



HHS Public Access

Author manuscript

Exp Dermatol. Author manuscript; available in PMC 2020 August 01.

Published in final edited form as:

Exp Dermatol. 2019 August ; 28(8): 886–891. doi:10.1111/exd.13978.

Contribution of Fibroblasts to Tunnel Formation and Inflammation In Hidradenitis Suppurativa/ Acne Inversa

JW Frew¹, K Navrazhina¹, M Marohn², CP Lu², JG Krueger¹

¹Laboratory of Investigative Dermatology, Rockefeller University, New York, NY

²The Hansjörg Wyss Department of Plastic Surgery, Department of Cell Biology New York University

Abstract

The precise pathogenic mechanisms in the development, persistence and worsening of Hidradenitis Suppurativa (HS) remain ill-defined. This chronic inflammatory dermatosis displays a strong Th1 and Th17 inflammatory signature with elevated levels of TNF- α , IL-1 β , IL-17 and IFN γ in lesional and perilesional tissue. HS significantly differs to other chronic inflammatory dermatoses due to the development of hypertrophic scarring and dermal tunnels. The development of scarring and tunnels suggests that fibroblastic stromal cells (including myofibroblasts, fibroblasts, pericytes etc.) may be involved in the development and progression of disease. Heterogeneous populations of fibroblasts have been identified in other inflammatory disorders and malignancy which contribute to inflammation and present novel therapeutic targets for fibrotic disorders. Findings in HS are consistent with these fibroblast subpopulations and may contribute to tunnel formation, aggressive squamous cell carcinoma and the phenotypic presentation of familial HS variants. We describe the existing knowledge regarding these mechanistic pathways and methods to confirm their involvement in the pathogenesis of HS.

Keywords

Hidradenitis Suppurativa; Acne Inversa; Pathogenesis; Fibroblasts; Scarring

1. Introduction:

The precise pathogenic mechanisms in the development, persistence and worsening of Hidradenitis Suppurativa (HS) remain ill-defined¹. This chronic inflammatory dermatosis displays a strong Th1 and Th17 inflammatory signature with elevated levels of TNF- α , IL-1 β , IL-17 and IFN γ in lesional and perilesional tissue². HS significantly differs to other chronic inflammatory dermatoses³ due to the development of hypertrophic scarring and dermal tunnels⁴. The development of which suggest that fibroblastic stromal cells (including

Corresponding Author: Dr John W Frew, Laboratory of Investigative Dermatology, The Rockefeller University, 1230 York Avenue, New York, NY, United States of America. Ph: +1-212-327-7153, Fax: +1-212-327-8232, jfrew@rockefeller.edu.

Author Contribution Statement: JWF, KN, MM, CPL, JGK performed the research; JWF, KN, JGK designed the study, JWF, KN, JGK analyzed the data and wrote the paper

All Authors have read and approved the final manuscript.

Conflicts of Interest: The authors have no conflicts of interest to declare

myofibroblasts, fibroblasts, pericytes etc.) may be involved in the development and progression of disease⁵. The recent investigations by Sanchez et al⁵ experimentally demonstrate that biologically active peptides and inflammatory mediators from the extracellular matrix (ECM) engage in inflammatory crosstalk with keratinocytes in HS. This paper begins to lay the foundation for future investigations into the interaction between ECM, fibroblasts and keratinocytes in HS. Such insights may help explain why certain patients are prone to hypertrophic scarring, tunneling and tract formation, and the mechanisms underlying these presentations. This may also lead to the identification of novel therapeutic targets which overlap with pipeline drugs for fibrotic diseases⁶. A large body of literature exists regarding epidermal-stromal interactions in the setting of cutaneous melanoma⁷ and other inflammatory disorders such as Rheumatoid Arthritis (RA)⁸ and Inflammatory Bowel Disease (IBD)⁹, however, investigations in inflammatory skin disease are limited. Our background knowledge in these conditions may help generate testable hypotheses regarding the mechanisms at play in inflammatory dermatoses including HS.

2. Mesenchymal Stromal Cells as a Heterogeneous Population:

Fibroblast populations in chronic inflammation and malignancy are heterogeneous. Fibroblasts are identified by their spindle shaped cellular morphology in combination with common mesenchymal markers such as vimentin⁹⁻¹² and develop from multiple sources including resident tissue stromal cells, bone marrow derived stromal cells, Epithelial Mesenchyme Transition (EMT) and Endothelial Mesenchyme Transition (endoMT) from epithelial and endothelial cells, respectively.^{10,11} The source of these stromal cells can be identified through examination of specific cell markers (such as PDGFR α , PDGFR β and Podoplanin)⁹⁻¹², however, no individual marker is known to be predictive of function. Transcriptomics and single cell RNA sequencing (scRNAseq) have recently been able to identify and characterize subpopulations of stromal cells^{9,11} and provide insights into their putative function in malignancy and inflammation. Factors including cellular origin, spatial location, microenvironment and degree of differentiation all contribute to the functional phenotype (i.e. matrix re-modelling, epithelial maintenance) of fibroblasts identified in these disorders⁹⁻¹². This results in a heterogeneous milieu of fibroblasts which are proposed to be differentially dysregulated in different forms of aberrant wound healing, fibrosis, hypertrophic scarring, malignancy and metastases⁹⁻¹². Common fibroblast subpopulations have been identified within chronic inflammatory conditions, chronic wound healing, malignancy and metastasis⁹⁻¹². This indicates common pathways may be at play which are potential therapeutic targets in these various disorders.

