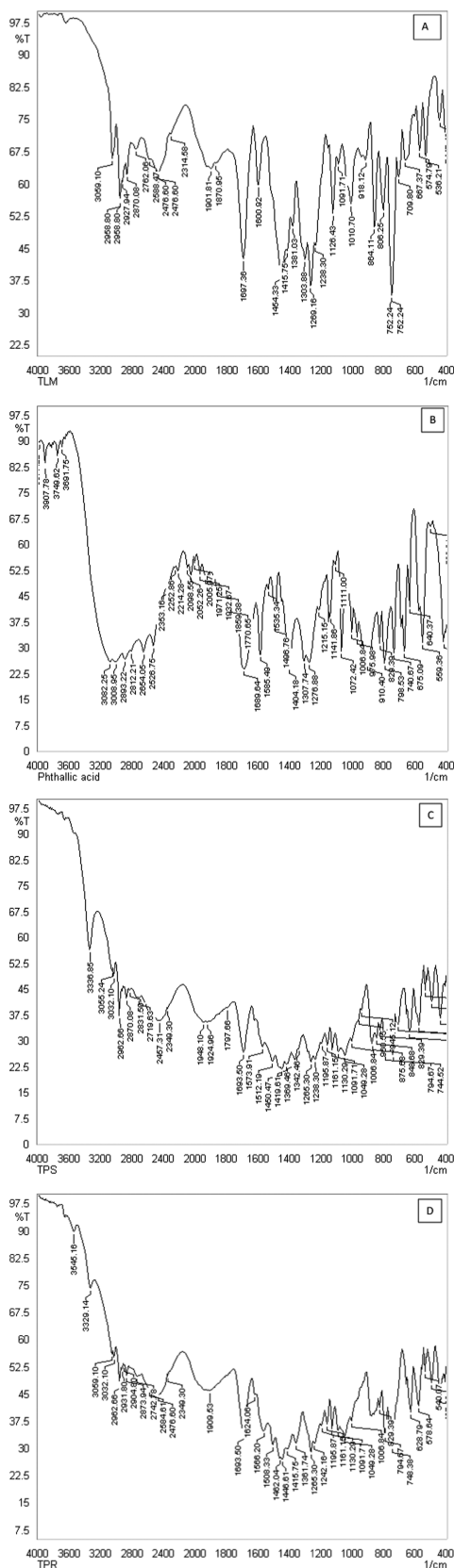


Figure 6. DSC thermogram of TPR (heating–cooling cycle).

1650  $\text{cm}^{-1}$  (asymmetric) and around 1400  $\text{cm}^{-1}$  (symmetric).<sup>6,12,39,40</sup> The carbonyl stretches of carboxylic acid of TLM were observed at 1697  $\text{cm}^{-1}$ , which is not affected and appeared at 1693  $\text{cm}^{-1}$  in TPS and TPR. Furthermore, one of the C=O stretches of PA was also merged within 1693  $\text{cm}^{-1}$ . The second C=O stretch of PA appeared at 1585  $\text{cm}^{-1}$ , which downshifted to 1573 and 1566  $\text{cm}^{-1}$  in TPS and TPR, respectively, due to the formation of H-bonding in the cocrystal system. The identification of C–N stretching is rather difficult because the mixing of bands is possible in this region.



**Figure 7.** FTIR spectra obtained for (A) TLM (B) PA (C) TPS, (D) TPR.

Hence, the FTIR bands at 1303 and 1381  $\text{cm}^{-1}$  have been attributed to the C–N stretching in TLM, which is observed at 1369 and 1361  $\text{cm}^{-1}$  for TPS and TPR, respectively.

The appearance of an additional OH stretching vibration at 3032  $\text{cm}^{-1}$  in TPS and TPR implies the presence of ethanol in the crystal lattice. The ethanol molecule possibly interacts only with C=O of PA, as there is no change in the C=O stretch of TLM. Thus, the FTIR spectra corroborate with  $^1\text{H}$  NMR and SCXRD, confirming that the new solid form (TPS, TPR) is a solvated cocrystal.

