

Crizotinib inhibition of *ROS1*-positive tumours in advanced non-small-cell lung cancer: a Canadian perspective

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ABSTRACT

The *ROS1* kinase is an oncogenic driver in non-small-cell lung cancer (NSCLC). Fusion events involving the *ROS1* gene are found in 1%–2% of NSCLC patients and lead to deregulation of a tyrosine kinase-mediated multi-use intracellular signalling pathway, which then promotes the growth, proliferation, and progression of tumour cells. *ROS1* fusion is a distinct molecular subtype of NSCLC, found independently of other recognized driver mutations, and it is predominantly identified in younger patients (<50 years of age), women, never-smokers, and patients with adenocarcinoma histology.

Targeted inhibition of the aberrant *ROS1* kinase with crizotinib is associated with increased progression-free survival (PFS) and improved quality-of-life measures. As the sole approved treatment for *ROS1*-rearranged NSCLC, crizotinib has been demonstrated, through a variety of clinical trials and retrospective analyses, to be a safe, effective, well-tolerated, and appropriate treatment for patients having the *ROS1* rearrangement.

Canadian physicians endorse current guidelines which recommend that all patients with nonsquamous advanced NSCLC, regardless of clinical characteristics, be tested for *ROS1* rearrangement. Future integration of multigene testing panels into the standard of care could allow for efficient and cost-effective comprehensive testing of all patients with advanced NSCLC. If a *ROS1* rearrangement is found, treatment with crizotinib, preferably in the first-line setting, constitutes the standard of care, with other treatment options being investigated, as appropriate, should resistance to crizotinib develop.

Key Words *ROS1*; oncogenic drivers; non-small-cell lung cancer, advanced; NSCLC, advanced; targeted therapy; crizotinib; molecular testing; NSCLC, nonsquamous

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INTRODUCTION

Non-small-cell lung cancer (NSCLC) is the most common malignant tumour and a leading cause of death worldwide¹, with an estimated 1.6 million new global diagnoses annually². Most patients are diagnosed with advanced-stage disease, which is characterized by a poor survival rate³. Until recently, NSCLC was approached therapeutically as a single-entity disease. The standard first-line treatment for advanced (unresectable or metastatic) NSCLC that had the most efficacy was platinum-based

doublet chemotherapy, which resulted in median survival durations of 10–12 months^{4–6}. Subsequent recognition of the genetic diversity and heterogeneity of NSCLC changed the focus to identifying new molecular subsets of NSCLC, with emphasis placed on identifying driver oncogenes and novel biomarkers^{3,7}. Identification of those driver mutations and the capability to analyze the molecular profiles of NSCLC tumours dramatically altered the treatment

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paradigm by identifying actionable target mutations^{7–9}, because a potentially targetable genetic driver alteration is present in nearly half of all cases of metastatic adenocarcinoma⁶. Those targeted treatments have proved to be more effective than standard doublet chemotherapy (either platinum- or non-platinum-based) in increasing PFS, and the resultant increases in quality of life (QOL) and survival have led to the adoption of screening for predictive biomarkers as a standard of care^{1,9}. To date, the most prevalent targetable mutations identified in NSCLC predominantly involve the deregulation of tyrosine kinase receptor-mediated signalling (as seen in *EGFR* and *ALK* mutations), which drives both the initiation and progression of cancer cells^{3,10}.

The *ROS1* oncogene, which is mutated in a variety of solid tumours and which also results in the deregulation of a tyrosine kinase-mediated signalling pathway, was identified specifically in NSCLC in 2007¹¹. Interchromosomal—and occasionally intrachromosomal—rearrangements of *ROS1* result in gene fusions involving the 3' region of *ROS1*, including the kinase domain, and several different 5' fusion partners^{2,6,12}, of which 26 have been identified to date¹³. All *ROS1* fusions show conservation of the *ros1* kinase domain^{2,12} and lead to tyrosine kinase activation^{2,12,13}, a multi-use intracellular pathway involved in the upregulation of SHP-1 and SHP-2 and resultant activation of the PI3K/AKT/mTOR, JAK/STAT, and MAPK/ERK pathways, which act in concert to promote cell survival and proliferation^{7,14}.

ROS1 fusions exist as a distinct molecular subtype of NSCLC and rarely overlap with other oncogenic drivers such as *EGFR*, *KRAS*, *HER2*, *RET*, *MET*, and *ALK*¹⁵. Specifically, *ROS1* and *ALK* are mutually exclusive, with no evidence of co-expression, but are phylogenetically related^{7,15,16}, sharing 70% homology and 77% similar amino acid identity within ATP binding sites¹⁷. *ROS1*- and *ALK*-positive patients also share many clinicopathologic features: female sex, younger age at diagnosis (<50 years), propensity toward Asian ethnicity, never-smoking history, adenocarcinoma histology, and advanced nonresectable (compared with advanced resectable) disease at diagnosis are frequent characteristics of patients positive for either *ALK* or *ROS1*^{7,8,15}. Unique to patients with *ROS1* rearrangement is the observation that *ros1* expression is higher in recurrent tumours than the primary tumour (28% vs. 19%)¹⁸, and that patients who are *ROS1*-positive, compared with those who are *ALK*-positive, have lower rates of extrathoracic metastases, including lower rates of brain metastases at initial metastatic diagnosis, and a cumulative lower incidence of brain metastases^{18,19}.

