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Novel Approaches to Ovarian Cancer Screening

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Abstract

Purpose of Review: Both conventional and novel approaches to early detection of ovarian cancer are reviewed in the context of new developments in our understanding of ovarian cancer biology.

Recent Findings—While CA125 as a single value lacks adequate specificity or sensitivity for screening, large studies have shown that a 2 stage strategy which tracks CA125 change over time and prompts transvaginal ultrasound (TVS) for a small subset of women with abnormally rising biomarker values achieves adequate specificity and detects a higher fraction of early stage disease. Sensitivity could clearly be improved in both blood tests and in imaging. Metastasis can occur from ovarian cancers too small to increase blood levels of protein antigens and a significant fraction of ovarian cancers arise from the fimbriae of fallopian tubes that cannot be imaged with TVS. Autoantibodies, miRNA, ctDNA, DNA methylation in blood and cervical mucus might improve sensitivity of the initial phase and magnetic relaxometry and auto-fluorescence could improve imaging in the second phase.

Summary—Enhancing the sensitivity of two stage strategies for early detection could reduce mortality from ovarian cancer.

Keywords

CA 125; ovarian cancer screening; novel approaches; HE4; fallopian tube cytology; TP53; miRNA; ctDNA

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Compliance with Ethical Standards

Conflict of Interest

Denise R. Nebgen declares that she has no conflict of interest.

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Introduction to Ovarian Cancer

The lifetime risk of ovarian cancer is approximately 1.3% in the general population; however, it has the highest mortality of all gynecologic malignancies.[1] Ovarian cancer incidence increases with age, particularly after age 45, with the median age at diagnosis of 63 years.[2, 3] In 2018, there were approximately 22,240 new cases of ovarian cancer and 14,070 deaths in the United States,[4] with 295,414 new cases and 184,799 deaths worldwide.[5] Ovarian cancer is the fifth leading cause of cancer deaths among US women and the eighth leading cause of death among women worldwide.[6]

If ovarian cancer is detected early, the 5-year survival rates are 90% if confined to the ovary (Stage I) or 70% if confined to the pelvis (Stage II). However, most ovarian cancer are diagnosed at Stages III (51%) and IV (29%),[1] when 5 year survival rates are less than 30%.[7, 1] Overall 5-year survival ranges between 30% and 40% worldwide, and has increased little (2–4%) over the last two decades.[8] Additionally, 70% of patients with advanced epithelial ovarian cancer will have cancer recurrence, after which time survival is extremely low.

Biology of Ovarian Cancer

Ovarian cancers are divided into epithelial ovarian cancers (EOCs), which are the most common type and make up 90% of cases, and non-epithelial cancers representing only 10% of cases. Epithelial ovarian cancers arise from the simple flattened surface epithelial cells that cover the ovary, subserosal inclusion cysts, and/or the fimbriated end of the fallopian tubes.[9, 10] These epithelial cells transform into different histotypes including cells that resemble the lining of the fallopian tube (serous, 52%), endometrium (endometrioid, 10%), endocervical glands (mucinous, 6%), or vaginal rest cells (clear cells, 6%); unspecified or rare histotypes account for the remaining quarter of EOCs.[1] EOCs are further classified into Type I and Type II based on clinical and pathologic features. Type I EOCs are usually low-grade tumors that are diagnosed in early stages (I/II) and show slow growth with low mortality including low-grade serous and endometriod. Type I EOCs usually develop from benign lesions, like endometriosis, that implant into the ovary and through a series of mutations result in malignant transformation.[10, 11]

Type II EOCs are usually high-grade, present at later stages (III/IV), and include high-grade serous, carcinosarcoma, and undifferentiated histotypes. They show aggressive growth, and have low survival rates. Most type II EOCs are associated with *TP53* mutations. In addition to the pelvic tumor, the tumor growth is usually abundant in the omentum and mesentery. [10–13]

Non-epithelial ovarian cancers (10%) include germ cell tumors, sex cord-stromal tumors, ovarian sarcoma, and small cell carcinomas.

Fallopian Tube Theory of Ovarian Carcinogenesis

Serous tubal intraepithelial carcinomas are now known as the precursor lesions for high grade serous carcinomas and spread from the open fimbriated ends of the fallopian tubes

throughout the peritoneal cavity. [14, 9] Widespread use of the Sectioning and Extensively Examining the Fimbriated End (SEE-FIM) protocol by pathologists has improved detection of serous tubal intraepithelial carcinomas.[15] The idea that many ovarian cancers originate from these tubal lesions is supported by their co-existence in up to 60% of all high-grade serous ovarian carcinomas.[16] As this is 60%, at least 40% of high grade serous cancers don't arise directly from the fimbriae.

Risk and Protective Factors for Ovarian Cancer

Genetic risk factors.

Compared to women without known hereditary risk factors for ovarian cancer, women with deleterious genetic mutations are at markedly higher risk for ovarian cancer and are diagnosed at younger ages. Most hereditary ovarian cancers are linked to mutations in *BRCA1* or *BRCA2*, which confer a mean cumulative risk for ovarian cancer by age 70 of 40% and 18%, respectively.[17, 18] A recent international registry of 5689 women from 78 participating centers in 12 countries yielded similar estimates; the cumulative risk of ovarian cancer to age 80 was 49% for *BRCA1* and 21% for *BRCA2* mutation carriers.[19] Women with Lynch syndrome, though at relatively higher risk for colorectal and endometrial cancers, also have elevated risk of ovarian cancer. The cumulative risks for ovarian cancer by age 70 for the Lynch syndrome mutations in *MLH1* and *MSH2* are 11% and 15%, respectively, whereas no ovarian cancer-specific risk has been identified for mutations in *MSH6* or *PMS2*.[20, 21]

Over the last decade, several moderate penetrance mutations have also been found to increase ovarian cancer risk. Mutations in *RAD51C* and *RAD51D* are estimated to increase risk for ovarian cancer by approximately 5-fold[22–24] and 12-fold,[25, 23, 24] respectively. In *BRIP1* mutation carriers, the cumulative lifetime risk of developing ovarian cancer by age 80 is estimated at 5.8%.[26, 23, 27]

Protective factors.

Non-genetic factors that confer protection against ovarian cancer include pregnancy (risk decreases with each pregnancy),[28] breastfeeding (risk decreases with duration),[28] oral contraceptive pill use (risk decreases with duration),[29] and bilateral tubal ligation (risk decreases by ~50%).[30]

Standard Approaches to Ovarian Cancer Screening

Screening Trials for Women at Normal Risk of Ovarian Cancer

Eighty-five percent of ovarian cancers occur in women without increased hereditary risk. Since the outcome of screening for ovarian cancer requires a surgical procedure, a positive predictive value (PPV) of 10% is recommended to balance the benefits of screening against the harms of unnecessary procedures.[31] Given the low prevalence of ovarian cancer ($\frac{1}{2}500$ postmenopausal women), a screening test for ovarian cancer requires a sensitivity for asymptomatic disease of > 75% and a specificity of >99.6% to meet this criterion.[32–34]

Efforts to detect ovarian cancer in early stage have utilized ultrasound imaging and blood tests, notably CA125. Transabdominal and, in more recent years, transvaginal sonography (TVS) have been evaluated in several large trials (see Table 1.) with women at normal risk. [35, 36] CA125 (MUC 16) is a high molecular weight (5 MDa) heavily glycosylated transmembrane cell surface protein that is expressed by human epithelial ovarian cancers [37]and by normal endometrium, lung and cornea. CA125 is cleaved and shed into body fluids where it can be measured by immunoassays.[38, 39] CA125 can be detected in blood from 90% of patients with stage III-IV and 50–60% of patients with stage I-II ovarian cancer.[38]

