

# CIC-DUX4 oncoprotein drives sarcoma metastasis and tumorigenesis via distinct regulatory programs

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**Transcription factor fusion genes create oncoproteins that drive oncogenesis and represent challenging therapeutic targets. Understanding the molecular targets by which such fusion oncoproteins promote malignancy offers an approach to develop rational treatment strategies to improve clinical outcomes. Capicua–double homeobox 4 (CIC-DUX4) is a transcription factor fusion oncoprotein that defines certain undifferentiated round cell sarcomas with high metastatic propensity and poor clinical outcomes. The molecular targets regulated by the CIC-DUX4 oncoprotein that promote this aggressive malignancy remain largely unknown. We demonstrated that increased expression of ETS variant 4 (*ETV4*) and cyclin E1 (*CCNE1*) occurs via neomorphic, direct effects of CIC-DUX4 and drives tumor metastasis and survival, respectively. We uncovered a molecular dependence on the CCNE-CDK2 cell cycle complex that renders CIC-DUX4-expressing tumors sensitive to inhibition of the CCNE-CDK2 complex, suggesting a therapeutic strategy for CIC-DUX4-expressing tumors. Our findings highlight a paradigm of functional diversification of transcriptional repertoires controlled by a genetically aberrant transcriptional regulator, with therapeutic implications.**

## Introduction

The biological regulation of transcription factors and repressor proteins is an essential mechanism for maintaining cellular homeostasis, and is often dysregulated in human cancer (1). Indeed, chromosomal rearrangements involving transcriptional regulatory genes that lead to transcriptional dysregulation are present in many cancers, including approximately 30% of all soft tissue sarcomas (2). The majority of these oncogenic fusions involve transcription factors or regulators that are not readily “druggable” in a direct pharmacologic manner and thus have proven difficult to therapeutically target in the clinic. A prime example is the CIC-DUX4 fusion oncoprotein, which fuses capicua (*CIC*) to the double homeobox 4 gene, *DUX4*. CIC-DUX4-positive soft tissue tumors are an aggressive subset of undifferentiated round cell sarcomas that arise in children and young adults. Despite histological similarities to Ewing sarcoma, CIC-DUX4-positive sarcomas are clinically distinct and typically characterized by the rapid development of lethal metastatic disease and chemoresistance (3). By revealing downstream molecular targets that relay the functional output of CIC-DUX4, we can potentially identify and develop a more rational therapeutic strategy to improve patient outcomes.

CIC is a transcriptional repressor protein (4). The CIC-DUX4 fusion structurally retains greater than 90% of native CIC, yet it

functions as a transcriptional activator instead of a transcriptional repressor (5, 6). This property suggests that the C-terminal DUX4 binding partner may confer neomorphic transcriptional regulatory properties to CIC, while retaining WT CIC DNA binding specificity. *ETV4* is one of the most well-characterized transcriptional targets of CIC-DUX4 (5, 7, 8). More than 90% of CIC-DUX4-expressing tumors show *ETV4* upregulation by IHC, which distinguishes them from other small round blue cell sarcomas (8). We and others have demonstrated a prometastatic function for *ETV4* overexpression in tumors with inactivation of WT CIC (9, 10). The functional role of *ETV4* in CIC-DUX4-positive tumors is unknown.

Beyond *ETV4*, the identity and function of other CIC target genes are less well defined. Intriguingly, recent studies showed increased expression of cell-cycle-regulatory genes in CIC-rearranged sarcomas, although the functional relevance of this observation for oncogenesis and cancer growth in this context is unclear (6, 7).

Here, we developed a range of in vitro and in vivo cancer model systems to define the mechanism by which CIC-DUX4 controls capicua-regulated transcriptional pathways to promote hallmark features of malignancy, including tumor cell survival, growth, and metastasis.

## Results and Discussion

We recently reported that inactivation of native CIC derepresses an *ETV4*-MMP24-mediated prometastatic circuit (9). Since *ETV4* is a direct transcriptional target of CIC-DUX4 (Supplemental Figure 1A; supplemental material available online with this article; <https://doi.org/10.1172/JCI126366DS1>) (6, 7), we hypothesized that the high metastatic rate observed in patients harboring CIC-DUX4 fusions was dependent on *ETV4* expression. To explore this hypothesis, we developed an orthotopic soft tissue metastasis mod-

