

Ventral Pallidum Output Pathways in Context-Induced Reinstatement of Alcohol Seeking

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Ventral pallidum (VP) is a well-established locus for the reinforcing effects of drugs of abuse and reinstatement of drug seeking. However, VP neurons are at the origin of multiple output pathways, with strong projections to ventral tegmental area (VTA), subthalamic nucleus (STN), lateral hypothalamus, among others, and the roles of these VP output pathways in reinstatement of drug seeking remain poorly understood. Here we addressed these issues using a combination of neuroanatomical tracing and chemogenetic approaches. First, using dual-retrograde tracing, we show that VP neurons projecting to either VTA or STN are recruited during context-induced reinstatement of extinguished alcohol seeking in rats. Then, using chemogenetics, we show modulation of context-induced reinstatement and reacquisition of alcohol seeking via designer receptors exclusively activated by designer drugs excitation or inhibition of the VP. To determine the causal roles of VP → VTA and VP → STN pathways in context-induced reinstatement and reacquisition we used a chemogenetic disconnection approach and show that silencing either the VP → VTA or VP → STN pathways is sufficient to reduce both reinstatement and reacquisition of alcohol seeking. Moreover, these disconnections also each reduced responding and motivation during a progressive ratio test but had no effect on locomotor activity. Together, these results show that multiple ventral pallidal output pathways contribute to relapse to alcohol seeking.

Key words: DREADD; reinstatement; relapse; ventral pallidum

Significance Statement

Ventral pallidum (VP) serves important roles in reward and motivation and is a critical node in the neural circuitry for reinstatement of drug seeking. Despite being a common locus for different forms of reinstatement, fundamental aspects of neural circuitry for these VP contributions to reinstatement of drug seeking remain unknown. Here we used a combination of neuroanatomical tracing and chemogenetic approaches to map the VP output pathways for context-induced reinstatement and reacquisition of alcohol seeking. We show that VP output pathways to the subthalamic nucleus and also to the ventral tegmental area are necessary for these forms of reinstatement.

Introduction

The ventral pallidum (VP), a major basal ganglia output structure (Heimer et al., 1982; Zahm et al., 1987, 1996; Zahm and Heimer, 1990), serves a critical role in reward and motivation (Smith et al., 2009; Root et al., 2015; Richard et al., 2016). VP is a well-established locus for the reinforcing effects of drugs of abuse and reinstatement of drug seeking. For example, microinjections of mu opioid receptor antagonists (Tang et al., 2005; Perry and

McNally, 2013), pharmacological inactivation (McFarland and Kalivas, 2001; McFarland et al., 2004), or chemogenetic silencing (Mahler et al., 2014) of VP prevent a variety of forms of reinstatement to drug seeking. Multiple striatal, cortical, and amygdala inputs converge on VP during reinstatement (Perry and McNally, 2013), with projections from nucleus accumbens core (AcbC) especially important. Pharmacological disconnection of dorsal medial prefrontal cortex and VP (McFarland and Kalivas, 2001) or optogenetic silencing of the AcbC → VP pathway prevents reinstatement of cocaine or reacquisition of alcohol seeking (Stefanik et al., 2013; Khoo et al., 2015). These findings support VP as a key component of a cortical-striatal-pallidal pathway for reinstatement of drug seeking (Kalivas and Volkow, 2005, 2011).

However, fundamental aspects of VP contributions to reinstatement of drug seeking remain poorly understood. Of relevance here, little is known regarding the critical VP outputs for reinstatement. Mahler et al. (2014) identified a role for the VP →

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ventral tegmental area (VTA) projection in reinstatement of cocaine seeking in rats. VP neurons projecting to VTA (ventromedial VP or “rostral” VP) were recruited during cue-induced reinstatement of cocaine seeking and chemogenetic silencing of VP terminals in VTA or disconnection of VP from VTA dopamine neurons prevented this reinstatement. However, the role of the VP → VTA pathway in reinstatement to seeking other drugs is unknown. It is also unknown whether this role is unique to the VP → VTA pathway or whether it is shared with VP efferents to thalamus, hypothalamus, and midbrain. For example, ventromedial VP projects to VTA but also lateral hypothalamus (Zahm, 1989; Groenewegen et al., 1993; Zahm et al., 1996; Tripathi et al., 2013), a region strongly implicated in reinstatement of drug seeking (Marchant et al., 2009, 2012, 2014), whereas dorsolateral VP projects extensively to subthalamic nucleus (STN) and substantia nigra (Zahm, 1989; Groenewegen et al., 1993; Zahm et al., 1996; Tripathi et al., 2013). VP projections to STN are of considerable interest because STN manipulations reduce motivation for cocaine as well as alcohol (Baunez et al., 2005; Lardeux and Baunez, 2008; Rouaud et al., 2010) and reduce reinstatement of methamphetamine seeking (Baracz et al., 2015), but it is unknown whether this role for STN in motivation and reinstatement of drug seeking is linked to its VP inputs.

Here we addressed these issues using a combination of neuro-anatomical tracing and chemogenetic approaches. First, using dual-retrograde tracing, we asked whether VP neurons projecting to VTA or STN were recruited during context-induced reinstatement (ABA renewal) (Crombag and Shaham, 2002; Hamlin et al., 2007; Bouton and Todd, 2014) of alcohol seeking in rats. Then, using chemogenetics, we asked whether modulation of VP or VP → VTA and VP → STN pathways affected context-induced reinstatement and reacquisition of alcohol seeking.

Materials and Methods

Subjects. Subjects were experimentally naive male Sprague Dawley rats (300–350 g), obtained from the Animal Resources Centre. Rats were housed in groups of four in ventilated racks maintained in a climate-controlled colony room. The colony room was maintained on a 12 h light/dark cycle (lights on 7:00 A.M.). Experiments were conducted during the light cycle. Food and water were available *ad libitum* until 2 d before behavioral training, after which rats were allowed 1 h access to food and water following daily training sessions. The University of New South Wales Animal Care and Ethics Committee approved the procedures.

Apparatus. Training, extinction, and test were conducted in standard operant chambers (Med Associates), enclosed in sound- and light-attenuating cabinets equipped with fans providing constant ventilation and low-level background noise. Inside each chamber were two nose-pokes containing a white cue-light and a recessed magazine. Responding on one (active) nosepoke delivered 4% (v/v) decarbonated alcoholic beer to the magazine, whereas responding on the other (inactive) nosepoke had no consequences. For experiments that involved manipulation of context, these chambers were arranged in two sets of four that differed in their visual (illuminated by an overhead LED vs dark), tactile (Perspex vs grid floors), and olfactory (rose oil vs peppermint essence) properties, as described previously. The two sets of chambers were fully counterbalanced to serve as contexts A and B.

Locomotor activity was assessed in Plexiglas chambers (Med Associates) 43.2 cm (width) × 43.2 cm (length) × 30.5 cm (height) for 40 min. Movement was tracked through the use of three 16 beam infrared arrays. Infrared beams were located on both the *x*- and *y*-axes for positional tracking.

Viral vectors and retrograde tracers. Adeno-associated viral vectors encoding the hM3Dq or hM4Di designer receptors exclusively activated by designer drugs (DREADDs) (Urban and Roth, 2015; Roth, 2016) or eYFP were obtained from University of North Carolina Vector Core (Chapel

Hill, NC). The vectors used in these experiments were AAV5-hSyn-HA-hM3Dq-IRES-mCitrine (2×10^{12} vp/ml), AAV5-hSyn-HA-hM4Di-IRES-mCitrine (2×10^{12} vp/ml), and AAV5-hSyn-eYFP (4×10^{12} vp/ml). The retrograde tracers were cholera toxin b subunit (CTb) conjugated to AlexaFluor-488 (CTb-488) or AlexaFluor-555 (CTb-555) (ThermoFisher Scientific). CNO was obtained from the RTI International. CNO was dissolved in 5% DMSO (Sigma) and 0.9% sterile saline solution.