When used alone on a single occasion neither CA125 nor TVS have had adequate sensitivity nor specificity for early detection. The Prostate, Lung, Colorectal, and Ovarian (PLCO) Cancer Screening Trial was one of the first large trials of ovarian cancer screening for the general population. This study randomized 78,216 postmenopausal women ages 55 to 74 to receive either annual transvaginal ultrasound (TVS) with annual serum CA125 or conventional care.[40] Based on TVS or CA125 considered independently, multiple operations were performed for benign disease and there was no demonstrated mortality benefit of screening after a median follow up of 14.7 years.[41]

Greater specificity and sensitivity has been attained with two stage strategies where rising CA125 has triggered TVS in 1–2% of participants. The United Kingdom Collaborative Trial of Ovarian Cancer Screening (UKCTOCS) is the largest randomized clinical trial to evaluate ovarian cancer screening's impact on mortality in the general population.[42] The trial randomized 202,638 postmenopausal women age >50 to multimodal screening (MMS) (50,640), ultrasound screening (USS), (50,639), and 101,359 women to no screening between 2001 and 2005. The MMS arm analyzed the trend of annual CA125 values analyzed with a Bayesian risk of ovarian cancer algorithm (ROCA), comparing each participants current CA125 value with results of previous assays to detect a change-point that could occur within or outside the range of normal values <35U/mL.[43] Women whose CA125 did not change returned in a year. A marked increase in CA125 prompted TVS[44] and abnormal TVS surgical exploration. Intermediate change in CA125 leads to a repeat CA125 in 3 months. This approach attained a specificity of >99.6% with only 4.4 operations per case of ovarian cancer detected.[45] The overall survival analysis was not statistically significant, but a mortality reduction was observed over the first 14 years for the MMS arm of 15% and USS of 11%. In a pre-specified subset analysis where prevalent cases were excluded, the overall average mortality reduction in incident cases with MMS versus no screening (P=0.021) was 20% (-2 to 40) with a reduction of 8% (-27 to 43) in years 0-7 and 28% (-3 to 49) in years 7-14, in favor of MMS. With wide confidence limits around these estimates, additional time will be required for the trial to mature and additional analysis will be performed. Sensitivity for early stage disease was improved where 52.6% (70 of 133 women) were diagnosed using the ROCA with CA125 levels in the normal range (35 U/ml), and 41.4% (55 of 133 women) were diagnosed in early stages I/II,[45] reflecting a stage shift.

Similar specificity has been observed in the Normal Risk of Ovarian Cancer screening study (NROSS) of 4,051 postmenopausal women in the United States who had yearly CA125

values interpreted using the ROCA using a protocol identical to the MMS arm of the UKCTOCS. Over the last 18 years, in parallel with the UKCTOCS, 21 women have undergone surgery based on the ROCA algorithm. Thirteen have had epithelial ovarian malignancies with 11 invasive cancers and 2 borderline tumors including 9 cases in stage I/II. Like the UKCTOCS, specificity and PPV were high with 3 procedures for each case of ovarian cancer detected.[46]

With an update of the UKCTOCS still pending, the US preventative services task force (USPSTF) concluded that screening for ovarian cancer for normal risk women does not reduce mortality and is not recommended.[47]

Screening Trials for Women with Elevated Risk of Ovarian Cancer

The National Comprehensive Cancer Network (NCCN) clinical practice guidelines recommend risk-reducing salpingo-oophorectomy (RRSO) upon completion of childbearing or between 35 and 40 years for *BRCA1* and between 40 and 45 years for *BRCA2* mutations carriers. Despite the proven benefit of RRSO for ovarian cancer risk reduction and mortality, [48] uptake of RRSO is variable among high-risk women. Many complex factors influence the decision to undergo RRSO, in these premenopausal women including fear of menopause, fear of cancer, family history of cancer, and perceived risks and benefits of surgery.[49] Despite uncertain benefits, current NCCN guidelines allow for screening with TVS combined with CA125, starting at 30–35 years, as an alternative for high-risk women who have not yet undergone RRSO.[26]

The United Kingdom Familial Ovarian Cancer Screening Study (UKFOCSS) enrolled 4348 women with an elevated risk of EOC or fallopian tube cancer with a median follow up time of 4.8 years. Women underwent ROCA screening every 4 months with annual TVS if normal. If ROCA values were elevated, TVS was recommended within 2 months. After a median follow-up time of 4.8 years, 19 cases of EOC or fallopian tube cancer were diagnosed, with ten (52.6%) at stage I/II, indicating a stage shift.[50] Whether these encouraging early findings will translate into lower mortality is not yet known, as data collection is ongoing.

Skates and colleagues reported in 2017 on the early detection of ovarian cancer using ROCA screening with every 3 month CA125 testing in 3692 high risk women by combining data from the Gynecologic Oncology Group (GOG)[51] and Cancer Genetics Network (CGN) trials. Similar to the UKFOCSS trial, about half of incident cancers were diagnosed at an early stage, and ROCA detected 3/6 (50%) before CA125 exceeded 35 U/mL. Screening yielded a relatively high specificity of 92%.[52]

Novel Approaches to Early Detection

In patients at average and elevated risk of ovarian cancer, two stage approaches to early detection appear promising where rising levels of blood biomarkers over time prompt imaging in a small fraction of patients, enhancing specificity and reducing stress, cost and possible morbidity. Clearly, however, there is an unmet need for greater sensitivity in both stages of screening.

Protein Biomarkers

Over the last 3 decades, more than 100 blood biomarkers have been evaluated for their ability to detect early stage ovarian cancer, individually and in combination with CA25. Human epididymis protein 4 (HE4) has two whey acidic protein (WAP) domains and a 4-disulfide core (WFDC2) and is secreted by epithelial ovarian cancer cells. HE4 is increased in at least 70% of ovarian cancers, but unlike CA125 is less frequently elevated by benign disease and can detect a fraction of cases with normal CA125.[53] CA72.4 is an antigenic determinant on a 200–400 KD glycoprotein that can detect cases of ovarian cancer missed by CA125 and HE4.

Moore et al. showed that when combined, HE4 and CA125 produced the highest sensitivity at 76.4% (specificity 95%) of other combinations of markers and increased the sensitivity of CA125 alone for differentiating benign verses cancer in pelvic masses.[54] Cramer et al. tested a panel of biomarkers on serum samples from the PLCO study. The top markers were CA125 (86%), HE4 (73%), transthyretin, CA72.4, and CA15.3, with sensitivities declining > 6 months from diagnosis.[55] Four biomarkers including CA125, HE4, CA72.4, and CA15.3 were tested in 810 invasive EOCs cases and controls from the European Prospective Investigation into Cancer and Nutrition (EPIC) study. Receiver operator performance was highest for CA125 (0.92), followed by HE4 (0.84), CA72.4 (0.77), and CA15.3 (0.73), with lower performance in earlier staged disease and with increasing time between sampling and diagnosis.[56] Using serum samples from the UKCTOCS trial, HE4 and CA72.4 detected 16% of cases missed by CA125, but did not provide additional lead-time.[57]

Autoantibodies

The limited ability of protein biomarkers to detect early ovarian cancer may be due to the small size of early cancers and/or to the low expression or shedding of biomarkers.. Autoantibodies could, however, be stimulated by small volumes of cancer.[58] The tumor suppressor gene *TP53* is mutated in almost all high grade serous ovarian cancers.[59] TP53 autoantibody levels are increased in more than 20% of pre-treatment sera from patients with both early and late stage ovarian cancer. Notably, TP53 autoantibody levels were elevated in 16% of patients whose cancer was not detected with ROCA-interpreted CA125 during the UKCTOCS trial. TP53 autoantibody levels rose 8 months prior to CA125 when it was elevated and 22 months prior to diagnosis when CA125 did not rise.[60]

A recent systematic review of the world literature by Fortner[61] identified additional evidence for the TP53 autoantibody as well as several other candidate autoantibodies for early detection of ovarian cancer that were elevated in a significant fraction of cancers, including anti-homeobox gene A7 (HOXA7)[61] and anti-Interleukin 8 (IL8).[62] Panels of autoantibodies are currently being evaluated by the National Cancer Institute Early Detection Research Network to complement CA125 and to enhance the sensitivity of two stage strategies where rising blood biomarkers prompt TVS.

