

The study of CTLA-4 and vitamin D receptor polymorphisms in the Romanian type 1 diabetes population

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Abstract

Several studies suggested that part of the genetic susceptibility for Type 1 diabetes (T1DM) is encoded by some polymorphisms of CTLA-4 gene (2q33) and of Vitamin D Receptor gene (VDR; 12q12-14). Our aim was to assess their contribution to T1DM genetic susceptibility in the Romanian population. We typed CTLA-4 49 A/G and VDR *FokI* (F/f), *ApaI* (A/a) and *TaqI* (T/t) polymorphisms by Sequence Specific Primer PCR (SSP-PCR) in 204 Romanian diabetic families (756 individuals: 212 T1DM probands and 544 unaffected parents and siblings). We studied alleles transmission using the Transmission Disequilibrium Test (TDT). We found an increased transmission of CTLA-4 49G allele to diabetics (54.8%, $p=0.11$). The transmission of F (56.1%, $p=0.063$), a (55.7%, $p=0.061$) and T (51.8%, $p=0.37$) alleles of VDR gene to diabetics was increased but did not reach statistical significance. In conclusion we found the same increased transmission of CTLA-4 49 G allele to diabetics as previously reported. VDR *FokI* F allele seems to be predisposing while *TaqI* T allele seems to be protective.

Keywords: type I diabetes • genetic • CTLA-4 • vitamin D receptor • Romania

Introduction

Type 1 diabetes mellitus (T1DM) or primary insulin dependent diabetes mellitus (IDDM) is a common chronic disease characterized by the autoimmune destruction of insulin secreting pancreatic cells. The

destruction is complete and results in absolute insulin dependency. T1DM is a multifactorial disease the etiology of which involves both genetic and environmental factors [1, 2, 3].

Genetically, T1DM is a complex, polygenic disease, with multiple susceptibility and protective alleles interacting with each other. The study of candidate genes identified the two major susceptibility genes for T1DM: *IDDM1* encoded in the HLA region of the Major Histocompatibility

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Complex (MHC) on chromosome 6p21 [1, 4, 5] and mapped to the *DRB1*, *DQB1* and *DQA1* loci [6, 7] and *IDDM2* encoded by the insulin gene region mapped to the Variable Number of Tandem Repeats (*VNTR*) region 5' of the insulin gene on chromosome 11p15 [8]. This approach also revealed the association of a third locus with T1DM, at least in some populations: *IDDM12* in the *CTLA-4* gene region on chromosome 2q [9, 10]. Using linkage analysis strategies by whole genome study, at least 20 other regions of the human genome were linked with T1DM [11, 12, 13] but none of these genes has been identified yet. All these "minor" genes have smaller contributions to T1DM genetic susceptibility. One of the two main T1DM susceptibility genes (*IDDM1* and *IDDM2*) is present in up to 90% of T1DM patients. This indicates that these main susceptibility genes are necessary for diabetogenesis. However they are not sufficient since around 8% of non-diabetic subjects carry the most diabetogenic haplotype (*IDDM1* and *IDDM2* positive). This suggests that in order to become diabetics, either some environmental factors must act on carriers of *IDDM1/IDDM2* or some of the "minor" genes should be involved.

We previously reported the *IDDM1* and *IDDM2* status for the Romanian population, characterized by one of the lowest T1DM incidences in Europe [14]. In this study on the Romanian population we aimed to study the role of two "minor genes: *CTLA-4* on chromosome 2q33 and *Vitamin D Receptor* (VDR) gene on chromosome 12q12-14. We tested for association with T1DM one *CTLA-4* gene and three VDR gene polymorphisms. For this, we typed 204 Romanian type 1 diabetic families using a unified method of genotyping multiple HLA and non-HLA polymorphisms under identical conditions employing the Sequence Specific Primer - SSP-PCR technique [15, 16, 17].

