

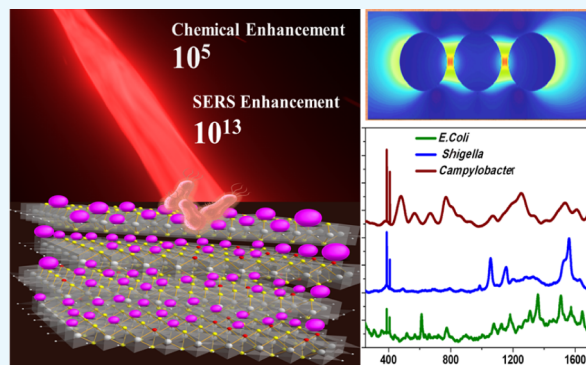
Giant Chemical and Excellent Synergistic Raman Enhancement from a 3D MoS_{2-x}O_x–Gold Nanoparticle Hybrid

Avijit Pramanik,¹ Ye Gao, Kaelin Gates, Salma Begum, and Paresh Chandra Ray*¹

Department of Chemistry and Biochemistry, Jackson State University, Jackson 39217, Mississippi, United States

S Supporting Information

ABSTRACT: Raman spectroscopy fingerprinting features many technological applications. For this purpose, the weak Raman signals need to be boosted dramatically by surface-enhanced Raman spectroscopy (SERS), which provides immense Raman enhancement via plasmonic and chemical mechanisms (CM). In this manuscript, we reveal the giant chemical as well as extremely high SERS enhancement from a three-dimensional MoS_{2-x}O_x–gold nanoparticle (GNP) hybrid, which has capability for ultrasensitive label-free sensing of chemical and biological molecules. Notably, reported data show that the chemical enhancement for the MoS_{2-x}O_x surface is $\sim 10^5$, which is comparable with the plasmonic enhancement factor (EF) by GNP. Reported data show that the total Raman EF is $\sim 10^{13}$ from the GNP–MoS_{2-x}O_x hybrid. Intriguingly, combined experimental and theoretical finite difference time domain stimulation modeling findings show that the synergistic effect of electromagnetic mechanism and CM is responsible for huge SERS enhancement. Experimental results demonstrate that a proposed hybrid SERS platform can be used for fingerprint sensing of different multiple drug resistance bacteria at 5 cfu/mL concentration. Importantly, the current manuscript provides a good strategy for manipulating the SERS sensitivity to 13 orders of magnitude, which is instrumental for next-generation technological applications of Raman spectroscopy.



1. INTRODUCTION

Raman spectroscopy is highly promising for fingerprint identification of chemical and biological molecules.^{1,2} Because of the above unique ability and multiplex detection capability, the Raman technique is highly valuable in forensics, homeland security, and medical diagnosis industry.^{3,4} However, because of the inherently low cross section of Raman scattering, it has not been used as an analytical tool for practical applications.^{5,6} In last few decades, it has been reported that extremely weak Raman signals can be dramatically enhanced by surface-enhanced Raman scattering (SERS) via plasmonic and chemical boosting mechanisms.^{7,8} In SERS, plasmonic enhancement occurs in the presence of plasmonic nanoparticles via electromagnetic mechanism (EM).^{9,10} On the other hand, chemical enhancement occurs via chemical mechanism (CM), which originated from the charge transfer between the Raman active molecule and the SERS substrate.^{11,12} In the last 2 decades, we and other groups have reported different types of SERS materials which are based on the noble plasmonic metal nanoparticle, where Raman intensity can be enhanced several orders of magnitude (10^6 or higher) via EM.^{13,14} On the other hand, for most of the reported SERS substrate, the reported chemical enhancement factor (EF) is $\sim 10^2$.^{15,16} Recently, the SERS substrate based on two-dimensional (2D) transition-metal dichalcogenides has been reported,^{17–25} where the chemical EF can be much

higher than 10^2 , and in this case, the laser excitation can be resonant to charge transfer and exciton transitions in an analyte 2D system.^{26–32} For real-life applications, an SERS probe should possess strong electromagnetic as well as strong chemical enhancement capability for providing excellent sensitivity. Herein, we report huge chemical (CM) and electromagnetic (EM) enhancements from a three-dimensional (3D) MoS_{2-x}O_x–gold nanoparticle (GNP) hybrid. Experimental data reported here indicate that the chemical EF is $\sim 10^5$ from the MoS_{2-x}O_x surface, which is comparable with the plasmonic enhancement by GNP. Reported data demonstrated that oxygen incorporation on MoS₂ can effectively improve the SERS performance via a strong chemical enhancement mechanism. On the other hand, the total SERS enhancement from the GNP–MoS_{2-x}O_x hybrid was observed to be $\sim 10^{13}$. Our experimental and theoretical finite difference time domain (FDTD) stimulation modeling^{22,23} shows that the synergistic effect of EM and CM is responsible for huge SERS enhancement. To demonstrate that MoS_{2-x}O_x–GNP-based ultrasensitive SERS is versatile for fingerprinting biological analysis, we have shown that an SERS platform can be used for fingerprint sensing of different

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multiple drug resistance bacteria such as carbapenem-resistant *Escherichia coli*, drug-resistant *Shigella*, and *Campylobacter*, at 5 cfu/mL concentration.

