

## Basic Study

## MiR-205 mediated APC regulation contributes to pancreatic cancer cell proliferation

Rui-Feng Qin, Jia Zhang, Hao-Ran Huo, Zeng-Jiang Yuan, Jia-Dong Xue

**ORCID number:** Rui-Feng Qin (0000-0001-6608-732X); Jia Zhang (0000-0002-5271-7906); Haoran Huo (0000-0002-7900-237X); Zeng-Jiang Yuan (0000-0003-3132-6719); Jia-Dong Xue (0000-0003-0916-4709).

**Author contributions:** Qin RF designed the research; Qin RF, Zhang J, Huo HR, Yuan ZJ, and Xue JD performed the research; Qin RF and Zhang J analyzed the data; Qin RF wrote the paper.

**Institutional review board statement:** The study was reviewed and approved by the Handan Central Hospital Institutional Review Board.

**Conflict-of-interest statement:** The authors declare no conflict of interest.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Unsolicited manuscript

**Received:** March 27, 2019

**Peer-review started:** March 27, 2019

**Rui-Feng Qin, Jia Zhang, Hao-Ran Huo, Zeng-Jiang Yuan, Jia-Dong Xue,** Third Department of General Surgery, Handan Central Hospital, Handan 056000, Hebei Province, China

**Corresponding author:** Rui-Feng Qin, MSc, Associate Chief Physician, Third Department of General Surgery, Handan Central Hospital, No. 15, Zhonghua South Street, Handan 056000, Hebei Province, China. [ruifengqin89@163.com](mailto:ruifengqin89@163.com)

**Telephone:** +86-133-8300-0998

**Fax:** +86-310-2110011

## Abstract

### BACKGROUND

Pancreatic cancer is a deadly malignancy with aggressive properties. MicroRNAs (miRNAs) participate in the pathogenesis of a variety of diseases and molecular processes by targeting functional mRNAs. Nevertheless, the regulatory role of miRNAs in signaling pathways involved in pancreatic cancer remains largely unknown.

### AIM

To explore the molecular regulation involved in pancreatic cancer and potential mechanisms of miR-205.

### METHODS

Microarray analysis was performed to investigate the expression profile of miRNAs in pancreatic cancer. Expression of miR-205 was validated by qRT-PCR. Target prediction and functional enrichment analysis were employed to seek potential target genes of miR-205 and potential functions of these genes. The target binding of miR-205 and adenomatous polyposis coli (APC) was validated by luciferase reporter assay. APC protein expression in pancreatic cancer was validated by qRT-PCR and Western blot. Proliferation was evaluated by MTT and colony formation assays.

### RESULTS

A large number of miRNAs with altered expression were identified in pancreatic cancer. MiR-205 was significantly up-regulated. APC was found to be a validated target of miR-205 and down-regulated in pancreatic cancer. Proliferation experiments showed that miR-205 could promote cell proliferation in pancreatic cancer by targeting APC.

### CONCLUSION

The above findings suggested that miR-205 mediated APC regulation contributes

**First decision:** May 16, 2019  
**Revised:** June 7, 2019  
**Accepted:** June 22, 2019  
**Article in press:** June 23, 2019  
**Published online:** July 28, 2019

**P-Reviewer:** Avci E, Bordonaro M, Mikulic D

**S-Editor:** Yan JP

**L-Editor:** Wang TQ

**E-Editor:** Zhang YL



to pancreatic cancer development, which could be considered as a novel prognostic biomarker for clinical care.

**Key words:** Pancreatic cancer; Microarray; MiR-205; Adenomatous polyposis coli; Proliferation

©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** A large number of microRNAs with altered expression were identified in pancreatic cancer. MiR-205 was found to be significantly up-regulated in pancreatic cancer. Adenomatous polyposis coli (APC) was found to be a validated target of miR-205 and down-regulated in pancreatic cancer. Proliferation experiments showed that miR-205 could promote cell proliferation in pancreatic cancer by targeting APC.

**Citation:** Qin RF, Zhang J, Huo HR, Yuan ZJ, Xue JD. MiR-205 mediated APC regulation contributes to pancreatic cancer cell proliferation. *World J Gastroenterol* 2019; 25(28): 3775-3786

**URL:** <https://www.wjgnet.com/1007-9327/full/v25/i28/3775.htm>

**DOI:** <https://dx.doi.org/10.3748/wjg.v25.i28.3775>

## INTRODUCTION

Pancreatic cancer is a severe malignancy with a five-year survival rate of less than 5% worldwide, due to its late detection, metastasis, and resistance to treatment such as chemotherapy and radiation therapy<sup>[1-3]</sup>. There are over 60 types of genetic mutations in pancreatic cancer according to previous studies, and these mutations have been related to several signaling pathways. Nevertheless, the regulation of these pathways in pancreatic cancer remains largely unknown<sup>[4-6]</sup>. Therefore, the exploration of molecular mechanisms involved in pancreatic cancer will be necessary and useful to improve its current therapy status.

In recent years, microRNAs (miRNAs) are emerging as important regulators in cancer development and play important roles in many physiological and pathological processes such as tumor growth, biosynthesis, immune regulation, and drug resistance<sup>[7-9]</sup>. MiR-205 has been found to play a role in various cancers<sup>[10,11]</sup>, but there has been no relevant report in pancreatic cancer. Adenomatous polyposis coli (APC) is a tumor suppressor protein that is involved in biological processes including transcriptional activation, apoptosis, cell migration, and cell adhesion<sup>[12,13]</sup>. Besides, APC has been found to have an important role in pancreatic cancer in a previous study<sup>[14]</sup>.

In the present study, microarray analysis was used to explore the differentially expressed miRNAs in pancreatic cancer. It was found that miR-205 was up-regulated in both pancreatic cancer tissue samples and cell lines, whereas APC was significantly down-regulated. Target prediction of miR-205 showed that there are potential targets that are involved in various signaling pathways. Furthermore, we confirmed that miR-205 could promote cell proliferation in pancreatic cancer by targeting APC *in vitro*. Taken together, our findings suggest that miR-205 might be used as a potential therapeutic target in human pancreatic cancer.

## MATERIALS AND METHODS

### **Study participants and cell lines**

Human pancreatic cancer tissue samples before treatment and adjacent healthy pancreatic tissue samples were obtained from Handan Central Hospital with patients' consent and were histologically verified. This study was approved by Handan Central Hospital and was performed in accordance with the Helsinki Declaration. HPDE6 (control) and SW1990/PANC-1/BXPC-3 cell lines were purchased from Tianjin Saier Biotechnology Company (Tianjin, China) and maintained in identical conditions with 10% fetal bovine serum (FBS) (Sigma, MO, United States).

