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In Situ Immobilized Silver Nanoparticles on Rubia tinctorum Extract-**Coated Ultrasmall Iron Oxide Nanoparticles: An Efficient** Nanocatalyst with Magnetic Recyclability for Synthesis of Propargylamines by A³ Coupling Reaction

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ABSTRACT: This research suggests a green method for synthesizing hybrid magnetic nanocomposites that can be used as a reductant and a stabilizing agent for immobilizing metal nanoparticles (NPs). The central idea is the modification of magnetic NPs using Rubia tinctorum extract, which consists of numerous carbonyl and phenolic hydroxyl functional groups to increase adsorption of metals and chelate silver ions, and decrease the adsorption of silver ions by Ag NPs, in situ. Thus, the suggested catalyst preparation process does not require toxic reagents, additional reductants, and intricate instruments. To show the effectiveness of the plant extract in reducing and immobilizing Ag NPs, the structural, morphological, and physicochemical features of the particles are studied using Fourier-transform infrared spectroscopy, inductively coupled plasma atomic emission spectroscopy, field emission scanning electron microscopy, energy-dispersive X-ray spectroscopy, high-resolution transmission electron microscopy, vibrating sample magnetometry, X-ray diffraction analysis, and X-ray photoelectron spectroscopy. One of the advantages of the suggested method is to reduce the size of the magnetic NPs from



15-20 to 2-5 nm, in the presence of the extract. Additionally, the prepared Fe₃O₄@R. tinctorum/Ag nanocatalyst is demonstrated to exhibit a very high activity in the catalysis of the three-component reaction of aldehydes, amines, and alkynes $(A^3 \text{ coupling})$ with good to high yields of diverse propargylamines. Moreover, the nanocatalyst can be recovered several times with no considerable leaching or loss of performance.

■ INTRODUCTION

Propargylamines (PPAs) play many main roles in the area of organic chemistry. They can be utilized as the precursors and building units of numerous nitrogen-containing heterocyclic compounds; meanwhile, they can also be used as the key intermediates in synthesizing bioactive medicines and natural products.1 Furthermore, numerous PPAs can cure neuropsychiatric disorders such as Alzheimer's and Parkinson's diseases.² Owing to the importance of PPAs, diverse methods, for example, A³ coupling reaction, have been improved for their preparation.³ Among all suggested PPA synthesis approaches, the A^3 coupling method is considered as the most efficient and direct method. In this technique, a homogeneous or heterogeneous transition-metal catalyst is used for coupling the three reaction components, that is, an amine, an aldehyde, and a terminal alkyne.⁴ In recent years, diverse homogeneous and heterogeneous catalysts have been applied to synthesize PPA via A^3 -coupling reaction based on transition metals containing $Zr^{4+,5}$ $Mn^{2+,6}$ $Re^{1+,7}$ $Fe^{2+,8}$ $Ru^{3+,9}$ $Co^{2+,10}$ $Ir^{1+,11}$ $Ni^{2+,12}$ $Pd^{2+,13}$ $Cu^{2+,14}$ $Ag^{1+,15}$ $Au^{0,16}$ $Zn^{2+,17}$ $Cd^{2+,18}$ and $Hg^{2+,19}$ The used transition-metal catalysts can be in the form of nanoparticles (NPs) resulting in a great surfaceto-volume ratio, excellent selectivity, and good activity for the coupling reaction.^{20,21}

The NPs can be a more attractive option if they are magnetic as magnetic NPs (MNPs) ease elimination and recovery of nanocatalysts from reaction mixtures.²² Diverse core-/shell-structured MNPs have been applied to reduce the cost of catalyzed reactions.²³⁻²⁸ However, Fe₃O₄-based core/ shell NPs have been the most extensively used nanocatalysts because of their ease in magnetic separation and noticeable catalytic activity. Importantly, Fe₃O₄ systems can be functionalized with numerous functional groups for carrying noble metals. In this regard, polymers are a suitable option as they can tolerate a large number of functional groups in their chains for presenting excellent compatibility with the interest metals. The other benefit of anchoring polymers on Fe₃O₄ MNPs is that the existence of the polymers avoids accumulation of the magnetic core/shell structures, which is a common issue in preparing Fe₃O₄ core/shell nanocatalysts. For this reason, numerous studies have emphasized on the encapsulation of Fe₃O₄ NPs by diverse polymers. Several examples of the reported Fe₃O₄-polymer composites are Fe₃O₄@polypyrrole, Fe₃O₄@polyaniline, and Fe₃O₄@polydoapmine, which have shown excellent catalytic potential.²⁹⁻³²