After the discovery of the *ROS1* fusion gene as an oncogenic driver in NSCLC, and in light of the close homology between the *ALK* and *ROS1* tyrosine kinase domains, the utility of crizotinib as a *ros1* inhibitor was explored^{19,20}. Oral crizotinib, an ATP-competitive small-molecule tyrosine kinase inhibitor, was developed as a c-MET inhibitor; it was later found to have activity against *ALK*-rearranged tumours^{6,21} when a phase I single-arm analysis of crizotinib (PROFILE 1001) yielded a response rate of 60% and PFS of 9.7 months²¹.

Based on those results and preliminary data from a single-arm phase II study (PROFILE 1005), accelerated regulatory approval for the use of crizotinib in *ALK*-positive

locally advanced or metastatic NSCLC, was awarded by the U.S. Food and Drug Administration and Health Canada in 2011 and 2012 respectively^{6,8,14,22,23}. Subsequently, both *in vivo* and *in vitro*, crizotinib was found to be a highly robust inhibitor of the *ros1* fusion protein, showing up to 5 times greater potency in the suppression of *ros1* activity and downstream signalling—and resultant superior inhibition of *ros1*-driven tumour growth—than what had been observed in *ALK*-rearranged tumours¹⁹. Subsequent clinical trials of crizotinib in *ROS1*-rearranged NSCLC yielded response rates of 70%–80%, and approval for crizotinib in the management of *ROS1*-positive locally advanced or metastatic NSCLC was granted in 2016 by the U.S. Food and Drug Administration and in 2017 by Health Canada for use in the first- and subsequent-line settings²⁴. To date, crizotinib remains the only approved targeted agent for *ROS1*-rearranged advanced NSCLC^{14,20}, and *ROS1*-rearranged NSCLC is now the 3rd genetically distinct population of NSCLC that can be managed through approved, effective targeted therapy^{7,25}.

ROS1 TESTING

Testing Method

Reliable and efficient detection of tumours harbouring *ROS1* fusions is required to identify patients whose treatment protocols should include *ros1* inhibition. Currently, no companion diagnostic that reliably selects patients with *ROS1* alterations has been approved.

At present, *ROS1* fusion in tumour cells can be detected using a variety of techniques: fluorescence *in situ* hybridization (FISH), immunohistochemistry (IHC), reverse transcriptase polymerase chain reaction (RT-PCR), and next-generation sequencing of RNA and DNA^{26–28}.

ROS1 break-apart FISH is currently considered the “gold standard” and is used globally for many *ROS1*-crizotinib studies because of its low tissue requirement and high sensitivity and specificity^{29–31}. However, FISH has some limitations: it is labour-intensive and more costly than IHC, and interpretation of the results requires experience²⁵ because false-negative results can occur when the *ROS1* fusion partner gene is located within several megabases of the *ROS1* gene on chromosome 6^{32,33}. Next-generation sequencing and RT-PCR both show utility. The former allows for multiplex testing, has the potential to identify the *ROS1* fusion partner, and can detect novel fusions, but has a higher tissue requirement, is relatively more expensive than IHC or FISH, and yields more information than is often clinically relevant²⁹. The latter is limited given the requirement for multiple primer sets and an incapacity to identify novel or rare *ROS1* fusions³². In comparison, IHC is widely used in routine pathology practice, is less expensive and usually automated³², and generally shows good sensitivity (compared with FISH results) for *ROS1* screening when IHC uses the commercially available D4D6 antibody clone²⁵. However, IHC positive staining has greater discordance with FISH, because some tumours can yield samples that are IHC-positive, but that test negative for rearrangement by FISH^{8,29,34}. The Canadian ROS Initiative, which involves 14 pathology laboratories in Canada and 1 in Japan, is working to validate IHC and FISH testing for *ROS1* translocations in

NSCLC tumour samples³⁵ and is using a strategy of IHC as a screening test, followed by confirmation of IHC-positive cases by FISH^{26,34}. The high level of optimization and validation for a specific purpose, as it applies to all predictive assays, also applies to *ROS1* IHC testing^{28,36–39}. Looking to the future, effective screening methods for *ROS1* rearrangements that hinge on inexpensive, rapid, sensitive, reliable methods and development of a minimally invasive method that can also identify the fusion partner, secondary mutations, or tumour heterogeneity would be of considerable clinical utility^{40,41}.