### **Cell culture and transfection**

Cells were cultured in RPMI 1640 (Hyclone, CA, United States) supplemented with FBS, streptomycin, and penicillin in a humidified chamber at 37 °C. Transfection was performed using Lipofectamine 2000 reagent in accordance with the manufacturer's instructions. The transfection efficiency was assessed by detecting the fluorescence of red fluorescent protein expressed by the transfection vector.

### **Microarray and bioinformatics analysis**

The cancer samples and controls were analyzed through Agilent microRNA microarray (v14.0). First, the raw data presented as gProcessedSignal were normalized. The differential expression was filtered by fold change > 2 and *P*-value < 0.05, and all bioinformatics computations were performed using R script<sup>[15]</sup>. The prediction of miRNA targets was performed with miRDB<sup>[16]</sup> and targetScan<sup>[17]</sup>. Functional analysis was performed with KOBAS<sup>[18]</sup> software.

### **Luciferase reporter assay**

The 3'-untranslated region (UTR) of APC containing miR-205 binding sites was cloned into the pGL3-Basic luciferase vector (Promega, Madison, United States) to generate APC wild type (wt) and mutated to generate APC mutant (mut). After transfection, cells were incubated in suitable conditions for 24 h. The activity of luciferase was measured using the Luciferase Reporter Assay Kit (Promega, Madison, United States) in accordance with the manufacturer's instructions.

### **Quantitative real-time reverse-transcription PCR**

In accordance with the manufacturer's protocol, total RNA was isolated from each sample with Trizol (Invitrogen, CA, United States) and treated with DNase I (Invitrogen, CA, United States) to remove residual DNA. Then, RNA was reverse-transcribed into cDNA. MiRNAs were reverse-transcribed using the Mir-XTM miRNA Synthesis Kit (Clontech, Shanghai, China) according to the manufacturer's protocol. For miRNA targets, the primary transcript levels were determined with the SYBR Green Master. The sequence-specific primers for the target genes were retrieved from PrimerBank database. MiRNAs and mRNAs were normalized into U6 and GAPDH, respectively.

### **Western blot assay**

Pancreatic cancer cells were lysed in ice-cold lysis buffer using the protein-extraction reagent (Roche, Mannheim, Germany). To determine the concentration of total protein, BCA Protein Assay (ThermoFisher, Shanghai, China) was used. According to the manufacturer's protocol, Western blot analysis was performed with antibodies specifically against APC and GAPDH (Proteintech, China). The experiment was independently repeated three times.

### **MTT and colony formation assays**

After 24 h transfection of miR-205 mimic and inhibitor, cells were put into 96-well plates for 12 h at a density of  $3 \times 10^3$  viable cells/well. MTT assay was conducted on a spectrophotometer (Shimadzu, Columbia, SC, United States) at 490 nm to observe cell viability. For colony formation assay, the treated cells were plated and cultured in 6-well plates for 2 wk. After paraformaldehyde (Saier Biotechnology, Tianjin, China) was applied to these cells, the number of colonies was counted as cells were stained with crystal violet (Saier Biotechnology, Tianjin, China).

## **RESULTS**

### **Overview of patient subjects**

A total of 52 pancreatic cancer patients who received examination during their stay in Handan Central Hospital from July 2014 to November 2017 were collected. After ruling out other relevant diseases, 41 patients aged < 87 years were included in the experiment group, including 16 women and 25 men. The age, height, weight, and tumor stage were recorded as shown in [Table 1](#).

### **Expression profiling of miRNAs in pancreatic cancer tissues shows that miR-205 is up-regulated**

To explore the pancreatic cancer-related miRNAs, microarray analysis was performed on five cancer tissue samples and five healthy control samples. The data demonstrated that a list of miRNAs were differentially expressed in pancreatic cancer (*P* < 0.05, fold change  $\geq 2$ ) ([Figure 1A](#)), including 79 up-regulated and 128 down-regulated miRNAs. Furthermore, we sorted out top 20 differentially expressed miRNAs with relatively large differences, including miR-205-3p, miR-216a, miR-216b,

**Table 1 Basic characteristics of pancreatic cancer patients**

Variable	Patients
Age (yr)	57.21 ± 4.36
Height (cm)	165.08 ± 3.12
Weight (kg)	56.89 ± 6.94
Tumor stage (%)	
T1-T2	91 ( <i>n</i> = 37)
T3-T4	8 ( <i>n</i> = 4)

Note: The data above are expressed as the mean ± SD.

miR-148b, miR-30a, miR-6130, miR-155, miR-5704, miR-222, miR-221, miR-21, miR-631, miR-181a, miR-6754-3p, miR-6817-5p, miR-3928-5p, miR-6805-5p, miR-199a-3p, miR-4726-5p, and miR-6845-3p. The hierarchical clustering analysis was performed to demonstrate their expression patterns, as shown in [Figure 1B](#). Among these miRNAs, miR-205 expression level was significantly increased in pancreatic cancer and there has been no relevant report about its mechanisms of action.

### ***Tumor-promoting effect of miR-205 in pancreatic cancer cells in vitro***

To further verify the miR-205 expression level, qRT-PCR was performed on patient samples and three pancreatic cancer cell lines as described in the Methods section. MiR-205 showed increased expression in cancer samples compared with healthy tissues (*n* = 41 each; [Figure 2A](#)). Similarly, miR-205 also showed an increased expression level in three pancreatic cancer cell lines compared with HPDE6 cells ([Figure 2B](#)). MTT assay was performed in PANC-1 cells and control cells, and the results demonstrated that miR-205 overexpression in PANC-1 cells could notably increase the growth rate while an opposite result was observed in miR-205 down-regulated PANC-1 cells, as shown in [Figure 2C](#). We then performed colony formation assay to assess its potential impact on cell proliferation. The results showed that proliferation of PANC-1 cells was increased after transfection with miR-205 mimic, as shown in [Figure 2D](#). Collectively, these findings demonstrated that miR-205 may exert a tumor-promoting effect in pancreatic cancer.