MicroRNAs (miRNAs)

MicroRNAs circulate in many body fluids as stable entities bound to proteins (e.g., Argonaute 2) or packaged in exosomes, microvesicles, or apoptotic bodies.[63] miRNAs are

single stranded, short (~22 nucleotide), non-coding RNA that regulate post-transcriptional gene expression, typically leading to translational repression.[64, 65] Since circulating miRNAs change based on pathologic states, they are considered potential cancer biomarkers, [64, 66] and several miRNA profiles show significant promise as EOC biomarkers. Four serum miRNAs (miR-182, miR-200a, MiR-200b, miR-200c) are elevated in blood from serous EOC patients, and miR-200b combined with miR-200c performed well in distinguishing serous EOC from controls (AUC=0.784).[67] The combination of miR-205 and let-7f, showed excellent accuracy for EOC (AUC = 0.831 (95% CI, 0.772 - 0.880) with high sensitivity (62.4%) and specificity (92.9%), especially in patients with stage I disease. [68] An eight miRNA panel, also detected early stage EOC from benign tumors with 86% sensitivity and 83% specificity.[69] Combining small RNA sequencing from 179 human serum samples with a neural network, Elias and colleagues produced an miRNA algorithm for the diagnosis of EOC with the most accurate performance to date (AUC 0.90; 95% CI: 0.81-0.99.[70] Although miRNA has significant potential as a biomarker, comparison and combination studies with CA125, HE4, and TVS in a prospective trial are lacking.

Circulating Tumor DNA (ctDNA)

Circulating tumor DNA (ctDNA) is the fraction of cell free DNA that contains tumorspecific somatic mutations that are found in the circulation.[71] Analysis of serum ctDNA ("liquid biopsy"[72]) provides a potential noninvasive method of tumor detection and monitoring[73, 72] and is especially advantageous for cancers that are difficult to biopsy. In a small sample of 7 patients with ovarian cancer, Bettegowda et al. found 100% had detectable ctDNA.[73] ctDNA has also been identified following uterine lavage in 24 (80%) of 30 patients with ovarian cancer and 5 of 5 with endometrial cancers; however, mutations were also found in 8 (27%) of 27 patients with benign lesions.[74] Detection of ctDNA has improved with targeted deep sequencing, which in one study identified mutations at allelic frequencies of 2% with >97% sensitivity and specificity.

Cohen and colleagues created CancerSEEK, which concurrently evaluates 8 protein biomarkers, including CA125, and the presence of mutations in 1933 genomic positions including single base substitutions, insertions, or deletions.[75] In a case-control study that included eight cancer types, CancerSEEK was most accurate in ovarian cancer, with a sensitivity of 98% and specificity of >99% with only 7 positives out of 812 patients without known cancers. However, a majority of patients with ovarian cancer had stage III disease where CA125, one component of CancerSEEK, would detect 90% of cases. Some of the drawbacks to ctDNA include detection of mutations in non-cancer patients and poor early stage detection.

DNA Methylation

DNA promotor hypermethylation of tumor suppressor genes is now known to occur in most cancers, resulting in the inactivation of tumor suppressor genes.[76] Promoter methylation usually occurs early during carcinogenesis, and is therefore a potential early tumor marker. [77, 76] A panel of 3 DNA methylation serum markers was tested in 250 EOC patients undergoing chemotherapy, and a large cohort of patients from the UKCTOCS trial. Serum

DNA methylation markers detected over half of the ovarian cancer cases in serum samples acquired up to two years prior to diagnosis.[78] Prospective studies are still needed.[77, 79]

Imaging

TVS is the gold standard imaging modality for viewing the adnexa and uterus. In the PLCO trial, TVS had a PPV of 1% and CA125 (>35 U/mL) had a PPV of 2.6%. A combination of abnormal TVS and elevated CA125 increased the PPV to 23.5%, but 60% of ovarian cancers would have been missed. The main drawbacks to TVS are the inability to visualize small lesions on the ovaries and the failure to visualize the fimbriae of the fallopian tubes.[80] In the UKCTOCS, the ROCA detected an early rise in the CA125 level in a significant number of screened subjects destined to develop ovarian cancer where the TVS was normal, resulting in a delay in surgery. Early stage ovarian lesions are now estimated to be < 3mm and may persist at this size for several years, requiring imaging techniques that can detect very small tumors.[81, 82]

Both magnetic resonance imaging (MRI) and magnetic relaxometry (MRX) can detect and locate magnetic nanoparticles without ionizing radiation.[83] Superconducting quantum interference device (SQUID) technology can detect delays in magnetic relaxation when nanoparticles conjugated to a monoclonal antibody reactive with ovarian cancer are bound to cancer cells, differentiating them from unbound normal cells.[83] This technique has been used in several tissue models including breast cancer and leukemia and holds potential for the detection of early ovarian cancer.[83, 84] Williams et al. engineered an optical sensor made up of an HE4 antibody-carbon nanotube complex which measures HE4 in different body fluids. The sensors were then implanted into four mouse models of ovarian cancer and were capable of noninvasive cancer biomarker detection.[85]

Doppler ultrasound is currently used to detect central ovarian blood flow when differentiating benign from ovarian malignancies, but it is unable to detect small ovarian cancers.[80, 86] The use of microbubble contrast agents with ultrasound has allowed visualization of neovascularity in some ovarian masses.[87] In a study by Xiang et al., 3-D microbubble contrast-enhanced TVS differentiated benign from malignant small ovarian masses with high sensitivity (100%) and specificity (98%).[80, 88] However, this method does not overcome the challenge of imaging the fallopian tubes or detecting small ovarian lesions.