**pH-Dependent Solubility Studies.** The pH-dependent solubility of telmisartan is attributed to its three  $\text{pK}_a$  values 3.5, 4.1, and 6.0 corresponding to its carboxylic (acidic) and two benzimidazole (basic) functional groups, respectively.<sup>19,41,42</sup> At acidic pH, the N of benzimidazole of TLM gets ionized, forming a cationic center, and at basic pH, the carboxylic group of TLM is ionized, forming an anionic center, which is responsible for its maximum solubility at extremely high and low pH. Even though TLM exhibits a good solubility profile at extremely high and low pH, it exhibits low solubility in the pH range of 3–7, i.e., in the physiological pH range. When the cocrystal gets dissolved in aqueous solvent, both the drug and cofomer ionize in aqueous solvent; therefore, the solution pH is an important factor in determining its solubility as well as stability. It is important to determine the thermodynamic stability with respect to pH for understanding the transformation of the multicomponent form of telmisartan to its free form. This result will help us achieve the desired dissolution rate and bioavailability over a wide pH range. Thus, the solubility measurement of TPS and TPR was carried out at various pH values in phosphate buffer.

In Figure 8, solubility–pH profiles of telmisartan, TPS, and TPR have been presented. A “U”-shaped solubility–pH profile was obtained for telmisartan due to its amphoteric nature. The ionizing nature of the cofomer imparted modification in the solubility–pH profile of telmisartan and increased its solubility at a lower pH. The solubilities of TPS and TPR have been greatly enhanced by 11-fold and 22-fold at solution pH 5, thereby showing its pH dependency, which can be visualized in Figure 9. The systemic absorption of TLM is found to be highest in small intestine where the pH lies in the range of 5–7.<sup>23,43</sup> Thereby, enhanced solubility of telmisartan at its best site of systemic absorption has been achieved through its cocrystalline form.

Due to the acidic nature of the cofomer, the solution pH was found to be decreased. Therefore, the equilibrium pH was noted upon completion of the solubility experiment and is presented in Table S2, Supporting Information.

**Determination of  $\text{pH}_{\text{max}}$ .** Another important aspect in the solubility–pH profile of an ionizable drug (acidic/basic/amphoteric) is the determination of  $\text{pH}_{\text{max}}$ .  $\text{pH}_{\text{max}}$  aids in understanding the stability of the new solid form with respect to pH and subsequently its in vivo and in vitro performance.<sup>44</sup>  $\text{pH}_{\text{max}}$  is referred to as the intersection point of two independent curves, where one curve represents the drug as the saturating or equilibrium species and the other curve represents the salt as the equilibrium species.<sup>45</sup> The same concept holds for determining the  $\text{pH}_{\text{max}}$  of the cocrystal. From Figure 8,  $\text{pH}_{\text{max}}$  of TPS and TPR can be observed at just below pH 8.

Theoretically, when pH is below  $\text{pH}_{\text{max}}$  the cocrystal is the saturation species; above that, the saturation species is the free base/drug; and  $\text{pH}_{\text{max}}$  is the pH where both the phases coexist. For analyzing this, the residual solids collected from the solubility study of TPS and TPR at different pH were characterized by PXRD and is shown in Figures 10 and 11.