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As stated, polymer-iron oxide composites have the ability to carry noble metals. The metal which is of interest in the current research is silver. So far, numerous investigations have reported the synthesis of Ag NPs for different purposes. As an example, Ag NPs have been used as extremely active surfaceenhanced Raman scattering substrates,³³ electrochemical and biosensors,^{34,35} antibacterial coatings,³⁶ and extremely effective catalysts of organic reactions.³⁷ As catalysts, Ag NPs have been used in oxidation of phenylsilanes, alcohol dehydrogenation, Diels-Alder cycloadditions, and reduction of aromatic compounds.³⁸⁻⁴⁰ Although the inadequacy of Ag NPs is that their surface atoms destabilize over reactions. Opportunely, immobilization of the Ag NPs onto supports can overcome this problem.^{41,42} In the current research, the support material is polymer-immobilized MNPs because of their various advantages. As mentioned above, one of the key advantages of using MNPs is that their supermagnetic and insoluble features allow the isolation of MNP-supported noble metals from reaction mixtures by means of an appropriate magnet and with no need of filtration or centrifugation. Moreover, immobilizing Ag NPs onto the surface of polymer-Fe₃O₄ particles develops dispersing Ag NPs and their ability to promote biological separation, biological targeting, catalysis, high-density magnetic recording, and targeting treatment.43

Regarding the wide variety of Ag-Fe₃O₄ applications, numerous approaches have been developed for their preparation. Zhang et al. studied the one-pot water-phase preparation of Ag-Fe₃O₄ MNPs with the FeCl₃ and AgNO₃ metal precursors and the ethylene glycol reductant.⁴⁴ Using the same technique, Zhu et al. synthesized Ag-Fe₃O₄ MNPs in the presence of polyvinylpyrrolidone.⁴⁵ Chen et al. used a solvothermal technique to attain Ag/Fe₃O₄ nanowires. In their research, a polyol synthesis method was used to prepare Ag nanowires and Fe₃O₄ particles were coated on the wires via the solvothermal technique.⁴⁶ Lopes et al. prepared Ag-Fe₃O₄ colloidal dimer NPs via a two-step chemical technique. First, they synthesized Ag NPs by heating a phenyl-ether solution of oleyamine, oleic acid, and C54H45NO3P3Ag at a temperature of 250 °C. Then, the obtained phenyl-ether mixture containing colloidal Ag NPs, Fe(acac)₃, oleic acid, 1,2-hexadecanediol, and oleyamine was heated to obtain heterodimer Ag-Fe₃O₄ NPs.⁴⁷ Li et al. utilized an electroless silver plating method to synthesize Ag/Fe₃O₄ MNPs via bringing a solution of AgNO₃ and formaldehyde (methanal) in contact with Fe₃O₄ NPs.⁴ The point is that these introduced preparation methods have numerous disadvantages such as operation at high temperatures and pressures, comprising complex chemical preparation, and using expensive surfactants. Consequently, these approaches should be substituted with an alternative simple, fast, and low-cost method, such as plant-mediated NP preparation. This biosynthesis method is a cost-effective and single-step method that needs nontoxic reagents and environmental friendly solvents. Accordingly, this technique is safe for humans and the environs.⁴⁹⁻⁵⁵

In the current study, the *Rubia tinctorum* plant (Figure 1) is selected for biosynthesizing the MNPs. This plant belongs to the Rubiaceae family, which has 6000 species and 500 genera,⁵⁶ and is named "madder" in English and "ronas" in Persian.⁵⁷ Over all, this plant has been originated from Caucasus and Near East; however, it is cultivated in the north of Africa, the Mediterranean regions from Spain to Asia, and, moreover, in some regions of Asia. In Iran, it is planted in some central and western regions, for example, Yazd.⁵⁸ This plant



Figure 1. R. tinctorum image.

comprises numerous anthraquinones containing rubiadin, purpurin, lucidin, and alizarin. $^{59-63}$

Today, ecological concerns have encouraged chemists to shift toward green chemistry.⁶⁴ Therefore, recoverable heterogeneous catalysts can involve in eliminating or reducing chemical wastes, mainly in the case of large-scale and industrial processes.⁶⁵ Regarding the outlined topics and in continuation of our research in this area,⁶⁶ this research emphasizes on preparation of PPAs through A^3 coupling of amines, aldehydes, and alkynes, from a green perspective. The suggested synthetic method is catalyzed by the Ag NPs supported on the ultrafine surface-modified Fe₃O₄ MNPs. In this technique, the *R tinctorum* extract is used as a stabilizer and a reductant agent that modifies the surface of the magnetic Fe₃O₄ MPs. The key advantage of the used Fe₃O₄@*R*. *tinctorum*/Ag nanocomposite is that it can be recovered and reused several runs with no remarkable loss of performance.