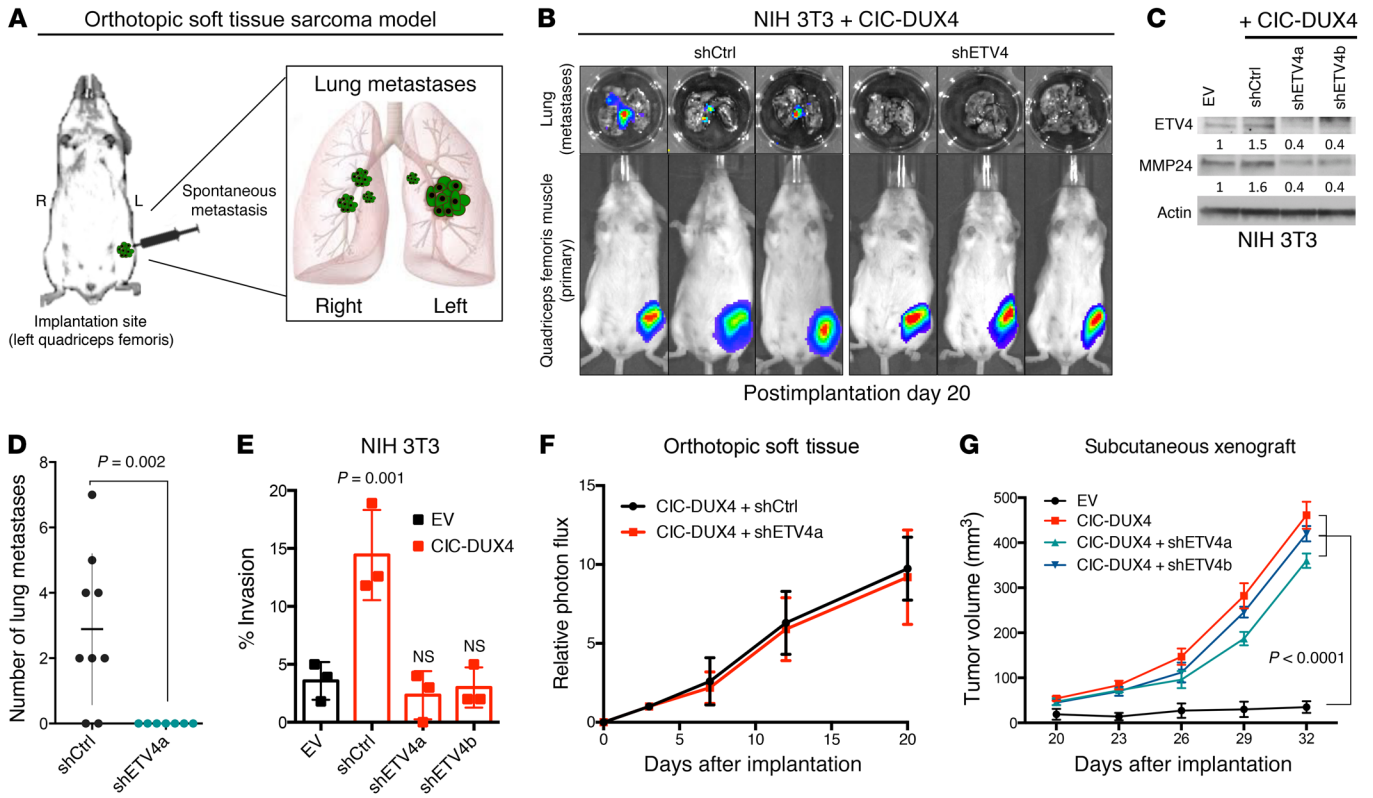
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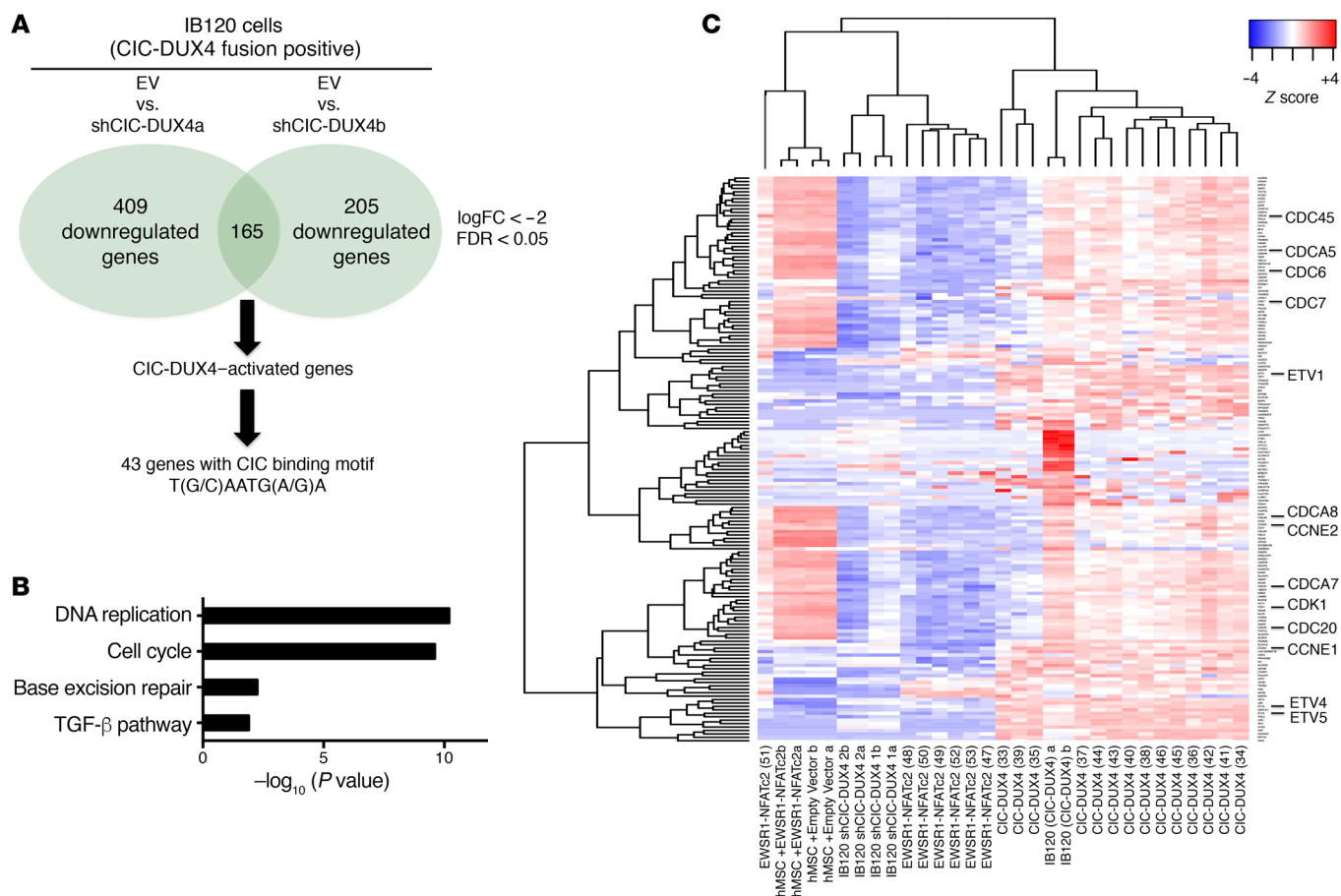