2. RESULTS AND DISCUSSION

As shown in Figure 1, we used a three-step method for the synthesis of a 3D MoS_{2-x}O_x-GNP hybrid. In the first step, a

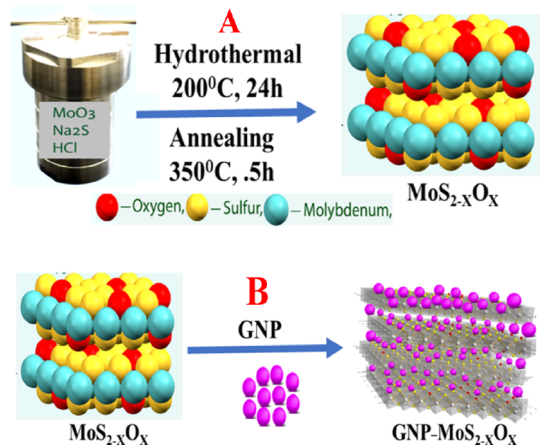


Figure 1. (A) Scheme shows the synthetic route we have used to develop 2D MoS_{2-x}O_x via a hydrothermal process as well as an annealing process. (B) Scheme shows the synthetic route we have used to develop GNP-MoS_{2-x}O_x nanocomposites.

facile hydrothermal synthetic method was used for the synthesis of MoS₂ nanosheets. Experimental details have been reported in the [Experimental Section](#). In brief, in the first step, molybdenum(VI) oxide powder, sodium sulfide (Na₂S), and HCl were mixed and kept into a Teflon-lined stainless steel autoclave. After that, the mixture was heated for 200 °C overnight. A black precipitate was obtained by centrifugation from the final reaction products and then the 2D MoS₂ was washed with distilled water and ethanol. Because it is now well-known that oxygen incorporation is the effective way to improve the SERS performance of nonmetal oxide semiconductors,^{20–22} in the next step, we have synthesized a 2D MoS_{2-x}O_x nanosheet. For this purpose, we have developed a MoS_{2-x}O_x nanosheet, via annealing of 2D MoS₂ at 350 °C temperature in air. In the third step, we have developed a 3D MoS_{2-x}O_x-GNP hybrid. For this purpose, we have used GNP as a linker between 2D MoS_{2-x}O_x nanosheets to form the 3D MoS_{2-x}O_x-GNP hybrid, via a -Mo-S-Au- bond.

After that, we have used different electron microscopic and spectroscopic techniques^{6,11–16} to characterize the 3D MoS_{2-x}O_x-GNP hybrid as reported in [Figures 2A–C](#) and [S1A–G](#). The elemental molar ratios of the 3D MoS_{2-x}O_x-GNP hybrid were determined using energy-dispersive X-ray (EDX), X-ray diffraction (XRD), and Raman data. [Figure S1A](#) reports the transmission electron microscopy (TEM) image which indicates that the size of the GNP we have synthesized is about 25 nm. [Figure S1B](#) reports the scanning electron microscopy (SEM) which shows the morphology of 2D MoS_{2-x}O_x nanosheets. [Figure 2A](#) reports the SEM image which shows the morphology of the 3D MoS_{2-x}O_x-GNP hybrid, which indicates that a porous structure is developed with a pore diameter varying from 10 to 400 nm. Both [Figures 2A](#) and [S1C](#) show the formation of a “hot spot” by the GNP on the MoS_{2-x}O_x nanosheets. [Figure S1E](#) shows the EDX data