RESULTS AND DISCUSSION

The current research emphasizes on a green and environmental friendly technique that uses a plant extract to modify MNPs for fabricating a novel adsorbent for adsorbing metal ions and reducing them to stabilized metal NPs. The used extract is the *R. tinctorum* extract, which can deposit Ag⁺ ions onto the Fe₃O₄ MNPs surface, reduce the ions to Ag NPs, and stabilize the reduced particles effectively. It is worth noting that the biomolecules of R. tinctorum have a significant tenacity against accumulation of the Fe₃O₄ MNPs, besides reducing silver salts strongly. Moreover, the phenolic compounds of the R. tinctorum extract consist of hydroxyl and ketonic groups that can connect to metals and show chelating impact. In the current research, the NPs of Fe₃O₄@R. tinctorum/Ag were prepared via the two-stage modification of iron oxide (Fe_3O_4) MNPs using the R. tinctorum extract and, then, adsorbing Ag⁺ ions onto the modified MNPs and reducing and stabilizing the adsorbed ions by the extract. One of the benefits of this preparation method is reducing the dimension of the MNPs from 15-20 to 2-5 nm, as the adsorbed extract can reduce and stabilize the adsorbed Ag⁺ ions to Ag NPs, in situ (Scheme 1). After preparation of the Fe₃O₄@R. tinctorum/Ag MNPs, the particles were collected using an external magnet. Atomic absorption spectroscopy showed that the Fe₃O₄@R. tinctorum/ Ag nanocomposite comprises 0.105 mmol g^{-1} of silver. The structure of the nanocatalyst was characterized using diverse analytical methods of X-ray diffraction (XRD), high-resolution transmission electron microscopy (HRTEM), field emission scanning electron microscopy (FESEM), energy-dispersive Xray (EDX) spectroscopy, inductively coupled plasma atomic emission spectroscopy (ICP-AES), X-ray photoelectron spectroscopy (XPS), vibrating sample magnetometry (VSM), and Fourier-transform infrared (FT-IR) spectroscopy.

FT-IR spectroscopy was performed to assess the variations in the structure of the Fe₃O₄ NPs upon modifying with the extract and formation of Fe₃O₄@*R. tinctorum*/Ag. Figure 2 Scheme 1. Schematic Fabrication of Fe₃O₄@R. tinctorum/Ag NPs Nanocatalyst





Figure 2. FT-IR spectrum of (a) Fe_3O_4 , (b) *R. tinctorum* extract, (c) $Fe_3O_4@R$. *tinctorum* NPs, and (d) $Fe_3O_4@R$. *tinctorum*/Ag NPs.

shows the FT-IR spectra corresponding to the Fe₃O₄ NPs, the R. tinctorum extract, the Fe₃O₄@R. tinctorum MNPs, and the Fe₃O₄@R. tinctorum/Ag nanocomposite. On the basis of Figure 2a, the FT-IR spectrum corresponding to the Fe₃O₄ NPs shows two broad peaks at 1615 and 3423 cm⁻¹ corresponding to the adsorbed water molecules and the OH groups; however, the bands observed at 439 and 581 cm⁻¹ are associated with the bending and stretching vibrations of the Fe–O function group, respectively, and the band at 629 cm⁻¹ confirms the presence of a pure Fe₃O₄ structure. On the other hand, the FT-IR spectrum corresponding to the aqueous R. tinctorum extract (Figure 2b) shows the 3424 (OH), 2924 (C-H), 1718 (CO), 1636 (C=C), 1285 (C-O), 1054 (C-O-C), and 815 cm^{-1} (O–H) vibrational modes that are related to polyols, carbonyls, and C=C bonds and propose the existence of flavonoids and terpenoids in the extract. When the Fe₃O₄

tinctorum NPs, the related FT-IR spectrum (Figure 2c) is accompanied with several new vibrational bands that are not found in Figure 1a,b. These bands are directly resulted by coating the NPs with the extract's compounds and appeared at 1723 and 1087 cm^{-1} corresponding to the stretching vibrations of the C=O and C-O bonds, respectively,⁶⁷ and the bands appeared at 2924 and 2887 cm⁻¹ rising from the C-H stretching vibrations.⁶⁸ Moreover, numerous bands have been observed at 1400-1600 cm⁻¹ that are ascribed to the stretching vibrations of C=C bonds in aromatics rings.⁶⁹ These bands show that the hydrophilic nature of the modified NP surface because of the effective coating of the R. tinctorum extract on the Fe₃O₄ NPs surface. Finally, the FT-IR spectrum corresponding to the Fe₃O₄@*R. tinctorum*/Ag NPs (Figure 2d) is the same as that of the Fe₃O₄@R. tinctorum NPs. However, as it can be observed, the 1087 $\rm cm^{-1}$ band of the C–O stretching vibration is moved to 1084 cm⁻¹, the C=C stretching band is shifted from 1633 to 1625 cm⁻¹, and the O-H stretching band is moved from 3425 to 3420 cm⁻¹. These shifts show that the extract components are chemically bonded to the Ag NPs on the MNP surface. According to the achieved spectra, the hydroxyl functional groups corresponding to the flavonoids and other phenolic compounds of the extract are the active moieties that cause reduction of the adsorbed Ag⁺ ions to Ag⁰ and stabilization of the Ag NPs. Actually, because of the antioxidant nature of flavonoids and phenolic compounds, donating an electron (accompanied with a hydrogen nucleus) from the OH groups of the phenolic rings should be involved in reducing Ag⁺ ions to Ag^{0.70}

NPs are functionalized by the extract, that is, the Fe₃O₄@R.