Testing Recommendations

Screening for actionable mutations in NSCLC are recommended by the U.S. National Comprehensive Cancer Network's clinical practice guidelines in oncology, the European Society for Medical Oncology's guidelines, the American College of American Pathologists, the International Association for the Study of Lung Cancer, the Association for Molecular Pathology, the Expert Committee of Lung Cancer Canada, and the American Society of Clinical Oncology. Unanimously, those groups recommend that *ROS1* testing be performed for all patients with advanced lung adenocarcinoma^{9,26,35,42,43}. The 2018 updated joint guideline from the American College of American Pathologists, the International Association for the Study of Lung Cancer, and the Association for Molecular Pathology²⁶ (endorsed by the American Society of Clinical Oncology⁴³) now advises that all patients with advanced-stage adenocarcinoma, regardless of other clinical characteristics, be offered either a comprehensive lung panel [*EGFR*, *ALK*, *ROS1*, *BRAF*, *MET*, *ERBB2 (HER2)*, *KRAS*, and *MET*] or targeted testing for genes in the "must test" category (*EGFR*, *ALK*, *ROS1*), with the option of offering expanded panels that include additional genes [*BRAF*, *MET*, *ERBB2 (HER2)*, and *RET*] to patients who are clinical trial candidates, the latter possibly after testing negative in single-panel *KRAS* testing²⁶.

Because targeted massively parallel or DNA- or RNA-based next-generation sequencing panels that enable the simultaneous analysis of a large number of genes and of multiple actionable fusion transcripts, including *ROS1*, are integrated into the health care setting, comprehensive testing of patients presenting with advanced lung adenocarcinoma might prove to be efficient and cost-effective. It would allow for extensive molecular characterization of limited amounts of tumour tissue and achieve the mandate for *ROS1* testing to be integrated for all patients with advanced lung adenocarcinoma as the standard of care^{44,45}.

EVIDENCE OF BENEFIT WITH CRIZOTINIB FOR TARGETED INHIBITION OF *ROS1*-REARRANGED TUMOURS

Clinical Trials

PROFILE 1001: *ROS1*-Positive Expansion Cohort

Originally designed as a single-arm, multi-cohort, multi-centre phase I study to determine the efficacy and safety of crizotinib to treat *ALK*-rearranged locally advanced or metastatic NSCLC², the PROFILE 1001 trial was amended to add an expansion cohort of *ROS1*-positive NSCLC patients

after *in vitro* evidence showed that crizotinib was an effective suppressor of *ROS1* activity, leading to decreased downstream signalling and inhibition of tumour growth^{6,7}. The expansion cohort contained 53 *ROS1*-positive patients, determined by FISH, with no previous use of *ALK* or *c-MET* inhibitors. The objective response rate by independent radiology review was 70% [95% confidence interval (CI): 56% to 82%], with a median duration of response of 17.6 months¹⁷, median PFS of 19.3 months (95% CI: 14.8 months to not reached), and a demonstrated 91% (95% CI: 79% to 96%) survival probability at 6 months⁴⁶. The safety profile of crizotinib in patients with *ROS1*-positive disease was similar to that seen in the *ALK*-positive treatment environment: grades 1 and 2 adverse events including nausea, vomiting, edema, diarrhea, and vision disturbances were experienced by 38%–85% of patients. Grade 3 adverse events such as hypophosphatemia (13%), neutropenia (9%), and elevated transaminases (4%) were present, but no grade 4 adverse events or deaths attributable to treatment were reported⁴⁶. Additionally, response to crizotinib in *ROS1*-positive disease was achieved regardless of previous lines of therapy⁶ and independent of the percentage of *ROS1*-rearranged cells detected⁴⁶.

In parallel to the results from the crizotinib-treated *ALK*-positive disease in the same trial, crizotinib treatment in *ROS1*-positive disease similarly demonstrated that crizotinib is associated with a well-tolerated, rapid, and durable response¹⁰. The outcome measures from the study were the first to confirm the clinically meaningful benefit and safety of crizotinib in patients with *ROS1*-altered advanced NSCLC⁴⁶ and led to the approval of crizotinib use in that population⁴⁷.

EUCROSS

A collaboration between the Lung Cancer Group in Cologne and the Spanish Lung Cancer Group resulted in the development of a prospective phase II trial to evaluate the use of crizotinib in *ROS1*-positive lung adenocarcinoma, regardless of previous lines of treatment. The study enrolled 34 patients identified as *ROS1*-positive by FISH, who were treated with crizotinib. Of the 34 patients, 20 had sufficient tumour tissue to perform CAGE (cap analysis of gene expression) to verify *ROS1* status, identifying the exact break-apart point and fusion genes⁴⁸. *ROS1* fusion was confirmed in 18 patients; the 2 remaining patients were ultimately determined to be negative for *ROS1* rearrangement and quickly experienced primary progression. Analysis of the 18 patients with dually confirmed *ROS1* rearrangement showed an objective response rate of 83% (95% CI: 67.7% to 94.2%). The assessment of safety considered all 34 patients, and adverse events (any grade) were reported in just under 50% of the group⁴⁸.

The study confirmed that crizotinib is a safe treatment and, in the subset of validated *ROS1*-positive patients, highly effective. The lack of concordance observed between FISH and CAGE sequencing of *ROS1* in 2 of 20 patients who underwent validation of their *ROS1* status, and the failure of crizotinib to show clinical benefit in those deemed *ROS1* wild-type through CAGE sequencing, highlights the efficacy of CAGE sequencing in the identification of clinically sensitive *ROS1* gene rearrangements, and the need for orthogonal validation of *ROS1* status⁴⁸.