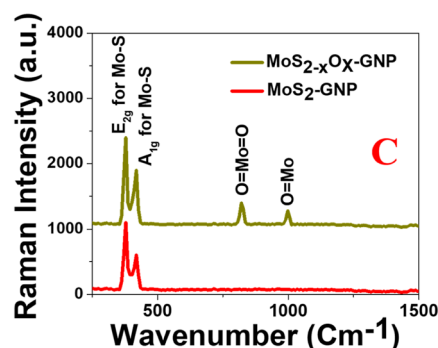
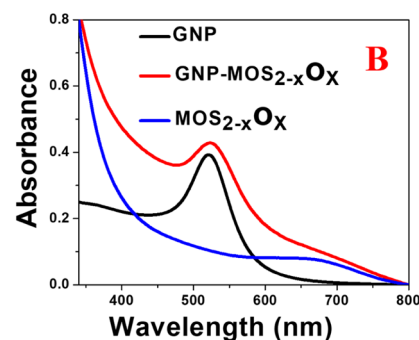
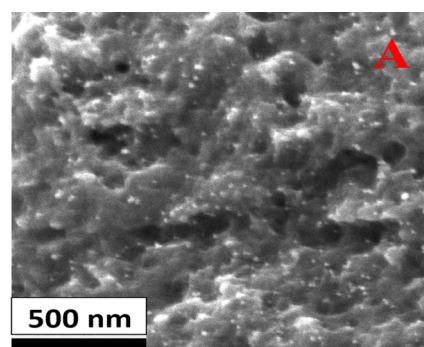


Figure 2. (A) SEM image shows the morphology of MoS_{2-x}O_x-GNP hybrid. (B) Extinction spectra from MoS_{2-x}O_x-GNP hybrid, GNP, and MoS_{2-x}O_x. (C) Raman spectra from MoS_{2-x}O_x-GNP hybrid and MoS₂-GNP hybrid.

from the MoS_{2-x}O_x-GNP hybrid, which clearly shows the presence of Mo, S, O, and Au. Similarly, [Figure S1F](#) shows the EDX data from MoS₂, before annealing, which clearly shows the presence of Mo and S.

[Figure S1G](#) shows the powder XRD data from the MoS_{2-x}O_x-GNP hybrid, which shows the presence of (002), (100), (104), and (201) reflection for MoS₂,^{20–23} (020) reflection for MoO₃,^{16–19} and (111) and (3111) reflection for GNP. [Figure 2B](#) shows the absorption spectra for GNP, 2D MoS_{2-x}O_x nanosheets, and 3D MoS_{2-x}O_x-GNP hybrid. Reported data clearly indicate a broad plasmon band from the 3D MoS_{2-x}O_x-GNP hybrid which is due to the aggregation of GNP on the 3D hybrid, as shown in the TEM and SEM images reported in [Figures S1C](#) and [2A](#). To compare the Raman spectra between the MoS_{2-x}O_x-GNP hybrid and the MoS₂-GNP hybrid, we have also synthesized the MoS₂-GNP hybrid. For this purpose, we have used a GNP as a linker between 2D MoS₂ nanosheets to form the MoS₂-GNP hybrid, via a -Mo-S-Au- bond. Experimental details have been reported in the [Supporting Information](#). The TEM image from the MoS₂-GNP hybrid, as reported in [Figure](#)

SID, clearly shows the aggregation of GNP on the MoS₂ surface. Figure 2C shows that the Raman spectra from the 3D MoS_{2-x}O_x-GNP hybrid and MoS₂-GNP hybrid clearly indicate the presence of an in-plane (E_{2g}) Raman band at ~384 cm⁻¹ and an out-of-plane (A_{1g}) Raman band at ~409 cm⁻¹, which is due to Mo-S vibration of MoS₂.²⁰⁻²³ We have observed a (E_{2g}) Raman band and a (A_{1g}) Raman band for MoS_{2-x}O_x-GNP hybrid, as well as for MoS₂-GNP hybrid. Similarly, as reported in Figure 2C, we have also observed Raman peaks at ~820 and ~996 cm⁻¹, which are due to the Mo=O vibration.¹⁹⁻²² Raman peaks at ~820 and ~996 cm⁻¹ are only observed for the MoS_{2-x}O_x-GNP hybrid, which has been developed after annealing of MoS₂, as we have discussed previously.

Because Raman EF is most important for a Raman substrate, we have measured Raman EF using a 4-aminothiophenol (4-ATP) and Rh-6G dye. For the Raman EF measurement, we have used a portable Raman probe, where a continuous wavelength 670 nm diode-pumped solid-state laser was used as the excitation source. We have used fiber optics probe for excitation and data collection.^{6,11-16} Experimental details are reported in the Experimental Section. Figure 3A shows strong Raman spectra from 4-ATP (10⁻⁶ M) on MoS_{2-x}O_x surface. On the other hand, in the same condition, we have not observed any Raman peak from 4-ATP (10⁻⁶ M), when a bulk sample was used. Reported Raman data reported in Figure 3A show that dominated vibrational peaks are due to the a₁ vibrational mode peaks and these are ν(CC + NH₂ bend) at ~1590 cm⁻¹ and ν(CS) at ~1078 cm⁻¹.⁵⁻⁸ As reported in Figure 3A, we have also observed Raman peaks due to b₂ modes, at ~1435 cm⁻¹ due to the CC str in Ph ring + NH₂ rock, and at ~1170 cm⁻¹ due to CH bend vibration.¹⁰⁻¹³