Light-induced endogenous fluorescence (autofluorescence) has been used to detect precancerous lesions in other organ systems, including the cervix. Autofluorescent imaging of surgically removed fallopian tubes had a sensitivity of 73%, a specificity of 83%, and a PPV of 57% for ovarian or serous tubal intraepithelial carcinomas. A next step in the development of this technology will be *in vivo* screening via falloposcopy.[89]

Fallopian Tube Cytology and Tumor DNA detection in Pap smears

Recognition that many ovarian cancers arise in the fallopian tube has stimulated development of tissue sampling methods specifically for the fallopian tube. Although reproductive endocrinologists once used falloposcopy to visualize the interior of the fallopian tubes, this practice has been mostly abandoned.[90] More recently, in a small

feasibility study Lum et al. attempted cytologic sampling of the fallopian tubes using hysteroscopic brush cytology without visualization. Falloposcopy was attempted and while unsuccessful for the hysteroscopic portion, was successful for the laparoscopic assessment. [91] A recent study proposal by Gizzo et al. will utilize a 4.9 mm integrated in office hysteroscope to cannulate the fallopian tube using a 4 French (1.3 mm) sterile ureteric drainage catheter, with the goal of reaching the fimbriated ends. Following instillation of normal saline, the fluid containing the end-luminal fallopian cells will be aspirated and analyzed with cytology.[92] Several groups have started to characterize the cytology of the fallopian tube as well as endometrial cytology for early detection of ovarian cancer, although further analysis and standardization is needed.[93–96]

Liquid based cervical Pap smear testing, which includes both cytological analysis and DNA detection, has dramatically reduced cervical cancer mortality and has recently been studied as a screening tool for other cancers. Kinde et al. identified mutations from DNA in liquid Pap smear specimens in 100% of endometrial cancers and 41% of ovarian cancers.[97] Another approach is PapSEEK, which detected endometrial and ovarian cancers based on DNA analysis from routine Pap tests by the detection of 18 genes and aneuploidy. The sensitivity for ovarian cancer was 63%, with a specificity of ~100%, but required the use of an endometrial brush rather than routine Pap brush.[98] Analyzing ctDNA from the cervix and blood has detected 55% of early stage ovarian cancers.[99]

Challenges and Future Directions

Despite extensive investigation, a single marker for early stage ovarian cancer remains elusive, and there is no currently accepted method for ovarian cancer screening. Examination of CA125 over time as opposed to a single time point still demonstrates the best sensitivity and specificity as a marker, although its value for future screening purposes may require complementary markers. Ongoing data collection will help determine whether earlier detection of ovarian cancer through multimodal screening yields reductions in mortality over the long term, as suggested by late trends in the UKCTOCs study. The standard of care worldwide is RRSO by age 40 for BRCA1 and 45 for BRCA2. Until high-risk women undergo surgery, an affective screening strategy would be helpful. The low incidence of ovarian cancer remains a challenge for identifying effective early detection strategies. Specificity needs to be > 99.6% in order to avoid harm in healthy women. Imaging needs to be able to visualize early stage I/II lesions yet remain cost effective. The number of women required for a prospective trial with the gold standard of mortality reduction as an end-point is also a challenge.

In the meantime, the treatment of women at high genetic risk of ovarian cancer are highly variable across the world. Many countries recommend RRSO, particularly within Europe and the UK, without screening options.[100] Within the US, RRSO is also recommended, but the NCCN includes screening with CA125 (> 35U/mL) and TVS as an option prior to RRSO.

Several protein biomarkers show potential to enhance screening and early detection. The addition of HE4 and CA 72.4 to CA125 detected 16% more than CA125 alone. The TP53

autoantibody was able to detect 16% of EOC patients that CA125 missed, indicating a possible use as an adjunct to conventional CA125 screening. A panel of autoantibodies might be even more effective in complementing CA125. The possibility of a "liquid biopsy" is within closer reach as detection of circulating tumor DNA has improved over time. Other tests based on DNA methylation assays, microRNA algorithms, and Pap-like cytologic analysis show potential. Of these, PapSEEK holds the greatest potential for reaching the largest number of patients, however its value hinges on a validation study, cost, and performance with the liquid Pap, since available worldwide. The UKCTOCs has a serum bank that may allow validation of these newer markers.

Conclusion

There remains intense interest from patients and the medical community to identify an effective strategy for the detection of early ovarian cancer. There are a number of promising strategies focusing on new biomarkers and new imaging techniques. Current two-stage multimodal ovarian cancer screening algorithms incorporating ROCA interpreted CA125 with TVS have shown a stage shift to earlier (Stage I/II) cancers in high-risk women and may ultimately demonstrate long-term effects on mortality. In the absence of a superior replacement, it is reasonable to offer two-stage screening to high-risk women who have not yet undergone RRSO. The addition of imaging to detect small fallopian tube or peritoneal lesions along with the further development of molecular techniques hold promise for supplementing or surpassing the two-stage screening test.

References

Papers of particular interest, published recently, have been highlighted as:

* Of importance

- Torre LA, Trabert B, DeSantis CE, Miller KD, Samimi G, Runowicz CD et al. Ovarian cancer statistics, 2018. CA: a cancer journal for clinicians. 2018;68(4):284–96. doi:10.3322/caac.21456. [PubMed: 29809280]
- Morice P, Gouy S, Leary A. Mucinous Ovarian Carcinoma. The New England journal of medicine. 2019;380(13):1256–66. doi:10.1056/NEJMra1813254. [PubMed: 30917260]
- 3. Institute NC. Cancer stat facts: ovarian cancer. 2019 http://seer.cancer.gov/statfacts/html/ovary.html. Accessed 4/6/2019 2019.
- 4. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. CA: a cancer journal for clinicians. 2018;68(1):7–30. doi:10.3322/caac.21442. [PubMed: 29313949]
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA: a cancer journal for clinicians. 2018;68(6):394–424. doi:10.3322/caac.21492. [PubMed: 30207593]
- Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer. 2015;136(5):E359–86. doi:10.1002/ijc.29210. [PubMed: 25220842]
- Peres LC, Cushing-Haugen KL, Kobel M, Harris HR, Berchuck A, Rossing MA et al. Invasive Epithelial Ovarian Cancer Survival by Histotype and Disease Stage. Journal of the National Cancer Institute. 2019;111(1):60–8. doi:10.1093/jnci/djy071. [PubMed: 29718305]
- 8. Allemani C, Weir HK, Carreira H, Harewood R, Spika D, Wang XS et al. Global surveillance of cancer survival 1995–2009: analysis of individual data for 25,676,887 patients from 279 population-

based registries in 67 countries (CONCORD-2). Lancet (London, England). 2015;385(9972):977–1010. doi:10.1016/S0140-6736(14)62038-9.