ACSe Study

A multicentric trans-tumour study, the phase II ACSe trial (NCT0163950 at <http://ClinicalTrials.gov/>), designed by the French National Cancer Institute, is considering the efficacy and safety of crizotinib as monotherapy in patients with *ALK*-, *ROS1*- (by FISH), or *MET*-positive tumours experiencing progression after at least 1 standard treatment (unless performance status has precluded first-line chemotherapy). The trial was designed to include 23 unique “cohorts,” including a *ROS1*-rearranged NSCLC cohort, with the goal of avoiding uncontrolled off-label use and allowing for nationwide safe access to crizotinib for patient populations demonstrating clinical benefit from this agent⁴⁹.

Preliminary results from the 29 patients in the *ROS1*-rearranged NSCLC cohort (secondarily confirmed by IHC) demonstrated an objective response rate of 63% (95% CI: 41% to 81%) and a 53% disease control rate at 6 months. Grade 1 adverse events were recorded in approximately 50% of patients, and grade 3 or greater adverse events were recorded in 31% of patients. Study completion and updated trial results were anticipated in spring 2019⁴⁹.

The preliminary results of ACSe reinforce the importance of integrating *ROS1* biomarker screening as part of routine care, because crizotinib has been demonstrated to be a safe, effective treatment with clinical benefit for patients harbouring *ROS1* rearrangements⁴⁹.

OxOnc Development Study

OxOnc (NCT01945021 at <http://ClinicalTrials.gov/>) was a phase II trial conducted as an open-label, multinational, and multicentre single-arm study of crizotinib in East Asian patients with advanced (locally advanced or metastatic) *ROS1*-positive NSCLC, not previously receiving targeted therapy for *ALK* or *ROS1*^{47,50}.

Of 127 patients with *ROS1*-positive disease (detected by RT-PCR) enrolled, 72% (95% CI: 63% to 79%) achieved an objective response. Median time to objective response was 1.9 months (range: 1.5–15.8 months), and the median duration of response was 19.7 months (95% CI: 14.1 months to not reached). Median PFS was 15.9 months (95% CI: 12.9 months to 24 months), with a disease control rate of 80% (95% CI: 72% to 87%) after 16 weeks on treatment, and a survival probability of 83% (95% CI: 75% to 89%) after 12 months of treatment⁴⁷. Treatment-related adverse events were noted in 96.1% of patients, mostly grade 1 or 2 in severity, and included elevated transaminases, vision disorders, nausea, diarrhea, and vomiting. Grades 3 and 4 events were reported in 25.2% of patients and included neutropenia and elevated transaminases. Dose reductions or interruptions attributable to grade 1 or 2 and grade 3 or 4 adverse events occurred in 15.7% and 22.8% of patients respectively, with 1 patient discontinuing crizotinib because of a grade 1 adverse event (diarrhea)⁴⁷. Assessments of QOL using the European Organisation for Research and Treatment of Cancer 30-question core Quality of Life Questionnaire and the 13-question lung cancer module revealed either stable (37%) or improved (46.8%) global QOL scores, compared with baseline scores, after 20 cycles of treatment, with statistically significant and clinically meaningful improvements in many lung cancer-related symptoms reported during those first 20 cycles, although

significant deterioration from baseline was observed for gastrointestinal symptoms⁴⁷.

This study provided clinical confirmation of the benefit of crizotinib through a high overall response rate, a rapid and durable response, and overall QOL improvement, confirming the known safety profile of crizotinib. On the basis of the study results, crizotinib was approved for the treatment of *ROS1*-positive NSCLC in Japan, Taiwan, Korea, and China in 2017⁴⁷.

Retrospective Reviews

EUROS1

The EUROS1 European retrospective review (France, Switzerland, Italy, Germany, Poland, Netherlands) was designed to characterize the outcomes of patients with *ROS1*-positive (identified by FISH) stage IV NSCLC with an adenocarcinoma histology, who had undergone documented (off-label) crizotinib therapy and 0 to 3 or more prior lines of therapy^{6,51}. In the 32 patients identified as meeting the study criteria, median PFS was 9.1 months, with an objective response rate of 80%, a disease control rate of 86.6%, and no reports of unexpected or serious adverse events⁵¹.

The review confirmed that *ROS1*-rearranged NSCLC is very sensitive to crizotinib¹. In the retrospective EUROS1 trial, unlike the prospective clinical trials, comorbidities or health status did not unselect patients for inclusion, and yet the response rate was similar to that in the PROFILE 1001 *ROS1*-positive expansion cohort. Results from EUROS1 demonstrated that the findings from the highly selected patient populations in the phase I clinical trials of crizotinib could be replicated in the real-world general population of patients with *ROS1*-rearranged NSCLC⁵¹.