Materials and methods

Families selection

204 Romanian Type 1 diabetic families were genotyped in this study. We chose the Romanian population because of its very low reported incidence of Type 1 diabetes [18, 19]. Informed consent was obtained prior to blood collection for DNA extraction from all subjects involved and the local ethics committee

approved the study. The ascertainment of families was made according to EURODIAB Study protocol as previously described [14]. Most were simplex families (196) but there were 3 multiplex families and 5 families with one parent also affected. Most were 4 member families (144) and the other 60 were 3 member families. Overall the study group comprised 756 individuals; 371 M (49.07%)/385 F (50.93%) of which 212 type 1 diabetic patients (106 M/106 F) with the onset of disease between 9 months and 43 years. The median age at the onset of disease was 12.1±6.7 years. Most patients (166 - 78.3%) had diabetes onset under 17 years age. There were 544 unaffected individuals (parents and siblings), 265 M (48.71%)/279 F (51.29%).

Selection of genes

We selected for typing on the Romanian families the 49 A/G polymorphism of *CTLA-4* gene (chromosome 2q33) and *FokI* (F/f), *ApaI* (A/a) and *TaqI* (T/t) polymorphisms of *Vitamin D Receptor* gene (chromosome 12q12-14). All these polymorphisms were previously described [16, 17].

Primer design

The primers used for typing of *CTLA-4* and VDR gene polymorphisms were as previously reported [16, 17]. For all alleles we used an internal control for validation of amplification: *HLA-DRB1* intron 3 amplicon of 796 bp, forward primer TgCCAAgTggAgCACCCAA with annealing position exon 3 amino acid 173-179; reverse primer gCATCTTgCTCTgTgCAGAT with annealing position in exon 4 amino acid 193-200, both 12 ng/PCR amplification.

DNA extraction, PCR amplification and electrophoresis

DNA extraction was done as previously described [14]. PCR amplification was done in 13 µl per reaction using the SSP-PCR technique as previously described [15, 17, 20]. Subsequently, PCR products were electrophoresed through 1% agarose gels containing 0.5 g/ml ethidium bromide in 0.5xTBE buffer (89 mmol Tris base, 89 mmol boric acid, 2 mmol EDTA, pH 8.0) for 20 - 30 min at 15 V/cm. Following electrophoresis, the products were visualised with UV illumination and the gel photographed with a Polaroid camera. Gel interpretation was simple, requiring the scoring of the presence or absence of an amplicon. For validation of PCR amplification, all reactions included an internal control, a 796 bp product depending on the HLA amplicon size, as detailed previously [15, 17, 20]. An example is given in Fig. 1.

Statistical analysis

Data was initially analyzed using the Transmission Disequilibrium Test (TDT) [21]. The TDT tests the possibility that transmission of alleles from heterozygous parents to affected siblings is not the expected 50%. The TDT relative risk (RR) was calculated by dividing the number of times an allele was transmitted by the number not transmitted. After analysis we found a trend of association with some polymorphisms of Vitamin D receptor and CTLA-4 [22].

Results

The results of the TDT analysis for CTLA-4 49 A/G polymorphism are shown in Table 1 while those for VDR polymorphisms are shown in Table 2. Our data show an increased transmission of 49G allele of CTLA-4 to diabetics compared to the unaffected siblings (54.84%, $p=0.11$). The percentage is similar to those reported for other Caucasian populations [9] but with $p=0.11$ the result does not reach statistical significance on this data set. We found an increased transmission of *FokI* F allele to diabetics (56.14%, $p=0.063$) compared to the unaffected siblings (48.84% transmission) suggesting a possible predisposing effect. We also found an increased transmission of *TaqI* T allele to unaffected siblings (58.27%, $p=0.038$) compared to diabetics (51.82% transmission). This sustains that this allele could be protective. The transmission of *ApaI* alleles to diabetics was not different compared to the transmission to the unaffected siblings.

Discussion

In this study we used a candidate gene approach to ascertain the genetic component of T1DM susceptibility on the Romanian population, using 204 Romanian Type 1 diabetic families. For this we used an SSP-PCR technique, which enabled us to test HLA and non-HLA polymorphisms under identical PCR amplification conditions. Previously we tested the Romanian families for *IDDM1* and *IDDM2* susceptibility genes in order to validate genetically for Type 1 diabetes this population. The results confirmed the major role of DR/DQ alleles and of Ins-VNTR alleles in T1DM susceptibility for the Romanian population [14].