**Figure 1. ETV4 promotes metastasis in CIC-DUX4-expressing sarcoma.** (A) Schematic of the orthotopic soft tissue metastasis model. (B) Representative bioluminescence (BLI) images from mice bearing CIC-DUX4-expressing NIH 3T3 cells with shCtrl ( $n = 9$ ) or shETV4 ( $n = 7$ ). (C) Immunoblot of ETV4 and MMP24 from NIH 3T3 cells expressing empty vector (EV), shCtrl, shETV4a, or shETV4b. (D) Number of lung metastases in mice bearing CIC-DUX4-expressing NIH 3T3 cells with shCtrl ( $n = 9$ ) or shETV4 ( $n = 7$ ).  $P$  value, Student's  $t$  test. (E) Transwell invasion assay comparing CIC-DUX4-expressing NIH 3T3 cells with EV, shCtrl, shETV4a, or shETV4b.  $P$  value, 1-way ANOVA. Error bars represent SEM. (F) Relative photon flux from mice orthotopically implanted with CIC-DUX4-expressing NIH 3T3 cells expressing either shCtrl or shETV4. Error bars reflect SEM. (G) Subcutaneously implanted NIH 3T3 cells with EV, CIC-DUX4, or CIC-DUX4 plus shETV4a or shETV4b.  $P$  value, 1-way ANOVA. Error bars represent SEM.

el utilizing luciferase-based imaging to track tumor dissemination in vivo (Figure 1A). This system produces rapid pulmonary metastases, accurately recapitulating metastatic tumor dissemination in patients (3). Using this in vivo system, we engineered NIH 3T3 mouse fibroblasts, which offer the advantage of a genetically controlled system (as with the study of other oncoproteins), to express the CIC-DUX4 fusion oncoprotein (11–13). We observed rapid primary tumor formation at the site of implantation in 100% of the injected mice (Figure 1B). Mice with genetic silencing of *ETV4* showed decreased expression of its established target, MMP24 (9) (Figure 1C, and Supplemental Figure 1, B and C), and impaired metastatic efficiency in vivo and invasive capacity in vitro compared with mice bearing a control silencing vector (Figure 1, B–E). While ETV4 suppression decreased pulmonary metastases, it did not have a profound impact on tumor growth when CIC-DUX4-expressing cells were implanted either orthotopically or subcutaneously into the flank of immunocompromised mice (Figure 1, F and G). These findings suggest that the primary function of CIC-DUX4-mediated *ETV4* upregulation is to promote invasion and metastasis, but not tumor cell proliferation or tumor growth per se.