Surgery. Before surgery, rats received an intraperitoneal injection of a mixture of 100 mg/ml ketamine (Ketapex, Apex Laboratories) and 0.3 ml/kg xylazine (Rompun; Bayer) and an injection of the anti-inflammatory carprofen (Rimadyl, 3 mg/kg). Once anesthetized, rats were placed in a stereotaxic apparatus (model 942, Kopf) and shaved to expose the skin surface of the head. Before incision, rats received a subcutaneous injection of carprofen (5 mg/kg) and an injection of 0.5% bupivacaine (Cenvet) just under the surface of incision site. Following incision, a hand drill was used to make craniotomies above the target region and a 5 μ l, 30-gauge conical tipped microinfusion syringe (Hamilton) was used to infuse 0.75 μ l of AAV vectors into VP (−0.12 AP, \pm 2.5 ML, −8.4 DV), 0.50 μ l into STN (−3.6 AP, \pm 2.5 ML, −8.8 DV), and 0.75 μ l into VTA (−5.6 AP, \pm 2.25 [10-degree angle] ML, −8.3 DV) over a 3 min period at a rate of 0.25 μ l/min (UMP3 with SYS4 Microcontroller; World Precision Instruments). The syringe was left in place for 5–7 min to permit diffusion of the injected vectors. CTb-488 and CTb-555 (0.1 μ l) were infused unilaterally into the VTA (−5.6 AP, \pm 2.25 [10-degree angle] ML, −8.3 DV) and contralateral STN (−3.6 AP, \pm 2.5 ML, −8.8 DV) (counterbalanced across hemispheres) using a 30 gauge conical tipped microinfusion syringe at a rate of 0.5 μ l/min (UMP3 with SYS4 Microcontroller; World Precision Instruments). All stereotaxic coordinates are distances in millimeters from bregma (Paxinos and Watson, 2007). Bone wax (Coherent Scientific) was used to seal the opening of the skull. After surgery, rats were injected intraperitoneally with 0.3 ml of 300 mg/ml solution of procaine penicillin (Benicillin; Illium) and subcutaneously with 0.3 ml of a 100 mg/ml solution of cephazolin (Hospira). Daily postoperative and recovery procedures, including weight and infection management, were conducted for the remainder of the experiment. All behavioral procedures commenced a minimum of 2 weeks after surgery.

Behavioral testing procedures. On the first 2 d, the animals received 20 min magazine training sessions in both chambers. During these sessions, there were 10 noncontingent deliveries of 0.6 ml of the reward (4% alcohol v/v decarbonated beer) at time intervals variable around a mean of 1.2 min. On the next 10 days, rats received self-administration training, in context A for 1 h per day. During this time, responding on the active nosepoke extinguished the nosepoke cue light and triggered delivery of 0.6 ml of beer to the magazine on an FR-1 schedule followed by a 24 s timeout. Responses on the inactive nosepoke were recorded but had no programmable consequences. On the next 4 d, rats received extinction training, in context B for 1 h per day. During this training, responses on the active nosepoke extinguished the cue light and triggered the pump, but no beer was delivered. All rats were tested starting 24 h after the last extinction session. A minimum of 24 h separated each test. These parameters were chosen based on our past research, which shows robust acquisition, extinction, and renewal of responding under these conditions (Hamlin et al., 2006, 2007, 2008).

Experiment 1: VP output pathways recruited during context-induced reinstatement of alcohol seeking. To assess recruitment of the VP → VTA and VP → STN pathways during context-induced reinstatement, we applied the retrograde tracers CTb-488 and CTb-555 (counterbalanced) to one VTA and to the contralateral STN. There were three groups: Group ABB ($n = 8$), Group ABA ($n = 8$), and Group AB0 ($n = 8$). All rats were trained to respond for alcoholic beer in context A and extinguished in context B. Rats were tested once, for 1 h, 24 h after the last extinction session. Group ABB was tested in the extinction context. Group ABA was tested in the training context. Group AB0 was not tested; instead, they were simply transported to the laboratory on test and received an equivalent period of handling. This group served as a control for the behavioral and pharmacological training histories. All rats were perfused 1 h after

test (or at the equivalent time for Group AB0). Their brains were later processed for expression of the c-Fos protein and the CTb tracers.

Experiment 2: VP chemogenetic manipulation on context-induced reinstatement and reacquisition of alcohol seeking. In Experiment 2a, there were two groups, eYFP ($n = 7$) and hM4Di ($n = 7$), which refer to the AAV vector expressed in the VP. In Experiment 2b, there were two groups, eYFP ($n = 7$) and hM3Dq ($n = 6$), which refer to the AAV vector expressed in the VP. These experiments were run at different times. In both experiments, rats were trained and extinguished as described above. In both experiments, testing commenced 24 h after extinction. Rats were tested for 1 h in the extinction context (ABB) and for 1 h in the training context (ABA) for expression of extinction and renewal (context-induced reinstatement), respectively. These tests were separated by 24 h, and the order of tests was counterbalanced within subjects. Rats were tested 24 h later for 1 h reacquisition of alcoholic beer seeking in the training context. Our past research has shown no impact of the prior order of ABA and ABB testing on responding during reacquisition. Rats received a single intraperitoneal injection of CNO 90 min before each of these tests, which is consistent with the pharmacokinetics of CNO and studies showing long-lasting behavioral and electrophysiological effects (Urban and Roth, 2015; Roth, 2016). For Experiments 2a and 3, the dose of CNO was 20 mg/kg (Mahler et al., 2014); whereas for Experiment 2b, it was 3 mg/kg (Yau and McNally, 2015; Sengupta et al., 2016).

Experiment 3: Chemogenetic disconnection of the VP → VTA and VP → STN pathways on context-induced reinstatement and reacquisition of alcohol seeking. There were three groups: ipsilateral, VP → VTA disconnection, and VP → STN disconnection. The ipsilateral group ($n = 12$) received hM4Di applied to the VP in one hemisphere (counterbalanced across the left and right hemispheres) and also hM4Di applied to the ipsilateral VTA ($n = 6$) or the ipsilateral STN ($n = 6$). The VP → VTA disconnection group ($n = 8$) received hM4Di applied to the VP in one hemisphere (counterbalanced across the left and right hemispheres) and also hM4Di applied to the contralateral VTA. The VP → STN disconnection group ($n = 8$) received hM4Di applied to the VP in one hemisphere (counterbalanced across the left and right hemispheres) and also hM4Di applied to the contralateral STN. All rats were then trained, extinguished, and tested for expression of extinction (ABB), context-induced reinstatement (ABA), and reacquisition as per Experiment 2.

After reacquisition test, rats rested for 2 d in their home cage. Then they received a single session of retraining, identical to self-administration, to ensure that all groups reached similar levels of responding. Rats were tested the following day in a progressive ratio (PR) test. During these tests, active nose pokes were rewarded on a PR schedule. The schedule was derived from the function where response ratio equals to $[5^{e(\text{reinforcer number} \times 0.2)}] - 5$, rounded to the nearest integer (Richardson and Roberts, 1996). The next day, rats were placed in locomotor chambers for 30 min to habituate them to the chambers. Finally, 24 h later, they were tested for locomotor activity for 30 min. Rats received an intraperitoneal injection of CNO (20 mg/kg) (Mahler et al., 2014) 90 min before ABB, ABA, PR, and locomotor tests.

Immunohistochemistry. At the conclusion of the experiments, rats were deeply anesthetized with sodium pentobarbital (100 mg/kg, i.p.) and perfused transcardially with 150 ml of 0.9% saline, containing heparin (5000 IU/ml), followed by 400 ml of 4% PFA in 0.1 M phosphate buffer (PB), pH 7.4. Brains were postfixed for 1 h in the same fixative and placed in 20% sucrose solution overnight. Brains were frozen and sliced to 40 μm coronal sections. Four serially adjacent sets from the VP were obtained from each brain and stored in 0.1 M sodium azide in 0.1 M PBS, pH 7.2.