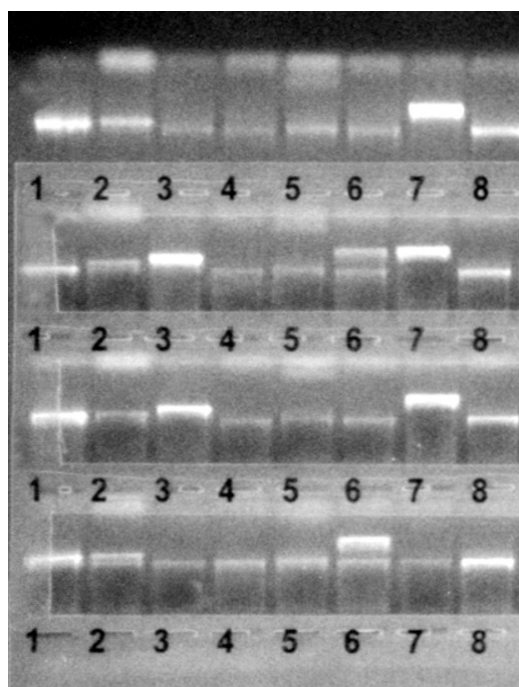


Fig. 1 The polaroid photograph of an electrophoresis gel visualised with UV illumination. PCR products for VDR polymorphisms on each row in positions 1, 2, 3, 4, 5, 6 and of CTLA4 polymorphisms in positions 7 and 8.

Using the genome wide scan technique, a region of linkage was described on the long arm of chromosome 2 (2q33), which was later assigned as *IDDM12*. In fact, in a 23 cM region of 2q31-2q35 at least three loci linked with T1DM were described: *IDDM7*, *IDDM12* and *IDDM13* [9, 23, 24]. CTLA-4 (Cytotoxic T Lymphocyte associated Antigen 4) gene on 2q33 is an ideal candidate gene for T1DM as an autoimmune disease. CTLA4 molecule is a member of the immunoglobulins superfamily and has multiple immunomodulatory actions. It mediates T cell apoptosis and negatively regulates T cell activation. It also downregulates expression of interleukin 2, an autocrine and paracrine growth factor responsible for continued T-cell proliferation.

An A/G transition in position 49 of exon 1 results in a Threonine to Alanine substitution in the CTLA-4 peptide. Initial studies showed that 49G allele is associated with T1DM in Italian and Spanish families but not for UK or USA [9]. Subsequently, the association was confirmed on a larger family collection from France, Spain and USA (Caucasian and Mexican-American) [25].

Table 1 TDT for CTLA 4 alleles on Romanian families.

	Polymorphism	T	NT	%T	p
Affected	CTLA-4 49 A	84	102	45.16%	0.89
Unaffected	CTLA-4 49 A	56	63	47.06%	0.71
Affected	CTLA-4 49 G	102	84	54.84%	0.11
Unaffected	CTLA-4 49 G	63	56	52.94%	0.29

T - Transmitted, NT - Not-transmitted, %T - Transmission percentage.

Another polymorphism in the 3' untranslated region of CTLA-4 (which shows linkage disequilibrium with 49 A/G polymorphism) showed a tendency of association with T1DM on UK and USA families [26]. An association of CTLA-4 polymorphisms with Hashimoto and Graves diseases was also shown [27, 28], supporting the contribution of this gene in the pathogeny of autoimmune diseases.

In our study on 204 Romanian diabetic families, we found the same increased transmission of

CTLA-4 49 G allele to diabetics (55%) as previous reported in other populations. This result, if replicated on a larger data set, will reach statistical significance and will confirm the role of CTLA-4 (*IDDM 12*) in T1DM genetic susceptibility for the Romanian population.

Vitamin D (1, 25 dihydroxy D3) has important immunoregulatory functions, including monocyte activation, inhibition of monocyte immunoglobulin receptor expression, inhibition of lymphocyte

Table 2 TDT for Vitamin D receptor alleles on Romanian families.