We next investigated transcriptional targets and programs that could regulate other key aspects of tumor biology beyond metastasis, including tumor cell proliferation and growth. Prior studies suggest-

ed that the CIC-DUX4 fusion can regulate cell cycle gene expression (6, 7). We established that ectopic expression of CIC-DUX4 increased NIH 3T3 cell growth in vitro and in vivo through enhanced cell cycle progression (Supplemental Figure 2, A–E). The CIC-DUX4 fusion increased the number of cells progressing through S-phase, as reflected by an increased  $G_2/M$  fraction compared with control (Supplemental Figure 2F). In order to identify CIC-DUX4-regulated cell cycle genes, we leveraged a publicly available microarray-based data set (GSE60740) to perform a comparative transcriptional analysis between CIC-DUX4-replete (IB120 EV cell line) and 2 independent CIC-DUX4-knockdown (KD) patient-derived cell lines (IB120 shCIC-DUX4a and IB120 shCIC-DUX4b) (14). Notably, in the context of the CIC-DUX4 fusion, CIC retains its binding specificity but acquires activating properties (5). Hence, we focused our attention on genes that were downregulated upon CIC-DUX4 KD. We observed 165 shared downregulated (and 105 upregulated) genes between the 2 independent shCIC-DUX4 data sets (IB120 shCIC-DUX4a and IB120 shCIC-DUX4b) (Figure 2A, and Supplemental Tables 1 and 2). Functional clustering of the 165 putative CIC-DUX4 target genes revealed enrichment for genes that regulate DNA replication and cell cycle machinery (Figure 2B) (15). We compared the expression of these 165 genes in 14 CIC-DUX4 and 7 EWS-NFATC2 patient-derived tumors that were profiled on the same Affymetrix array (GSE60740). We observed a



**Figure 2. CIC-DUX4 regulates cell cycle genes.** (A) Schematic algorithm to identify putative CIC-DUX4 target genes. (B) Functional annotation of CIC-DUX4-activated genes in IB120 cells. (C) Heatmap depicting the 165 CIC-DUX4-activated genes identified in A across 14 CIC-DUX4 and 7 EWSR1-NFATc2 patient-derived tumors. Cell cycle gene symbols are magnified.

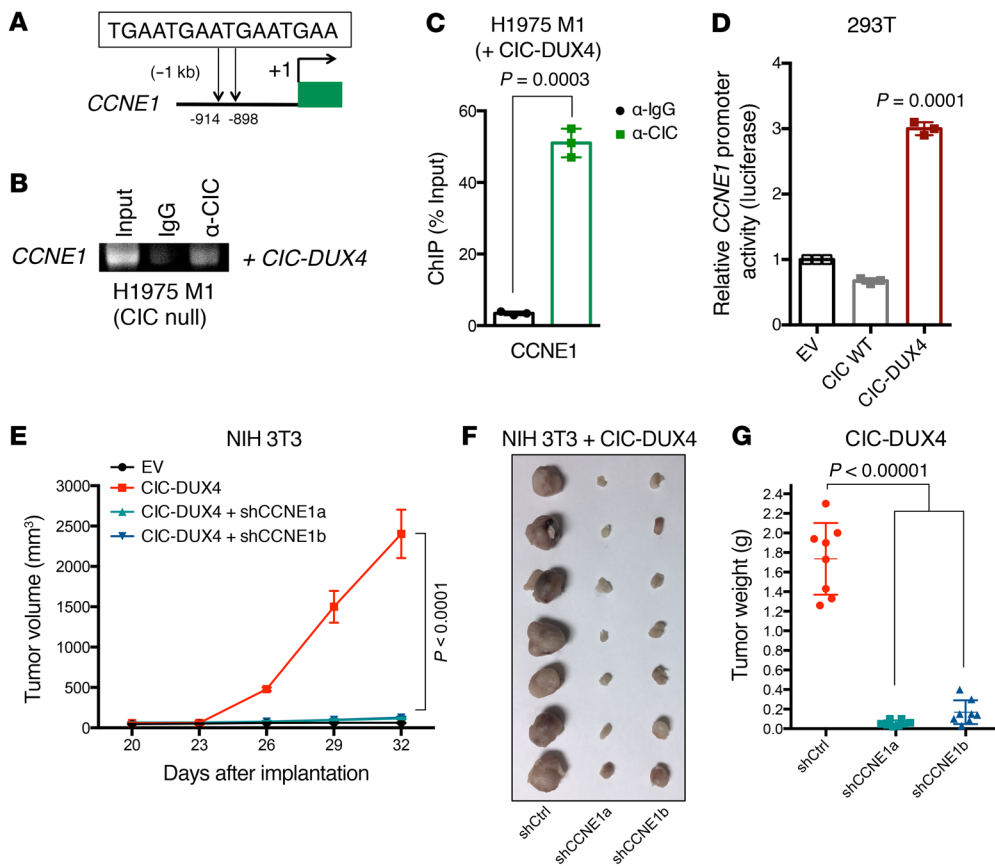
significant increase in the expression of multiple cell-cycle-regulatory genes in CIC-DUX4-expressing tumors (Figure 2C). Our findings extend recent studies (6, 7) and indicate that targeting the cell cycle in CIC-DUX4-expressing tumors is a potential therapeutic approach.