3. The role of Fibroblasts in chronic inflammation and epithelial integrity:

Fibroblast dysregulation has been implicated in impaired wound healing¹², inflammatory disorders such as pyoderma gangrenosum¹⁴ (associated with HS in the PASH and PAPASH syndromes)¹⁵, and in the development of tertiary lymphoid follicles¹² in RA and IBD.¹⁵ Dysregulation is proposed to involve priming signals in the stromal environment during acute inflammation (mediated by granulocytes) to epigenetically modulate fibroblasts, leading to expression of markers including α -SMA, S100A4, FAP- α ¹⁶. Such activation is purported to result in epigenetic modifications¹⁷ given the persistence of the activated

phenotype after isolation of cells and *in vitro* culture^{12,15,17}. This activation leads to upregulation of ICAM-1, VCAM-1, CXCL13, CCN2, IL-6^{18,19} as well as NLRP3 and Caspase-1 inflammasome activation leading to further IL-1 β , IL-18 release, matrix metalloproteinase (MMP) production and release^{9,10,15}, and leucocyte recruitment^{18,19}. MMPs are required for the activation of pro IL-1 β , pro TNF- α , pro TGF- β as well as potentiating the action of IL-8²⁰. In normal tissue, the presence of a second maturation and stabilization signal (including activation of the TNFRSF3 Pathway and ROR γ pathway)¹⁵ ensues, however, blockade of these signals leads to a disorganized collection of mixed inflammatory cells reminiscent of the mixed inflammation in established HS.

Bidirectional stromal-epithelial signaling via the Wnt pathway is integral to maintaining the epidermal stem cell compartment¹³. Dysregulation of this signaling leads to poor epidermal regeneration and is implicated in impaired wound healing and ulceration in Crohn's disease⁹. A specific subpopulation of fibroblasts, found adjacent to the involved epithelium, has been identified which mediates this bidirectional signaling via CD44^{9-12,16}, however, communication can occur without direct cell to cell contact²¹ via microvesicles¹⁸ or other not-yet-identified mechanisms. Upregulation of Wnt signaling has been shown to accelerate both fibrosis and re-epithelialization in animal and human models²², with Wnt/B-Catenin signaling being suppressed in cutaneous wounds of diabetic patients^{22,23}, a common comorbidity in HS.

4. Fibroblast Like Stromal Cells in Malignancy and Metastases

In the setting of malignancy, dysregulated fibroblasts are known as cancer-associated fibroblasts (CAFs)^{10,11,16,24,25}. CAFs are associated with increased tumor growth and metastatic potential as well as extra cellular matrix remodeling^{10,11,16,24,25}. CAFs contribute to the pro-inflammatory nature of the tumor microenvironment²⁵ and remodeling of the ECM^{11,16}. Bidirectional crosstalk with tumor cells has been shown to increase the metastatic potential of melanoma, breast, pancreatic and lung tumours^{11,16,24,25}. Stromal-derived IL-6 and TGF- β have been documented to suppress cell cycle progression in melanoma²⁷, controlling tumor spread in early stage disease. However, in the setting of malignancy with activated fibroblasts, the altered CAF secretome results in exposure of the tumor cells to growth factors and metabolites which promote tumor propagation²⁸. CAFs have been shown to harbor immunomodulatory mechanisms with upregulation of production of cytokines and chemokines, including PDGF, vascular endothelial growth factor A (VEGFA), prostaglandin E2, IL-6, TNF, nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), IL-8, hepatocyte growth factor (HGF), and CXCL12^{28,29}. This inflammation results in an autocrine signaling capability in CAFs contributing to ongoing inflammation. Activated CAFs, in a similar fashion to other fibroblasts, produce multiple MMPs including MMP1-3, MMP9, and MMP13-14, as well as tissue inhibitors of metalloproteinases (TIMPs), which together result in ECM remodeling. In malignancy, this remodeling facilitates tumor invasion and metastasis²⁹. Additionally, activated stromal cells can induce EMT and motility in malignant cells via CD44 signaling¹⁶. The cause of this aberrant activation is secondary to the pre-existing inflammatory milieu, including the local microenvironment, EMT and EndoEMT programs, as well as the influx of heterogeneous populations of mesenchymal cells primed for activation from bone marrow. Concurrently,

expression of CCN2 and production of miRNAs can activate stromal cells to a CAF phenotype^{24,25}, demonstrating the extensive cross talk in the setting of malignancy.