Immunoreactivity for mCitrine (AAV5-hSyn-eYFP, AAV5-hSyn-HA-hM3Dq-IRES-mCitrine, AAV5-hSyn-HA-hM4Di-IRES-mCitrine) was performed using an eGFP antibody. Sections were washed in 0.1 M PB, followed by 50% ethanol, 50% ethanol with 3% hydrogen peroxidase, then 5% normal horse serum in PB (30 min each). Sections were then incubated in rabbit antiserum against eGFP (1:2000; Invitrogen, catalog #AA11122) in a PB solution blocking buffer (2% normal horse serum and 0.2% Triton X-10) (48 h at 4°C). The sections were then washed and incubated in biotinylated donkey anti-rabbit (1:1000; catalog #11,065,152, Jackson ImmunoResearch Laboratories, 24 h at 4°C). Finally, the sections were incubated in avidin-biotinylated HRP complex (Vector Elite kit: 6 $\mu\text{l/ml}$ avidin and 6 $\mu\text{l/ml}$ biotin; Vector Laboratories,

2 h at room temperature), washed in PB, and then incubated (15 min) in a DAB solution containing 0.1% 3,3'-diaminobenzidine, 0.8% D-glucose and 0.016% ammonium chloride. Immunoreactivity was catalyzed by the addition of 0.2 $\mu\text{l/ml}$ glucose oxidase (24 mg/ml, 307 U/mg, Sigma-Aldrich). Brain sections were then washed in PB, pH 7.4.

For detection of c-Fos (Experiment 1 only), brain sections were then incubated in rabbit anti-c-Fos (1:500, sc52; Santa Cruz Biotechnology) for 24 h at 4°C. The primary antibodies were diluted in blocking buffer. After washing off unbound primary antibody, sections were incubated overnight at 4°C in biotinylated donkey anti-rabbit IgG (1:500; catalog #711065152, Jackson ImmunoResearch Laboratories) diluted in blocking buffer. After washing off unbound secondary antibody, sections were incubated for overnight at 4°C in streptavidin, AlexaFluor-350 conjugate (1:300, S11249, Invitrogen). Brain sections were then washed in PB, pH 7.4, and mounted using mounting media. VP sections were delineated according to Paxinos and Watson (2007) and imaged at 20 \times using a transmitted light microscope (Olympus BX51). Counts of all neurons immunoreactive (IR) for c-Fos and CTb-488 or CTb-555 native fluorescence across 3 VP sections (80 μm apart) were made using Photoshop (Adobe).

Data analyses. The numbers of nose pokes were recorded during training, extinction, and tests. For locomotor activity, the number and location of beam breaks were counted. These data, as well as the total counts of c-Fos-IR, CTb, and c-Fos-IR/CTb neurons in VP, were analyzed by means of planned contrasting testing procedure. A multivariate approach to repeated measures was adopted (Boik, 2016). The Type I error rate (α) was controlled at 0.05 for each contrast tested (Harris, 2004).

Results

Experiment 1: VP output pathways recruited during context-induced reinstatement of alcohol seeking

First, we examined recruitment of VP projections to the VTA and STN during context-induced reinstatement of alcohol seeking. Rats with CTb-488 and CTb-555 applied to one VTA and the contralateral STN were trained to respond for alcoholic beer in one context (A) before extinction in a second context. One group (AB0) remained in their home cage on test and received handling. A second group (ABB) was tested in the extinction context. The third group (ABA) was tested in the training context. We examined recruitment of the VP → VTA and VP → STN pathways during these tests by assessing expression of the c-Fos protein in the VP and its colocalization with CTb-488 and CTb-555.

Histology

Four animals were excluded due to incorrect placement of CTb outside the boundaries of the STN or VTA. Some animals (ABA, $n = 1$; ABB, $n = 2$; AB0, $n = 1$) had correct CTb placement in one region but not the other. These animals were included in analyses for the correct CTb placement but excluded for the incorrect CTb placement.

Behavior

All rats acquired alcohol self-administration. Averaged across the last 3 d of acquisition training, the mean \pm SEM nose pokes for ABA were 99.67 ± 12.26 (active) and 1.57 ± 0.81 (inactive), ABB were 105.43 ± 12.23 (active) and 1.38 ± 0.20 (inactive), and for AB0 were 107.50 ± 17.00 (active) and 1.72 ± 0.58 (inactive). There were significantly more active than inactive nose pokes ($F_{(1,17)} = 165.95, p < 0.05$) but no overall difference between AB0 and ABB, ABA ($F_{(1,17)} = 0.09, p > 0.05$) groups. During extinction training (Fig. 1A), rats made significantly more active than inactive nose pokes ($F_{(1,17)} = 113.56, p < 0.05$). Responding decreased across days of extinction ($F_{(1,17)} = 61.48, p < 0.05$). This decrease was greater for the active than the inactive nose pokes ($F_{(1,17)} = 77.96, p < 0.05$). However, there were no interactions