As reported in Figure 3A, we have observed a clear 1078 cm⁻¹ vibrational band from ATP in the bulk and on all surfaces, and as a result, we have used 1078 cm⁻¹ vibrational band intensity for Raman EF calculation. From the Raman spectra, we have measured the Raman EF using the following equation.¹⁰⁻¹²

$$G = [I_{\text{MoS}_{2-x}\text{O}_x}] / [I_{\text{bulk}}] \times [M_{\text{bulk}}] / [M_{\text{ads}}] \quad (1)$$

where $I_{\text{MoS}_{2-x}\text{O}_x}$ is the intensity of 1078 cm⁻¹ vibrational mode from 4-ATP on MoS_{2-x}O_x surface. Similarly, I_{bulk} is the intensity of 1078 cm⁻¹ vibrational band in the absence of MoS_{2-x}O_x surface. M_{bulk} is the number of 4-ATP used in the bulk experiment without MoS_{2-x}O_x surface and M_{ads} is the number of 4-ATP used for the Raman experiment on MoS_{2-x}O_x surface. For bulk experiment, a Si/SiO₂ wafer was used as the normal Raman reference. For Raman experiment on all different surfaces, we have assumed that the analytes were distributed uniformly on the surface. The number of molecules is calculated using a laser spot size of 40 μm. From the experimental data on MoS_{2-x}O_x surface as reported in Figure 3A, we found out that the EF is ~1.3 × 10⁵. Because of the lack of surface plasmons in the visible light for MoS_{2-x}O_x surface, the observed huge Raman enhancement on MoS_{2-x}O_x surface can be attributed to the chemical enhancement mechanism.

The huge chemical enhancement on MoS_{2-x}O_x surface is due to the photon-induced charge transfer from the MoS_{2-x}O_x surface to the adsorbed 4-ATP molecule. As reported in Figure S2C in the Supporting Information, the GNP-adsorbed 4-ATP molecule exhibits a new absorption peak with λ_{max} around 670

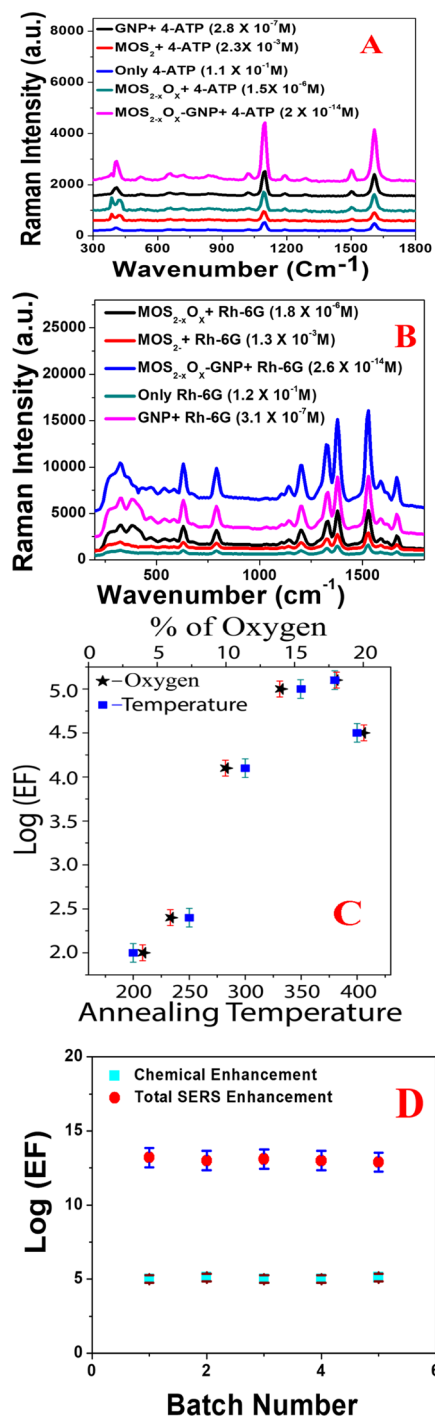


Figure 3. (A) Raman profile of 4-ATP in the presence of GNP, MoS₂, MoS_{2-x}O_x, GNP-MoS_{2-x}O_x surface, and without any surface. (B) Raman profile of Rh-6G on MoS₂, MoS_{2-x}O_x, GNP, GNP-MoS_{2-x}O_x surface, and without any surface. (C) Plot shows how the Raman EF for 4-ATP on MoS_{2-x}O_x surface varies with annealing temperature, as well as with the percentage of oxygen incorporation. (D) Plot shows how the chemical EF and total Raman EF factor vary with samples made in different batches.