Table 1. Relevant Bands of TLM, PA, TPS, and TPR in Their FTIR Spectra

|                           | TLM        | PA         | TPS              | TPR              |
|---------------------------|------------|------------|------------------|------------------|
| C–O stretching vibrations | 1269       | 1276       | 1265, 1238       | 1265, 1242       |
| C–N stretching vibrations | 1303, 1381 |            | 1369             | 1361             |
| C=O stretching vibrations | 1697       | 1689, 1585 | 1693, 1573       | 1693, 1566       |
| O–H stretching vibrations | 3059       | 3691, 3008 | 3336, 3055, 3032 | 3329, 3059, 3032 |

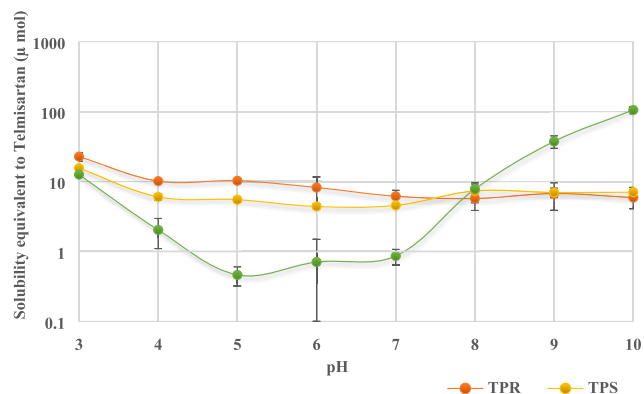


Figure 8. Solubility of TPS, TPR, and TLM at varying pHs (3–10).

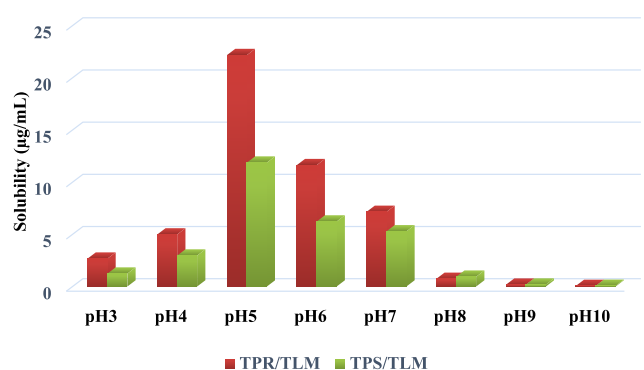


Figure 9. Solubility comparison of TPS, TPR, and TLM at varying pHs (3–10).

The diffraction peaks corresponding to the TPS cocrystal, i.e., at 9.82, 13.96, 19.6, and 20.16, were almost retained in all residual samples collected from the solubility study within the pH of 3–7. Moreover, the diffraction peaks corresponding to the TPR cocrystal at  $2\theta$ , 13.63, 17.01, and 20.63, were almost retained in all residual samples collected from the solubility study within the pH of 3–7. In between pHs 6 and 7, for TPS and TPR, both cocrystals and telmisartan were found to be present in the residual solid, confirmed by the appearance of characteristic peaks of telmisartan at  $2\theta$  6.78 and 14.18° in the PXRD diffractogram along with the peaks of TPS and TPR. This may be concluded that the  $\text{pH}_{\text{max}}$  for TPS and TPR lies around pH 7. When pH is above the  $\text{pH}_{\text{max}}$ , the saturated species would be free base, telmisartan. The appearance of characteristic peaks of telmisartan and disappearance of the characteristic peak of TPS and TPR from the PXRD diffractogram from pH 8 (which is above  $\text{pH}_{\text{max}}$ ) onwards confirmed that the saturation species is telmisartan. Therefore, the PXRD result corroborated well with the concept of  $\text{pH}_{\text{max}}$  in the case of the cocrystal of basic drugs.

## CONCLUSIONS

In conclusion, this work demonstrates the finding of a novel solid form of telmisartan through the crystal engineering approach. Of several cofomers used for screening, phthalic acid formed a new multicomponent solid phase with telmisartan. The new multicomponent solid forms were prepared by two methods, namely, the solution crystallization method (TPS) and reaction crystallization method (TPR). Both the forms were characterized by powder X-ray diffraction (PXRD),  $^1\text{H}$  NMR, differential scanning calorimetry (DSC), thermogravimetric analyses (TGA), and Fourier Transform Infrared Spectroscopy (FTIR). TPS and TPR were found to be