The shape, distribution of size, surface morphology, and major physical features of the produced particles were assessed using FESEM. Because the FESEM images of the Fe₃O₄ and Fe₃O₄@*R*. *tinctorum*/Ag MNPs (Figure 3) show that the Fe₃O₄ particle size in the Fe₃O₄@*R*. *tinctorum*/Ag MNPs is smaller compared to the initial Fe₃O₄ particles while both specimens consist quasi-spherical NPs, it is obvious that coating Fe₃O₄ NPs with the extract causes a significant decrease in its particle size. Furthermore, the considerable difference between the Ag NPs and the iron oxide particles in Figure 3b shows that the Ag⁺ ions have been effectively decreased in situ for immobilizing Ag NPs onto the Fe₃O₄@*R*. *tinctorum* NP surface. Furthermore, the FESEM images display



Figure 3. FE-SEM images of (a) Fe_3O_4 and (b) $Fe_3O_4@R$. *tinctorum*/ Ag NPs.

that both the Ag and iron oxide NPs have crystalline structures with dimension ranging in 20-40 and 2-5 nm, respectively.

Figure 4 demonstrates the elemental composition corresponding to the magnetic samples. According to the achieved



EDX outcomes, silver species are dispersed on the extract shell homogeneously and the Fe₃O₄ NPs have been fully coated with the extract. Furthermore, the Fe, C, N, S, O, and Ag elements are in the Fe₃O₄@*R*. *tinctorum*/Ag nanocatalyst. The S signal that represents the alizarin molecules of the *R*. *tinctorum* extract approves effective modification of the Fe₃O₄ NPs using the extract while the presence of the Ag signal confirms successful functionalization of Ag NPs and synthesis of the desired Fe₃O₄@*R*. *tinctorum*/Ag NPs. Also, its elemental maps are displayed in Figure 5, which confirmed the welldispersed Ag NPs in the sample.

Moreover, the morphology of the as-prepared NPs was analyzed using HRTEM. Comparing the HRTEM images

corresponding to the Fe₃O₄ (Figure 6a), Fe₃O₄@R. tinctorum (Figure 6b), and $Fe_3O_4@R$. tinctorum/Ag (Figure 6c) NPs unravels that the spherical Fe_3O_4 cores of the $Fe_3O_4 @R$. tinctorum/Ag and Fe₃O₄@R. tinctorum MNPs are very smaller compared to the initial Fe₃O₄ particles and the magnetic cores are well distributed following coating them with the biocomponents of the R. tinctorum extract. It seems that the appeared thin layer of biomolecules on the Fe₃O₄@R. tinctorum particles is responsible for reduction and stabilization of silver on the magnetic support. Moreover, comparison of Figure 6b and c shows the existence of the functionalized Ag NPs on the Fe₃O₄ MNP surface and proposes that the Ag NPs are almost 20-40 nm in scale within a narrow size distribution. The agreement of the HRTEM and FESEM outcomes leaves no doubt about the ability of the *R. tinctorum* layer of $Fe_3O_4@$ R. tinctorum in reduction and deposition of Ag NPs with an excellent particle distribution on the MNPs surface. Figure 6d shows the HRTEM image corresponding to the Ag NPs at 5 nm magnification. On the basis of Figure 6d, the Ag NPs include lattice fringes, which confirm the excellent crystallinity of these NPs. Meanwhile, the fast Fourier transform (FFT) image corresponding to the Ag NPs (Figure 6e) shows bright diffraction spots and ring patterns that signify the crystalline structure of the Ag atoms on the surface of $Fe_3O_4@R$. tinctorum.

Besides confirming the deposition of Ag NPs onto the Fe₃O₄@R. tinctorum surface using the FESEM and HRTEM methods, the nanocatalyst was studied using XRD analysis to additionally approve the coated particles and unravel their crystal features. The obtained XRD patterns corresponding to the Fe₃O₄ and Fe₃O₄@R. tinctorum/Ag NPs are shown in Figure 7. The (220), (311), (400), (422), (511), and (440) crystal planes of the Fe₃O₄ NPs indicate the cubic structures of Fe₃O₄ MNPs (JCPDS no. 19-0629). However, these crystal planes also appeared in the XRD pattern corresponding to Fe₃O₄@R. tinctorum/Ag; the additional bands observed at 38.2° (111), 44.3° (200), 64.5° (220), and 78.2° (311) show that the cubic magnetite crystals are combined with facecentered cubic silver crystals (JCPDS no. 87-720). This evidence approves effective synthesis of the Ag NPs on the surface of Fe₃O₄@R. tinctorum MNPs. The last point of the crystal structures of the synthesized NPs is that the surface functionalization and silver deposition procedures are unsuccessful on the crystal structure of the Fe₃O₄ NPs. Thus, the specific XRD bands of the pristine Fe₃O₄ NPs also appear in the XRD pattern corresponding to the Fe₃O₄@R. tinctorum/Ag NPs.