nm. We believe that the charge-transfer resonance from the MoS_{2-x}O_x surface to the adsorbed 4-ATP molecule coupled with exciton resonance as well as with molecular resonances, and as a result, we have observed huge chemical enhancement.¹⁶⁻¹⁹ As we have discussed previously, a recent report indicates that oxygen incorporation can effectively improve the

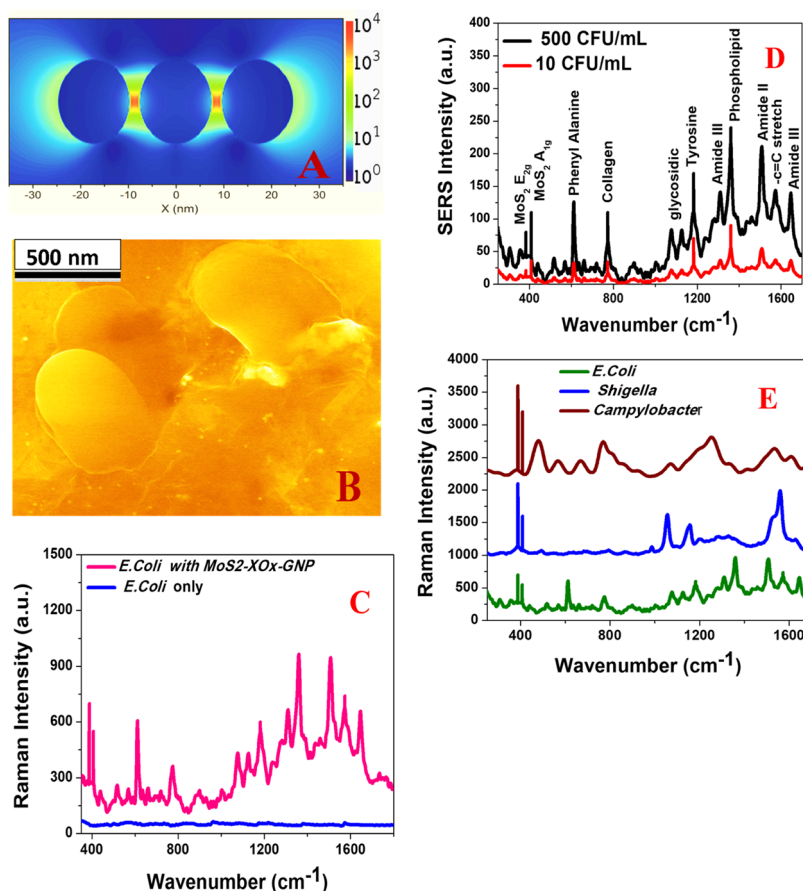


Figure 4. (A) 3D FDTD simulated electric field enhancement square ($|E|^2$) profiles for GNP assembly containing three nanoparticles. (B) SEM image shows that the drug-resistant *Campylobacter* bacteria are on the surface of 3D $\text{MoS}_{2-x}\text{O}_x$ -GNP. (C) Raman spectrum shows a huge signal from drug-resistant *E. coli* (3000 cfu/mL) on 3D $\text{MoS}_{2-x}\text{O}_x$ -GNP, whereas we have not observed any Raman signal from drug-resistant *E. coli* (3000 cfu/mL) in the absence of 3D $\text{MoS}_{2-x}\text{O}_x$ -GNP surface. (D) Plot shows Raman spectrum from drug resistant *E. coli* at different concentrations on 3D $\text{MoS}_{2-x}\text{O}_x$ -GNP. (E) Raman spectrum shows that 3D $\text{MoS}_{2-x}\text{O}_x$ -GNP-based Raman can be used as a fingerprint for different bacteria such as *E. coli*, *Shigella*, and *Campylobacter*.

SERS performance of a semiconductor.^{22–25} To understand better, we have also performed Raman experiment on the MoS_2 surface. Reported experimental data in Figure 3A clearly indicate that Raman EF is much higher on $\text{MoS}_{2-x}\text{O}_x$ surface than that of MoS_2 surface. From the experimental data, we found out that the Raman EF for MoS_2 surface is $\sim 2.3 \times 10^2$, whereas the EF is $\sim 1.3 \times 10^5$ for $\text{MoS}_{2-x}\text{O}_x$. To understand how oxygen incorporation affects Raman intensity, we have developed $\text{MoS}_{2-x}\text{O}_x$ surface from MoS_2 by annealing at different temperatures, where the x value should be higher at higher temperature. As reported in Figure 3C, experimental data show that the Raman EF increases as the annealing temperature increases, which is due to the high amount of oxygen incorporation. Reported data in Figure 3C also indicate that the Raman EF decreases above 400 °C, which is mainly due to the phase change as we have noted from the XRD study. We have measured the S and O ratio using EDX, XRD, and Raman data and we found out that the oxygen percentage increases from 4% at 200 °C to 20% at 350 °C. As reported in Figure 3C, experimental data show that the Raman EF increases with the high amount of oxygen incorporation. As reported in Figure 3B, from experimental data using Rh-6G as a bulk and on the surface, we have found out that the Raman EF for MoS_2 surface is $\sim 1.3 \times 10^2$ and the EF is $\sim 1.8 \times 10^5$ for $\text{MoS}_{2-x}\text{O}_x$, which is very similar to the observed data with 4-