	Polymorphism	T	NT	%T	p
Affected	VDR ex2 f	75	96	43.86%	0.94
Unaffected	VDR ex2 f	66	63	51.16%	0.43
Affected	VDR ex2 F	96	75	56.14%	0.063
Unaffected	VDR ex2 F	63	66	48.84%	0.57
Affected	VDR intr8 a	113	90	55.67%	0.061
Unaffected	VDR intr8 a	79	62	56.03%	0.089
Affected	VDR intr8 A	90	113	44.33%	0.94
Unaffected	VDR intr8 A	62	79	43.97%	0.91
Affected	VDR ex9 t	66	71	48.18%	0.63
Unaffected	VDR ex9 t	53	74	41.73%	0.96
Affected	VDR ex9 T	71	66	51.82%	0.37
Unaffected	VDR ex9 T	74	53	58.27%	0.038

T - Transmitted, NT - Not-transmitted, %T - Transmission percentage.

activation and proliferation, suppression of immunoglobulin production, cytokine synthesis, MHC class II and CD4 expression on different cells [29]. There are several arguments for Vitamin D involvement in the pathogenesis of both murine and human T1DM. Thus, in NOD mice the development of diabetes can be prevented by administration of 1, 25(OH)2D3 which inhibits lymphocyte activation and restores the altered ratio of CD4/CD8 cells [30]. The data of EURODIAB epidemiologic multicentric study showed that the administration of Vitamin D supplements in newborns and young children is associated with a reduction of T1DM incidence [31].

Vitamin D exerts its metabolic and immunomodulatory actions via the nuclear Vitamin D Receptor (VDR). VDR belongs to the steroid receptor super-family and is widely expressed in many cell types, including lymphocytes, macrophages and pancreatic cells. So VDR is a natural candidate gene for T1DM. VDR gene is located on chromosome 12q12-14, it was completely sequenced and several polymorphisms were described. A C/T transition at the junction of intron 1 and exon 2 creates an initiation codon (ATG) three codons proximal to downstream start site, leading to a protein variant with three additional amino-acids [32]. This transition introduces a site recognized by *FokI* restriction enzyme. The two alleles of this polymorphism are F and f where f is recognized by *FokI*. *In vitro* studies demonstrated an increased transcription rate (1.7 fold) of the VDR gene in cells with the FF haplotype [32]. Other polymorphisms induced by restriction enzymes *BsmI* (alleles b/B) and *ApaI* (alleles a/A) in intron 8 and a „silent” T/C substitution in exon 9 which creates a *TaqI* restriction site (alleles t/T).

There are several studies regarding the association of VDR *FokI*, *BsmI*, *ApaI* and *TaqI* polymorphisms with T1DM. In 1997, McDermott reports an association of „bAT” haplotype with T1DM on a collection of Indian ancestry families living in UK. „BAT” haplotype was protective [33]. These results were reconfirmed on a family collection from Germany [34], but in this population „bAT” haplotype seems to be also protective rather than predisposing. However, the association was not confirmed on some US families [35].

In our study on 204 Romanian T1DM families, *FokI* F allele seems to be predisposing (56.14% transmission, p=0.063) while *TaqI* T allele seems to be protective (58.27% transmission, p=0.038). However these results must be replicated on a second family set ascertained in the same conditions as that examined in the present study.

In conclusion, our data sustain the role of CTLA-4 (*IDDM 12*) and Vitamin D Receptor in Type 1 diabetes genetic susceptibility for the Romanian population.

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References

1. **Cudworth A., Woodrow J.**, HL-A system and diabetes mellitus, *Diabetes*, **24**: 345-349, 1974
2. **Todd JA.**, A protective role of the environment in the development of type 1 diabetes?, *Diabetic Med.*, **8**: 906-910, 1991
3. **Kyvik K.O., Green A., Beck-Nielsen H.**, Concordance rates of insulin dependent diabetes mellitus: a population based study of young Danish twins, *B.M.J.*, **311**: 913-917, 1995
4. **Singal D.P., Blajchman M.A.**, Histocompatibility antigens, lymphocytotoxic antibodies and tissue antibodies in patients with diabetes mellitus, *Diabetes*, **22**: 429-432, 1973
5. **Nerup J., Platz P., Anderson O.O., Christy M., Lyngsoe J., Poulsen J.E., Ryder L.P., Nielsen L.S., Thomsen M., Svejgaard A.**, HLA antigens and diabetes mellitus, *Lancet*, **2**: 864-866, 1974
6. **Todd J.A., Bell J.I., McDevitt H.O.**, HLA-DQ beta gene contributes to susceptibility and resistance to insulin-dependent diabetes mellitus, *Nature*, **329**: 599-604, 1987
7. **Sheehy M.J., Scharf S.J., Rowe J.R., Neme D.G.M., Meske L.M., Erlich H.A., Nepom B.S.**, A diabetes-susceptible HLA haplotype is best defined by a combination of HLA-DR and -DQ alleles, *J. Clin. Invest.*, **83**: 830-835, 1989