5. Mesenchymal Subpopulations Mediate Disparate Functional Pathways in Inflammation, Fibrosis and Fistula Formation in Inflammatory Bowel Disease

Dysregulated fibroblasts are implicated in the development of inflammation, fibrosis and fistula formation in IBD⁹, several inflammatory characteristics of RA^{12,15}, and enhanced metastatic potential in breast carcinoma¹¹. Specific subpopulations of fibroblasts are associated with the loss of epithelial integrity, ECM remodeling, angiogenesis and development of tertiary lymphoid structures^{9,11,12,15}. PDFGRA+ cells with differentially elevated expression of DCN, LUM, VCAN, Col14A1, FBLN1, FBLN2, SMOC, LOX, LOX1 and CXCL14 display an ECM and EMT related transcriptomic signatures associated with ECM remodeling and stimulation of EMT in epidermal tissues^{9,10,11,12,15}. PDFGRB-fibroblasts with differentially elevated expression of POSTN, BMP2, BMP5, WNT5A, WNT5B, SOX6, SCRG1, SOX9, SOX10, MFAP5 and CCN2 were described as enriched in inflammation^{9,10,11,12,15}, and associated with fibroblasts derived from epidermal tissues via EMT. Podoplanin + fibroblasts were identified as a subgrouping of PDFGRA+ fibroblasts with expression suggestive of an EndoMT origin^{9,10,11,12,15}. This subset has been implicated in the development of tertiary lymphoid follicles in chronic inflammation including IBD and RA^{9,12,15}. NR2F2 expressing fibroblasts with elevated expression of Notch3, Epas1, and COL18A1 have transcriptomic profiles suggestive of a pro-angiogenic profile with a subset of these cells also expressing high levels of cell cycle related genes^{9,10,11,12,15}. In IBD, an increase proportion of PDGFRA+ cells are implicated in T cell recruitment and barrier dysfunction⁹. This subpopulation was significantly expanded in IBD patients compared with healthy controls⁹. PDFGRB- fibroblasts were decreased in IBD and this subpopulation was implicated in epithelial maintenance⁹.

6. The Inflammatory Signature of Hidradenitis Suppurativa Suggests a Possible Contribution of Fibroblasts

Existing studies in HS demonstrate intriguing evidence of fibroblast-related pathways being activated in lesional tissue. Elevations in IL-1 β , IL-6, IL-8, MMP2 and MMP9 as well as extensive matrix remodeling are core manifestations of HS^{1,2,5}. IHC studies have identified co-localization of MMP2 to dermal fibroblasts³⁰. The presence of isolated keratinocytes in the dermis is a longstanding phenomenon in HS and can be explained by EMT and the motility of keratinocyte derived MSCs which still partially express keratinocyte markers³⁰. The Invasive Proliferative Gelatinous Mass (IPGM), found attached to the epithelium of the sinus tracts of HS, has been characterized as an active inflammatory component of the disease and comprised of Neutrophil Extracellular Traps (NETs)³¹. Such NETs develop through a process of NETosis which is stimulated by IL-8, IL-6, TNF- α ³². NETs have been documented to stimulate EMT and EndoMT in other autoimmune diseases such as lupus nephritis and rheumatoid arthritis³³. Microbiota such as porphyromonas sp., which are

associated with HS and colonize epithelialized dermal tunnels, are known triggers of NETosis^{32,34}. The extensive tunnel formation in HS may be partially explained by PDFGRA + fibroblasts stimulating matrix remodeling and tunnel formation, whilst the epithelialization of dermal tunnels may be partially explained by the role of PDFGRB- fibroblasts in stimulating re-epithelialization as a form of aberrant wound healing. Such epithelialized tunnels may then perpetuate Th17-mediated keratinocyte feed forward inflammation, as well as stimulating fibroblasts to produce IL-6 and IL-8. This may partially explain the dramatic reduction in draining fistula counts with IL-23 blockade³⁵.