- Kindelberger DW, Lee Y, Miron A, Hirsch MS, Feltmate C, Medeiros F et al. Intraepithelial carcinoma of the fimbria and pelvic serous carcinoma: Evidence for a causal relationship. The American journal of surgical pathology. 2007;31(2):161–9. doi:10.1097/01.pas. 0000213335.40358.47. [PubMed: 17255760]
- 10. Koshiyama M, Matsumura N, Konishi I. Recent concepts of ovarian carcinogenesis: type I and type II. BioMed research international. 2014;2014:934261. doi:10.1155/2014/934261.
- Kurman RJ. Origin and molecular pathogenesis of ovarian high-grade serous carcinoma. Annals of oncology : official journal of the European Society for Medical Oncology. 2013;24 Suppl 10:x16– 21. doi:10.1093/annonc/mdt463. [PubMed: 24265397]
- Kurman RJ, Shih Ie M. The Dualistic Model of Ovarian Carcinogenesis: Revisited, Revised, and Expanded. The American journal of pathology. 2016;186(4):733–47. doi:10.1016/j.ajpath. 2015.11.011. [PubMed: 27012190]
- Romero I, Bast RC, Jr. Minireview: human ovarian cancer: biology, current management, and paths to personalizing therapy. Endocrinology. 2012;153(4):1593–602. doi:10.1210/en.2011-2123. [PubMed: 22416079]
- Crum CP, Drapkin R, Miron A, Ince TA, Muto M, Kindelberger DW et al. The distal fallopian tube: a new model for pelvic serous carcinogenesis. Curr Opin Obstet Gynecol. 2007;19(1):3–9. doi:10.1097/GCO.0b013e328011a21f. [PubMed: 17218844]
- Medeiros F, Muto MG, Lee Y, Elvin JA, Callahan MJ, Feltmate C et al. The tubal fimbria is a preferred site for early adenocarcinoma in women with familial ovarian cancer syndrome. The American journal of surgical pathology. 2006;30(2):230–6. [PubMed: 16434898]
- Gockley AA, Elias KM. Fallopian tube tumorigenesis and clinical implications for ovarian cancer risk-reduction. Cancer Treat Rev. 2018;69:66–71. doi:10.1016/j.ctrv.2018.06.004. [PubMed: 29909222]
- Chen S, Parmigiani G. Meta-analysis of BRCA1 and BRCA2 penetrance. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2007;25(11):1329–33. doi:10.1200/JCO.2006.09.1066. [PubMed: 17416853]
- Network NCC. NCCN Clinical Practice Guidelines in Oncology. Genetic/Familial High-Risk Assessment: Breast and Ovarian (Version 3.2019). 2019 https://www.nccn.org/professionals/ physician_gls/pdf/genetics_screening.pdf. Accessed 4/7/2019.
- Kotsopoulos J, Gronwald J, Karlan B, Rosen B, Huzarski T, Moller P et al. Age-specific ovarian cancer risks among women with a BRCA1 or BRCA2 mutation. Gynecologic oncology. 2018;150(1):85–91. doi:10.1016/j.ygyno.2018.05.011. [PubMed: 29793803]
- Moller P, Seppala T, Bernstein I, Holinski-Feder E, Sala P, Evans DG et al. Cancer incidence and survival in Lynch syndrome patients receiving colonoscopic and gynaecological surveillance: first report from the prospective Lynch syndrome database. Gut. 2017;66(3):464–72. doi:10.1136/ gutjnl-2015-309675. [PubMed: 26657901]
- Network NCC. NCCN Clinical Practice Guidelines in Oncology. Genetic/Familial High-Risk Assessment: Colorectal (Version 1.2018). 2018 https://www.nccn.org/professionals/ physician_gls/pdf/genetics_colon.pdf. Accessed 4/7/2019.
- Loveday C, Turnbull C, Ruark E, Xicola RM, Ramsay E, Hughes D et al. Germline RAD51C mutations confer susceptibility to ovarian cancer. Nature genetics. 2012;44(5):475–6; author reply 6. doi:10.1038/ng.2224. [PubMed: 22538716]
- Norquist BM, Harrell MI, Brady MF, Walsh T, Lee MK, Gulsuner S et al. Inherited Mutations in Women With Ovarian Carcinoma. JAMA oncology. 2016;2(4):482–90. doi:10.1001/jamaoncol. 2015.5495. [PubMed: 26720728]
- Song H, Dicks E, Ramus SJ, Tyrer JP, Intermaggio MP, Hayward J et al. Contribution of Germline Mutations in the RAD51B, RAD51C, and RAD51D Genes to Ovarian Cancer in the Population. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2015;33(26):2901–7. doi:10.1200/jco.2015.61.2408. [PubMed: 26261251]

- Loveday C, Turnbull C, Ramsay E, Hughes D, Ruark E, Frankum JR et al. Germline mutations in RAD51D confer susceptibility to ovarian cancer. Nature genetics. 2011;43(9):879–82. doi: 10.1038/ng.893. [PubMed: 21822267]
- Network NCC. Genetic/Familial High-Risk Assessment: Breast and Ovarian (Version 3.2019). 2019.
- 27. Ramus SJ, Song H, Dicks E, Tyrer JP, Rosenthal AN, Intermaggio MP et al. Germline Mutations in the BRIP1, BARD1, PALB2, and NBN Genes in Women With Ovarian Cancer. Journal of the National Cancer Institute. 2015;107(11). doi:10.1093/jnci/djv214.
- 28. Sung HK, Ma SH, Choi J-Y, Hwang Y, Ahn C, Kim B-G et al. The Effect of Breastfeeding Duration and Parity on the Risk of Epithelial Ovarian Cancer: A Systematic Review and Metaanalysis. Journal of preventive medicine and public health = Yebang Uihakhoe chi. 2016;49(6): 349–66. doi:10.3961/jpmph.16.066. [PubMed: 27951628]
- 29. Havrilesky IJ MP, Lowery WJ, Gierisch JM, Coeytaux RR, Urrutia RP, et al. . Oral contraceptive pills as primary prevention for ovarian cancer: a systemic review and meta-analysis. . Obstet Gynecol. 2013(122).
- Cibula D, Widschwendter M, Majek O, Dusek L. Tubal ligation and the risk of ovarian cancer: review and meta-analysis. Hum Reprod Update. 2011;17(1):55–67. doi:10.1093/humupd/dmq030. [PubMed: 20634209]
- 31. Jacobs I, Bast RC Jr., The CA 125 tumour-associated antigen: a review of the literature. Human reproduction (Oxford, England). 1989;4(1):1–12. * established the specificity of 99.6% and PPV of 10% for OC screening test.
- Das PM, Bast RC Jr., Early detection of ovarian cancer. Biomarkers in medicine. 2008;2(3):291– 303. doi:10.2217/17520363.2.3.291. [PubMed: 20477415]
- Elias KM, Guo J, Bast RC Jr., Early Detection of Ovarian Cancer. Hematol Oncol Clin North Am. 2018;32(6):903–14. doi:10.1016/j.hoc.2018.07.003. [PubMed: 30390764]
- Skates SJ, Xu FJ, Yu YH, Sjovall K, Einhorn N, Chang Y et al. Toward an optimal algorithm for ovarian cancer screening with longitudinal tumor markers. Cancer. 1995;76(10 Suppl):2004–10. [PubMed: 8634992]
- Kobayashi H, Yamada Y, Sado T, Sakata M, Yoshida S, Kawaguchi R et al. A randomized study of screening for ovarian cancer: a multicenter study in Japan. Int J Gynecol Cancer. 2008;18(3):414– 20. doi:10.1111/j.1525-1438.2007.01035.x. [PubMed: 17645503]
- van Nagell JR Jr., Miller RW, DeSimone CP, Ueland FR, Podzielinski I, Goodrich ST et al. Longterm survival of women with epithelial ovarian cancer detected by ultrasonographic screening. Obstet Gynecol. 2011;118(6):1212–21. doi:10.1097/AOG.0b013e318238d030. [PubMed: 22105249]
- Bast RC, Feeney M, Lazarus H, Nadler LM, Colvin RB, Knapp RC. Reactivity of a monoclonal antibody with human ovarian carcinoma. J Clin Invest. 1981;68(5):1331–7. [PubMed: 7028788]
- Bast RC Jr., , Klug TL, St John E, Jenison E, Niloff JM, Lazarus Het al. A radioimmunoassay using a monoclonal antibody to monitor the course of epithelial ovarian cancer. The New England journal of medicine. 1983;309(15):883–7. doi:10.1056/nejm198310133091503. [PubMed: 6310399]
- Yin BW, Lloyd KO. Molecular cloning of the CA125 ovarian cancer antigen: identification as a new mucin, MUC16. The Journal of biological chemistry. 2001;276(29):27371–5. doi:10.1074/ jbc.M103554200. [PubMed: 11369781]
- Buys SS, Partridge E, Black A, Johnson CC, Lamerato L, Isaacs C et al. Effect of screening on ovarian cancer mortality: the Prostate, Lung, Colorectal and Ovarian (PLCO) Cancer Screening Randomized Controlled Trial. JAMA. 2011;305(22):2295–303. doi:10.1001/jama.2011.766. [PubMed: 21642681]
- Pinsky PF, Yu K, Kramer BS, Black A, Buys SS, Partridge E et al. Extended mortality results for ovarian cancer screening in the PLCO trial with median 15years follow-up. Gynecologic oncology. 2016;143(2):270–5. doi:10.1016/j.ygyno.2016.08.334. [PubMed: 27615399]
- 42. Jacobs IJ, Menon U, Ryan A, Gentry-Maharaj A, Burnell M, Kalsi JK et al. Ovarian cancer screening and mortality in the UK Collaborative Trial of Ovarian Cancer Screening (UKCTOCS):

a randomised controlled trial. Lancet (London, England). 2016;387(10022):945–56. doi:10.1016/s0140-6736(15)01224-6.