### ***MiR-205 target prediction and functional analysis***

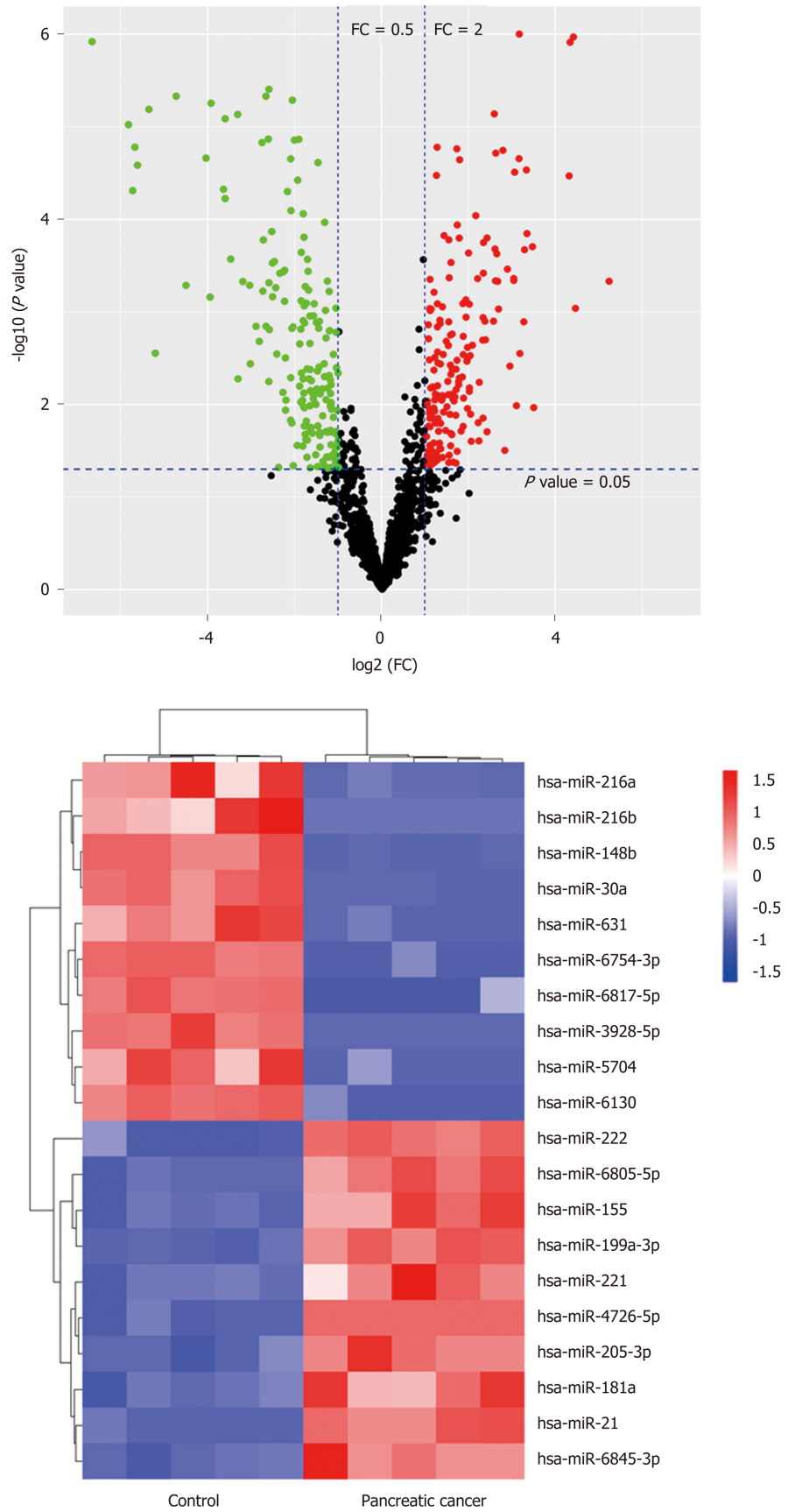
In order to investigate the molecular mechanism of miR-205, we performed target prediction and functional enrichment analysis of the targets using bioinformatics tools. A total of 168 targets were mapped to KOBAS database and subjected to GO/pathway analysis. According to the GO enrichment results, these genes participate in the regulation of cellular components, biological processes, and molecular functions. The bar chart shows that these genes are related to plasma membrane part, cell projection, and neuron part in the cellular components category, and are involved in signaling, cell communication, and developmental process in the biological processes category. Moreover, the molecular functions category showed that these genes mainly play a role in signaling and transmembrane receptor activity ([Figure 3](#)). According to the pathway analysis results, the target genes are related to cytokine-cytokine receptor interaction, regulation of IFNA signaling, and the TGF- $\beta$  signaling pathway ([Figure 4](#)). Furthermore, we noticed that APC is a potential target of miR-205, which has been discovered to play an important role in the development of pancreatic cancer.