7. Stromal Cell MMP secretion linked to gamma secretase associated pathways:

Gamma secretase associated polymorphisms have been identified in a minority of patients with familial HS³⁶. These polymorphisms involve three of the four subunits of the gamma secretase complex - Nicastrin, Presenilin 1 and the Presenilin enhancer³⁶. The existing hypothesis is that gamma secretase polymorphisms alter Notch signaling which leads to hyperproliferative keratinocytes in the follicular infundibulum leading to follicular occlusion¹, however, the possibility of alterations in Notch signaling representing a generalizable inflammatory phenomenon (rather than a mechanistic pathway specific to HS) have not been experimentally tested³⁷. Nicastrin (NCSTN) haploinsufficiency is shown to decrease expression of Interferon related genes in HEK293 cells³⁸, however, does not alter cytokine profiles of stimulated PBMCs³⁹. An only recently appreciated aspect of the gamma secretase complex is its role in fibrosis⁴⁰⁻⁴². Presenilin-1 is the proteolytic component of gamma secretase which is responsible for degradation of CD44 as well as other adhesion molecules such as N-cadherin⁴⁰. Cao et al's Nicastrin knockdown model also revealed downregulated N-Cadherin and upregulation of CXCL14³⁸, which in addition to its immune surveillance properties, is an autocrine factor associated with activated fibroblasts⁴³. Given the role of Nicastrin as the 'gatekeeper' of the catalytic activity of the gamma secretase complex^{41,42}, polymorphisms in Nicastrin leading to overactivity of gamma secretase can theoretically lead to profibrotic activity through upregulation of EMT and cellular motility⁴⁰⁻⁴². Presenilin 1 also upregulates TGF- β activity, MMP secretion and the β catenin/Wnt pathway^{40,44}. Upregulation of extracellular proteolysis would also explain the excessive fibrotic extensive matrix remodeling and significant scarring seen in Chinese HS patients with documented Nicastrin mutations³⁷, as opposed to the typical axillary-mammary inflammatory form of the disease¹.

8. Stromal Cell Contribution May Explain Aggressive Behaviour of SCC in HS

One of the most serious complications of long-term HS is the development of aggressive, metastatic squamous cell carcinoma⁴⁵⁻⁴⁷. Despite their well differentiated nature, these tumors rapidly metastasize and can be fatal. Suggested contributing factors include human papillomavirus infection⁴⁸ as well as the propensity for longstanding chronic wounds to develop SCCs⁴⁹. Despite this well documented association between chronic non-healing wounds, the exact mechanisms remain unclear. Evidence from Epidermolysis Bullosa

(Recessive Dystrophic, Generalized Severe subtype)⁴⁷ implicates a unique dermal microenvironment (including elevated Wnt5A and TSP1 signaling along with a dense population of inflammatory cells and activated fibroblasts) in the development and rapid progression of cutaneous SCC, which are often fatal⁴⁷. Additionally, gram negative flagellated bacteria may activate TLR5 in RDEB contributing to the pro-tumorigenic environment⁴⁷. These same upregulated factors are identified in PDGFRB- fibroblasts in Crohn's disease, and are associated with epithelial maintenance⁹. This implies a potential common mechanism in the development of highly aggressive RDEB and other diseases including Crohn's disease and possibly HS.

9. Common mesenchymal pathways may explain the association of Pyoderma Gangrenosum/ IBD/ Arthritis and Neutrophilic Alopecias

Monogenic Autoinflammatory Syndromes manifest in both HS and other inflammatory disorders such as Pyoderma Gangrenosum and Inflammatory Arthropathies including the PASH and PAPASH syndromes⁵⁰. HS also has a strong epidemiological association with IBD^{1,51}. The coexistence of these conditions implies a possible degree of commonality in their pathogenesis. Dysregulated fibroblasts have been implicated in inflammatory arthropathies and play a significant role in impaired wound healing^{19,22,23,52}. The pro-inflammatory actions of activated fibroblasts are NLRP3 and Caspase-1 inflammasome mediated^{9,10,15}, lending credence to the suggestion that they may be involved in the pathogenesis of PAPA and PASH syndromes, given the association of these syndromes with pathogenic sequence variants in these pathways³⁶. The corollary of this is that other manifestations of these disorders (such as Pyoderma Gangrenosum) may also involve fibroblast dysregulation as a manifestation of aberrant wound healing. The hypothesis would include a role for the increase of PDGFRB+ fibroblasts due to the extensive degree of ECM remodeling and a decrease in PDGFRB- fibroblast which are responsible for epithelial integrity in a similar fashion to colonic ulceration in IBD⁹. Two other cutaneous conditions bear striking histological, clinical and immunological resemblance to HS, Dissecting Cellulitis of the Scalp and Complex Pilonidal Disease, and the involvement of fibroblasts in these disorders warrants further consideration.

10. IL-17, IL-1, CXCL14, JAK-STAT, Cadherin-11, CDK, miR-203, FRP2/ALX, podoplanin, CCN2 pathways are potential therapeutic targets of Fibroblasts in HS.