ATP. Figure 3A indicates that the Raman EF on $\text{MoS}_{2-x}\text{O}_x$ surface is comparable with the GNP surface. From experimental data using 4-ATP, we have found out that the Raman EF is $\sim 2.2 \times 10^6$ for GNP, which is around an order of magnitude higher than the $\text{MoS}_{2-x}\text{O}_x$ surface. As reported in Figure S2A in the Supporting Information, the GNP-adsorbed 4-ATP molecule exhibits a new absorption peak with λ_{max} around 670 nm, which is due to the GNP aggregation because of the interaction between GNP and 4-ATP. Very interestingly, Raman data reported in Figure 3A,B indicate that the Raman EF is $\sim 4.5 \times 10^{13}$ for GNP- $\text{MoS}_{2-x}\text{O}_x$ surface, which is around 8 orders of magnitude higher than the $\text{MoS}_{2-x}\text{O}_x$ surface and 7 orders of magnitude higher than the GNP surface. The observed extremely high Raman EF from GNP- $\text{MoS}_{2-x}\text{O}_x$ surface is due to the strong electromagnetic as well as strong chemical enhancement capability, which provide excellent enhancement.

For our GNP- $\text{MoS}_{2-x}\text{O}_x$ surface, the GNP enhances the Raman signal via EM, and on the other hand, $\text{MoS}_{2-x}\text{O}_x$ enhances the Raman signal via a chemical enhancement mechanism. As we have reported in this manuscript, the Raman EF for only GNP is 2.2×10^6 and the same for $\text{MoS}_{2-x}\text{O}_x$ is $\sim 1.2 \times 10^5$, whereas the Raman EF is $\sim 4.5 \times 10^{13}$ for the GNP- $\text{MoS}_{2-x}\text{O}_x$ surface. We observed 2 orders of magnitude higher Raman EF for GNP. The $\text{MoS}_{2-x}\text{O}_x$ surface

is mainly due to the formation of a “hot spot” by a GNP in 3D interior and exterior surfaces, as shown in the SEM and TEM images reported in Figures 2A and S1C. To find out the origin of a synergistic enhancement mechanism, we have performed the 3D FDTD simulation to understand the “hot spot”-based plasmon coupling, which allows a huge EM enhancement mechanism.^{6,11–13,35,36} Calculation details are reported in the Experimental Section and also reported previously by our group.^{6,11–16} As reported in Figure 4A, FDTD simulation data show that the field enhancement for GNP aggregates in “hot spots” can be more than an order of magnitude higher than that of the individual GNP.

Because Raman EF varies with the square of field EF, we expect to increase the Raman enhancement around 2–3 orders of magnitude because of the “hot spot” formation. It is well documented that the reproducibility and stability of Raman EF are very important criteria for applications.^{1–5} For this purpose, we have developed a GNP–MoS_{2-x}O_x surface and only MoS_{2-x}O_x surface in different batches and then monitored the reproducibility of the Raman EF using 4-ATP. Figure 3D reports the reproducibility data for Raman EF and chemical enhancement, which indicate very good reproducibility with a relative standard deviation around 6.2%.