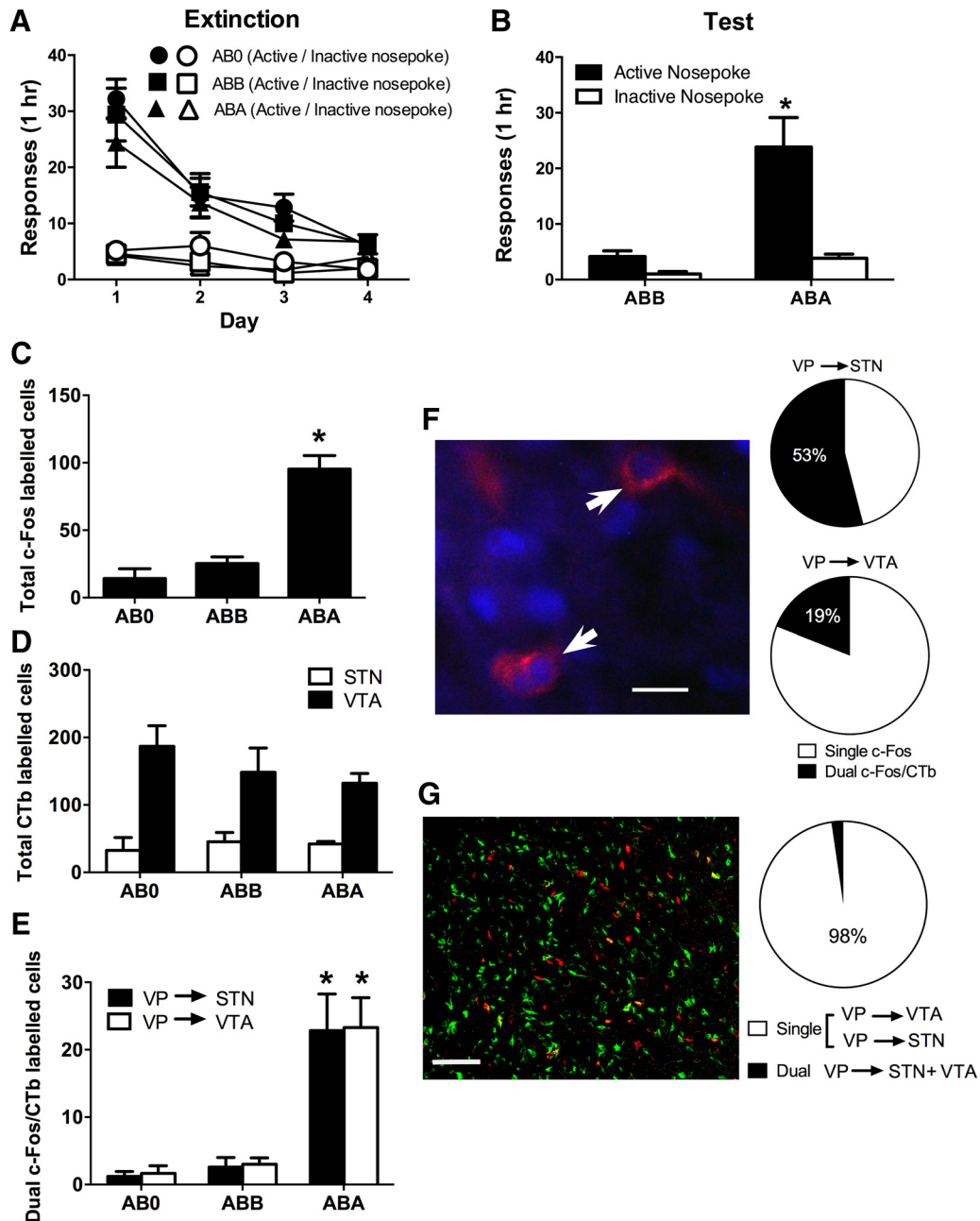


Figure 1. *A*, Mean (\pm SEM) responding during extinction training. *B*, Mean (\pm SEM) responding during test. There was context-induced reinstatement of alcohol seeking when animals were tested in the training (ABA) context. *C*, Mean (\pm SEM) counts of c-Fos-IR in VP during expression of context-induced reinstatement (ABA, $n = 7$), extinction (ABB, $n = 7$), or in controls trained and extinguished but never tested (AB0, $n = 6$). There was reinstatement-associated c-Fos in Group ABA. *D*, Mean (\pm SEM) counts of VP neurons retrogradely labeled from either VTA or STN. There were no differences between Groups ABA, ABB, and AB0 in number of retrograde-labeled neurons. *E*, Mean (\pm SEM) counts of dual c-Fos/CTb-labeled VP neurons, with significantly more dual-labeled VP neurons from either STN or VTA in Group ABA. *F*, Example of dual-labeled VP neuron (blue represents c-Fos; red represents CTb), and proportion of retrograde-labeled VP neurons expressing c-Fos for each pathway. *G*, Example of segregation of VP neurons projecting to the STN (red) or VTA (green) and proportion of labeled neurons projecting to either STN or VTA (single) versus projecting to both STN and VTA (dual). * $p < 0.05$, compared with ABB (*B*) or ABB and AB0 (*C,E*). Scale bars: *F*, 31.25 μ m; *G*, 125 μ m.

between extinction and any of the differences between groups ($F_{(1,17)} = 1.02, p > 0.05$).

On test (Fig. 1*B*), there was more active than inactive responding ($F_{(1,12)} = 17.81, p < 0.05$). There was overall more responding in the training context (ABA) compared with the extinction context (ABB) ($F_{(1,12)} = 17.22, p < 0.05$), and this increase in responding upon test in the training context was greater for the active compared with the inactive nosepoke ($F_{(1,12)} = 17.81, p < 0.05$). Hence, there was evidence for context-induced reinstatement.

Immunohistochemistry

Context-induced reinstatement was associated with significant c-Fos-IR in the VP (Perry and McNally, 2013). Group ABA expressed significantly more c-Fos-IR than Groups ABB and AB0 ($F_{(1,17)} = 64.60; p < 0.05$), which did not differ from each other ($F_{(1,17)} < 1; p > 0.05$) (Fig. 1*C*). There were no significant differences between groups in the number of VP neurons retrogradely labeled from VTA ($F_{(1,17)} < 1; p > 0.05$) or STN ($F_{(1,13)} < 1; p > 0.05$) (Fig. 1*D*). For both the VP → STN and VP → VTA projections, there were significantly more dual-labeled c-Fos-IR/CTb

neurons (Fig. 1E,F) in Group ABA compared with Groups ABB and AB0 VTA ($F_{(1,17)} = 37.53, p < 0.05$) and STN ($F_{(1,13)} = 165.67, p < 0.05$) (Fig. 1D), and Groups ABB and AB0 did not differ from each other (VTA: $F_{(1,17)} < 1, p > 0.05$; STN $F_{(1,13)} < 1, p > 0.05$). A total of 53% of VP neurons retrograde-labeled from STN, and 19% of VP neurons retrograde-labeled from VTA, expressed c-Fos-IR during context-induced reinstatement (Fig. 1F). So, context-induced reinstatement was associated with recruitment of the VP → VTA and VP → STN pathways.

Our procedure did not allow examination of the extent of colocalization of the VP → VTA and VP → STN pathways because each tracer was applied to a different hemisphere. So, in a separate group ($n = 10$), we applied CTb-488 and CTb-555 to one VTA and the ipsilateral STN. We examined VP for colabeled neurons indicative of the same VP neurons at the origins of the VP → VTA and VP → STN pathways. Consistent with past reports that the VP → STN and VP → VTA pathways are segregated with distinct topographical organization (Zahm, 1989; Zahm and Heimer, 1990; Groenewegen et al., 1993), we saw little colocalization of the two tracers in VP neurons retrogradely labeled from these two sites (Fig. 1G). Approximately 98% of labeled neurons were retrograde-labeled from either the VTA or STN, and only ~2% of neurons retrograde-labeled from both. Consistent with past research, STN-projecting VP neurons tended to be located in dorsolateral portions of the VP, whereas VTA-projecting neurons tended to be located in ventromedial medial portions; however, there was considerable intermingling of these populations.

Experiment 2: effects of chemogenetic inhibition or excitation of VP neurons on context-induced reinstatement and reacquisition of extinguished alcohol seeking

Having established recruitment of key VP output pathways during context-induced reinstatement, we next examined the effects of chemogenetic inhibition or excitation of VP neurons on reinstatement of extinguished alcohol seeking. In separate experiments, we expressed the inhibitory hM4Di DREADD or eYFP (Experiment 2a) or the excitatory hM3Dq DREADD or eYFP (Experiment 2b) in VP neurons. We trained and extinguished animals, then tested for expression of extinction (ABB), context-induced reinstatement (ABA), and reacquisition of alcohol-seeking. We used two different forms of reinstatement (context and reacquisition) because they model different features of relapse. Whereas context-induced reinstatement, such as cue- and stress-induced reinstatement, assesses reinstatement in the absence of the reinforcer, reacquisition uniquely models features of relapse that depend on contingent contact with the reinforcer. Reacquisition and context-induced reinstatement depend on different inputs to VP (Khoo et al., 2015), and it remains to be determined whether they share common VP substrates or output pathways.

hM4Di histology

Figure 2A shows the maximal extent of eYFP-IR across all rats included in the analyses, with each rat represented at 20% opacity. One rat was excluded because of unilateral expression of hM4Di.

hM4Di behavior

All rats acquired self-administration. Averaged across the last 3 d of acquisition training, the mean \pm SEM nose pokes for eYFP were 151.14 ± 26.83 (active) and 2.14 ± 0.39 (inactive) and for hM4Di were 117.95 ± 1.67 (active) and 11.07 ± 0.51 (inactive). There were significantly more active than inactive nose pokes

($F_{(1,12)} = 83.05, p < 0.05$) but no overall difference between eYFP and hM4Di ($F_{(1,12)} < 1.35, p > 0.05$) groups. During extinction training (Fig. 2B, inset), all rats made significantly more active than inactive nose pokes ($F_{(1,12)} = 44.19, p < 0.05$). Overall, responding decreased across extinction ($F_{(1,12)} = 15.16, p < 0.05$), which was greater for the active than the inactive nose poke ($F_{(1,12)} = 18.07, p < 0.05$). However, there were no interactions between extinction and any of the differences between groups ($F_{(1,12)} = 0.22, p > 0.05$).

For tests in the extinction (ABB) and training context (ABA) contexts, the hM4Di group responded significantly less than the eYFP group ($F_{(1,12)} = 9.73, p < 0.05$) (Fig. 2B). There was also significantly more active than inactive responding ($F_{(1,12)} = 32.80, p < 0.05$), significantly more responding in the training context (ABA) compared with the extinction context (ABB) ($F_{(1,12)} = 9.26, p < 0.05$), and a context \times manipulanda interaction ($F_{(1,12)} = 15.64, p < 0.05$), providing evidence for context-induced reinstatement. There was significantly less reinstatement in the hM4Di group because the three-way DREADD \times context \times manipulanda interaction was significant ($F_{(1,12)} = 6.46, p < 0.05$). So, chemogenetic inhibition of VP attenuated context-induced reinstatement of extinguished alcohol seeking.