China: Efficacy of Crizotinib and Pemetrexed-Based Therapy in Chinese Patients with *ROS1*-Rearranged NSCLC

This retrospective review of 51 Chinese patients with *ROS1*-rearranged disease (determined by RT-PCR) who received either crizotinib, pemetrexed, or non-pemetrexed therapy demonstrated statistically significant differences in PFS, with crizotinib demonstrating the highest PFS (294 days), followed by pemetrexed-based chemotherapy (179 days) and non-pemetrexed chemotherapy (110 days).

Those findings corroborate previous results showing that, compared with patients having other identified driver mutations and receiving pemetrexed, patients with *ROS1* rearrangement experience increased clinical benefit from pemetrexed chemotherapy²⁵, suggesting that *ROS1* rearrangement might be a marker of increased pemetrexed sensitivity¹. Further, despite the efficacy of pemetrexed in this population of patients with *ROS1* rearrangement, those results reinforce the superior efficacy of crizotinib in the treatment of Chinese patients with *ROS1*-rearranged NSCLC.

MEETING THE CHALLENGE OF PROGRESSIVE DISEASE

Acquired Resistance

Development of acquired resistance to crizotinib in *ROS1*-rearranged tumours poses a serious clinical challenge,

given that most patients treated using this agent will acquire resistance¹⁹ and that the duration of response to crizotinib cannot yet be predetermined and seems to have no relation to the *ROS1* fusion partner⁵². Resistance to crizotinib, and resulting disease progression, comes about by a variety of mechanisms: development of secondary mutations within the kinase domain, which impedes drug binding¹⁴; epithelial-to-mesenchymal transition^{47,53}; or upregulation and activation of compensatory pathways¹⁴ such as *EGFR*, *RAS*, and *KIT*¹⁹.

Development of secondary crizotinib-resistant mutations appears to account for most acquired resistance, and the molecular changes involved in crizotinib resistance show a high level of heterogeneity⁵³. The most common secondary mutation, G2032R [c.6094G>A (p.Gly2032Arg)], accounts for 41% of identified secondary mutations¹⁹, and it is unclear whether crizotinib use selects for pre-existing resistant clones or whether the evolution of crizotinib-resistant cells occurs during a period of exposure¹⁹. Given the diverse mechanisms that lead to crizotinib resistance, sequential treatment targeting crizotinib-resistant cells, or dual inhibition of *ROS1* and potentially upregulated pathways, might show efficacy in minimizing and managing resistance to crizotinib¹⁴.

Although secondary mutations in *ROS1* and *ALK* show overlapping sensitivity profiles⁴⁰, sequential therapy using second-generation *ALK* inhibitors to combat crizotinib resistance in *ROS1*-rearranged tumours seems limited in *ROS1*-positive NSCLC. Secondary mutations in *ROS1* tend to harbour off-target mechanisms of resistance, such as bypass tracks²⁰, and most show decreased sensitivity to second-generation *ALK* inhibitors¹⁹. Indeed, the second-generation *ALK* inhibitors—ceritinib, brigatinib, and entrectinib (*STARTRK-1*, *STARTRK-2*, and *ALKA-372-001* trials⁵⁴)—have been associated with clinically meaningful responses in crizotinib-treated patients with *ROS1*-rearranged tumours and with increased disease control rates for intracranial disease²⁰; however, none has shown effective inhibition against *ROS1*-rearranged tumours harbouring the common secondary G2032R mutation¹⁹, limiting use of those agents as second-line therapy²⁰.

Therapeutic Options Beyond Progression

Targeted agents such as DS-605-1, repotrectinib [TPX-005 (see NCT03093116 at <http://ClinicalTrials.gov/>)], lorlatinib (NCT01970865), cabozantinib, and foretinib have demonstrated anti-*ROS1* activity in the second-line setting, including activity against G2032R, with all but the latter two agents demonstrating good tolerability, with safety and efficacy data that are being confirmed in ongoing clinical trials^{19,47,55,a}. Cabozantinib has been shown to be effective, but to be associated with higher toxicity, and it is therefore limited as a therapeutic agent for some patients^{3,14,16,47}. Foretinib has been withdrawn from the market (NCT02034097).

With a current paucity of suitable second-line treatments for use in crizotinib-resistant *ROS1*-rearranged tumours, two methods of management have shown promise

as second-line treatments. The conventional cytotoxic chemotherapy agent pemetrexed has been associated with an objective response rate of 40%–58% and a PFS of 6.8–7.5 months in various lines of treatment and is therefore a viable treatment option for patients with *ROS1*-rearranged crizotinib-resistant disease^{1,25}. Alternatively, crizotinib resistance resulting from crizotinib-mediated upregulation of bypass signalling pathways (*EGFR*, *RAS*, and *KIT*)¹⁹ could be managed through targeted agents designed to modulate those upregulated systems, such as afatinib or PF29984 (*EGFR*)⁵³ and ponatinib (*KIT*)¹⁴.

As the options for treatment beyond crizotinib are explored, it remains true that desirable treatments post-crizotinib have to be highly potent agents with central nervous system penetrability and activity against *ROS1* G2032R²⁰. Appropriate treatments and management strategies for patients with *ROS1*-rearranged disease could then rely on a personalized approach in which repeat molecular characterization, both temporally and spatially, which captures the heterogeneity of *ROS1*-rearranged tumours and tailors therapies appropriately, should be engaged¹⁴.