- *UKCTOCS trial demonstrated a possible mortality reduction
- 43. Skates SJ. Ovarian cancer screening: development of the risk of ovarian cancer algorithm (ROCA) and ROCA screening trials. International journal of gynecological cancer : official journal of the International Gynecological Cancer Society. 2012;22 Suppl 1(Suppl 1):S24–S6. doi:10.1097/IGC. 0b013e318256488a. [PubMed: 22543916]
- 44. Barrett J, Jenkins V, Farewell V, Menon U, Jacobs I, Kilkerr J et al. Psychological morbidity associated with ovarian cancer screening: results from more than 23,000 women in the randomised trial of ovarian cancer screening (UKCTOCS). BJOG. 2014;121(9):1071–9. doi: 10.1111/1471-0528.12870. [PubMed: 24865441]
- 45. Menon U, Ryan A, Kalsi J, Gentry-Maharaj A, Dawnay A, Habib M et al. Risk Algorithm Using Serial Biomarker Measurements Doubles the Number of Screen-Detected Cancers Compared With a Single-Threshold Rule in the United Kingdom Collaborative Trial of Ovarian Cancer Screening. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2015;33(18):2062–71. doi:10.1200/jco.2014.59.4945. [PubMed: 25964255]
- 46. Lu KH, Skates S, Hernandez MA, Bedi D, Bevers T, Leeds L et al. A 2-stage ovarian cancer screening strategy using the Risk of Ovarian Cancer Algorithm (ROCA) identifies early-stage incident cancers and demonstrates high positive predictive value. Cancer. 2013;119(19):3454–61. doi:10.1002/cncr.28183. [PubMed: 23983047]
- Henderson JT, Webber EM, Sawaya GF. Screening for Ovarian Cancer: Updated Evidence Report and Systematic Review for the US Preventive Services Task Force. Jama. 2018;319(6):595–606. doi:10.1001/jama.2017.21421. [PubMed: 29450530]
- Rebbeck TR, Kauff ND, Domchek SM. Meta-analysis of risk reduction estimates associated with risk-reducing salpingo-oophorectomy in BRCA1 or BRCA2 mutation carriers. Journal of the National Cancer Institute. 2009;101(2):80–7. doi:10.1093/jnci/djn442. [PubMed: 19141781]
- Mai PL, Piedmonte M, Han PK, Moser RP, Walker JL, Rodriguez G et al. Factors associated with deciding between risk-reducing salpingo-oophorectomy and ovarian cancer screening among highrisk women enrolled in GOG-0199: An NRG Oncology/Gynecologic Oncology Group study. Gynecol Oncol. 2017;145(1):122–9. doi:10.1016/j.ygyno.2017.02.008. [PubMed: 28190649]
- 50. Rosenthal AN, Fraser LSM, Philpott S, Manchanda R, Burnell M, Badman P et al. Evidence of Stage Shift in Women Diagnosed With Ovarian Cancer During Phase II of the United Kingdom Familial Ovarian Cancer Screening Study. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2017;35(13):1411–20. doi:10.1200/jco.2016.69.9330.
- *demonstrates a stage shift from screening for patients at high-risk

[PubMed: 28240969]

- 51. Greene MH, Piedmonte M, Alberts D, Gail M, Hensley M, Miner Z et al. A prospective study of risk-reducing salpingo-oophorectomy and longitudinal CA-125 screening among women at increased genetic risk of ovarian cancer: design and baseline characteristics: a Gynecologic Oncology Group study. Cancer epidemiology, biomarkers & prevention : a publication of the American Association for Cancer Research, cosponsored by the American Society of Preventive Oncology. 2008;17(3):594–604. doi:10.1158/1055-9965.epi-07-2703.
- 52. Skates SJ, Greene MH, Buys SS, Mai PL, Brown P, Piedmonte M et al. Early Detection of Ovarian Cancer using the Risk of Ovarian Cancer Algorithm with Frequent CA125 Testing in Women at Increased Familial Risk - Combined Results from Two Screening Trials. Clinical cancer research : an official journal of the American Association for Cancer Research. 2017;23(14):3628–37. doi: 10.1158/1078-0432.ccr-15-2750.
- *additional experience using two-stage screening for women at high risk of ovarian cancer

[PubMed: 28143870]

 Hellstrom I, Raycraft J, Hayden-Ledbetter M, Ledbetter JA, Schummer M, McIntosh M et al. The HE4 (WFDC2) protein is a biomarker for ovarian carcinoma. Cancer research. 2003;63(13):3695– 700. [PubMed: 12839961]

- 54. Moore RG, Brown AK, Miller MC, Skates S, Allard WJ, Verch T et al. The use of multiple novel tumor biomarkers for the detection of ovarian carcinoma in patients with a pelvic mass. Gynecologic oncology. 2008;108(2):402–8. doi:10.1016/j.ygyno.2007.10.017. [PubMed: 18061248]
- 55. Cramer DW, Bast RC Jr., Berg CD, Diamandis EP, Godwin AK, Hartge P et al. Ovarian cancer biomarker performance in prostate, lung, colorectal, and ovarian cancer screening trial specimens. Cancer prevention research (Philadelphia, Pa). 2011;4(3):365–74. doi: 10.1158/1940-6207.capr-10-0195.
- 56. Terry KL, Schock H, Fortner RT, Husing A, Fichorova RN, Yamamoto HS et al. A Prospective Evaluation of Early Detection Biomarkers for Ovarian Cancer in the European EPIC Cohort. Clinical cancer research : an official journal of the American Association for Cancer Research. 2016;22(18):4664–75. doi:10.1158/1078-0432.ccr-16-0316. [PubMed: 27060155]
- 57. Simmons AR, Fourkala EO, Gentry-Maharaj A, Ryan A, Sutton MN, Baggerly K et al. Complementary longitudinal serum biomarkers to CA125 for early detection of ovarian cancer. Cancer Prev Res (Phila). 2019. doi:10.1158/1940-6207.capr-18-0377.
- Macdonald IK, Parsy-Kowalska CB, Chapman CJ. Autoantibodies: Opportunities for Early Cancer Detection. Trends in cancer. 2017;3(3):198–213. doi:10.1016/j.trecan.2017.02.003. [PubMed: 28718432]
- 59. Ahmed AA, Etemadmoghadam D, Temple J, Lynch AG, Riad M, Sharma R et al. Driver mutations in TP53 are ubiquitous in high grade serous carcinoma of the ovary. The Journal of pathology. 2010;221(1):49–56. doi:10.1002/path.2696. [PubMed: 20229506]
- 60. Yang WL, Gentry-Maharaj A, Simmons A, Ryan A, Fourkala EO, Lu Z et al. Elevation of TP53 Autoantibody Before CA125 in Preclinical Invasive Epithelial Ovarian Cancer. Clinical cancer research : an official journal of the American Association for Cancer Research. 2017;23(19):5912– 22. doi:10.1158/1078-0432.ccr-17-0284.
- * Anti-TP53 autoantibodies produce lead time over CA125

[PubMed: 28637689]