### ***APC is down-regulated in pancreatic cancer***

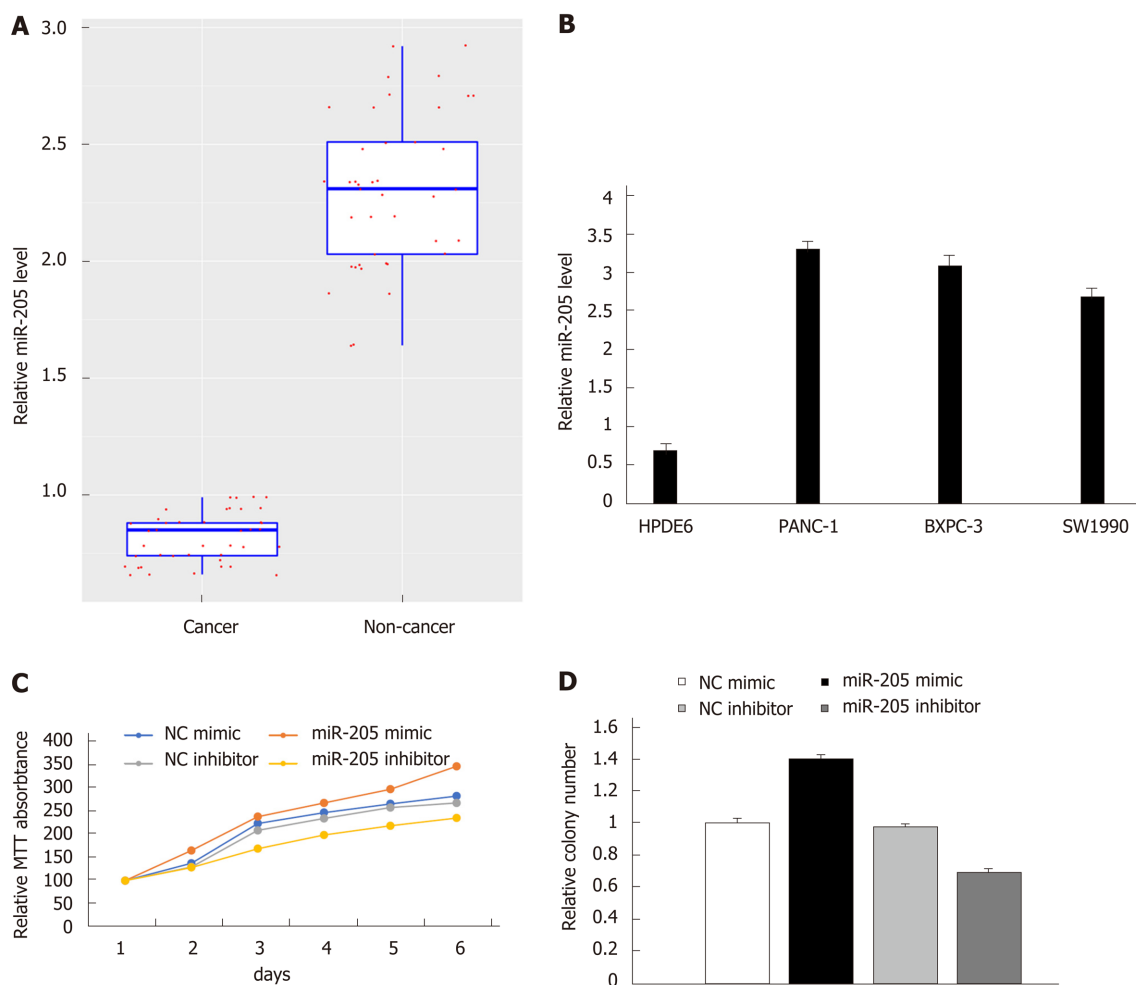
We evaluated the expression level of APC in HPDE6 and three pancreatic cancer cell lines by qRT-PCR assay. Lower expression of APC was observed in pancreatic cancer cell lines but not in HPDE6 cell line ([Figure 5A](#)). Meanwhile, we also evaluated the protein level of APC in these cell lines using Western blot analysis, and the results demonstrated that APC protein level was notably decreased in pancreatic cancer cell lines ([Figure 5B](#)). Since APC showed an opposite expression pattern with miR-205, it might be a potential target of miR-205.

### ***MiR-205 promotes cell proliferation in pancreatic cancer cells by targeting APC***

To confirm that APC is a target of miR-205, luciferase reporter assay was performed. The 3'-UTR of APC wild-type or mutant was cloned, as shown in [Figure 6A](#). The results indicated that miR-205 mimic could dramatically decrease the luciferase activity of wild-type vector, but had no effect on the mutant ([Figure 6B](#)) in PANC-1



**Figure 1** The miRNA expression profile in pancreatic cancer. A: The volcano plot was constructed using *P*-values and fold change values; B: The clustered heatmap showed the top 20 differentially expressed miRNAs between the control and pancreatic cancer groups. Red indicates the up-regulated expression and green/blue indicates the down-regulated expression.



**Figure 2** Detection of expression of miR-205 in pancreatic cancer tissues and cell lines by qRT-PCR. A: MiR-205 was up-regulated in pancreatic cancer tissues ( $n = 41$ ); B: MiR-205 was up-regulated in pancreatic cancer cell lines (PANC-1, BXPC-3, and SW1990) compared with normal human pancreatic duct epithelial cell line HPDE6 ( $P < 0.05$ ); C: MTT assay. Cell viability was significantly increased in cells with overexpression of miR-205 compared with control cells in a time-dependent manner; D: Colony formation assay. The effect of miR-205 overexpression on cell proliferation was evaluated.

cells. Furthermore, we found that APC mRNA level in miR-205 mimic treated PANC-1 was decreased while an opposite result was observed in miR-205 inhibitor treated cells (Figure 6C). Since APC protein was reported to participate in the development of pancreatic cancer, MTT assay was then carried out in APC overexpressing PANC-1 cells, and the results demonstrated that the promoting effect of miR-205 on pancreatic cancer cell proliferation was reduced (Figure 6D and E). These results indicated that miR-205 directly inhibits APC expression by binding to its 3'-UTR in pancreatic cancer cells. Moreover, miR-205 promotes the proliferation of pancreatic cancer cells by targeting APC.

## DISCUSSION

Pancreatic cancer, an aggressive malignancy with a five-year survival rate of less than 5%<sup>[2]</sup>, is a multi-genic disease that develops in a stepwise manner. The treatment of pancreatic cancer is often very difficult due to the impact of drug resistance and metastasis<sup>[1]</sup>. MiRNAs are a group of small, non-coding RNA molecules, which have been investigated as novel therapeutic targets in cancer<sup>[19]</sup>. The main mechanism for miRNAs to exert their function is down-regulating the expression or inducing degradation of target genes by binding to the 3'-UTR of target genes in mammals<sup>[8,17]</sup>.

Microarray assay is a convenient and efficient technique to explore the genome-wide gene expression<sup>[20]</sup>. In this study, 207 miRNAs were found to be abnormally expressed in pancreatic cancer by microarray analysis, including 79 up-regulated and 128 down-regulated miRNAs. These aberrantly expressed miRNAs may provide valuable resources for cancer research. Among these miRNAs, several miRNAs have