Next, animals were tested for reacquisition (Fig. 2B). Overall, there were more active than inactive nose pokes ($F_{(1,12)} = 50.18, p < 0.05$) and an overall significant difference between eYFP and hM4Di groups ($F_{(1,12)} = 7.89, p < 0.05$). There was also a DREADD \times manipulanda interaction ($F_{(1,12)} = 6.98, p < 0.05$) showing that chemogenetic VP inhibition reduced reacquisition.

hM3Dq histology

Figure 2A shows the extent of eYFP-IR across all rats included in the analyses with each rat represented at 20% opacity. Three rats were excluded due to expression of the eYFP or hM3Dq being located either unilaterally or regions outside VP.

hM3Dq behavior

All rats acquired alcohol self-administration. Averaged across the last 3 d of acquisition training, the mean \pm SEM nose pokes for eYFP were 82.52 ± 6.12 (active) and 2.67 ± 0.80 (inactive) and for hM3Dq were 93.61 ± 5.06 (active) and 2.00 ± 0.66 (inactive). There were significantly more active than inactive nose pokes ($F_{(1,11)} = 394.12, p < 0.05$) but no overall difference between eYFP and hM3Dq ($F_{(1,11)} < 1.31, p > 0.05$) groups. During extinction training (Fig. 2C, inset), there was no overall difference between groups ($F_{(1,11)} < 1, p > 0.05$). All rats made significantly more active than inactive nose pokes ($F_{(1,11)} = 59.29, p < 0.05$), and this was greater for the hM3Dq compared with the eYFP group ($F_{(1,11)} = 5.70, p < 0.05$). Responding decreased across days of extinction ($F_{(1,11)} = 28.93, p < 0.05$). This decrease was greater for the active nose poke ($F_{(1,11)} = 40.25, p < 0.05$), and there was a three-way DREADD \times manipulanda \times day interaction ($F_{(1,11)} = 10.12, p < 0.05$). This was unexpected but shows that the decrease in active nose poke responding during extinction was greater in the eYFP versus hM3Dq group.

For tests in the extinction (ABB) and training contexts (ABA) (Fig. 2C), there was more responding in the hM3Dq compared with the eYFP group ($F_{(1,11)} = 6.02, p < 0.05$). There was also significantly more active than inactive responding ($F_{(1,11)} = 46.68, p < 0.05$) that was equivalent across groups ($F_{(1,11)} < 1, p < 0.05$). There was significantly more responding in the training context (ABA) compared with the extinction context (ABB) ($F_{(1,11)} = 71.66, p < 0.05$), and this increase was significantly greater for the hM3Dq compared with the eYFP group ($F_{(1,11)} =$

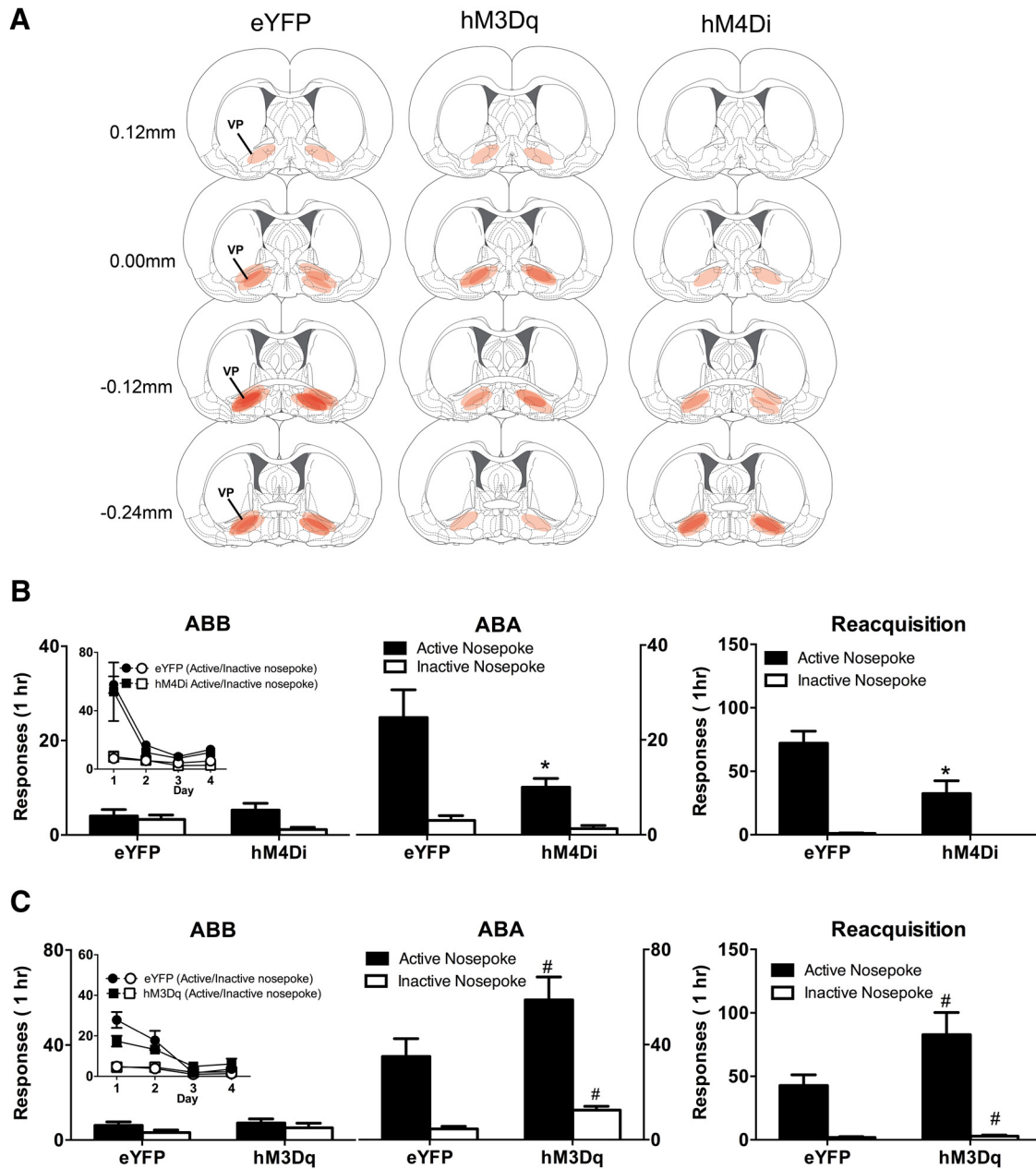


Figure 2. *A*, Location of eYFP, hM3Dq, and hM4Di expression in VP. *B*, Mean (\pm SEM) responding on tests in the extinction (ABB) and training (ABA) contexts as well as during reacquisition for hM4Di ($n = 7$) versus eYFP ($n = 7$) groups. Inset, Responding during extinction training. There was significant context-induced reinstatement that was reduced in the hM4Di groups, and this chemogenetic silencing also significantly reduced reacquisition of alcohol seeking. *C*, Mean (\pm SEM) responding on tests in the extinction (ABB) and training (ABA) contexts as well as during reacquisition for hM3Dq ($n = 6$) versus eYFP ($n = 7$) groups. Inset, Responding during extinction training. There was significant context-induced reinstatement, but the hM3Dq manipulation nonselectively increased responding during this reinstatement and during reacquisition. * $p < 0.05$, significant interaction between context, manipulanda, and DREADD. # $p < 0.05$, significant main effect of DREADD.