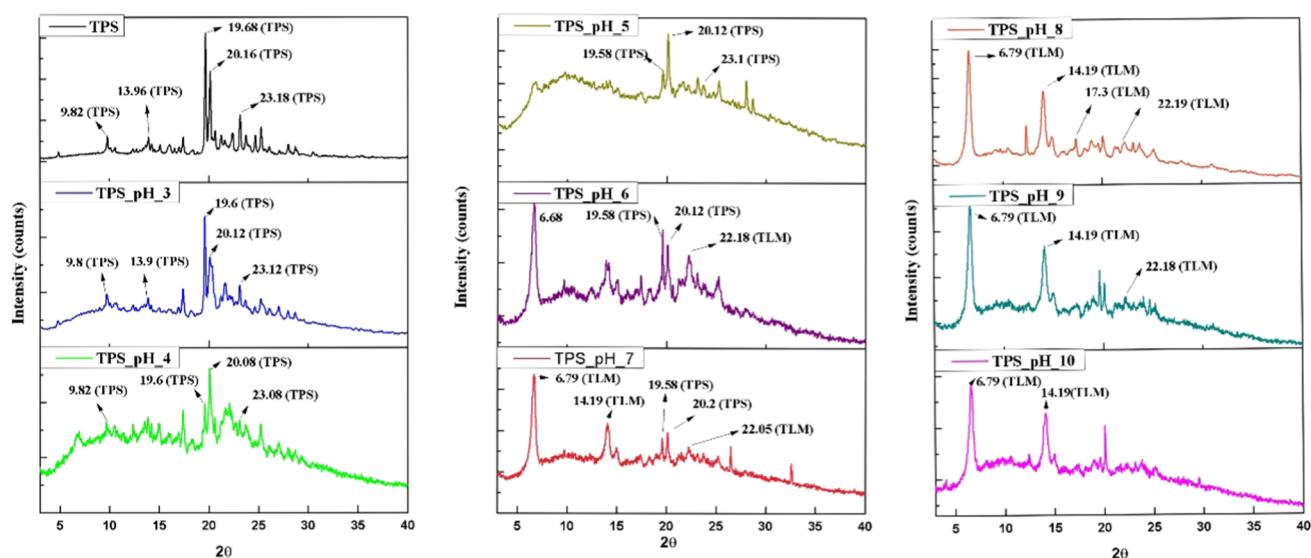
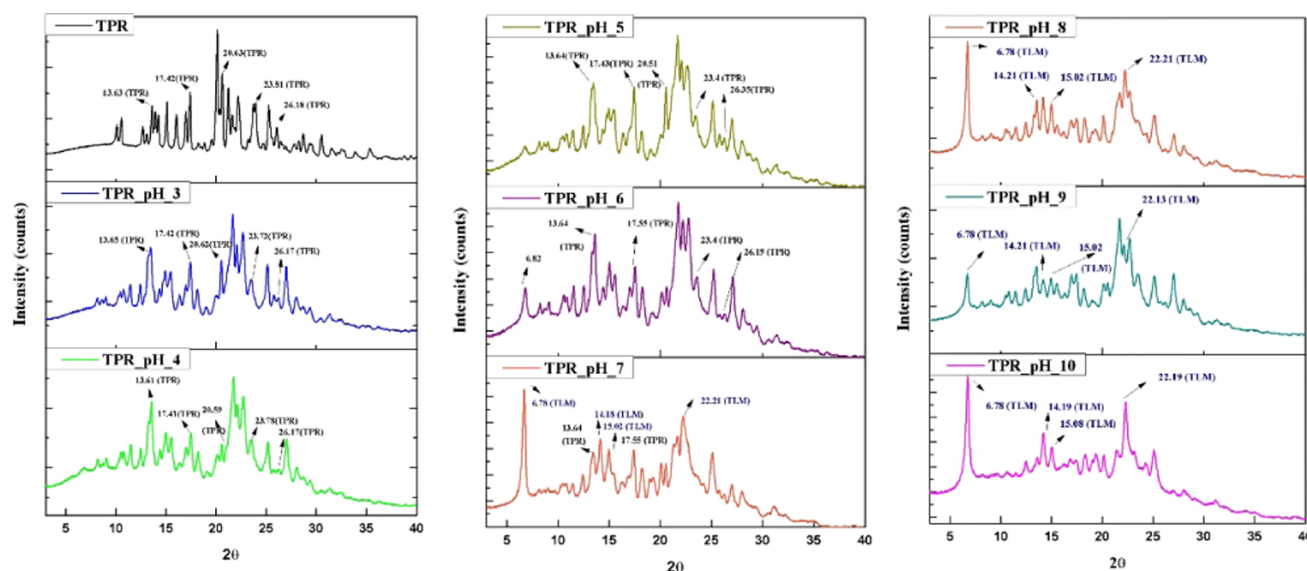