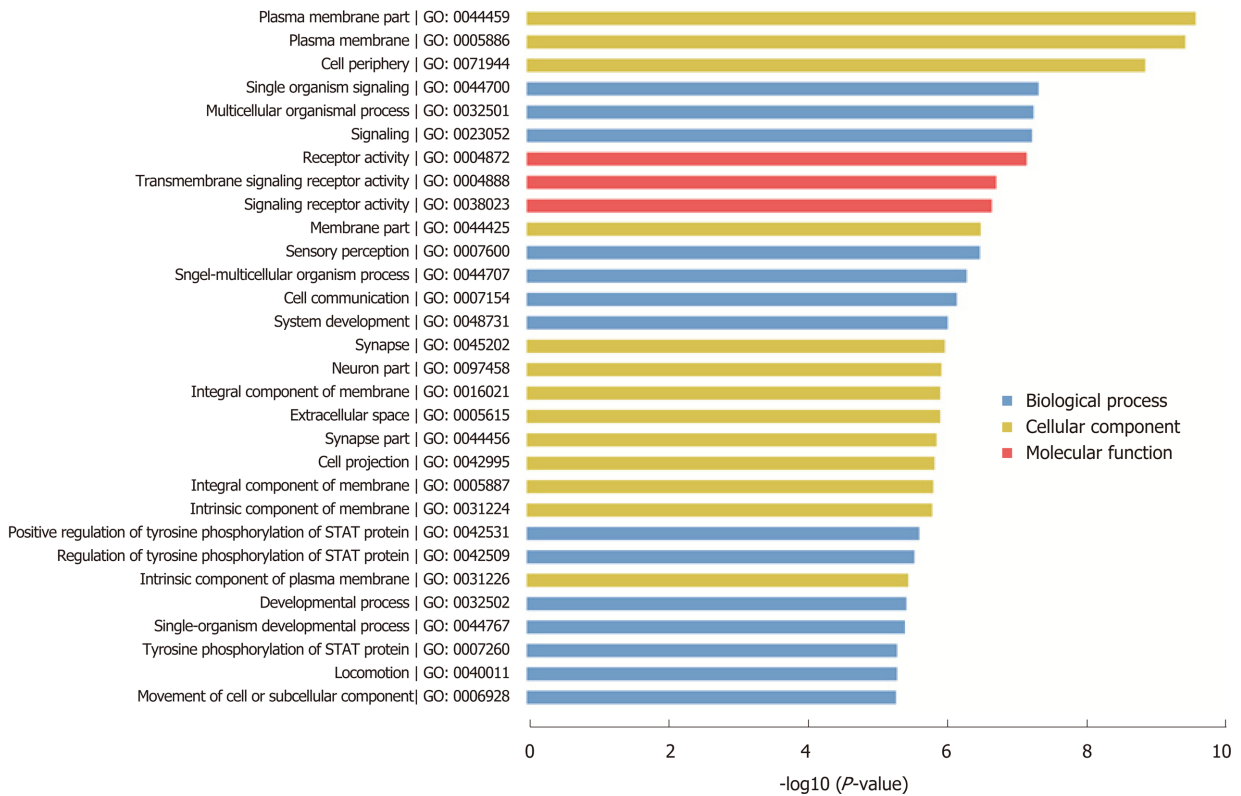


Figure 3 GO pathway analysis of the target genes of miR-205.

been studied in previous research. For instance, miR-216 was identified to be down-regulated in pancreatic cancer, which is consistent with previous result<sup>[21]</sup>, and miR-221 was found to promote proliferation and enhance cell cycle progression<sup>[22]</sup>. MiR-205 was found to play a role in various cancers, including breast cancer and prostate cancer<sup>[10,11]</sup>, but there has been no relevant report in pancreatic cancer. We found that miR-205 showed increased expression in pancreatic cancer and could promote cell proliferation *in vitro*. The target prediction of miR-205 and functional enrichment analysis of the targets identified the genes involved in various functions and pathways, including cytokine-cytokine receptor interaction, regulation of IFN $\alpha$  signaling, the Wnt signaling pathway, and the TGF- $\beta$  signaling pathway. Among them, the Wnt signaling pathway was demonstrated to regulate crucial aspects of cell fate determination, cell migration, and organogenesis. A previous study found that mutations cause Wnt pathway activation in human cancers<sup>[23]</sup>. As one of the miR-205 targets, APC acts as a tumor suppressor protein and participates in transcriptional activation, apoptosis, cell migration, and cell adhesion<sup>[12,13]</sup>. We confirmed that miR-205 promoted cell proliferation by targeting APC in pancreatic cancer.

In summary, our results revealed the expression profile of miRNAs in pancreatic cancer. The expression of miR-205 was up-regulated in pancreatic cancer whereas APC was significantly down-regulated. Target prediction of miR-205 and KOBAS enrichment analysis identified the genes involved in various functions and pathways. Furthermore, we confirmed that miR-205 could promote cell proliferation in pancreatic cancer by targeting APC. Our findings provide informative knowledge for the potential therapeutic use of miR-205 in human pancreatic cancer.