There is an urgent need for novel therapies in HS and therefore, identification of novel druggable targets is critical. The presence of distinct fibroblast subpopulations has been identified in lesional and perilesional HS tissue which mirror the expression markers (PDGFRA+, PDGFRA-, NR2F2, Podoplanin) seen in other inflammatory conditions such as Crohn's disease (Lu et al, unpublished data), although their functions in vivo remain to be established. A number of fibroblast-targeted therapies exist within the rheumatological and immunological therapeutic pipeline⁵³. Along with IL-17, IL-1 and JAK-STAT blockade, (currently in Phase 2 clinical trials)⁵⁴, other druggable targets include IL-6R, Cadherin 11,

cyclin dependent kinases (CDK1, CDK2, CDK4, CDK6), SYK (reducing IL-6 production via the MAPK-PKC pathway), CXCL14, CCN2 as well as Formyl peptide receptor 2 (FRP2/ALX) as an upstream mediator of Stat 1, IL-6 and Podoplanin. One potential complication of note is the disparity between responses to therapy between joint-derived versus skin-derived fibroblasts⁵⁵ which is reflected in the imperfect efficacy correlation between skin and joint targets in psoriasis and psoriatic arthritis⁵⁶.

Acknowledgements:

J.W.F. was supported in part by grant # UL1 TR001866 from the National Center for Advancing Translational Sciences (NCATS), National Institutes of Health (NIH) Clinical and Translational Science Award (CTSA) program. K.N. was supported by a MSTP grant from the National Institute of General Medical Sciences of the NIH under award number T32GM007739 to the Weill Cornell/Rockefeller/Sloan Kettering Tri-Institutional MD-PhD Program.

References:

1. Vossen ARJV, van der Zee HH and Prens EP. Hidradenitis Suppurativa: A Systematic Review Integrating Inflammatory Pathways Into a Cohesive Pathogenic Model. *Front. Immunol* 2018 9:2965. doi: 10.3389/fimmu.2018.02965 [PubMed: 30619323]
2. Frew JW, Hawkes JE and Krueger JG. A systematic review and critical evaluation of inflammatory cytokine associations in hidradenitis suppurativa [version 1; referees: awaiting peer review]. *F1000Research* 2018, 7:1930 [PubMed: 30828428]
3. Czarnowicki T, He H, Leonard A, Kim HJ et al. Blood endotyping distinguishes the profile of vitiligo from that of other inflammatory and autoimmune skin diseases *J Allergy Clin Immunol* 2019;
4. Scheinfeld N An Atlas of the morphological manifestations of hidradenitis suppurativa *Dermatol Online J* 2014;20:22373 [PubMed: 24746309]
5. Sanchez J, Le Jan S, Muller C Matrix Remodeling and MMP expression/activation is associated with hidradenitis suppurativa skin inflammation. *Exp Dermatol* 2019; doi: 10.1111/exd.13919
6. Henrot P, Truchetet ME, Fisher G, Taieb A, Cario M CCN proteins as potential actionable targets in scleroderma *Exp Dermatol* 2019;28:11–18 [PubMed: 30329180]
7. Izar B, Joyce CE, Goff S, Cho NL et al. Bidirectional cross talk between patient-derived melanoma and cancer-associated fibroblasts promoted invasion and proliferation *Pigment Cell Melanoma Res.* 2016;29:656–668 [PubMed: 27482935]
8. Hirota K, Hashimoto M, Ito Y, Matsuura M et al. Autoimmune Th17 cells induced Synovial Stromal and Innate Lymphoid Cell Secretion of the Cytokine GM-CSF to Initiate and Augment Autoimmune Arthritis *Immunity* 2018;48(6):1220–1225 [PubMed: 29802020]
9. Kinchen J, Chen HH, Parikh K, Antanaiciute A et al. Structural Remodeling of the Human Colonic Mesenchyme in Inflammatory Bowel Disease *Cell* 2018;175:372–386 [PubMed: 30270042]
10. Sasaki K, Sugai T, Ishida K, OSAkabe M et al. Analysis of cancer-associated fibroblasts and the epithelial mesenchymal transition in cutaneous basal cell carcinoma, squamous cell carcinoma and malignant melanoma *Human Pathol* 2018;79:1–8 [PubMed: 29555579]
11. Bartoschek M, Oskolkov N, Bocci M, Lovrot J et al. Spatially and functionally distinct subclasses of breast cancer -associated fibroblasts revealed by single cell RNA sequencing. *Nature Comm* 2018;9:5150
12. Dorraji SE, Hovd AK, Kanapathipillai P, Bakland G Mesenchymal stem cells and T Cells in the formation of Tertiary Lymphoid Structures in Lupus Nephritis. *Scientific Reports* 2018;8:7861 [PubMed: 29777158]
13. Philippeos C, Telerman SB, Oules B, Pisco AO et al. Spatial and Single Cell Transcriptional Profiling Identifies Functionally Distinct Human Dermal Fibroblast Subpopulations *J Invest Dermatol* 2018;138:811–825 [PubMed: 29391249]