Next, to understand whether our 3D GNP–MoS_{2-x}O_x can be used for trace-level fingerprint sensing of multi-drug-resistant superbugs, we have used carbapenem-resistant *E. coli*, drug-resistant *Shigella*, and *Campylobacter*. For this purpose, we have added different superbugs such as carbapenem-resistant *E. coli*, drug-resistant *Shigella*, and *Campylobacter* at different concentrations (cfu/mL) to the GNP–MoS_{2-x}O_x surface. As shown in Figure 4B, drug-resistant *Campylobacter* bacteria are on the surface of 3D MoS_{2-x}O_x–GNP. Figure 4C,D shows strong Raman peak from carbapenem-resistant *E. coli* even at 5 cfu/mL level. On the other hand, we have not observed the Raman peak from carbapenem-resistant *E. coli* in the absence of GNP–MoS_{2-x}O_x surface, even at 3000 cfu/mL level. Although we have observed huge Raman enhancement using GNP–MoS_{2-x}O_x surface, as we have not observed any Raman peak from carbapenem-resistant *E. coli* in the absence of GNP–MoS_{2-x}O_x surface, as reported in Figure 4C, we are not able to determine the Raman EF using carbapenem-resistant *E. coli*. As reported in Figure 4D, in Raman spectra, we have observed in-plane (E_{2g}) Raman and out-of-plane (A_{1g}) Raman bands because of the MoS_{2-x}O_x surface. On the other hand, as reported in Table 1 and Figure 4D, we have observed amide I, II, and III bands, phenyl alanine, tyrosine, collagen, phospholipid, and glycosidic bands^{5–7,11,14–16,33,34} because of carbapenem-resistant *E. coli*. Raman spectra from carbapenem-resistant *E. coli*, drug-resistant *Shigella*, and *Campylobacter*, reported in Figure 4E and Table 1, clearly indicate that 3D GNP–MoS_{2-x}O_x can be used for fingerprint Raman detection of different superbugs. As reported in Figure 4E, phospholipid and amide-III bands are unique for carbapenem-resistant *E. coli*, which have not been observed for drug-resistant *Shigella* and *Campylobacter*. On the other hand, guanine, tyrosine, and adenine nucleic acid bands are more prominent for *Campylobacter*, which we have not been observed for carbapenem-resistant *E. coli* and drug-resistant *Shigella*. Similarly, lipid bands near 1480 cm⁻¹ are unique for *Shigella*.

3. CONCLUSIONS

In conclusion, our findings reveal that the 3D MoS_{2-x}O_x–GNP hybrid provides immense Raman enhancement via giant

Table 1. Fingerprint Raman Modes Observed from Drug-Resistant *E. coli*, *Shigella*, and *Campylobacter*^a

| Raman peak (cm ⁻¹) for <i>E. coli</i> | Raman peak (cm ⁻¹) for <i>Shigella</i> | Raman peak (cm ⁻¹) for <i>Campylobacter</i> | vibration mode |
|---|--|---|---------------------------------|
| 1646 | | | amide I |
| 1574 | | | adenine guanine ring stretching |
| | 1480 | | lipid |
| 1507 | | | amide II |
| 1301 | | | amide III |
| | | 1240 | DNA/RNA bases |
| | 1180 | | phospholipids |
| | 1110 | | fatty acid in lipids |
| | | 814 | –O–P–O– for DNA |
| 780 | | | Collagen |
| | | 747 | tyrosine |
| | | 676 | guanine |

^aAll the bands have been assigned using reported data from different microorganisms.^{5–7,11,14–16,33,34}

chemical enhancement mechanisms, as well synergistic plasmonic enhancement mechanism. Reported data show that because of the presence of MoS_{2-x}O_x in our hybrid, the chemical EF is ~10⁵, which is comparable with plasmonic EF by plasmonic nanoparticle. We have demonstrated that oxygen incorporation on MoS₂ can effectively improve the SERS performance via a strong chemical enhancement mechanism, which is due to the photon-induced charge transfer from the MoS_{2-x}O_x surface to the adsorbed molecule. On the other hand, the total Raman EF is ~10¹³ from GNP–MoS_{2-x}O_x hybrid because of the synergistic effect of electromagnetic and chemical enhancement mechanisms. The reported synergistic Raman EF is mainly due to the “hot spot” formation by GNP in 3D interior and exterior surfaces. Our experimental results show that the GNP–MoS_{2-x}O_x hybrid has the capability for ultrasensitive label-free sensing of carbapenem-resistant *E. coli*, drug-resistant *Shigella*, and *Campylobacter*, even at 5 cfu/mL concentration level.

4. EXPERIMENTAL SECTION

Molybdenum(VI) oxide powder, sodium sulfide (Na₂S) and HCl, different solvents, and other chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA). Superbugs such as carbapenem-resistant *E. coli*, drug-resistant *Shigella*, and *Campylobacter* and growth media were purchased from American Type Culture Collection (ATCC, Rockville, MD).

4.1. Synthesis of the Gold Nanoparticles. Spherical-shaped gold nanoparticles (AuNPs) were synthesized according to the previous work by our group.^{6,11–16} For this purpose, we have used 1.25 mL of 10 mM HAuCl₄ solution and 2 mL of 1% trisodium citrate dihydrate. At the end, the purified AuNPs were characterized by a microscopic technique, as reported in Figure S1A.