In order to identify direct transcriptional targets of the CIC-DUX4 fusion, we first surveyed the promoters (within -2 kb and +150 bp of the transcription start site) of all 165 putative response genes for the highly conserved CIC-binding motif (T[G/C]AATG[A/G]A) (5). Using this systematic approach, we identified 43 genes, including the known CIC target genes *ETV1*, -4, and -5 and multiple regulators of the cell cycle (Supplemental Table 3). Since ectopic expression of CIC-DUX4 promoted S-phase progression (Supplemental Figure 2, E and F), we focused on genes that directly regulate the G<sub>1</sub>/S transition. Of these genes, cyclin E1 (*CCNE1*) expression was consistently upregulated in CIC-DUX4-expressing tumors (Figure 2C). We therefore investigated whether CIC-DUX4 transcriptionally controls *CCNE1* expression. To explore this hypothesis, we first localized 2 tandem CIC-binding motifs within 1 kb of the *CCNE1* transcriptional start site (Figure 3A). We next performed ChIP-PCR analysis, which revealed *CCNE1* promoter occupancy by the CIC-DUX4 fusion (Figure 3, B and C). Additionally, a luciferase-based reporter assay demonstrated enhanced promoter activity by the ectopic expression of CIC-DUX4 compared with CIC WT and emp-

ty vector control (Figure 3D). These data show that *CCNE1* is a direct transcriptional target that is upregulated by CIC-DUX4.

To explore the functional role of *CCNE1* in CIC-DUX4-expressing tumors, we genetically silenced *CCNE1* in CIC-DUX4-expressing NIH 3T3 cells (a genetically controlled system) and observed decreased growth in vitro and in vivo compared with control (Figure 3, E-G, and Supplemental Figure 3, A-C). While there is no validated pharmacologic strategy to directly block *CCNE1*, inhibition of the *CCNE1* binding partner *CDK2* has therapeutic efficacy in other cancer types (16-18). We hypothesized that inhibiting *CDK2* in CIC-DUX4-expressing cells with the established small molecule drug dinaciclib (16) could limit tumor growth. To explore this hypothesis, we implanted CIC-DUX4-expressing NIH 3T3 cells subcutaneously into immunodeficient mice and treated them with low-dose dinaciclib (20 mg/kg/d); we observed decreased growth compared with vehicle control (Figure 4, A-C) (16). These findings suggest that pharmacologic inhibition of the *CCNE1*-*CDK2* complex is a potential therapeutic strategy in CIC-DUX4-expressing tumors.

There are few patient-derived models of CIC-DUX4-positive tumors. Nevertheless, in order to increase the clinical relevance of our findings we obtained rare, established patient-derived CIC-DUX4-expressing cells (NCC\_CDS1\_X1 and NCC\_CDS1\_X3) (19) to test the functional impact of *ETV4* KD and *CDK2* inhibition.



**Figure 3. CCNE1 is a molecular target of CIC-DUX4.** (A) Schematic of 2 CIC-binding motifs in the *CCNE1* promoter. (B and C) ChIP-PCR from H1975 M1 (CIC null) cells reconstituted with CIC-DUX4, showing CIC-DUX4 occupancy on the *CCNE1* promoter. *P* value, Student's *t* test. Error bars represent SEM. Performed in triplicate. (D) *CCNE1* luciferase promoter assay in 293T cells comparing EV with CIC WT or CIC-DUX4. *P* value, 1-way ANOVA. Error bars represent SEM. Performed in triplicate. (E) Subcutaneously implanted NIH 3T3 cells expressing EV, CIC-DUX4 (*n* = 7), or CIC-DUX4 plus shCCNE1a (*n* = 7) or shCCNE1b (*n* = 7). *P* value, 1-way ANOVA. Error bars represent SEM. (F) Tumor explants from mice in E. (G) Tumor weights from mice in E. *P* values, 1-way ANOVA.