7.41, $p < 0.05$). Finally, there was evidence for context-induced reinstatement (i.e., a context \times manipulanda interaction; $F_{(1,11)} = 29.67$, $p < 0.05$), but this did not interact with the DREADD manipulation ($F_{(1,11)} = 1.64$, $p > 0.05$). So, chemogenetic VP excitation increased responding during reinstatement, but it acted nonselectively during this test on both active and inactive nosepokes.

During reacquisition (Fig. 2*C*), VP chemogenetic excitation again increased responding ($F_{(1,11)} = 4.99$, $p < 0.05$), and there were significantly more active than inactive nosepokes ($F_{(1,11)} = 42.68$, $p < 0.05$). However, this chemogenetic increase in responding was again relatively nonselective because it did not dif-

fer between the active and inactive nosepokes ($F_{(1,11)} = 4.43$, $p > 0.05$).

These effects of the hM3Dq manipulation are worth addressing. Whereas the impact of VP silencing via the hM4Di was selective to the active nosepoke, the impact of VP excitation via the hM3Dq was not. Rather, VP excitation via the hM3Dq increased responding on both the active and inactive nosepoke. This could reflect a chemogenetic amplification of general arousal or appetitive motivation, or it could reflect chemogenetic increases in locomotor activity. Regardless, it suggests caution in interpreting the effects of the hM3Dq and highlights the utility of the control manipulanda in this experiment.

Experiment 3: chemogenetic disconnection of the VP → VTA and VP → STN pathways on context-induced reinstatement and reacquisition of alcohol seeking

Having established that the VP → VTA and VP → STN pathways are both recruited during reinstatement and that chemogenetic silencing of VP attenuates both context-induced reinstatement and reacquisition of alcohol seeking, we next asked whether the VP → VTA and VP → STN pathways contribute to reinstatement. To do so, we used a chemogenetic disconnection strategy. We expressed the inhibitory hM4Di in one VP and the contralateral VTA (VP → VTA) or in one VP and the contralateral STN (VP → STN). For our control group, we expressed the hM4Di in one VP and ipsilateral VTA or ipsilateral STN (ipsilateral). We trained the rats and then extinguished their lever responding, then tested for expression of extinction (ABB), context-induced reinstatement (ABA), and for reacquisition of alcohol-seeking.

We also assessed the effects of VP → VTA and VP → STN disconnection on motivation to respond for alcoholic beer on a PR schedule because deep brain stimulation in STN reduces motivation to work for cocaine and alcohol (Baunez et al., 2005; Rouaud et al., 2010), and we wanted to determine whether this depended on VP-STN interactions. Finally, we assessed the impact of VP → VTA and VP → STN disconnection on locomotor activity.

Histology

Four rats were excluded due to hM4Di misplaced (2 STN and 2 VTA misplacements). Figure 3A shows the extent of eYFP-IR across all rats included in the analyses with each rat represented at 20% opacity.

Behavior

All rats acquired self-administration. Averaged across the last 3 d of acquisition training, the mean ± SEM nosepekes for ipsilateral were 98.53 ± 9.00 (active) and 2.03 ± 0.55 (inactive), VP → STN were 72.75 ± 4.90 (active) and 1.42 ± 0.25 (inactive), and VP → VTA were 100.54 ± 10.90 (active) 0.79 ± 0.21 (inactive). There were significantly more active than inactive nosepekes ($F_{(1,25)} = 279.25, p < 0.05$) but no overall difference between ipsilateral disconnection compared with VP → STN and VP → VTA disconnection ($F_{(1,25)} = 1.39, p > 0.05$) groups or between VP → STN and VP → VTA disconnection ($F_{(1,25)} = 3.68, p > 0.05$).

During extinction training (Fig. 3B, inset), rats made significantly more active than inactive nosepekes ($F_{(1,25)} = 94.74, p < 0.05$). Overall, responding decreased across days of extinction ($F_{(1,25)} = 28.08, p < 0.05$). This decrease was greater for the active than the inactive nosepoke ($F_{(1,25)} = 43.62, p < 0.05$). There were no differences between groups during extinction ($F_{(1,25)} < 1, p > 0.05$). Figure 3B shows the mean ± SEM responses during tests in the extinction context (ABB) and the training context (ABA). There was an overall main effect of contralateral disconnection (ipsilateral vs VP → STN and VP → VTA; $F_{(1,25)} = 14.96, p < 0.05$) but no difference between the VP → STN and VP → VTA disconnection groups. Overall, there was more active than inactive responding ($F_{(1,25)} = 102.43, p < 0.05$), and this was greater for ipsilateral versus VP → STN and VP → VTA disconnection groups ($F_{(1,25)} = 9.72, p < 0.05$), which did not differ ($F_{(1,25)} < 1, p > 0.05$). There was evidence for context-induced reinstatement because there was significantly more responding in the training context (ABA) compared with the extinction context (ABB) ($F_{(1,25)} = 17.22, p < 0.05$), which was greater for the active compared with the inactive nosepoke ($F_{(1,25)} = 38.73, p < 0.05$). Importantly, this was significantly reduced among the VP → STN

and VP → VTA disconnection groups (three-way interaction: $F_{(1,25)} = 10.80, p < 0.05$). However, the VP → STN and VP → VTA disconnection groups did not differ from each other ($F_{(1,25)} < 1, p > 0.05$). So, chemogenetic disconnection of either VP → STN and VP → VTA pathways reduced context-induced reinstatement.

Next, we tested rats for reacquisition (Fig. 3B). During this test, there were significantly more active than inactive nosepekes ($F_{(1,25)} = 88.75, p < 0.05$). There was an overall effect of disconnection (ipsilateral vs VP → STN and VP → VTA) ($F_{(1,25)} = 4.90, p < 0.05$) and also a disconnection (ipsilateral vs VP → STN and VP → VTA) × manipulanda interaction ($F_{(1,25)} = 4.34, p < 0.05$). However, there was no overall difference between the VP → STN and VP → VTA disconnection groups ($F_{(1,25)} < 1, p > 0.05$) or an interaction with manipulanda ($F_{(1,25)} < 1, p > 0.05$). So, chemogenetic disconnection of the VP → STN and VP → VTA pathways reduced reacquisition.

We then gave all animals a reminder self-administration session to ensure that responding was equivalent. Responding on the active nosepoke was high during this session (ipsilateral mean = 127.4; SEM = 16.9; VP → VTA mean = 115.2, SEM = 9.5; VP → STN mean = 134.0, SEM = 13.6), inactive nosepekes were low (mean < 3 for each group), and there were no differences between groups ($F_{(1,25)} < 1, p > 0.05$). The following day, we tested animals for motivation to respond for and consume alcoholic beer via a PR test (Fig. 3C).

Overall, the disconnection groups (VP → STN and VP → VTA) made significantly fewer responses than the ipsilateral group during PR test ($F_{(1,25)} = 10.78, p < 0.05$), and this decrease was greater on the active than inactive nosepoke ($F_{(1,25)} = 9.64, p < 0.05$). So, disconnecting the VP → STN and VP → VTA pathways reduced responding. But, again, there were no differences between the VP → STN and VP → VTA disconnections ($F_{(1,25)} < 1, p > 0.05$). The reduction in responding caused by disconnecting the VP → STN and VP → VTA pathways was associated with a commensurate reduction in breakpoints (last ratio completed) (Fig. 3C) with the VP → STN and VP → VTA groups significantly lower than the ipsilateral control ($F_{(1,25)} = 12.77, p < 0.05$), and the VP → STN and VP → VTA groups did not differ ($F_{(1,25)} < 1, p > 0.05$). Finally, we tested the groups for locomotor activity in a novel environment (Fig. 3C). There was no effect of disconnection (ipsilateral vs VP → STN and VP → VTA; $F_{(1,25)} < 1, p > 0.05$) and no difference between the VP → STN and VP → VTA groups ($F_{(1,25)} < 1, p > 0.05$) for locomotor activity. So, chemogenetic disconnection of either the VP → VTA or VP → STN pathways reduces context-induced reinstatement and reacquisition of alcoholic beer seeking, reduces motivation to respond for alcoholic beer on a PR schedule, but does not disrupt locomotor activity.