RECOMMENDATIONS

As Canadian physicians involved in the management of patients with advanced lung cancer, we recommend molecular testing (inclusive of IHC), comprising detection of *ROS1* rearrangements, directly or indirectly by detecting *ROS1* chimeric RNA or fusion protein expression in tumours, because such testing is critical to the appropriate and timely therapeutic management of NSCLC. The testing should be offered as part of the standard of care to patients presenting with advanced disease, regardless of clinical characteristics³⁵. Given that *ROS1*-rearranged NSCLC represents a molecularly distinct subset of NSCLC, the ideal standard of care for these patients is targeted therapy with a *ROS1*-inhibiting agent.

Crizotinib has demonstrated clinical benefit and a favourable benefit–risk profile for patients with advanced NSCLC and *ROS1* rearrangement, and it is the first targeted agent approved for *ROS1*-positive tumours. Response rates achieved with crizotinib, regardless of treatment line (63%–83%), in this susceptible population are greatly superior to the 10%–35% and 5%–22% response rates obtained with use of the traditional cytotoxic therapies in the first-line and second-line settings respectively⁶. Low rates of *ROS1* rearrangement in the population make the initiation of phase III randomized clinical trials untenable at present, but the observed objective response rate, prolonged PFS, and similar efficacy across all lines of therapy as evidenced by a variety of phase I and II studies, retrospective analyses, and single-institution experiences in diverse patient populations with advanced NSCLC lend credence to the efficacy of crizotinib as an effective pharmaceutical to manage *ROS1*-altered lung cancer in larger patient populations. In light of current results and experiences, we support and recommend the use of crizotinib in this patient group.

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REFERENCES

- Zhang L, Jiang T, Zhao C, *et al.* Efficacy of crizotinib and pemetrexed-based chemotherapy in Chinese NSCLC patients with *ROS1* rearrangement. *Oncotarget* 2016;7:75145–54.
- Davies KD, Le AT, Theodoro MF, *et al.* Identifying and targeting *ROS1* gene fusions in non–small cell lung cancer. *Clin Cancer Res* 2012;18:4570–9.
- Quintanal-Villalonga A, Paz-Ares L, Ferrer I, Molina-Pinelo S. Tyrosine kinase receptor landscape in lung cancer: therapeutic implications. *Dis Markers* 2016;2016:9214056. [Erratum in: *Dis Markers* 2018;2018:3714684]
- Reck M, Popat S, Reinmuth N, De Ruysscher D, Kerr KM, Peters S on behalf of the ESMO Guidelines Working Group. Metastatic non-small-cell lung cancer (NSCLC): ESMO clinical practice guidelines for diagnosis, treatments and follow-up. *Ann Oncol* 2014;5(suppl 3):ii27–39.
- Ramalingam S, Belani C. Systemic chemotherapy for advanced non–small cell lung cancer: recent advances and future directions. *Oncologist* 2008;13(suppl 1):5–13.
- Kazandjian D, Blumenthal GM, Luo L, *et al.* Benefit–risk summary of crizotinib for treatment of patients with *ROS1* alteration–positive, metastatic non–small cell lung cancer. *Oncologist* 2016;21:974–80.
- Gainor JF, Shaw AT. Novel targets in non–small cell lung cancer: *ROS1* and *RET* fusions. *Oncologist* 2013;18:865–75.
- Marchetti A, Barberis M, di Lorito A, *et al.* *ROS1* gene fusion in advanced lung cancer in women: a systematic analysis, review of the literature, and diagnostic algorithm. *JCO Precis Oncol* 2017;1:1–9.
- Kerr KM, Bubendorf L, Edelman MJ, *et al.* Second ESMO Consensus Conference on Lung Cancer: pathology and molecular biomarkers for non-small-cell lung cancer. *Ann Oncol* 2014;25:1681–90.
- Melosky B, Agulnik J, Albadine R, *et al.* Canadian consensus: inhibition of *ALK*-positive tumours in advanced non-small-cell lung cancer. *Curr Oncol* 2016;23:196–200.
- Rikova K, Guo A, Zeng, Q, *et al.* Global survey of phosphotyrosine signaling identifies oncogenic kinases in lung cancer. *Cell* 2007;131:1190–203.
- Bubendorf L, Buttner R, Al-Dayel F, *et al.* Testing for *ROS1* in non–small cell lung cancer: a review with recommendations. *Virchows Arch* 2016;469:489–503.
- Uguen A, De Braekeleer M. *ROS1* fusions in cancer: a review. *Future Oncol* 2016;12:1911–28.
- Alrifai D, Forster MD, Janes SM. Emerging resistant pathways in lung cancer: what has *ROS-1* taught us? *Transl Lung Cancer Res* 2018;7(suppl 1):S9–12.
- Korpanty GJ, Graham DM, Vincent MD, Leighl NB. Biomarkers that currently affect clinical practice in lung cancer: *EGFR*, *ALK*, *MET*, *ROS-1* and *KRAS*. *Front Oncol* 2014;4:204.
- Katayama R, Kobayashi Y, Friboulet L, *et al.* Cabozantinib overcomes crizotinib resistance in *ROS1* fusion positive cancer. *Clin Cancer Res* 2015;21:166–74.
- Shaw AR, Ou SH, Bang YJ, *et al.* Crizotinib in *ROS1* rearranged non-small-cell lung cancer. *N Engl J Med* 2014;371:1963–71.
- Lee HJ, Seol HS, Kim JY, *et al.* *ROS1* receptor tyrosine kinase, a druggable target, is frequently overexpressed in non–small cell lung carcinomas via genetic and epigenetic mechanisms. *Ann Surg Oncol* 2013;20:200–8.
- Gainor JF, Tseng D, Yoda S, *et al.* Patterns of metastatic spread and mechanisms of resistance to crizotinib in *ROS1*-positive non-small-cell lung cancer. *JCO Precis Oncol* 2017;1:1–13.
- Dagogo-Jack I, Shaw AT. Expanding the roster of *ROS1* inhibitor. *J Clin Oncol* 2017;35:2595–7.
- Camidge DR, Bang YJ, Kwak EL, *et al.* Activity and safety of crizotinib in patients with *ALK*-positive non-small-cell lung cancer: updated results from a phase 1 study. *Lancet Oncol* 2012;13:1011–19.