Figure 10. PXRD of residual solids collected from the solubility study of TPS at pH range 3–10.



**Figure 11.** PXRD of residual solids collected from the solubility study of TPR at pH range 3–10.

a solvated cocrystal of telmisartan. The objective of this work of enhancing the solubility of telmisartan within the physiological range was achieved. TPS and TPR markedly increased the solubility of telmisartan by 11-fold and 22-fold at pH 5, the pH at which telmisartan gets absorbed from systemic circulation. The experimental solubility result suggests the selection of coformer to be a crucial step and its  $pK_a$  to modulate the physicochemical properties of an amphoteric API. Thus, by the cocrystallization technique it is possible to improve the pH-dependent solubility of telmisartan, which may eliminate the use of inorganic alkalizers in its marketed dosage form.

## EXPERIMENTAL SECTION

**Materials.** Polymorph A of telmisartan (TLM, 99.7% pure) was obtained as a gift sample from Unichem Laboratories Ltd. (Himachal Pradesh, India). Phthalic acid was procured from Sigma-Aldrich Chemicals and was used without any further purification. The solvents (chloroform, ethanol) used for crystallization were of high-performance liquid chromatography (HPLC) grade (Rankem, India).

**Synthesis of the New Solid Form by the Solution Crystallization Method (TPS).** Equimolar quantity of telmisartan (25 mg, 0.0485 mmol) and phthalic acid (8.05 mg, 0.0485 mmol) was ground in a mortar and pestle for 10 min and the ground mixture was dissolved in 6–7 mL of chloroform:ethanol (2:1 v/v) and allowed to evaporate at room temperature. After 6–7 days, single crystals of suitable size for XRD analysis were obtained. The formation of a new solid phase was preliminarily confirmed by PXRD and then analyzed by DSC, TGA, FTIR,  $^1\text{H}$  NMR, and SCXRD.

**Synthesis of the New Solid form by the Reaction Crystallization Method (TPR).** A saturated solution of telmisartan was prepared in chloroform and ethanol (1:1 v/v). To it, phthalic acid was added in an amount under its solubility limit. The solid precipitate was collected and initially confirmed by PXRD. It was then further analyzed by DSC, TGA, FTIR, and  $^1\text{H}$  NMR.

**Characterization of the New Solid Form. Powder X-ray Diffraction (PXRD).** PXRD patterns were collected on a Rigaku Smart Lab diffractometer system with a 1.5406 Å radiation

wavelength. The tube voltage and current were set at 45 kV and 200 mA, respectively. Each sample was placed in an aluminum sample holder and measured by a continuous scan between 3 and 40° in  $2\theta$  with a step size of 0.02°/min at room temperature and with a scan speed of 5°/min using a semiconductor detector. The experimental PXRD patterns were refined using Smart Lab Guidance software.