14. Wang EA, Steel A, Luxardi G, Mitra A, et al. Classic Ulcerative Pyoderma Gangrenosum Is a T Cell-Mediated Disease Targeting Follicular Adnexal Structures: A Hypothesis Based on Molecular and Clinicopathologic Studies. *Frontiers in Immunology*, 2018; 8, 1980. [PubMed: 29379508]
15. Barone F, Gardner DH, Nayar S, Steinthal N et al. Stromal Fibroblasts in Tertiary Lymphoid Structures: A Novel Target in Chronic Inflammation *Frontiers Immunol* 2016;7:477
16. Zhou L, Yang K, Andl T, Wickett RR et al. Perspective of Targeting Cancer-Associated Fibroblasts in Melanoma *J Cancer* 2015;6:717–726 [PubMed: 26185533]
17. Albregues J, Bertero T, Grasset E Bonon S Epigenetic switch drives the conversion of fibroblasts into pro-invasive cancer-associated fibroblasts *Nature Comm* 2015;6:10204
18. Huang P, Bi J, Owen GR, Chen W Keratinocyte Microvesicles Regulate the Expression of Multiple Genes in Dermal Fibroblasts *J Invest Dermatol* 2015;135:3051–3059 [PubMed: 26288358]
19. Georganas C, Liu H, Perlman H, Hoffman A et al. Regulation of IL-6 and IL-8 Expression in Rheumatoid Arthritis Synovial Fibroblasts: the Dominant Role for NF-κB but not C/EBPβ or c-Jun *J Immunol* 2000;165:7199–7206 [PubMed: 11120852]
20. Nissinen L, Kahari VM Matrix Metalloproteinases in inflammation *Biochimica et Biophysica Acta (BBA)- General Subjects* 2014;1840:2571–2580
21. Hutchenreuther J, Vincent KM, Carter DE, Postovit LM, Leask A CCN2 Expression by tumor Stroma is required for Melanoma Metastasis *J Invest Dermatol* 2015;135:2805–2813 [PubMed: 26168233]
22. McBride JD, Jankins AJ, Liu X, Zhang B Elevated circulation levels of an antiangiogenic SERPIN in patients with diabetic microvascular complications impair wound healing through suppression of Wnt signaling. *J Invest Dermatol* 2014;134:1725–1734 [PubMed: 24463424]
23. Zhang H, Nie X, Shi X, Zhao J Regulatory Mechanisms of the Wnt/B-Catenin Pathway in Diabetic Cutaneous Ulcers *Front Pharmacol* 2018;9:1114 [PubMed: 30386236]
24. Hutchenreuther J, Vincent K, NORley C, Racanelli M Activation of cancer-associated fibroblasts is required for tumor neovascularization in a murine model of melanoma *Matrix Biol* 2018;74:52–61 [PubMed: 29885461]
25. Hutchenreuther J, Vincent KM, Carter DE, Postovit LM, Leask A CCN2 Expression by Tumor Stroma is Required for Melanoma Metastasis *J Invest Dermatol* 2015;135: 2805–2813 [PubMed: 26168233]
26. Balachander GM, Talukdar PM, Debnath M, Rangarajan A, Chatterjee K Inflammatory Role of Cancer-Associated Fibroblasts in Invasive Breast Tumors Revealed Using a Fibrous Polymer Scaffold *ACS Appl Mater Interfaces* 2018;10:33814–33826 [PubMed: 30207687]
27. Alkalias T, Flaberg E, Kashuba V, Alexeyenko A Inhibition of tumor cell proliferation and motility by fibroblasts is both contact and soluble factor dependent *Proc Natl Acad Sci USA* 2014;111:17188–17193 [PubMed: 25404301]
28. Alkalias T, Moyano-Galceran L, Arsenian-Henrikson M, Lehti K Fibroblasts in the Tumor Microenvironment: Shield or Spear? *Int J Mol Sci* 2018;19:1532
29. Kalluri R The biology and function of fibroblasts in cancer. *Nat. Rev. Cancer* 2016;16:582–598. doi: 10.1038/nrc.2016.73. [PubMed: 27550820]
30. Frew JW, Hawkes JE and Krueger JG. A systematic review and critical evaluation of immunohistochemical associations in hidradenitis F1000Research 2018, 7:1923 [PubMed: 31281635]
31. Kidacki M, Cong Z, Flamm A, Helm K, Danby F and Nelson A (2019), 'Invasive proliferative gelatinous mass' of hidradenitis suppurativa contains distinct inflammatory components. *Br J Dermatol*. doi: 10.1111/bjd.17541
32. Yang H, Biermann MH, Brauner JM, Liu Y et al. New Insights into Neutrophil Extracellular Traps: Mechanisms of Formation and Role in Inflammation *Front Immunol* 2016;7:302
33. Lee KH, Kronbichler A, Park DDY, Park YM et al. Neutrophil Extracellular Traps (NETs) in autoimmune diseases: a comprehensive review *Autoimmunity Rev* 2017;16:1160–1173 [PubMed: 28899799]
34. Ring HC, Sigsgaard V, Thorsen J, Fursted K et al. The microbiome of tunnels in hidradenitis suppurativa patients *J Eur Acad Dermatol Venereol* 2019;doi: 10.1111/jdv.15597