8. **Bennett S.T., Lucassen A.M., Gough S.C.L., Powell E.E., Undlien D.E., Pritchard L.E., Merriman M.E. et al.**, Susceptibility to human type 1 diabetes at *IDDM2* is determined by tandem repeat variation at the insulin gene minisatellite locus, *Nature Genet.*, **9**: 284-292, 1995
9. **Nistico L., Buzzetti R., Pritchard L.E., Van der Auwera B., Giovannini C., Bosi E., Larrad M.T.M. et al.**, The *CTLA-4* gene region of chromosome 2q33 is linked to, and associated with, type 1 diabetes, *Hum. Mol. Genet.*, **5**: 1075-1080, 1996
10. **Todd J.A., Farrall M.**, Panning for gold: genome-wide scanning for linkage in type 1 diabetes, *Hum. Mol. Genet.*, **5**: 1443-1448, 1996
11. **Davies J.L., Kawaguchi Y., Bennett S.T., Copeman J.B., Cordell H.J., Pritchard L.E., Reed P.W., Gough S.C.L., Jenkins S.C., Palmer S.M., Balfour K.M., Rowe B.R., Farrall M., Barnett A.H., Bain S.C., Todd J.A.**, A genome-wide search for human type 1 diabetes susceptibility genes, *Nature*, **371**: 130-136, 1994
12. **Todd J.A.**, Genetic analysis of type 1 diabetes using whole genome approaches, *Proc. Natl. Acad. Sci. USA*, **92**: 8560-8565, 1995
13. **Mein C.A., Esposito L., Dunn M.G., Johnson G.C., Timms A.E., Goy J.V., Smith A.N., Sebag-Montefiore L., Merriman M.E., Wilson A.J., Pritchard L.E., Cucca F., Barnett A.H., Bain S.C., Todd J.A.**, A search for type 1 diabetes susceptibility genes in families from the United Kingdom, *Nat. Genet.*, **19**: 297-300, 1998
14. **Ionescu-Tîrgoviște C., Guja C., Herr M., Cucca F., Welsh K., Bunce M., Marshall S., Todd J.A.**, Low frequency of HLA DRB1*03 - DQB1*02 and DQB1*0302 haplotypes in Romania is consistent with the country's low incidence of type 1 diabetes, *Diabetologia*, **44**: B60-B66, 2001
15. **Bunce M., O'Neill C.M., Barnardo M.C.N.M., Krausa P., Browning M.J., Morris P.J., Welsh K.I.**, Phototyping: comprehensive DNA typing for HLA-A, B, C, DRB1, DRB3, DRB5 & DQB1 by PCR with 144 primer mixes utilizing sequence-specific primers (PCR-SSP), *Tissue Antigens*, **46**: 355-367, 1995
16. **McLaren A.J., Marshall S.E., Haldar N.A., Mullighan C.G., Fuggle S.V., Morris P.J., Welsh K.I.**, Adhesion molecule polymorphisms in chronic renal allograft failure, *Kidney International*, **55**: 1977-1982, 1999
17. **Mullighan C.G., Marshall S.E., Bunce M., Welsh K.I.**, Variation in immunoregulatory genes determines the clinical phenotype of common variable immunodeficiency, *Genes and Immunity*, **1**: 137-148, 1999
18. **Ionescu-Tîrgoviste C., Serban V., Guja C., Mota M., Creteanu G., Calin A., Morosanu M., Ferariu I., Halmagy I., Cristescu J., Strugariu M., Minescu A., Barbul R.**, Very low incidence of type 1 diabetes in children in Romania, *Diabetologia*, **42**: A85, 1999
19. **EURODIAB ACE Study Group.**, Variation and trends in incidence of childhood diabetes in Europe, *Lancet*, **355**: 873-876, 2000
20. **Bunce M.**, The development and applications of a single PCR-based method of HLA genotyping, PhD thesis, *Oxford Brookes University*, Oxford, 1996
21. **Spielman R.S., McGinnis R.E., Ewens W.J.**, Transmission test for linkage disequilibrium: the insulin gene region and insulin-dependent diabetes mellitus (IDDM), *Am. J. Hum. Genet.*, **52**: 506-516, 1993
22. **Guja C., Todd J.A., Smith A., Welsh K., Marshall S., Ionescu-Tîrgoviște C.**, Analysis of the *CTLA-4* and vitamin D receptor gene polymorphisms in Romanian type 1 diabetic families, *Diabetologia*, **43**: A7 (Abstract), 2000
23. **Copeman J.B., Cucca F., Hearne C.M., Cornall R.J., Reed P.W., Ronningen K.S. et al.**, Linkage disequilibrium mapping of a type 1 diabetes susceptibility gene (*IDDM7*) to human chromosome 2q31-q33, *Nat. Genet.*, **9**: 80-85, 1995
24. **Owerbach D., Gabbay K.H.**, The search for IDDM susceptibility genes: the next generation, *Diabetes*, **45**: 544-551, 1996
25. **Marron M.P., Raffel L.J., Garchon H.J., Jacob C.O., Serrano-Rios M., Larrad M.T.M et al.**, Insulin-dependent diabetes mellitus (IDDM) is associated with *CTLA4* polymorphisms in multiple ethnic groups, *Hum. Mol. Genet.* **6**: 1275-1282, 1997
26. **Esposito L., Hill N., Pritchard L.E., Cucca F., Muxworthy C., Merriman M.E. et al.**, Analysis of putative loci *IDDM7*, *IDDM12*, *IDDM13* and candidate genes *NRAMP1* and *IA-2* and the interleukin-1 gene cluster, *Diabetes*, **47**: 1797-1800, 1998
27. **Awata T., Kurihara S., Itaka M., Takei S.I., Inou I., Ishii C. et al.**, Association of *CTLA4* gene A-G polymorphism (*IDDM12* locus) with acute-onset and insulin depleted IDDM as well as autoimmune thyroid disease (Grave's disease and Hashimoto's thyroiditis) in the Japanese population, *Diabetes*, **47**: 128-129, 1998
28. **Buzzetti R., Petrone A., Mesturino G., Giorgi R., Fiori R., Nistico L. et al.**, Major role for *CTLA-4* gene in DR4-positive type 1 diabetic patients and in DR3 negative Grave's disease patients, *Diabetologia*, **42**: A74 (Abstract), 1999
29. **Thomasset M.**, Vitamin D and the immune system, *Pathol. Biol.*, **42**: 163-172, 1994
30. **Mathieu C., Waer M., Laureys J., Rutgeerts O., Bouillon R.**, Prevention of autoimmune diabetes in NOD mice by 1,25 dihydroxyvitamin D₃, *Diabetologia*, **37**: 552-558, 1994