**$^1\text{H}$  NMR Spectroscopy.**  $^1\text{H}$  NMR analysis of TPS and TPR was performed on a 400 MHz JEOL NMR spectrometer using  $\text{DMSO}-d_6$  as a solvent at 25 °C. The  $\text{D}_2\text{O}$  exchange experiment was conducted to identify the chemical shift of acidic protons. Stoichiometry of TPS and TPR cocrystal systems was estimated using relative integration of the distinguished  $-\text{CH}_2$  peak (Figure 1) of TLM in comparison to distinguished protons of phthalic acid and ethanol.

**Single-Crystal X-ray Diffraction Analysis of TPS.** X-ray diffraction data were recorded on a SuperNova Eos diffractometer using monochromatic  $\text{Mo K}\alpha$  radiation ( $\lambda = 0.71073$  Å). Data collection was performed at room temperature (293 K) for the samples studied. Olex2 was used to solve the structure using ShelXS and refined with the ShelXL refinement package using least-squares minimization. The observed disordered atoms C17 (C17A and C17B) and C18 (C18A and C18B) from the propyl side chain of TLM were refined using Olex2 and are shown in the ORTEP diagram (Figure 3). All of the nonhydrogen atoms were refined anisotropically, and the hydrogen atoms were placed on the basis of the mixed mode through Fourier difference maps and geometrical assignment.

**Differential Scanning Calorimetry (DSC).** DSC of all samples was conducted using a DSC 4000 instrument (PerkinElmer). Samples (3–5 mg) were crimped in non-hermetic aluminum pans and scanned from 30 to 300 °C at a heating rate of 5 °C/min under a continuously purged dry nitrogen atmosphere at a constant flow rate (20 mL/min). The data were processed using Pyris manager software.

**Thermogravimetric Analysis (TGA).** TGA was performed on a TGA 4000 (PerkinElmer) instrument. The samples (5–8 mg) were placed into a ceramic pan and heated from 30 to 700 °C at a rate of 10 °C/min under nitrogen purge at a flow rate

of 20 mL/min. The data were managed by Pyris manager software.

**Fourier Transform Infrared Spectroscopy (FTIR).** The infrared spectra of the drug, cofomer, and salts were recorded individually by a FTIR-8400S spectrophotometer (Shimadzu, Japan) using the potassium bromide (KBr) pellet method. The IR absorption spectra of samples with 50 scans were measured over the range of 4000–500  $\text{cm}^{-1}$  with a resolution of 4  $\text{cm}^{-1}$  for each sample. The data were analyzed using IR solution software.

**Solubility Measurement.** All solubility measurements were carried out in phosphate buffer at 25 °C. pH was adjusted with potassium hydroxide and hydrochloric acid. Aliquots (0.2 mL) were withdrawn at 72 and 96 h (for ensuring equilibrium conditions) and filtered through a 0.45  $\mu\text{m}$  syringe filter (AXIVA). The concentrations of telmisartan and phthalic acid were analyzed by an HPLC system (Waters TM 486, tunable absorbance detector) equipped with a UV/Vis detector. A C18 Nova-Pak column (5  $\mu\text{m}$ , 4.6  $\times$  250 mm) at ambient temperature with a flow rate of 1 mL/min was used to separate telmisartan and phthalic acid. An isocratic method with methanol and phosphate buffer (10 mM) mixed in a ratio of 72:28 (v/v) was opted for quantitative determination of telmisartan and phthalic acid at an optimum wavelength of 296 nm. Sample injection was 20  $\mu\text{L}$ . EMPOWER software was used for collection and processing the data. The concentrations of telmisartan and phthalic acid were calculated using the standard curve (linearity range 2–16  $\mu\text{g}/\text{mL}$  for both the analytes), which was prepared in a mobile phase. The samples were diluted by methanol prior to analysis. The experiments were conducted in triplicate and were expressed as mean  $\pm$  standard deviation.

## ■ ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsomega.8b02144.

Screening list information;  $^1\text{H}$  NMR spectra; and equilibrium solution pH of TLM, TPS, and TPR (PDF)

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### Notes

The authors declare no competing financial interest.

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