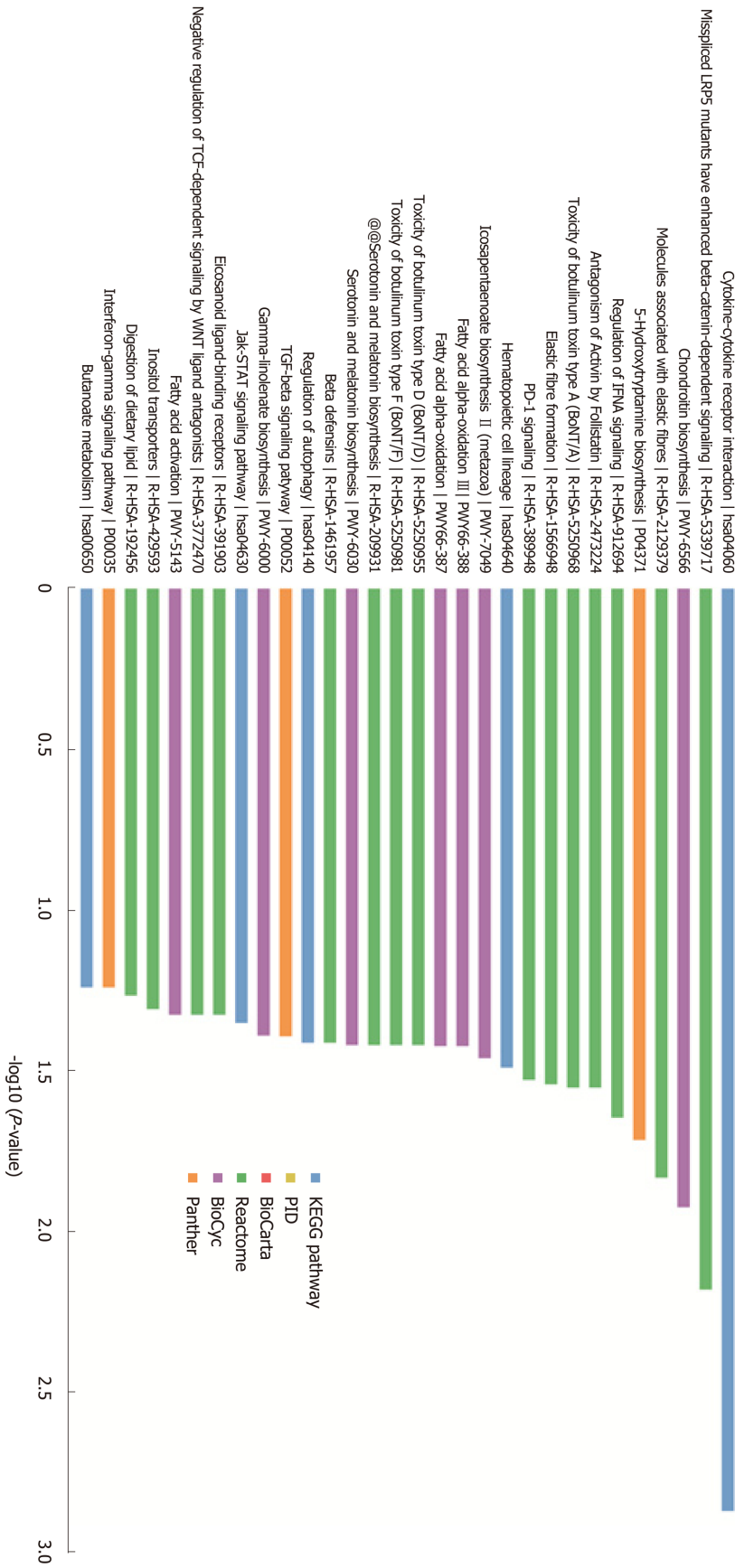
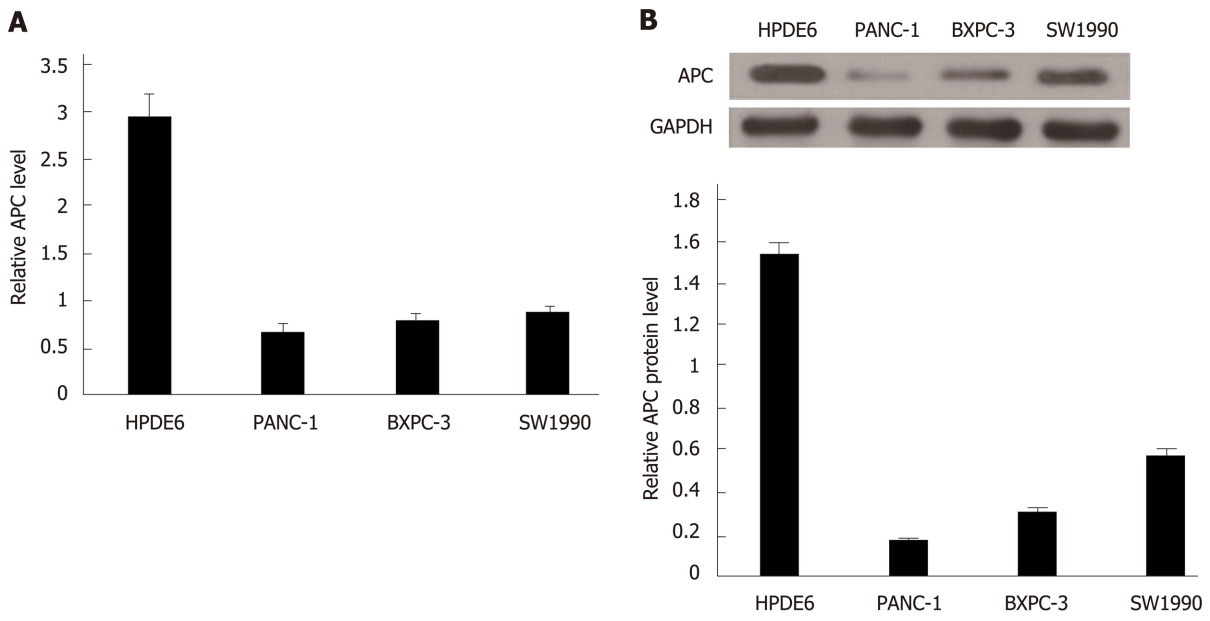
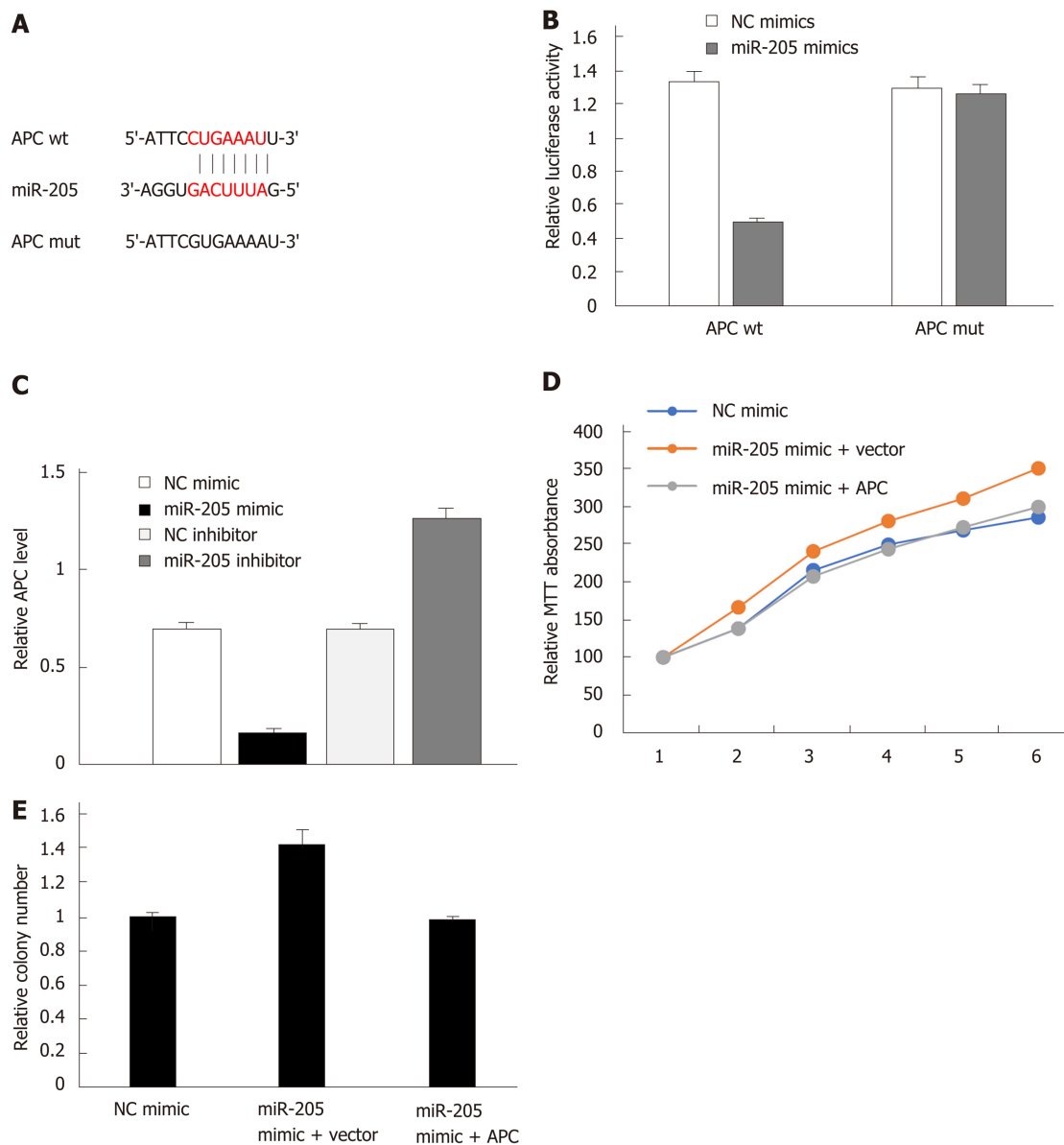