Discussion

We examined the role of VP, and its projections to the VTA and STN, in context-induced reinstatement and reacquisition of alcohol seeking. Consistent with our previous findings, we showed that return to the training context after extinction in a second different context was able to reinstate extinguished alcohol seeking and that this reinstatement was associated with significant recruitment of VP neurons, as shown by expression of the c-Fos protein (Perry and McNally, 2013). Using dual-retrograde tracing, we mapped some of the VP output pathways recruited during context-induced reinstatement and showed a significant recruitment of both the VP → VTA and the VP → STN pathways during this reinstatement. We were able to confirm that these

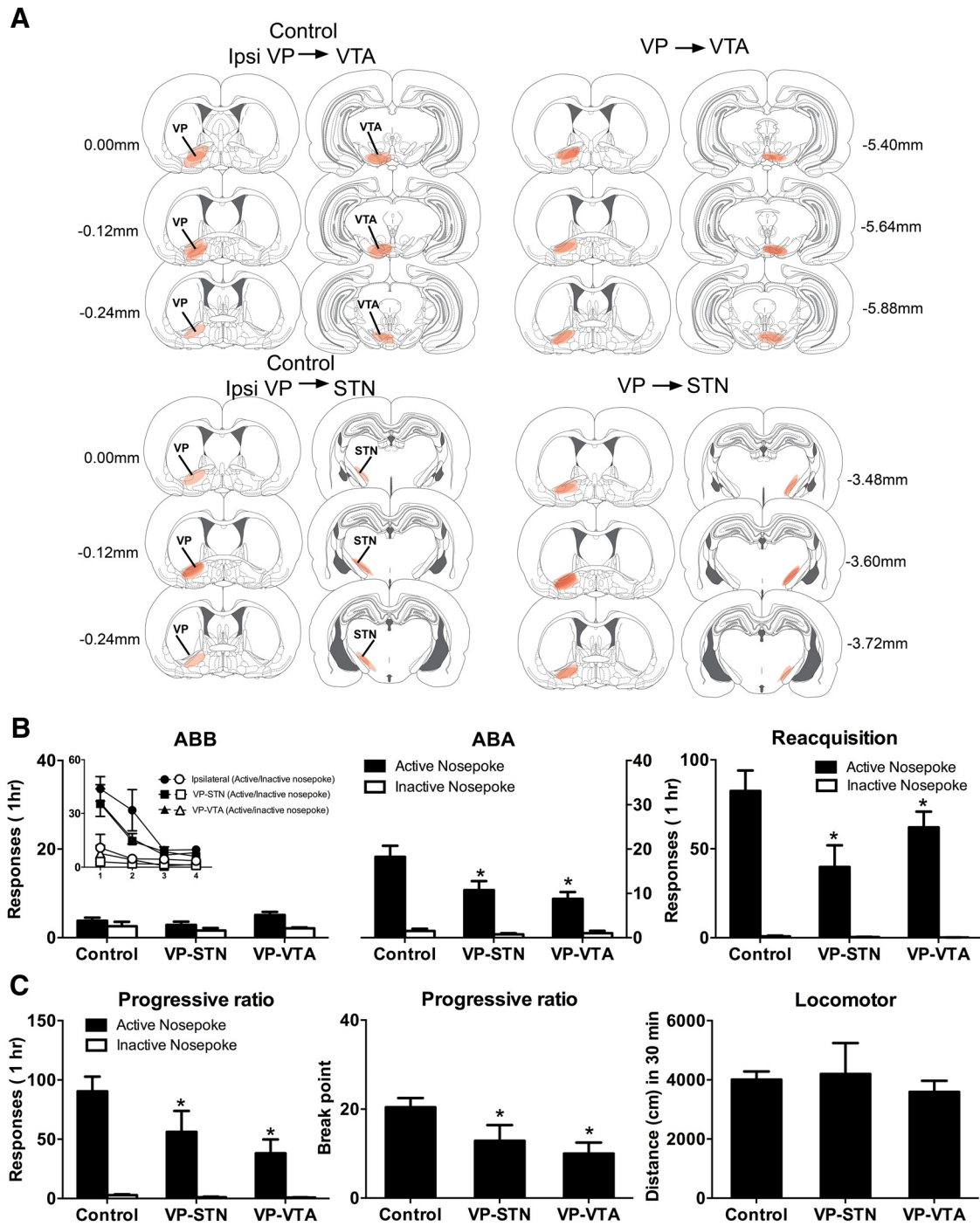


Figure 3. *A*, Location of hm4Di expression in VP, VTA, and STN for groups Control ($n = 12$), VP → VTA ($n = 8$), and VP → STN ($n = 8$). *B*, Mean (\pm SEM) responding on tests in the extinction (ABB) and training (ABA) contexts as well as during reacquisition. Inset, Responding during extinction training. There was significant context-induced reinstatement that was reduced in the VP → VTA and VP → STN disconnection groups, and this chemogenetic silencing also significantly reduced reacquisition of alcohol seeking. *C*, Mean (\pm SEM) responding on tests for PR and locomotor activity. Chemogenetic disconnection in the VP → VTA and VP → STN groups reduced responding and breakpoints but did not affect locomotor activity. * $p < 0.05$, significant difference between VP → VTA and VP → STN versus Control.

were distinct VP output pathways with few VP neurons projecting to both VTA and STN. Our findings on recruitment of the VP → VTA pathway during context-induced reinstatement extend previous reports of recruitment of this pathway during cue-induced reinstatement of cocaine seeking (Mahler and Aston-Jones, 2012). Importantly, we show that this recruitment is not unique. It is shared with another major VP output pathway, the VP → STN projection.

Multiple VP output pathways and reinstatement

Using a chemogenetic disconnection strategy, we then showed that both the VP → VTA and the VP → STN pathways were necessary for context-induced reinstatement and reacquisition of alcohol seeking because chemogenetic disconnection of either pathway was sufficient to attenuate reinstatement compared with ipsilateral disconnection. Interestingly, this role for VP → VTA and the VP → STN pathways extended to motivation to respond

for and consume alcoholic beer under a PR schedule, showing overlap in the neural substrates for reinstatement and self-administration at the level of VP and its major output pathways. Finally, despite the well-documented role of VP in locomotor activity (Root et al., 2015), there was no evidence here for alterations in activity following chemogenetic disconnection of the VP → VTA and the VP → STN pathways, although it remains possible that nonspecific effects, such as increased locomotor activity, did contribute to the impact of chemogenetic excitation of VP itself. Together, these experiments provide converging lines of evidence for the contribution of multiple VP output pathways to context-induced reinstatement and reacquisition of alcohol seeking.

Multiple VP inputs are recruited during context-induced reinstatement of alcohol-seeking (Perry and McNally, 2013) and context-induced reinstatement and reacquisition depend on different VP input pathways (Khoo et al., 2015). However, our first finding is that common and multiple VP output pathways contribute to context-induced reinstatement and reacquisition of alcohol seeking. These roles for VP → STN and VP → VTA pathways in multiple forms of reinstatement to alcohol seeking are consistent with the view that different pallidal channels control complementary aspects of motivated behavior (Root et al., 2015). VP neurons projecting to the STN are located predominantly in dorsolateral VP (VPdl) and receive extensive inputs from AcbC (Zahm et al., 1987, 1996; Groenewegen et al., 1993; Tripathi et al., 2013). This input is not necessary for context-induced reinstatement (Khoo et al., 2015) but is necessary for reacquisition of alcohol seeking (Khoo et al., 2015) as well as cue and primed reinstatement of cocaine seeking (Stefanik et al., 2013). In contrast, VP neurons projecting to the VTA are located predominantly in VPM and receive extensive inputs from accumbens shell (AcbSh) (Groenewegen et al., 1993; Zahm et al., 1996; Tripathi et al., 2010), a structure critical for context-induced reinstatement to seeking a variety of drugs of abuse (Bossert et al., 2006, 2007; Hamlin et al., 2006, 2007; Fuchs et al., 2008; Chaudhri et al., 2009; Marchant et al., 2009; Millan and McNally, 2012). The role of the AcbSh → VP pathway in context-induced reinstatement remains to be determined. Nevertheless, one attractive possibility supported by our data is that inputs from corticostriatal and amygdalostriatal circuits interface with segregated pallidal outputs, encompassing VPvm → lateral hypothalamus/VTA and VPdl → STN/substantia nigra (Zahm and Heimer, 1990; Groenewegen et al., 1993), to mediate reinstatement to drug seeking (Fig. 4).