22. Mullard A. 2011 FDA drug approvals. *Nat Rev Drug Discov* 2012;11:91–4.
23. Poon CC, Kelly JJ. Development of crizotinib, a rationally designed tyrosine kinase inhibitor for non–small cell cancer. *Int J Cancer* 2017;140:1945–54.
24. Pfizer Canada. Xalkori approved by Health Canada for the treatment of patients with ROS1-positive locally advanced or metastatic non–small cell lung cancer [online news release]. Kirkland, QC: Pfizer Canada; 2017. [Available at: <https://www.pfizer.ca/xalkori%C2%AE-approved-health-canada-treatment-patients-ros1-positive-locally-advanced-or-metastatic-1>; cited 9 May 2018]
25. Juan O, Popat S. Crizotinib for ROS1 patients: one small step in biomarker testing, one giant leap for advanced NSCLC patients. *Lung Cancer* 2017;104:131–3.
26. Lindeman NI, Cagle PT, Aisner DL, et al. Updated molecular testing guideline for the selection of lung cancer patients for treatment with targeted tyrosine kinase inhibitors. *J Mol Diagn* 2018;20:129–59.
27. Bergethon K, Shaw AT, Ou SH, et al. ROS1 rearrangements define a unique molecular subclass of lung cancers. *J Clin Oncol* 2012;30:863–70.
28. Torlakovic EE, Cheung CC, D'Arrigo C, et al. on behalf of the International Society for Immunohistochemistry and Molecular Morphology and the International Quality Network for Pathology. Evolution of quality assurance for clinical immunohistochemistry in the era of precision medicine. Part 3: Technical validation of immunohistochemistry (IHC) assays in clinical IHC laboratories. *Appl Immunohistochem Mol Morphol* 2017;25:151–9.
29. Lin JJ, Ritterhouse LL, Ali SM, et al. ROS1 fusions rarely overlap with other oncogenic drivers in non–small cell lung cancer. *J Thorac Oncol* 2017;12:872–7.
30. Varella-Garcia M, Yoshida A. ROS1 testing with FISH. In: Tsao MS, Hirsch FR, Yatabe Y, eds. *IASLC Atlas of ALK and ROS1 Testing in Lung Cancer*. 2nd ed. Aurora, CO: IASLC Editorial Rx Press; 2016: 53–62.
31. Scholl L, Yoshida A, Nicholson A, Lantuejoul S, Hirsch F. ROS1 testing with IHC. In: Tsao MS, Hirsch FR, Yatabe Y, eds. *IASLC Atlas of ALK and ROS1 Testing in Lung Cancer*. 2nd ed. Aurora, CO: IASLC Editorial Rx Press; 2016: 35–40.
32. Boyle TA, Masago K, Ellison KE, Yatabe Y, Hirsch F. ROS1 immunohistochemistry across major genotypes of non-small-cell lung cancer. *Clin Lung Cancer* 2015;16:106–11.
33. Rossi G, Jocolle G, Conti A. Detection of ROS1 rearrangement in non–small cell lung cancer: current and future perspectives. *Lung Cancer (Auckl)* 2017;8:45–55.
34. Selinger CI, Li BT, Pavlakis N, et al. Screening for ROS1 gene rearrangements in non–small cell lung cancers using immunohistochemistry with FISH confirmation is an effective method to identify this rare target. *Histopathology* 2017;70:402–11.
35. Meloksy B, Blais N, Cheema P, et al. Standardizing biomarker testing for Canadian patients with advanced lung cancer. *Curr Oncol* 2018;25:73–82.
36. Torlakovic EE, Cheung CC, D'Arrigo C, et al. on behalf of the International Society for Immunohistochemistry and Molecular Morphology and the International Quality Network for Pathology. Evolution of quality assurance for clinical immunohistochemistry in the era of precision medicine. Part 2: Immunohistochemistry test performance characteristics. *Appl Immunohistochem Mol Morphol* 2017;25:79–85.
37. Torlakovic EE, Neilsen S, Francis G, et al. Standardization of positive controls in diagnostic immunohistochemistry: recommendations from the International Ad Hoc Expert Committee. *Appl Immunohistochem Mol Morphol* 2015;23:1–18.
38. Torlakovic EE, Francis G, Garratt J, et al. on behalf of the International Ad Hoc Expert Panel. Standardization of negative controls in diagnostic immunohistochemistry: recommendations from the International Ad Hoc Panel. *Appl Immunohistochem Mol Morphol* 2014;22:241–52.