4.2. Synthetic Procedure of MoS₂ Nanosheets. A facile hydrothermal synthetic method was adopted for the synthesis of MoS_{2-x}O_x nanosheets. In a typical experiment, 0.72 g of molybdenum(VI) oxide (MoO₃) powder and 3.6 g of sodium sulfide (Na₂S) were mixed gently and the mixture was transferred into a Teflon-lined stainless steel autoclave of capacity 100 mL. Then, approximately 70 mL of 0.2 N HCl

was added to fill the autoclave up to 75–80% of the total volume. After that, the autoclave was tightly sealed, maintained at 200 °C overnight, and then cooled to room temperature. A black precipitate was obtained by centrifugation from the final reaction products and washed successively with distilled water and ethanol several times. Finally, the semisolid MoS_{2-x}O_x nanosheets were dried in vacuum at 60 °C for 6 h. The pure products are characterized by SEM and other spectroscopic and microscopic techniques, as reported in Figure S1.

4.3. Synthesis of the MoS₂–GNP Hybrid. To synthesize AuNP-decorated MoS₂ nanosheets, 5.0 mL of 10 nM gold nanoparticle solution was dropped in 10 mL of 5 ppm of dispersed in freshly prepared MoS₂ solutions in phosphate-buffered saline (PBS) buffer and the mixture was sonicated for 2 h at room temperature. The mixture was continuously stirred at very low speed overnight at room temperature for the completion of the reaction. To remove excess reagents and buffer solution, the mixed solution was washed with methanol two to three times by centrifugation at 5000 rpm for 15 min followed by decantation. Finally, the AuNP-decorated MoS₂ pellet was dried under vacuum at room temperature for a week. Figure S1D shows the TEM image of freshly prepared MoS₂–GNP hybrid.

4.4. Synthetic Procedure of MoS_{2-x}O_x Nanosheets. Because it is now well-known that oxygen incorporation is the effective way to improve the SERS performance of nonmetal oxide semiconductors,^{12–19} in the next step, we have synthesized a 2D MoS_{2-x}O_x nanosheet. For this purpose, we have developed a MoS_{2-x}O_x nanosheet, via annealing of 2D MoS₂ at 350 °C temperature in air. The pure products are characterized by SEM and other spectroscopic and microscopic techniques, as reported in Figure S1.

4.5. Synthesis of the MoS_{2-x}O_x–GNP Hybrid. To synthesize AuNP-decorated MoS_{2-x}O_x nanosheets, 5.0 mL of 10 nM gold nanoparticle solution was dropped in 10 mL of 5 ppm of dispersed in freshly prepared MoS_{2-x}O_x solutions in PBS buffer and the mixture was sonicated for 2 h at room temperature. The mixture was continuously stirred at very low speed overnight at room temperature for the completion of the reaction. To remove excess reagents and buffer solution, the mixed solution was washed with methanol two to three times by centrifugation at 5000 rpm for 15 min followed by decantation. Finally, the AuNP-decorated MoS_{2-x}O_x pellet was dried under vacuum at room temperature for a week. After that, the purified MoS_{2-x}O_x–GNP hybrid was characterized by powder XRD, high-resolution tunneling electron microscopy, EDX spectroscopy, and Raman spectroscopy, as reported in Figure S1A–G. The elemental molar ratios for MoS_{2-x}O_x–GNP hybrid were determined using inductively coupled plasma–mass spectrometer data and EDX data.

4.6. Superbug Sample Preparation. Carbapenem-resistant *E. coli*, drug-resistant *Shigella*, and *Campylobacter* superbugs were cultured according to the ATCC protocol, as we have reported previously.^{6,11–16}

4.7. Raman Experimental Details. We have used a portable Raman probe for the fingerprint detection of different superbugs, as we have reported previously.^{6,11–16} For the Raman experiments using MoS_{2-x}O_x–GNP hybrid and other materials, we have used 670 nm light as the excitation light source and a QE65000 spectrometer for Raman data collection.

4.8. 3D FDTD Simulation. We have used the 3D FDTD simulation age for full-field electromagnetic wave calculations,

as we have reported previously.^{6,11–16} For the calculation, we have used a gold nanoparticle of 30 nm diameter which is decorated on the MoS_{2-x}O_x nanosheet as we have observed experimentally. 670 nm was used as the incident wavelength and the entire process has been performed under 0.001 nm mesh resolution and 4000 fs duration.^{11–16}

■ ASSOCIATED CONTENT

📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsomega.9b00866.

Microscopic and spectroscopic characterization data (PDF)

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: paresh.c.ray@jsums.edu. Fax: +16019793674.

ORCID

Avijit Pramanik: 0000-0002-4623-2099

Paresh Chandra Ray: 0000-0001-5398-9930

Notes

The authors declare no competing financial interest.

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