Our second key finding is that the role of STN in reinstatement as well as alcohol seeking more generally depends, at least in part, on VP inputs. STN is gaining increased attention for its role in appetitive motivation and drug seeking (Uslaner et al., 2008; Baunez et al., 2011). STN manipulations reduce motivation for cocaine as well as alcohol (Baunez et al., 2005; Lardeux and Baunez, 2008; Rouaud et al., 2010) and reduce reinstatement of methamphetamine seeking (Baracz et al., 2015). Based on these findings, STN has been identified as a target for deep brain stimulation treatment of addiction (Baunez et al., 2011; Pelloux and Baunez, 2013) and related disorders of motivation (Bandini et al., 2007). However, the circuit level mechanisms for STN contributions to reinstatement, and appetitive motivation more generally, are poorly understood. Here we show that this role for STN depends substantially on inputs from VP. Chemogenetic disconnection of the VP → STN pathway not only reduced context-induced reinstatement and reacquisition but also significantly reduced motivation to respond in a PR task, showing overlap in

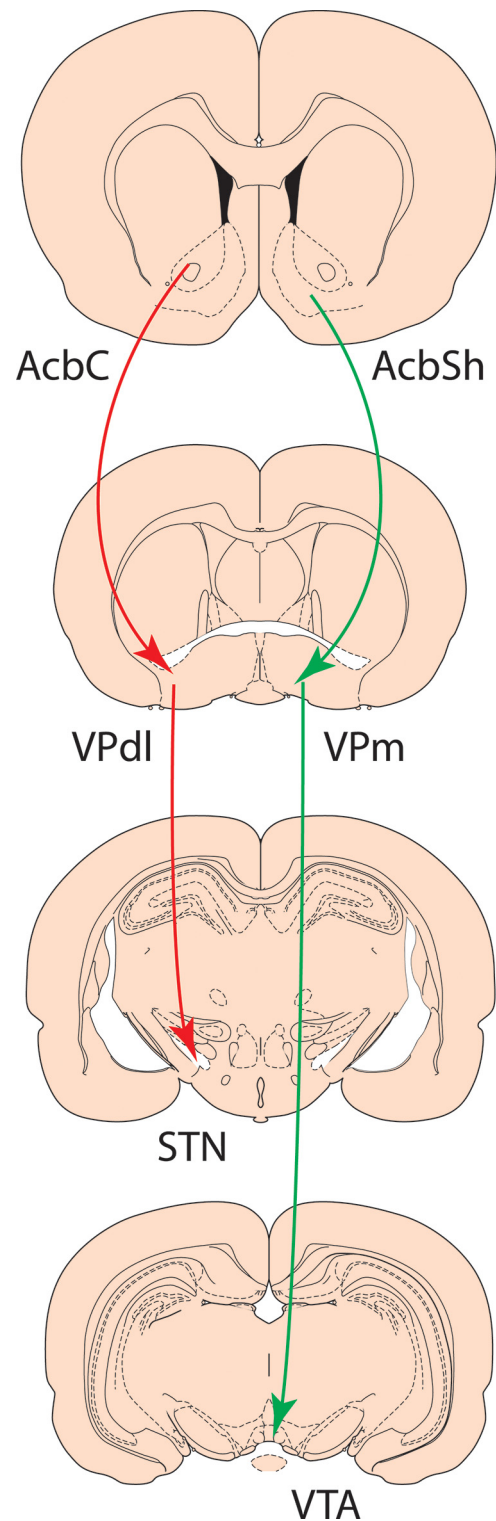


Figure 4. Organization of segregated pallidal channels, encompassing medial VP (VPM) → VTA and VPdl → STN, that interface with distinct ventral striatal, cortical, and amygdala inputs to mediate reinstatement of drug seeking.

the neural substrates for reinstatement and self-administration at the level of the VP → STN pathway. Importantly, there was no evidence here for a general suppressive effect of this chemogenetic disconnection on locomotor activity. There is extensive reciprocal connectivity between VPdl and STN (Zahm et al., 1987; Zahm and Heimer, 1990), and the role of STN → VP projections

in reinstatement remain to be determined, but our retrograde tracing data combined with chemogenetic disconnection provides strong evidence that STN contributions to reinstatement and motivation depend on VP inputs.

Further issues

Our findings leave several questions unanswered. First, it is not clear why these distinct VP output pathways contribute to both these forms of reinstatement. One possibility is that these pathways control distinct aspects of reinstatement behavior that our measures simply could not isolate. For example, consistent with recent distinctions between AcbC and AcbSh (Floresco, 2015), the VP → STN pathway may control approach, whereas the VP → VTA pathway may maintain task-related performance to ensure sustained responding in the reinstatement tasks (Floresco, 2015; Root et al., 2015). Reinstatement is likely assembled from these (and other) different functions, and their disruption would be sufficient to reduce context-induced reinstatement or reacquisition when measured via total responses, but our measures may lack the resolution to separately isolate these functions. Second, it is unclear to what extent the roles for these VP output pathways in reinstatement are specific to seeking drug rewards versus appetitive motivation more generally. The VP → VTA pathway has been implicated more broadly in the control over actions by environmental stimuli (Leung and Balleine, 2013), so it likely serves a general role across different natural and drug rewards. Our findings that chemogenetic silencing disrupted responding in the presence (reacquisition) or absence (renewal) of alcohol and also reduced motivation on a PR schedule support this possibility. Similarly, although the STN was initially identified as serving opposing roles in the motivation for cocaine versus natural rewards (Baunez et al., 2005), more recent work shows that STN contributions are linked to initial preference for reward rather than whether it is a natural or drug reward (Lardeux and Baunez, 2008). Finally, VP output pathways converge directly, or indirectly, on VTA and SNr, but the nature of VTA and SNr contributions to context-induced reinstatement and reacquisition remains poorly understood. Although VTA contributes to context-induced reinstatement of heroin seeking (Bossert et al., 2004), its role in context-induced reinstatement to seeking other drugs is unknown. Moreover, the critical VTA neuronal populations mediating context-induced reinstatement and reacquisition are unknown. VP inputs provide GABAergic control over VTA dopamine neuron activity and dopamine efflux in ventral striatum (Floresco et al., 2003; Grace et al., 2007; Grace, 2016). Mahler et al. (2014) showed that VTA TH neurons were necessary for cue-induced reinstatement of cocaine seeking but that this was not due to direct input from VP terminals, raising the possibility that complex disinhibitory intra-VTA circuits contribute to reinstatement.

In conclusion, here we show that multiple VP output pathways contribute to reinstatement of alcohol seeking. Much remains to be learned approximately how VP contributes to reinstatement of drug seeking. Most research has focused on roles of striatal inputs (McFarland and Kalivas, 2001; Torregrossa and Kalivas, 2008; Leung and Balleine, 2013; Stefanik et al., 2013; Khoo et al., 2015). However, the convergence of multiple activated striatal, thalamic, and amygdala inputs in VP (Perry and McNally, 2013) with recruitment of multiple VP output pathways during reinstatement, combined with recent findings on the timing of reward related VP neuron activity (Richard et al., 2016), suggests that the role for VP in reinstatement of drug seeking is likely to be more complex than currently appreciated.

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