The electronic features of the surface species of the MNPs containing their oxidation states, electron environs, and binding energies (BEs) were examined using XPS. Figures 8



Figure 5. (A–D) Corresponding elemental mapping of Fe₃O₄@R. tinctorum/Ag NPs, Fe, C, and Ag, respectively (scale bar is 500 nm).



Figure 6. HRTEM images of the as-prepared Fe₃O₄ NPs (a), Fe₃O₄@R. *tinctorum*/Ag NPs (b,c), HRTEM image of Ag NPs at 5 nm (d), and FFT image of Ag NPs corresponding to image of (d) (e).



Figure 7. XRD patterns of (a) Fe_3O_4 and (b) $Fe_3O_4@R$. tinctorum/Ag nanocomposites.

and 9 show the XPS elemental scans for the $Fe_3O_4@R$. *tinctorum*/Ag nanocatalyst. In agreement with the EDX outcomes, the XPS spectra show the existence of oxygen, carbon, nitrogen, silver, and iron. Moreover, the XPS outcomes show an intense doublet at the 374.33 and 368.32 eV BEs, which imply the Ag $3d_{3/2}$ and Ag $3d_{5/2}$ bands, respectively. The XPS outcomes show the presence of reduced Ag(0) species and exclude the possibility of +1 oxidation state for the coated Ag NPs. The well reduction of Ag(I) to Ag(0) approves the efficiency of the *R. tinctorum* layer.⁷¹

In Figure 9, the characteristic bands of carbon (C 1s) and iron (Fe 2p) are obvious, which shows that the *R. tinctorum*

extract is effectively deposited on the iron oxide MNPs. The XPS spectrum corresponding to Fe 2p displays two key bands that are associated with Fe $2p_{3/2}$ and Fe $2p_{1/2}$ and can be deconvoluted to the 712.4, 715.8, 719.3, 726.5, 729.8, and 733.4 eV bands. Four of these bands refer to Fe^{2+} (712.4 and 726.5 eV) and Fe^{3+} (715.8 and 729.8 eV) in the Fe_3O_4 phase while the two satellite bands (719.3 and 733.4 eV) relate with Fe^{3+} in the Fe_2O_3 phase. In other words, the study of the oxidation states points to the presence of few γ -Fe₂O₃ particles.⁷²⁻⁷⁴ Finally, fitting of the C 1s XPS spectra corresponding to the Fe₃O₄@R. tinctorum/Ag nanocatalyst shows nonoxygenated carbon at 285.2 eV, carbon in C-OH/ C-O-C at 288.2 eV, carbon in C=O at 289.2 eV, and the O-C=O group at 292.7 eV. Also, the UV analysis of the Fe_3O_4 @R. tinctorum/Ag NPs is demonstrated in Figure 10a. The peak absorption at 418 nm related to Ag NPs, which confirmed the presence of Ag NPs in the Fe₃O₄(@R. tinctorum/Ag NP nanocamposite.

As the final characterization stage, the magnetic features of the prepared Fe₃O₄ and Fe₃O₄@*R. tinctorum*/Ag NPs were examined with VSM. On the basis of Figure 10b, both of the Fe₃O₄ and Fe₃O₄@*R. tinctorum*/Ag samples show a usual superparamagnetic activity with almost zero remanence and coercivity. Furthermore, the VSM outcomes indicate that adding *R. tinctorum* extract and Ag layers reduces the saturation magnetization (M_s) of the Fe₃O₄ NPs. However, the M_s value for Fe₃O₄@*R. tinctorum*/Ag (40.3 emu/g) is significantly smaller compared with that of the Fe₃O₄@*R. tinctorum*/Ag is adequate for its magnetic recovery from diverse reaction mixtures. Consequently, the prepared nanocomposite has the ability to act as a magnetically recoverable nanocatalyst.

Once the Fe₃O₄@*R. tinctorum*/Ag NP structure had been recognized, as a nanocatalyst, the catalytic activity of Fe₃O₄@*R. tinctorum*/Ag NPs was evaluated in the three-component



Figure 8. XPS spectrum related to the elemental survey scan of Fe₃O₄@R. tinctorum/Ag and in the Ag 3d regions.