39. Cheung CC, D'Arrigo C, Dietel M, et al. on behalf of the International Society for Immunohistochemistry and Molecular Morphology and the International Quality Network for Pathology. Evolution of quality assurance for clinical immunohistochemistry in the era of precision medicine. Part 4: Tissue tools for quality assurance in immunohistochemistry. *Appl Immunohistochem Mol Morphol* 2017;25:227–30.
40. Facchinetti F, Loriot Y, Kuo MS, et al. Crizotinib-resistant ROS1 mutations reveal a predictive kinase inhibitor sensitivity model for ROS1 and ALK-rearranged lung cancers. *Clin Cancer Res* 2016;22:5983–91.
41. Sacher AG, Paweletz C, Dalhberg SE, et al. Prospective validation of rapid plasma genotyping for the detection of EGFR and KRAS mutations in advanced lung cancer. *JAMA Oncol* 2016;2:1014–22.
42. Riely GL. What, when and how of biomarker testing in non–small cell lung cancer. *J Natl Compr Canc Netw* 2017;15:686–8.
43. Kalemakerian GP, Narula N, Kennedy EB, et al. Molecular testing guideline for the selection of patients with lung cancer for treatment with targeted tyrosine kinase inhibitors: American Society of Clinical Oncology endorsement of the College of American Pathologists/International Association for the Study of Lung Cancer/Association for Molecular Pathology clinical practice guideline update. *J Clin Oncol* 2018;36:911–19.
44. Tsoulos N, Papadopoulou E, Metaxa-Mariatou V, et al. Tumor molecular profiling of NSCLC patients using next-generation sequencing. *Oncol Rep* 2017;38:3419–29.
45. Michels S, Wolf J. Stratified treatment in lung cancer. *Oncol Res Treat* 2016;39:760–6.
46. Shaw A, Riley GJ, Bang YJ, et al. Crizotinib in advanced ROS1-rearranged non–small cell lung cancer (NSCLC): updated results from PROFILE 1001. *Ann Oncol* 2016;27:416–54.
47. Wu YL, Yang CH, Kim DW, et al. Phase II study of crizotinib in East Asian patients with ROS1-positive advanced non-small-cell lung cancer. *J Clin Oncol* 2018;36:1405–11.
48. Michels S. EUCROSS: a European phase II trial of crizotinib in advanced adenocarcinoma of the lung harbouring ROS1 rearrangements—preliminary results [abstract MA07.05]. *J Thorac Oncol* 2017;12(suppl):S379–80.
49. Moro-Sibilot D, Falvre L, Zalcman G, et al. Crizotinib in patients with advanced ROS1-rearranged non–small cell lung cancer (NSCLC). Preliminary results of the acse phase II trial [abstract 8065]. *J Clin Oncol* 2015;33:. [Available online at: https://ascopubs.org/doi/abs/10.1200/jco.2015.33.15_suppl.8065; cited 15 July 2019]
50. Goto K, Yang JCH, Kim DW, et al. Phase II study of crizotinib in East Asian patients with ROS1-positive advanced non–small cell lung cancer [abstract 9022]. *J Clin Oncol* 2016;34:. [Available online at: https://ascopubs.org/doi/abs/10.1200/JCO.2016.34.15_suppl.9022; cited 15 July 2019]
51. Mazieres J, Zalcman G, Crino L, et al. Crizotinib therapy for advanced lung adenocarcinoma and a ROS1 rearrangement: results from the EUROS1 cohort. *J Clin Oncol* 2015;33:992–9.
52. Solomon B. Validating ROS1 rearrangements as a therapeutic target in non–small cell lung cancer. *J Clin Oncol* 2015;33:972–4.
53. Song A, Kim TM, Kim DW, et al. Molecular changes associated with acquired resistance to crizotinib in ROS1-rearranged non–small cell lung cancer. *Clin Cancer Res* 2015;21:2379–87.
54. Doebele RC, Ahn M, Siena S, et al. Efficacy and safety of entrectinib in locally advanced or metastatic ROS1 fusion-positive non–small cell lung cancer (NSCLC) [abstract OA02.01]. *J Thorac Oncol* 2018;13(suppl):S321–2.
55. Drilon A, Somwar R, Wagner JP, et al. A novel crizotinib-resistant solvent-front mutation responsive to cabozantinib in a patient with ROS1-rearranged lung cancer. *Clin Cancer Res* 2016;22:2351–8.