We found that genetic silencing of *ETV4*, but not *ETV1* or *ETV5* (known CIC-DUX4 target genes) (5), decreased invasiveness but did not impact tumor growth or apoptosis (Supplemental Figure 4, A-G). Additionally, we found that CIC-DUX4-expressing cells were exquisitely sensitive to nanomolar (nM) concentrations of 2 established, independent CDK2 inhibitors, dinaciclib and SNS-032 (16, 20) (Figure 4D and Supplemental Figure 5A). The effects on tumor viability were largely mediated through apoptotic cell death as measured by poly(ADP-ribose) polymerase (PARP) cleavage and caspase activity, again indicating that CIC-DUX4-expressing tumors are dependent on the CCNE-CDK2 complex for survival (Figure 4, E-G, and Supplemental Figure 5, B and C). To mitigate the off-target effects of dinaciclib and SNS-032, we performed genetic silencing of CDK7 and CDK9, which did not suppress NCC\_CDS1\_X1 growth (Supplemental Figure 5, D-G).

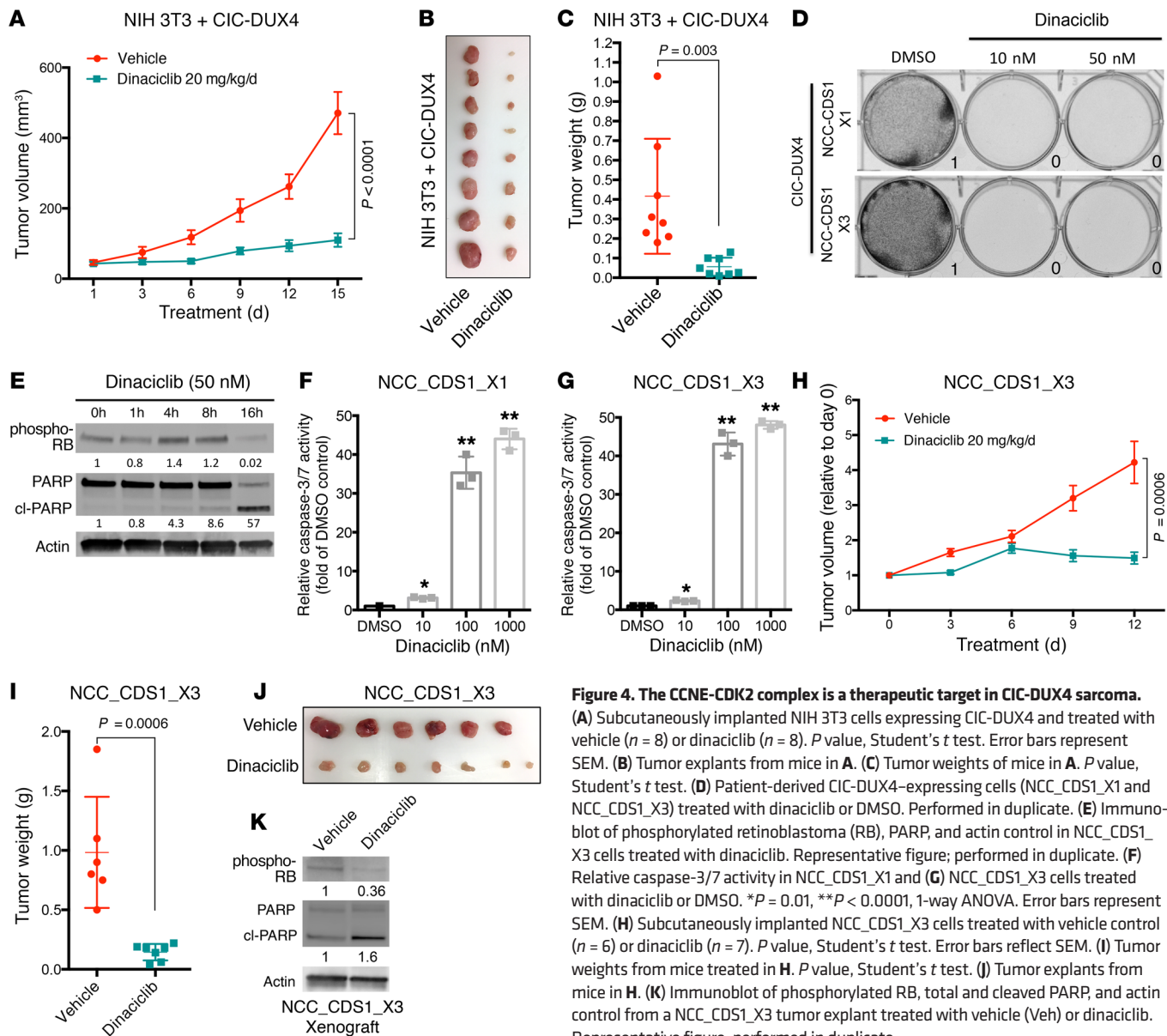
We next tested whether CDK2 inhibition could specifically suppress the growth of patient-derived CIC-DUX4-expressing cells in vivo. To address this hypothesis, we generated subcutaneous tumor xenografts of the patient-derived CIC-DUX4-expressing cells in immunodeficient mice and found that dinaciclib limited tumor growth compared with vehicle control treatment (Figure 4, H-J). The decreased tumor growth observed in dinaciclib-treated mice was accompanied by increased PARP cleavage in tumor explants, consistent with the apoptotic effect observed in vitro (Figure 4K). The impact on tumor growth following CDK2 inhibition was relatively specific for CIC-DUX4-expressing tumors, as we did not observe a significant response to dinaciclib in other sarcoma subtypes that harbor distinct transcription factor fusion oncoproteins (rhabdomyosarcoma or