- Fortner RT, Damms-Machado A, Kaaks R. Systematic review: Tumor-associated antigen autoantibodies and ovarian cancer early detection. Gynecologic oncology. 2017;147(2):465–80. doi:10.1016/j.ygyno.2017.07.138. [PubMed: 28800944]
- Lokshin AE, Winans M, Landsittel D, Marrangoni AM, Velikokhatnaya L, Modugno F et al. Circulating IL-8 and anti-IL-8 autoantibody in patients with ovarian cancer. Gynecologic oncology. 2006;102(2):244–51. doi:10.1016/j.ygyno.2005.12.011. [PubMed: 16434085]
- Nakamura K, Sawada K, Yoshimura A, Kinose Y, Nakatsuka E, Kimura T. Clinical relevance of circulating cell-free microRNAs in ovarian cancer. Mol Cancer. 2016;15(1):48. doi:10.1186/ s12943-016-0536-0. [PubMed: 27343009]
- Nakamura K, Sawada K, Yoshimura A, Kinose Y, Nakatsuka E, Kimura T. Clinical relevance of circulating cell-free microRNAs in ovarian cancer. Molecular cancer. 2016;15(1):48-. doi:10.1186/ s12943-016-0536-0. [PubMed: 27343009]
- 65. Palma Flores C, Garcia-Vazquez R, Gallardo Rincon D, Ruiz-Garcia E, Astudillo de la Vega H, Marchat LA et al. MicroRNAs driving invasion and metastasis in ovarian cancer: Opportunities for translational medicine (Review). International journal of oncology. 2017;50(5):1461–76. doi: 10.3892/ijo.2017.3948. [PubMed: 28393213]
- Wittmann J, Jack HM. Serum microRNAs as powerful cancer biomarkers. Biochimica et biophysica acta. 2010;1806(2):200–7. doi:10.1016/j.bbcan.2010.07.002. [PubMed: 20637263]
- Kan CW, Hahn MA, Gard GB, Maidens J, Huh JY, Marsh DJ et al. Elevated levels of circulating microRNA-200 family members correlate with serous epithelial ovarian cancer. BMC cancer. 2012;12:627. doi:10.1186/1471-2407-12-627. [PubMed: 23272653]
- Zheng H, Zhang L, Zhao Y, Yang D, Song F, Wen Y et al. Plasma miRNAs as diagnostic and prognostic biomarkers for ovarian cancer. PloS one. 2013;8(11):e77853. doi:10.1371/journal.pone. 0077853.

- Yokoi A, Yoshioka Y, Hirakawa A, Yamamoto Y, Ishikawa M, Ikeda SI et al. A combination of circulating miRNAs for the early detection of ovarian cancer. Oncotarget. 2017;8(52):89811–23. doi:10.18632/oncotarget.20688. [PubMed: 29163790]
- 70. Elias KM, Fendler W, Stawiski K, Fiascone SJ, Vitonis AF, Berkowitz RS et al. Diagnostic potential for a serum miRNA neural network for detection of ovarian cancer. eLife. 2017;6. doi: 10.7554/eLife.28932.
- * miRNA has potential for early detection of ovarian cancer
- Patel KM, Tsui DWY. The translational potential of circulating tumour DNA in oncology. Clinical Biochemistry. 2015;48(15):957–61. doi:10.1016/j.clinbiochem.2015.04.005. [PubMed: 25889059]
- 72. Cheng X, Zhang L, Chen Y, Qing C. Circulating cell-free DNA and circulating tumor cells, the "liquid biopsies" in ovarian cancer. J Ovarian Res. 2017;10(1):75. doi:10.1186/ s13048-017-0369-5. [PubMed: 29132396]
- Bettegowda C, Sausen M, Leary RJ, Kinde I, Wang Y, Agrawal N et al. Detection of circulating tumor DNA in early- and late-stage human malignancies. Science translational medicine. 2014;6(224):224ra24. doi:10.1126/scitranslmed.3007094.
- * Use of ctDNA for the detection of human cancers
- 74. Maritschnegg E, Wang Y, Pecha N, Horvat R, Van Nieuwenhuysen E, Vergote I et al. Lavage of the Uterine Cavity for Molecular Detection of Müllerian Duct Carcinomas: A Proof-of-Concept Study. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2015;33(36):4293–300. doi:10.1200/JCO.2015.61.3083. [PubMed: 26552420]
- Cohen JD, Li L, Wang Y, Thoburn C, Afsari B, Danilova L et al. Detection and localization of surgically resectable cancers with a multi-analyte blood test. Science (New York, NY). 2018;359(6378):926–30. doi:10.1126/science.aar3247.
- * Protein biomarkers and ctDNA were used to detect multiple cancers
- 76. Esteller M, Silva JM, Dominguez G, Bonilla F, Matias-Guiu X, Lerma E et al. Promoter hypermethylation and BRCA1 inactivation in sporadic breast and ovarian tumors. Journal of the National Cancer Institute. 2000;92(7):564–9. [PubMed: 10749912]
- 77. Bartlett TE, Chindera K, McDermott J, Breeze CE, Cooke WR, Jones A et al. Epigenetic reprogramming of fallopian tube fimbriae in BRCA mutation carriers defines early ovarian cancer evolution. Nature communications. 2016;7:11620. doi:10.1038/ncomms11620.
- Widschwendter M, Zikan M, Wahl B, Lempiainen H, Paprotka T, Evans I et al. The potential of circulating tumor DNA methylation analysis for the early detection and management of ovarian cancer. Genome medicine. 2017;9(1):116. doi:10.1186/s13073-017-0500-7. [PubMed: 29268796]
- 79. Zhang Q, Hu G, Yang Q, Dong R, Xie X, Ma D et al. A multiplex methylation-specific PCR assay for the detection of early-stage ovarian cancer using cell-free serum DNA. Gynecologic oncology. 2013;130(1):132–9. doi:10.1016/j.ygyno.2013.04.048. [PubMed: 23623832]
- Mathieu KB, Bedi DG, Thrower SL, Qayyum A, Bast RC, Jr. Screening for ovarian cancer: imaging challenges and opportunities for improvement. Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology. 2018;51(3):293–303. doi:10.1002/uog.17557.
- * Up to date review of imaging for early detection of ovarian cancer
- Brown PO PC. The preclinical natural history of serous ovarian cancer: defining the target for early detection. PLoS Med. 2009;6(7):e1000114.
- Lutz AM, Willmann JK, Cochran FV, Ray P, Gambhir SS. Cancer screening: a mathematical model relating secreted blood biomarker levels to tumor sizes. PLoS Med. 2008;5(8):e170. doi:10.1371/ journal.pmed.0050170.
- Adolphi NL, Butler KS, Lovato DM, Tessier TE, Trujillo JE, Hathaway HJ et al. Imaging of Her2targeted magnetic nanoparticles for breast cancer detection: comparison of SQUID-detected magnetic relaxometry and MRI. Contrast media & molecular imaging. 2012;7(3):308–19. doi: 10.1002/cmmi.499. [PubMed: 22539401]
- Jaetao JE, Butler KS, Adolphi NL, Lovato DM, Bryant HC, Rabinowitz I et al. Enhanced leukemia cell detection using a novel magnetic needle and nanoparticles. Cancer research. 2009;69(21): 8310–6. doi:10.1158/0008-5472.CAN-09-1083. [PubMed: 19808954]