Figure 9. Wide scan XPS spectrum of $Fe_3O_4@R$. tinctorum/Ag and in the Fe 2p and C 1s regions.

coupling reaction of aldehydes, amines, and alkynes (A³ coupling). Initial experiments using phenylacetylene, morpholine, and benzaldehyde were performed to optimize diverse parameters comprising solvent, time, temperature, and catalyst load. The results are listed in Table 1. First, the impact of catalyst was examined, and as expected, the desired product was not attained with no catalyst offering the vital role of $Fe_3O_4@R$. *tinctorum*/Ag NPs in the reaction mechanism (Table 1, entry 1). The reaction yield was attained 70% by using a 0.1 mol % catalyst loading in water at a temperature of 50 °C (Table 1, entry 2). Moreover, the reaction took place quantitatively by using 0.1 mol % of catalyst at a temperature of 80 °C (Table 1, entry 3). Smaller yields were attained by reducing the catalyst level to 0.06 and 0.03 mol % (Table 1, entries 4 and 5). However, using 0.03 mol % of catalyst in other solvents containing CH_2Cl_2 , dimethylformamide (DMF), EtOH, toluene, CH_3CN , and neat provided smaller reaction performances (Table 1, entries 6–11). There was no change in the reaction time and reaction performance by growing the amount of catalyst or reaction temperature (Table



Figure 10. UV–vis spectrum of Fe₃O₄@*R. tinctorum*/Ag NPs (a) and VSM of (a) Fe₃O₄ and (b) Fe₃O₄@*R. tinctorum*/Ag NPs.

Table 1. Optimization of Reaction Parameters for the Model Reaction Using Fe₃O₄@*R. tinctorum*/Ag NPs Catalyst^a



^{*a*}Reaction conditions: benzaldehyde (1.0 mmol), phenylacetylene (1.2 mmol), morpholine (1 mmol), Fe₃O₄@*R. tinctorum*/Ag NPs, and solvent (3.0 mL). ^{*b*}Yields are based on ¹H NMR.

1, entries 12 and 13). Therefore, we selected water as a safe and eco-friendly solvent, 0.1 mol % catalyst loading, and reaction temperature of 80 $^{\circ}$ C as the most efficient and optimal reaction conditions to study the generality of this A³ coupling.

Under the optimum reaction conditions, diverse aldehydes were interacted with other aliphatic amines and phenyl-

acetylene that provided the A³ coupling product with excellent yields and good turnover frequencies (TOFs) (Table 2). At the initial research of the aldehyde substrate scope, morpholine and phenylacetylene were applied as model reaction and diverse aldehydes were examined for the A³-coupling (Table 2, entries 1-20). The results display that aromatic aldehydes activity like functional groups such as -Br, -Cl, -OH, -OMe, or -Me could influence on the A³-coupling. Furthermore, minor electronic influence was observed that is associated with the reaction of aryl aldehydes with electron-withdrawing groups (Table 2, entries 2-4) and formation of the correlated products in high yields, while replacement of electron-rich groups (Table 2, entries 5-7) on the benzene ring decreased the efficiency and resulted less yields. Moreover, reaction of heterocyclic aldehyde like furan-2-carboxaldehyde or thiophene-2-carbaldehyde with phenylacetylene and morpholine proceeded effectively and the relating PPAs were archived in high yields (Table 2, entries 8 and 9). The aliphatic aldehyde (cyclohexanecarbaldehyde/butanaldehyde) presented high yields under this optimal condition (Table 2, entries 10 and 11). To develop the generality of amine compounds, a mixture containing phenylacetylene-benzaldehyde-amine was chosen, and diverse amines were examined (Table 2, entries 1 and 12-16). The results show that cyclic, heterocyclic (pyrrolidine/ morpholine/piperidine), and acyclic (dibenzyl/diethyl) secondary amines resulted in excellent yields of products (Table 2, entries 1 and 12-15). However, no product was obtained by aniline as a substrate (Table 2, entry 16). Curiously, the reaction of aliphatic alkyne 1-octyne, which is typically difficult to activate, reacted well and provided the product in high yield (Table 2, entries 17–20).

The proposed mechanism is demonstrated in Scheme 2. At first, phenyl acetylene molecules due to interaction p-complex attached to the AgNPs immobilized on $Fe_3O_4(@R. tinctorum involving.$ On the other hand, the aldehyde is condensed with the secondary amine to give an iminium ion. In the next step, the iminium molecules attach to the AgNPs. Finally, coupling between iminium and phenyl acetylene molecules was performed. The catalytic cycle continues until the completion of the reaction.

Recovery of heterogeneous noble catalysts is a very important point. Consequently, the current research is aimed to recycle the catalyst for the reaction system of benzaldehyde, morpholine, and phenylacetylene under the optimal parameter. For this purpose, after compilation of the reaction, the catalyst was separated from the reaction medium by a magnet and washed with H_2O and EtOH twice, and then it was recovered up to seven times without significant decrease in its catalytic performance (Figure 11). This recoverability shows the high stability and turnover of catalyst under working conditions.