Ewing sarcoma), in vitro or in vivo (Supplemental Figure 6, A and B). Consistent with these findings, we observed low levels of *CCNE1* in PAX3-FOXO1-positive RH30 rhabdomyosarcoma cells compared with CIC-DUX4-expressing NCC\_CDS1\_X1 cells (Supplemental Figure 6C). These findings further suggest that CIC-DUX4-expressing tumors are molecularly dependent on the CCNE-CDK2 complex, which is a potential therapeutic target.

To further demonstrate the therapeutic specificity of targeting CDK2, we used the clinical CDK4/6 inhibitor palbociclib, which has no substantial activity against CDK2, and found no impact on tumor growth or apoptosis in NCC\_CDS1\_X1 cells (Supplemental Figure 7A). Moreover, while genetic silencing of *CDK2* with siRNAs did not impact cell invasion, it did decrease cell number and viability in CIC-DUX4-expressing NCC\_CDS1\_X1 cells compared with control (Supplemental Figure 7, B-E). While we did not observe an effect on viability with *CCNE1* KD alone, combined *CCNE1* and *CCNE2* KD reduced viability to similar levels as siCDK2 cells (Supplemental Figure 7, C, D, F, and G). These findings are consistent with prior observations that both *CCNE1* and *CCNE2* converge on and activate CDK2, which our pharmacologic and genetic studies described above indicate is required for CIC-DUX4-expressing tumor survival (21).

Our data suggest that in human-derived NCC\_CDS1\_X1 cells, *CCNE2* can compensate for *CCNE1* loss. Consistent with this notion, we observed approximately 100-fold upregulation of *CCNE2* mRNA upon genetic silencing of *CCNE1* and through ChIP-Seq analysis identified CIC-DUX4 binding to the promoter of *CCNE1* but not *CCNE2* (Supplemental Figure 8, A-C) in human NCC\_CDS1\_X1 cells. In contrast to these findings, we did not observe a compensatory increase in *CCNE2* mRNA with *CCNE1* KD in NIH 3T3 (mouse) cells expressing CIC-DUX4 (Supplemental Figure 8, D and E). The lack of *CCNE2* upregulation in CIC-DUX4-expressing NIH 3T3 cells is a plausible explanation for the growth-suppressive effect of *CCNE1* KD in our initial NIH 3T3 system (Figure 3E), a phenotype that was





**Figure 4. The CCNE-CDK2 complex is a therapeutic target in CIC-DUX4 sarcoma.** (A) Subcutaneously implanted NIH 3T3 cells expressing CIC-DUX4 and treated with vehicle ( $n = 8$ ) or dinaciclib ( $n = 8$ ).  $P$  value, Student's  $t$  test. Error bars represent SEM. (B) Tumor explants from mice in A. (C) Tumor weights of mice in A.  $P$  value, Student's  $t$  test. (D) Patient-derived CIC-DUX4-expressing cells (NCC\_CDS1\_X1 and NCC\_CDS1\_X3) treated with dinaciclib or DMSO. Performed in duplicate. (E) Immunoblot of phosphorylated retinoblastoma (RB), PARP, and actin control in NCC\_CDS1\_X3 cells treated with dinaciclib. Representative figure; performed in duplicate. (F) Relative caspase-3/7 activity in NCC\_CDS1\_X1 and (G) NCC\_CDS1\_X3 cells treated with dinaciclib or DMSO.  $*P = 0.01$ ,  $**P < 0.0001$ , 1-way ANOVA. Error bars represent SEM. (H) Subcutaneously implanted NCC\_CDS1\_X3 cells treated with vehicle control ( $n = 6$ ) or dinaciclib ( $n = 7$ ).  $P$  value, Student's  $t$  test. Error bars reflect SEM. (I) Tumor weights from mice treated in H.  $P$  value, Student's  $t$  test. (J) Tumor explants from mice in H. (K) Immunoblot of phosphorylated RB, total and cleaved PARP, and actin control from a NCC\_CDS1\_X3 tumor explant treated with vehicle (Veh) or dinaciclib. Representative figure; performed in duplicate.