Figure 4 Pathway analysis of the target genes of miR-205.





**Figure 5** Expression of adenomatous polyposis coli in pancreatic cancer cell lines. A: mRNA level; B: Protein level,  $P < 0.05$ . APC: Adenomatous polyposis coli.



**Figure 6** MiR-205 promotes proliferation of pancreatic cancer cells by targeting adenomatous polyposis coli. A: Predicted binding sites of miR-205 in adenomatous polyposis coli (APC). The fragment of wild-type or mutant 3'-UTR of APC was cloned into the luciferase reporter vector; B: Luciferase activities measured at 48 h post transfection; C: APC mRNA expression was decreased in miR-205 mimic treated PANC-1 cells but increased in miR-205 inhibitor treated cells; D and E: The promoting effect of miR-205 on proliferation was reduced in pancreatic cancer cells overexpressing APC, as measured by MTT and colony formation assays. APC: Adenomatous polyposis coli; wt: Wild-type; mut: Mutant.

## ARTICLE HIGHLIGHTS

### Research background

Pancreatic cancer is known as a deadly malignancy in the world, and a sufficient treatment for the disease has not been found yet. We aimed to explore the regulatory mechanisms of microRNAs (miRNAs) in pancreatic cancer.

### Research motivation

As a group of non-coding RNAs, miRNAs were found to play important roles in human disease. In this study, we aimed to explore the miRNAs involved in pancreatic cancer and potential regulatory mechanisms of miR-205, which may contribute to pancreatic cancer treatment.

### Research objectives

MiRNA expression pattern in pancreatic cancer and potential regulatory mechanisms of miR-205 and adenomatous polyposis coli (APC) were analyzed. The findings may contribute to clinical care of pancreatic cancer.

### Research methods

Microarray analysis was used to explore the genome-wide miRNA expression profile in pancreatic cancer. QRT-PCR and Western blot were performed to validate gene expression. Bioinformatics analysis was performed to predict target genes and their potential functions. Dual luciferase reporter assay was used to validate the binding of miR-205 and APC. Proliferation was evaluated by MTT and colony formation assays.

### Research results

A large number of differentially expressed miRNAs were identified in pancreatic cancer. MiR-205 was significantly up-regulated while APC was down-regulated in pancreatic cancer. Dual luciferase reporter assay showed that APC is a validated target of miR-205. Moreover, miR-205 could promote cell proliferation in pancreatic cancer by targeting APC.

### Research conclusions

This study, for the first time, revealed that miR-205 mediated APC regulation contributed to pancreatic cancer development. Microarray analysis was used to fully disclose the genome-wide miRNAs expression profile in pancreatic cancer. Proliferation experiment showed that miR-205 could promote cell proliferation in pancreatic cancer cells by targeting APC, which could be considered as novel prognostic biomarkers for future clinical care.

### Research perspectives

The aberrantly expressed miRNAs identified in pancreatic cancer may provide valuable resources for future cancer research.

