



## Commentary

# Aminoglycoside resistance mechanism inference algorithm: Implication for underlying resistance mechanisms to aminoglycosides



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The discovery of antibiotics in 1928 and their subsequent large-scale production are considered to be one of the most important achievements in the history of medicine [1]. One of the most important discoveries after that of  $\beta$ -lactams was streptomycin, the first aminoglycoside discovered. The history of aminoglycosides was then marked by the successive introduction of a series of compounds (kanamycin, gentamicin, and tobramycin) for the treatment of infections due to Gram-negative bacilli [2].

The recent expansion of extensively drug-resistant (XDR) pathogens and particularly that of carbapenem-producing *Enterobacteriaceae* (CRE) has brought into light aminoglycosides, which may retain activity even in XDR isolates [3]. Specific indications for aminoglycoside therapy include amikacin and gentamicin administered intravenously for infections caused by MDR Gram-negative organisms [4].

Interpretative reading of antimicrobial susceptibility test results allows to analyze the susceptibility pattern and to predict the underlying resistance mechanisms [5]. Contrary to  $\beta$ -lactams antibiotics, correlations between resistance to aminoglycosides inferred based on the European Committee on Antimicrobial Susceptibility Testing (EUCAST) clinical breakpoints and expert rules are generally poor [6]. Therefore, the need for improvement of detection of aminoglycosides resistance mechanisms in routine is of great importance.

Recently in *EBioMedicine*, Mancini and colleagues presented an Aminoglycoside Resistance Mechanism Inference Algorithm (ARMIA) for the inference of resistance mechanisms from inhibition zone diameters [7]. This algorithm uses ECOFFs for gentamicin, tobramycin and kanamycin as well as a working separator cut-off for amikacin. They compared the performance of ARMIA and EUCAST CBPs/expert rules with that of whole-genome sequencing (WGS) in predicting aminoglycoside resistance. The results of this study showed that ARMIA-based inference of resistance mechanisms and WGS data were congruent in 96.3%. In contrast, there was a poor correlation between resistance mechanisms inferred using EUCAST CBPs/expert rules and WGS data (85.6%) [7].

When assessing the accuracy of various susceptibility testing methods as compared to standard reference methods, the terms very major errors (vME) have been used to describe false susceptible [8]. Thus, in the comparison made by Mancini and colleagues, they reported

that EUCAST produced 63 (12.9%) vME, compared to only 2 (0.4%) vME with ARMIA [7].

*In vitro* susceptibility rates may vary significantly, depending on the aminoglycoside resistance mechanisms, which are frequently co-transferred along with other resistance genes on mobile genetic elements [3]. It is reported that aminoglycoside resistance mechanisms, such as 16S rRNA methylase, coexist with other resistance mechanisms including extended-spectrum  $\beta$ -lactamase, carbapenemase, and plasmid-mediated quinolone resistance determinants [9]. Thus, it is important to detect the underlying aminoglycosides resistance mechanisms to prevent co-selection of these resistance mechanisms. The ARMIA developed by Mancini and colleagues would be useful for this purpose to avoid misidentification of the aminoglycoside resistance mechanisms.

## Declaration of Competing Interest

None to declare.

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