31. **EURODIAB Substudy 2 Study Group.**, Vitamin D supplement in early childhood and risk for Type 1 (insulin-dependent) diabetes mellitus, *Diabetologia*, **42**: 51-54, 1999
32. **Gross C., Krishnan A.V., Malloy P.J., Eccleshall T.R., Zhao X.Y., Feldman D.**, The vitamin D receptor gene start codon polymorphism: a functional analysis of FokI variants, *J. Bone Miner Res.*, **13**: 1691-1699, 1998
33. **McDermott M.F., Ramachandran A., Ogunkolade B.W., Aganna E., Curtis D., Boucher B.J., Snehalatha C., Hitman G.A.**, Allelic variation in the vitamin D receptor influences susceptibility to IDDM in Indian Asians, *Diabetologia*, **40**: 971-975, 1997
34. **Pani M.A., Knapp M., Donner H., Braun J., Baur M.P., Usadel K.H., Badenhop K.**, Vitamin D receptor allele combinations influence genetic susceptibility to type 1 diabetes in Germans, *Diabetes*, **49**: 504-507, 2000
35. **Malecki M.T., Klupa T., Moczulski D., Warram J.H., Krolewski A.S.**, No evidence of association between vitamin D receptor (VDR) gene polymorphisms and type 1 diabetes in Caucasians, *Diabetologia*, **43**: A7 (Abstract), 2000