- Williams RM, Lee C, Galassi TV, Harvey JD, Leicher R, Sirenko M et al. Noninvasive ovarian cancer biomarker detection via an optical nanosensor implant. Science Advances. 2018;4(4):eaaq1090. doi:10.1126/sciadv.aaq1090.
- Medeiros LR, Rosa DD, da Rosa MI, Bozzetti MC. Accuracy of ultrasonography with color Doppler in ovarian tumor: a systematic quantitative review. Int J Gynecol Cancer. 2009;19(7): 1214–20. doi:10.1111/IGC.0b013e3181a386e5. [PubMed: 19823057]
- Fleischer AC, Lyshchik A, Andreotti RF, Hwang M, Jones HW 3rd, , Fishman DA. Advances in sonographic detection of ovarian cancer: depiction of tumor neovascularity with microbubbles. AJR American journal of roentgenology. 2010;194(2):343–8. doi:10.2214/ajr.09.3446. [PubMed: 20093594]
- Xiang H, Huang R, Cheng J, Gulinaer S, Hu R, Feng Y et al. Value of three-dimensional contrastenhanced ultrasound in the diagnosis of small adnexal masses. Ultrasound in medicine & biology. 2013;39(5):761–8. doi:10.1016/j.ultrasmedbio.2012.11.008. [PubMed: 23453372]
- McAlpine JN, El Hallani S, Lam SF, Kalloger SE, Luk M, Huntsman DG et al. Autofluorescence imaging can identify preinvasive or clinically occult lesions in fallopian tube epithelium: a promising step towards screening and early detection. Gynecologic oncology. 2011;120(3):385– 92. doi:10.1016/j.ygyno.2010.12.333. [PubMed: 21237503]
- 90. Dechaud H, Daures JP, Hedon B. Prospective evaluation of falloposcopy. Human reproduction (Oxford, England). 1998;13(7):1815–8.
- 91. Lum D, Guido R, Rodriguez E, Lee T, Mansuria S, D'Ambrosio L et al. Brush cytology of the fallopian tube and implications in ovarian cancer screening. Journal of minimally invasive gynecology. 2014;21(5):851–6. doi:10.1016/j.jmig.2014.03.017. [PubMed: 24713115]
- Gizzo S, Noventa M, Quaranta M, Vitagliano A, Saccardi C, Litta P et al. A novel hysteroscopic approach for ovarian cancer screening/early diagnosis. Oncology letters. 2017;13(2):549–53. doi: 10.3892/ol.2016.5493. [PubMed: 28356928]
- 93. Chen H, Klein R, Arnold S, Chambers S, Zheng W. Cytologic studies of the fallopian tube in patients undergoing salpingo-oophorectomy. Cancer Cell International. 2016;16(1):78. doi: 10.1186/s12935-016-0354-x. [PubMed: 27733814]
- Chen H, Klein R, Arnold S, Wang Y, Chambers S, Zheng W. Tubal Cytology of the Fallopian Tube as a Promising Tool for Ovarian Cancer Early Detection. Journal of visualized experiments : JoVE. 2017(125). doi:10.3791/55887.
- Otsuka I, Kameda S, Hoshi K. Early detection of ovarian and fallopian tube cancer by examination of cytological samples from the endometrial cavity. British journal of cancer. 2013;109(3):603–9. doi:10.1038/bjc.2013.402. [PubMed: 23868002]
- Rodriguez EF, Lum D, Guido R, Austin RM. Cytologic findings in experimental in vivo fallopian tube brush specimens. Acta cytologica. 2013;57(6):611–8. doi:10.1159/000353825. [PubMed: 24107657]
- 97. Kinde I, Bettegowda C, Wang Y, Wu J, Agrawal N, Shih Ie M et al. Evaluation of DNA from the Papanicolaou test to detect ovarian and endometrial cancers. Science translational medicine. 2013;5(167):167ra4. doi:10.1126/scitranslmed.3004952.
- 98. Wang Y, Li L, Douville C, Cohen JD, Yen TT, Kinde I et al. Evaluation of liquid from the Papanicolaou test and other liquid biopsies for the detection of endometrial and ovarian cancers. Science translational medicine. 2018;10(433). doi:10.1126/scitranslmed.aap8793.
- 99. Bast RC Jr., Matulonis UA, Sood AK, Ahmed AA, Amobi AE, Balkwill FR et al. Critical Questions in Ovarian Cancer Research and Treatment: Report of an American Association for Cancer Research Special Conference. Cancer. 2019. doi:10.1002/cncr.32004.
- 100. Mourits MJ DBG. European/US comparison and contrasts in ovarian cancer screening and prevention in a high-risk population. 2017 ASCO Educational Book 2017:124–7.

Study Name Recruitment Years	RCT	Location	# Women	Screening Method	Aim	Outcome	Limitations/Strengths
Kentucky study [36]	No	USA Kentucky	37,293 Screened	Annual TVS	5-year OC survival:	Screened (74.8±6.6%) vs Unscreened (53.7±2.3%) P<0.001 Hard to interpret true effects of screening on disease mortality in non-RCT	Single-center Unscreened controls were diagnosed with OC in the center but not part of study Lead time effect Healthy volunteer effect
SCSOCS [35] 1985 to 1999	Yes	Japan Shizuoka 212 Hospitals	82,487 Total (41,688 Screened) (40,799 Control)	Annual CA125 (> 35U/mL) Annual TVS	Early stage I/II diagnosis:	Screened (63%) vs Control (38%) P=0.23	
PLCO [40, 41] 1993 to 2001	Yes	USA 10 Sites	78,216 Total (34,253 Screened) (34,304 Usual care)	Annual CA125 (> 35U/mL) (6 years) Annual TVS (first 4 years)	All-cause mortality: For invasive EOC, tubal and peritoneal cancer	12.4 year risk ratio (1.18, 95% CI 0.91–1.54) 14.7 year risk ratio (1.01, 95% CI 0.97–1.05)	High surgical complication rate (15%) Lack of stage shift (22.2%)
UKCTOCS [42] 2001 to 2005	Yes	UK 13 Sites	202,638 Total (50,639 USS) (50,640 MMS *) (101,359 Control)	* SMM	Mortality reduction: For invasive EOC, tubal and peritoneal cancer Early stage I/II diagnosis:	Primary analysis: Mortality reduction over years 0- II4 was MMS 15% (95% CI -3 to 30; p=0.10) USS 11% (-7 to 27; p=0.21) Analysis excluding prevalent cases: Overall mortality reduction 20% (-2 to 40) (p=0.021) with reduction of 8% (-27 to 43) with reduction of 8% (-27 to 43) with reduction of 8% (-7 to 43) with reduction of 8% (-7 to 43) with reduction over years $7-14$ in favor of MMS and 23%, (-3 to 49) mortality reduction over years $7-14$ in favor of MMS for 53.9%, but not with USS (P=.0001)	Screening group had higher worry and lower pleasure scores when >2 TVS performed as compared to no TVS [44] Good PPV of 4.4 surgeries per cancer detected [45]
NROSS [46] 2001 to 2011	No	USA 7 Sites	4,051 Total	*MMS	Specificity and PPV of OC screening in PM women:	PPV of 40% (95% CI 12.2% to 73.8%) Specificity 99.9% (95% CI 99.7% to 100%)	
*					- - - - -		

^{*}MMS (ROCA CA125 + TVS) CA125 interpreted using a risk of ovarian cancer algorithm (ROCA) comparing each participant's CA125 value with its previous value, and testing for a change-point even at values < 35U/mL. No change in CA125 led to annual CA125. Intermediate change in CA125 triggered a repeat CA125 in 3 months, larger changes led to a secondary screen with TVS.

Table 1

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Ovarian Cancer Screening Trials in Normal Risk Postmenopausal Women