Moreover, to study the catalyst heterogeneity, for the A^3 coupling of the above model substrates, a hot filtration test was performed with Fe₃O₄@*R. tinctorum*/Ag NPs under the optimum conditions. The reaction yield of 65% could be attained after 4 h, and then, the catalyst was isolated using an external magnet. No increase in the activity of the product was observed when the reaction was continued for an additional 4 h following the isolation of the catalyst. This outcome proved the heterogeneous property of the synthesized catalyst. The TEM and EDX data (Figure 12) of the catalyst following the seven cycles confirmed the protection of the catalyst's nanostructure.

Table 2. Reactions of Aldehydes, Amines, and Alkynes in the Presence of Fe₃O₄@R. tinctorum/Ag NP Catalyst^a

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$R^{3} + R^{1}CHO - Fe_{3}O_{4}@R. tinctorum/Ag NPs $ $R^{2}_{2}NH$ $R^{2}_{2}NH$									
entry	\mathbb{R}^1	amine	R ³	yield (%) ^b	TOF $(h^{-1})^c$	TON $(h^{-1})^d$			
1	Ph	morpholine	Ph	96	120	960			
2	4-ClC ₆ H ₄	morpholine	Ph	95	118.7	950			
3	3-ClC ₆ H ₄	morpholine	Ph	92	115	920			
4	$4-BrC_6H_4$	morpholine	Ph	96	120	960			
5	4-OHC ₆ H ₄	morpholine	Ph	90	112.5	900			
6	$4-MeC_6H_4$	morpholine	Ph	88	110	880			
7	4-OMeC ₆ H ₄	morpholine	Ph	85	106.2	850			
8	2-thiophenyl	morpholine	Ph	95	118.7	950			
9	2-furfuryl	morpholine	Ph	90	112.5	900			
10	cyclohexyl	morpholine	Ph	92	115	920			
11	C_3H_7	morpholine	Ph	88	110	880			
12	Ph	piperidine	Ph	96	120	960			
13	Ph	pyrrolidine	Ph	92	112.5	900			
14	Ph	diethyl	Ph	90	112.5	900			
15	Ph	dibenzyl	Ph	90	112.5	900			
16	Ph	aniline	Ph	0	0	0			
17	Ph	morpholine	$n - C_6 H_{13}$	85	106.2	850			
18	Ph	piperidine	$n - C_6 H_{13}$	80	106.2	800			
19	$4-ClC_6H_4$	morpholine	$n - C_6 H_{13}$	85	106.2	850			
20	4-OMeC ₆ H ₄	morpholine	$n - C_6 H_{13}$	70	87.5	700			

^{*a*}Reaction conditions: aldehyde (1.0 mmol), amine (1 mmol), alkyne (1.2 mmol), and $Fe_3O_4@R$. *tinctorum*/Ag NPs (10 mg, 0.1 mol %) were stirred in water (3.0 mL) at 80 °C for 8 h. ^{*b*}Yields are based on ¹H NMR. ^{*c*}TOF, turnover frequency (TOF = (yield/time)/amount of catalyst (mol)). ^{*d*}TON, turnover number (TON = yield/amount of catalyst (mol)).

Scheme 2. Proposed Mechanism for Synthesis of PPA in the Presence $Fe_3O_4@R$. *tinctorum*/Ag NP Catalyst





Figure 11. Recycling of the Fe $_3O_4$ @R. tinctorum/Ag NP catalyst.



Figure 12. (a) TEM and (b) EDX analysis of reused catalyst after the seventh run.

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Three catalysts were prepared with different loading amounts of Ag % (0.105, 0.29, and 0.44 mmol/g), which resulted in the increase of reaction rate with the increase of Ag in the Fe₃O₄@*R*. *tinctorum*/Ag composite (Table 3).

Table 3. Effect of Different Loading Amounts of Ag % on the A³-Coupling Reaction of Benzaldehyde, Morpholine, and Phenylacetylene

entry	Ag loading (mmol/g)	TOF (h^{-1})
1	0.105	120
2	0.29	205
3	0.44	294

To prove the ability of this technique for synthesis purposes, the scale-up production of PPAs was studied with 4-(1,3-diphenylprop-2-ynyl)morpholine (Table 2, entry 1) as a model reaction. The results are shown in Table 4. The A³-coupling

Table 4. Scale-Up Synthesis of the A³-Coupling Reaction of Benzaldehyde, Morpholine, and Phenylacetylene

scale (mmol)	isolated yield (%)
1	96
10	94
20	90
50	85
100	82
	scale (mmol) 1 10 20 50 100

reaction between benzaldehyde, phenylacetylene, and morpholine proceeded successfully, and the product was attained in high yields almost comparable to those in the small-scale reactions. The results of scale-up tests revealed that the purified product yield was 82% if the reaction scale was 100 mmol.