35. Kovacs M and Podda M (2019), Guselkumab in the treatment of severe hidradenitis suppurativa. *J Eur Acad Dermatol Venereol*, 33: e140–e141. doi: 10.1111/jdv.15368 [PubMed: 30480844]
36. Frew J, Vekic D, Woods J and Cains G (2017), A systematic review and critical evaluation of reported pathogenic sequence variants in hidradenitis suppurativa. *Br J Dermatol*, 177: 987–998. [PubMed: 28278367]
37. Frew JW We Need to Talk About Notch: Notch dysregulation as an epiphenomenon in inflammatory skin disease. *Br J Dermatol*. 2019 2;180(2):431–432
38. Cao L, Morales-Heil DJ, Roberson EDO Nicastrin haploinsufficiency alters expression of type-1 interferon stimulated genes in two immortalized human cell lines *Clin Exp Dermatol*. 2019 1 17. doi: 10.1111/ced.13906
39. Xu H, He Y, Hui Y, Xiao X et al. NCSTN mutations in hidradenitis suppurativa/acne inversa do not influence cytokine production by peripheral blood mononuclear cells *Br J Dermatol* 2017;176:270–280 [PubMed: 27479915]
40. Kryczka J and Boncela J Proteases Revisited: Roles and Therapeutic Implications in Fibrosis *Mediators INflamm* 2017:2570154
41. De Strooper B, Nicastrin: gatekeeper of the gamma-secretase complex *Cell* 2005;122(3):318–320 [PubMed: 16096051]
42. Shah S, Lee SF, Tabuchi K, Hao YH Nicastrin functions as a gamma-secretase substrate receptor *Cell* 2005;122(3):435–447 [PubMed: 16096062]
43. Lu J, Chatterjee M, Schmid H, Beck S, & Gawaz M (2016). CXCL14 as an emerging immune and inflammatory modulator. *Journal of inflammation (London, England)*, 13, 1. doi: 10.1186/s12950-015-0109-9
44. Patil PU, D’Ambrosio J, Inge LJ, Mason RW, and Rajasekaran AK, “Carcinoma cells induce lumen filling and EMT in epithelial cells through soluble E-cadherin-mediated activation of EGFR,” *Journal of Cell Science*, vol. 128, no. 23, pp. 4366–4379, 2015. [PubMed: 26483386]
45. Juviler PG, Patel AP, Qi Y. Infiltrative squamous cell carcinoma in hidradenitis suppurativa: A case report for early surgical intervention. *Int J Surg Case Rep*. 2019;55:50–53. doi: 10.1016/j.ijscr.2019.01.006. Epub 2019 Jan 19. [PubMed: 30685629]
46. Jourabchi N, Fischer AH, Cimino-Mathews A, Waters KM, Okoye GA. Squamous cell carcinoma complicating a chronic lesion of hidradenitis suppurativa: a case report and review of the literature. *Int Wound J*. 2017 4;14(2):435–438. doi: 10.1111/iwj.12671. Epub 2016 Sep 29. [PubMed: 27681476]
47. Kim M, Murrell DF. Update on the pathogenesis of squamous cell carcinoma development in recessive dystrophic epidermolysis bullosa. *Eur J Dermatol*. 2015 4;25 Suppl 1:30–2. doi: 10.1684/ejd.2015.2552. [PubMed: 26083672]
48. Scheinfeld N A case of a patient with Stage III Familial hidradenitis suppurativa treated with 3 courses of infliximab and died of metastatic squamous cell carcinoma. *Dermatol Online J* 2014;20:3
49. Reich-Schupke S, Doerler M, Wollina U, Dissemmond J, et al. Plattenepithelkarzinome in chronischen venösen Ulcera crurum. Daten aus dem deutschen Marjolin-Register und Übersichtsdarstellung. *JDDG: Journal der Deutschen Dermatologischen Gesellschaft* 2015; 13: 1006–1014.
50. Gasparic J, Theut Riis P, Jemec GB Recognizing syndromic hidradenitis suppurativa: a review of the literature *J Eur Acad Dermatol Venereol* 2017;31:1809–1816 [PubMed: 28696038]
51. Vilarrasa Rull E, Gonzalez Lama Y Clinical features of hidradenitis suppurativa and Crohns disease: what do these two entities have in common? *Actas Dermosifiliogr* 2016;107:S21–26
52. Caley MP, Martins VL, & O’Toole EA (2015). Metalloproteinases and Wound Healing. *Advances in wound care*, 4(4), 225–234. [PubMed: 25945285]
53. Dakin SG, Coles M, Sherlock JP, Powrie F et al. Pathogenic stromal cells as therapeutic targets in joint inflammation *Nat Rev Rheum* 2018;14:714–726
54. Van Straalen KR, Schneider-Burns S, Prens EP Current and Future treatment of hidradenitis suppurativa *Br J Dermatol* 2018;doi: 10.1111/bjd.16768