## REFERENCES

- 1 Falasca M, Kim M, Casari I. Pancreatic cancer: Current research and future directions. *Biochim Biophys Acta* 2016; **1865**: 123-132 [PMID: 26794394 DOI: 10.1016/j.bbcan.2016.01.001]
- 2 Garrido-Laguna I, Hidalgo M. Pancreatic cancer: From state-of-the-art treatments to promising novel therapies. *Nat Rev Clin Oncol* 2015; **12**: 319-334 [PMID: 25824606 DOI: 10.1038/nrclinonc.2015.53]
- 3 Kanji ZS, Gallinger S. Diagnosis and management of pancreatic cancer. *CMAJ* 2013; **185**: 1219-1226 [PMID: 23610017 DOI: 10.1503/cmaj.121368]
- 4 Roe JS, Hwang CI, Somerville TDD, Milazzo JP, Lee EJ, Da Silva B, Maiorino L, Tiriach H, Young CM, Miyabayashi K, Filippini D, Creighton B, Burkhart RA, Buscaglia JM, Kim EJ, Grem JL, Lazenby AJ, Grunkemeyer JA, Hollingsworth MA, Grandgenett PM, Egeblad M, Park Y, Tuveson DA, Vakoc CR. Enhancer Reprogramming Promotes Pancreatic Cancer Metastasis. *Cell* 2017; **170**: 875-888.e20 [PMID: 28757253 DOI: 10.1016/j.cell.2017.07.007]
- 5 Shukla SK, Purohit V, Mehla K, Gunda V, Chaika NV, Vernucci E, King RJ, Abrego J, Goode GD, Dasgupta A, Illies AL, Gebregiworgis T, Dai B, Augustine JJ, Murthy D, Attri KS, Mashadova O, Grandgenett PM, Powers R, Ly QP, Lazenby AJ, Grem JL, Yu F, Matés JM, Asara JM, Kim JW, Hankins JH, Weekes C, Hollingsworth MA, Serkova NJ, Sasson AR, Fleming JB, Oliveto JM, Lyssiotis CA, Cantley LC, Berim L, Singh PK. MUC1 and HIF-1 $\alpha$  Signaling Crosstalk Induces Anabolic Glucose Metabolism to Impart Gemcitabine Resistance to Pancreatic Cancer. *Cancer Cell* 2017; **32**: 392 [PMID: 28898700 DOI: 10.1016/j.ccell.2017.08.008]
- 6 Gaianigo N, Melisi D, Carbone C. EMT and Treatment Resistance in Pancreatic Cancer. *Cancers (Basel)* 2017; **9**: pii: E122 [PMID: 28895920 DOI: 10.3390/cancers9090122]
- 7 Rupaimoole R, Slack FJ. MicroRNA therapeutics: Towards a new era for the management of cancer and other diseases. *Nat Rev Drug Discov* 2017; **16**: 203-222 [PMID: 28209991 DOI: 10.1038/nrd.2016.246]
- 8 Thyagarajan A, Shaban A, Sahu RP. MicroRNA-Directed Cancer Therapies: Implications in Melanoma Intervention. *J Pharmacol Exp Ther* 2018; **364**: 1-12 [PMID: 29054858 DOI: 10.1124/jpet.117.242636]
- 9 Varshney A, Panda JJ, Singh AK, Yadav N, Bihari C, Biswas S, Sarin SK, Chauhan VS. Targeted delivery of microRNA-199a-3p using self-assembled dipeptide nanoparticles efficiently reduces hepatocellular carcinoma in mice. *Hepatology* 2018; **67**: 1392-1407 [PMID: 29108133 DOI: 10.1002/hep.29643]
- 10 Du YE, Tu G, Yang G, Li G, Yang D, Lang L, Xi L, Sun K, Chen Y, Shu K, Liao H, Liu M, Hou Y. MiR-205/YAP1 in Activated Fibroblasts of Breast Tumor Promotes VEGF-independent Angiogenesis through STAT3 Signaling. *Theanostics* 2017; **7**: 3972-3988 [PMID: 29109792 DOI: 10.7150/thno.18990]
- 11 Bhatnagar N, Li X, Padi SK, Zhang Q, Tang MS, Guo B. Downregulation of miR-205 and miR-31 confers resistance to chemotherapy-induced apoptosis in prostate cancer cells. *Cell Death Dis* 2010; **1**: e105 [PMID: 21368878 DOI: 10.1038/cddis.2010.85]
- 12 Lamlum H, Ilyas M, Rowan A, Clark S, Johnson V, Bell J, Frayling I, Efstathiou J, Pack K, Payne S, Roylance R, Gorman P, Sheer D, Neale K, Phillips R, Talbot I, Bodmer W, Tomlinson I. The type of somatic mutation at APC in familial adenomatous polyposis is determined by the site of the germline mutation: A new facet to Knudson's 'two-hit' hypothesis. *Nat Med* 1999; **5**: 1071-1075 [PMID: 10470088 DOI: 10.1038/125111]
- 13 Yamaguchi K, Nagayama S, Shimizu E, Komura M, Yamaguchi R, Shibuya T, Arai M, Hatakeyama S, Ikenoue T, Ueno M, Miyano S, Imoto S, Furukawa Y. Reduced expression of APC-1B but not APC-1A by the deletion of promoter 1B is responsible for familial adenomatous polyposis. *Sci Rep* 2016; **6**: 26011 [PMID: 27217144 DOI: 10.1038/srep26011]
- 14 Grant RC, Selander I, Connor AA, Selvarajah S, Borgida A, Briollais L, Petersen GM, Lerner-Ellis J, Holter S, Gallinger S. Prevalence of germline mutations in cancer predisposition genes in patients with pancreatic cancer. *Gastroenterology* 2015; **148**: 556-564 [PMID: 25479140 DOI: 10.1053/j.gastro.2014.11.042]
- 15 Dai Z, Sheridan JM, Gearing LJ, Moore DL, Su S, Wormald S, Wilcox S, O'Connor L, Dickens RA, Blewitt ME, Ritchie ME. edgeR: A versatile tool for the analysis of shRNA-seq and CRISPR-Cas9 genetic screens. *F1000Res* 2014; **3**: 95 [PMID: 24860646 DOI: 10.12688/f1000research.3928.2]
- 16 Wong N, Wang X. miRDB: an online resource for microRNA target prediction and functional annotations. *Nucleic Acids Res* 2015; **43**: D146-D152 [PMID: 25378301 DOI: 10.1093/nar/gku1104]

- 17 **Agarwal V**, Bell GW, Nam JW, Bartel DP. Predicting effective microRNA target sites in mammalian mRNAs. *Elife* 2015; 4 [PMID: 26267216 DOI: 10.7554/eLife.05005]
- 18 **Xie C**, Mao X, Huang J, Ding Y, Wu J, Dong S, Kong L, Gao G, Li CY, Wei L. KOBAS 2.0: A web server for annotation and identification of enriched pathways and diseases. *Nucleic Acids Res* 2011; 39: W316-W322 [PMID: 21715386 DOI: 10.1093/nar/gkr483]
- 19 **Catela Ivkovic T**, Voss G, Cornella H, Ceder Y. microRNAs as cancer therapeutics: A step closer to clinical application. *Cancer Lett* 2017; 407: 113-122 [PMID: 28412239 DOI: 10.1016/j.canlet.2017.04.007]
- 20 **Daimiel L**, Ordoñas JM, Dávalos A. miRNAs modified by dietary lipids in Caco-2 cells. A microarray screening. *Genom Data* 2015; 5: 171-172 [PMID: 26484250 DOI: 10.1016/j.gdata.2015.05.011]
- 21 **Wang S**, Chen X, Tang M. MicroRNA-216a inhibits pancreatic cancer by directly targeting Janus kinase 2. *Oncol Rep* 2014; 32: 2824-2830 [PMID: 25220761 DOI: 10.3892/or.2014.3478]
- 22 **Sarkar S**, Dubaybo H, Ali S, Goncalves P, Kollepara SL, Sethi S, Philip PA, Li Y. Down-regulation of miR-221 inhibits proliferation of pancreatic cancer cells through up-regulation of PTEN, p27(kip1), p57(kip2), and PUMA. *Am J Cancer Res* 2013; 3: 465-477 [PMID: 24224124 DOI: 10.1016/S1098-3015(10)70951-7]
- 23 **Saito-Diaz K**, Benchabane H, Tiwari A, Tian A, Li B, Thompson JJ, Hyde AS, Sawyer LM, Jodoin JN, Santos E, Lee LA, Coffey RJ, Beauchamp RD, Williams CS, Kenworthy AK, Robbins DJ, Ahmed Y, Lee E. APC Inhibits Ligand-Independent Wnt Signaling by the Clathrin Endocytic Pathway. *Dev Cell* 2018; 44: 566-581.e8 [PMID: 29533772 DOI: 10.1016/j.devcel.2018.02.013]



Published By Baishideng Publishing Group Inc  
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA  
Telephone: +1-925-2238242  
Fax: +1-925-2238243  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>