Numerous selected processes in reports and current process are compared in Table 5, which specify that $Fe_3O_4@R$. *tinctorum*/Ag NPs are equally or more efficient catalysts in terms of reaction time and yield compared to previously reported ones.

CONCLUSIONS

This research introduced an efficient, easy, green, and economical technique for synthesizing $Fe_3O_4@R$. *tinctorum* as a metal reductant and stabilizer agent and used it for the preparation of $Fe_3O_4@R$. *tinctorum*/Ag NPs. The character-

Table 5. Comparison Efficiency of Fe₃O₄@*R. tinctorum*/Ag NPs with Some Reported Methods for the A³ Coupling Reaction (Phenylacetylene/Benzaldehyde/Piperidine)

entry	reaction conditions	temp. (°C)	$\begin{array}{c} TOF \\ (h^{-1}) \end{array}$	yield %
1	$Fe_3O_4@R. tinctorum/Ag NPs, H_2O$	80	120	this work
2	AgI, H ₂ O, N ₂	100	3.3	76
3	Ag NPs, PEG	100	1.92	81
4	Ag–NaY, neat	100	0.97	82
5	Ag–CIN-1, H ₂ O	40	2.72	16c
6	ZnO-IL/Ag, H ₂ O, 3 h	reflux	51.1	83
7	PS-NHC-Ag(1), solvent	RT	4	84
8	g-C ₃ N ₄ /Ag(0), EtOH/H ₂ O,	80	245	85

ization of particles was performed using the FT-IR, SEM, energy-dispersive X-ray spectroscopy (EDS), TEM, XRD, XPS, and VSM methods. The achieved $Fe_3O_4@R$. *tinctorum*/Ag NPs were reported to be air-stable, economic, and effective for the preparation of PPAs through one-pot three-component A^3 coupling of amines, aldehydes, and alkynes. The products of the A^3 -coupling reaction were obtained in good to excellent yields and the catalyst was recovered for seven runs with no loss in its catalytic performance. The main properties of the suggested method do not need homogeneous catalysts, providing high product yields; use of a plant extract as a green and fast synthetic method for modifying MNPs; no need of surfactants, capping agents, or templates; easy immobilization of Ag NPs; and catalyst recoverability.

EXPERIMENTAL SECTION

Materials and Apparatus. This part is according to our previously work.⁸⁶

Preparations of Fe₃**O**₄*@R. tinctorum by R. tinctorum* **Extract.** *R. tinctorum* powder (0.5 g, Ronas) was poured into 50 ml of Milli-Q water and agitated at a temperature of 50 °C for 20 min. Next, the extract was filtered through Whatman no. 1 filter paper and centrifuged at 4000 rpm for 5 min to separate unwanted aggregates. The filtered extract is utilized for the following phase.

For the preparation of $Fe_3O_4@R$. *tinctorum* NPs in the first stage, magnetite NP (500 mg) was distributed in 100 mL of water and sonicated for 20 min. Then, the *R. tinctorum* extract was poured into the mixture. Subsequently, the solution was agitated for 3 days at ambient temperature, and then, the $Fe_3O_4@R$. *tinctorum* NPs precipitate achieved was isolated by magnetic decantation and washed several times with deionized water. Finally, it is dried in a vacuum oven at 40 °C for 12 h.

Preparation of the Fe₃O₄@*R. tinctorum***/Ag NPs.** The Fe₃O₄@*R. tinctorum* NPs (500 mg) were distributed in deionized water (200 mL) via sonication for 30 min. Then, a solution containing AgNO₃ (30 mg) in 20 mL of H₂O was poured into dispersion and the solution was agitated for 5 h at ambient temperature to guarantee full reduction of Ag(1) ions in the precursor solution. Next, the Fe₃O₄@*R. tinctorum*/Ag NPs were isolated by magnetic decantation and rinsed with H₂O and acetone to eliminate the unattached substrates. Scheme 1 showed the synthetic process of Fe₃O₄@*R. tinctorum*/Ag NPs. The last nanocatalyst was dried in vacuum at a temperature of 40 °C. The amount of silver was 0.105 mmol/g, which was measured by ICP-AES.

General Procedure for the Synthesis of PPAs Using $Fe_3O_4@R$. *tinctorum*/Ag NPs. The $Fe_3O_4@R$. *tinctorum*/Ag catalyst (10 mg, 0.1 mol %) was added to the mixture of aldehyde (1 mmol), phenylacetylene (1.2 mmol), and amine (1 mmol) in H₂O (3 mL) and the mixture was agitated at a temperature of 80 °C. At the end of the reaction, the catalyst was isolated using a magnet, rinsed with ethanol, and dried for another cycle. The product mixtures were isolated with EtOAc and concentrated to obtain the crude product. In the next step, the crude product was purified using a silica gel column chromatography (70:30, hexane/EtOAc). All compounds were identified and characterized using spectral analysis or melting points.^{74–85}

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Notes

The authors declare no competing financial interest.

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