55. Noack M, Miossec P P067 IL-17 resulting from cell interactions during chronic inflammation: comparison between joint-derived- and skin-derived-mesenchymal cells *Annals of the Rheumatic Diseases* 2018;77:A41–A42.
56. Hawkes Jason E., Yan Bernice Y., Chan Tom C., Krueger James G. Discovery of the IL-23/IL-17 Signaling Pathway and the Treatment of Psoriasis *J Immunol* 2018, 201:1605–1613; [PubMed: 30181299]

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

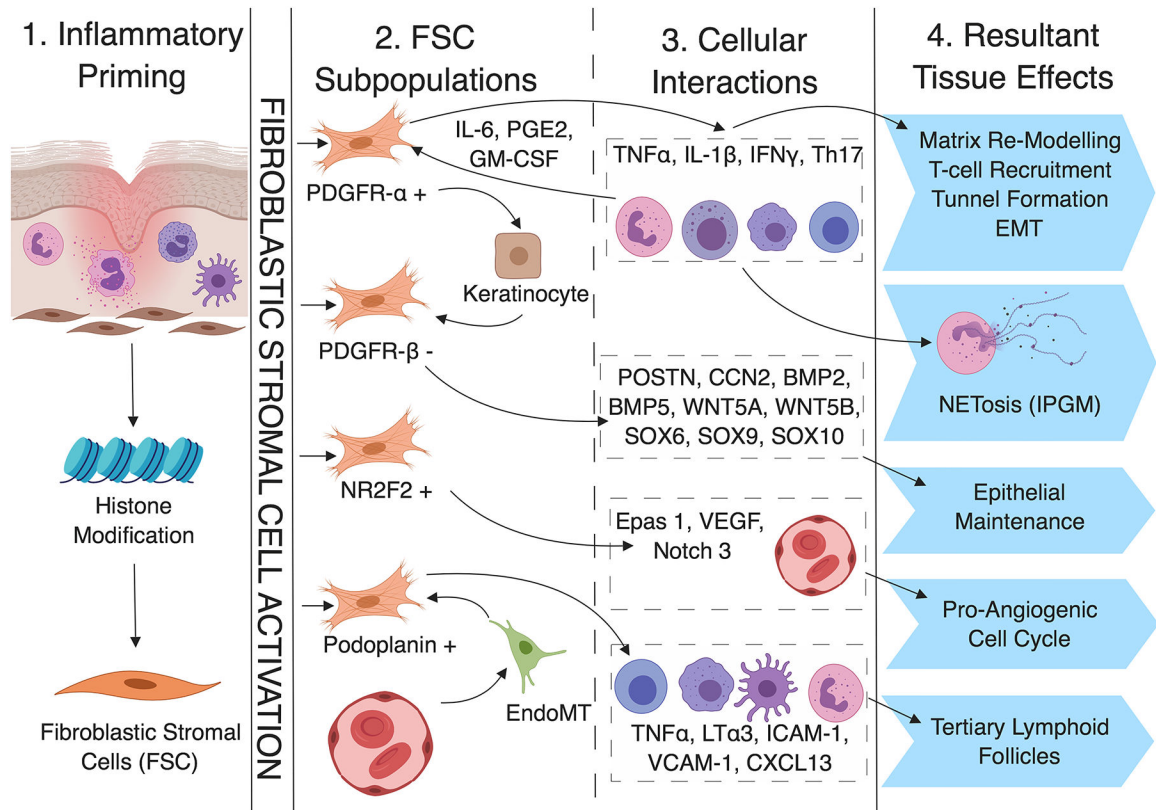


Figure 1: Proposed Role for Fibroblasts in Tunnel Formation and Inflammation in Hidradenitis Suppurativa. Acute inflammation may act as a priming signal for the activation of fibroblasts through epigenetic mechanisms (1. Inflammatory Priming) leading to the emergence of four discrete subpopulations (2. FSC Subpopulations). Each subpopulation demonstrates specific functional characteristics via actions involving specific inflammatory mediators, transcription factors and cellular pathways (3. Cellular Interactions). Such interactions are associated with (and proposed to be mechanistic causative in) the development of tunnels, epithelialized tracts and fibrosis in HS (4. Resultant Tissue Effects).