not shared with human-derived NCC\_CDS1 cells (Supplemental Figure 7, C and D). Importantly, we identified CIC-DUX4-binding sites in the promoter of mouse *CCNE1*, but not *CCNE2* (−4 kb to +100 of the transcription start site), suggesting that *CCNE1* (but not *CCNE2*) is a conserved transcriptional target in human (NCC\_CDS1) and mouse (NIH 3T3 expressing CIC-DUX4) cells (Supplemental Tables 3 and 4, and Supplemental Figure 8F). Our mechanistic dissection of CIC-DUX4-expressing tumors reveals a unique molecular and therapeutic dependence on the CCNE-CDK2 cell cycle complex. Our data show that this dependence can be exploited with clinical CDK2 inhibitors that limit tumor growth through apoptotic induction in a tumor type with few effective therapies.

The CIC-DUX4 fusion oncoprotein is a relatively understudied molecular entity that characterizes a rare but lethal subset of undifferentiated round cell sarcomas. We undertook a mechanistic dissection of the molecular function of the CIC-DUX4 fusion oncoprotein and report unique dependencies on specific transcriptional targets of the

fusion oncoprotein that promote either tumor growth or metastasis. We reveal that *ETV4* is a conserved target gene that enhances the metastatic capacity of CIC-DUX4-expressing tumors, without significantly impacting tumor growth. These findings extend previous data that demonstrate a prometastatic role for *ETV4* in certain human cancers (9, 22). Targeting an *ETV4*-mediated transcriptional program downstream of CIC-DUX4 can potentially limit tumor dissemination.

Further analysis of CIC-DUX4-bearing tumors coupled with our functional studies also identified a dependence on specific cell cycle machinery to enhance tumor growth but not invasive capacity. We reveal that the CCNE-CDK2 complex is a molecular target of the CIC-DUX4 oncoprotein that controls tumor growth and survival. While others have observed transcriptional upregulation of cell cycle genes in CIC-DUX4-expressing tumors (6, 7), our data establish the CCNE-CDK2 complex as a direct molecular target that can be pharmacologically exploited with clinically developed CDK2 inhibitors. The therapeutic impact may extend beyond the CIC-DUX4 fusion

to include other CIC-fused oncoproteins. Recent findings reveal a shared transcriptional program downstream of all known CIC fusions, including CIC-FOXO4 and CIC-NUTM1 (23). These findings suggest that many CIC-fused tumors retain their CIC-binding specificity while converting native CIC into a transcriptional activator instead of a repressor in a neomorphic manner. It would be compelling to explore whether the CCNE-CDK2 complex or other components of the cell cycle machinery drive tumor growth in these other CIC-fused tumor types, an area for future investigation.

Our mechanistic dissection of the downstream molecular targets of CIC-DUX4 provides therapeutically relevant insight for targeting ETV4-mediated metastasis and CCNE-CDK2-regulated tumor growth. Our data support a broader conceptual paradigm in which certain transcription factor fusion oncoproteins, such as CIC-DUX4, utilize neomorphic and distinct downstream regulatory programs to control divergent cancer hallmarks, such as proliferative capacity and metastatic competency. The data offer a potential mechanistic explanation for the pleiotropic functions of this important class of oncoproteins (i.e., transcription factor fusion oncoproteins such as CIC-DUX4).

Our findings highlight the clinical importance of molecular subclassification of morphologically similar tumor types, such as small round cell sarcomas. Identifying the different fusion oncoproteins present in clinical samples paves the way for oncoprotein fusion-specific therapeutic targeting to improve patient outcomes. Our study highlights the utility of elucidating the mechanistic features of tumors that are driven by transcription factor fusion oncoproteins to identify precision medicine-based molecular therapeutic strategies.

## Methods

Refer to Supplemental Methods for details.

**Statistics.** Experimental data are presented as mean ± SEM. *P* values derived for all in vitro experiments were calculated using 2-tailed Student's *t* test or 1-way ANOVA. *P* < 0.05 was considered statistically significant.

**Study approval.** For tumor xenograft studies, including orthotopic and subcutaneous models, specific pathogen-free conditions and facilities were approved by the American Association for Accreditation of Laboratory Animal Care. Surgical procedures were reviewed and approved by the UCSF IACUC, protocol AN107889-03A.

## Author contributions

RAO designed and performed the experiments, analyzed the data, and wrote the manuscript. RAO, VO, SN, YKL, RKP, RO, and TK performed in vitro and in vivo mouse studies. WW performed bioinformatics. TK performed experiments and provided cell lines. TGB directed the project, analyzed experiments, and wrote the manuscript.

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