J. Cell. Mol. Med. Vol 8, No 1, 2004 pp. 77-84

Mini Review

Role of human prostasomes in the activation of spermatozoa

G. Arienti [a] *, E. Carlini [a], C. Saccardi [b], C. A. Palmerini [b] *

[a] Dipartimento di Medicina Interna, Sez di Biochimica,
[b] Dipartimento di Scienze Biochimiche e Biotecnologie Molecolari,
Università di Perugia, Italia

Received: October 10, 2003; Accepted: January 15, 2004

Abstract

Prostasomes are small vesicles of prostatic origin contained in human semen. Their composition is peculiar under many aspects. Cholesterol is abundant and many proteins are endowed with enzymatic or other activities. The function of prostasomes has been amply debated and several hypotheses have been put forward. The liquefaction of semen, spermatozoa motility, antibacterial activity and immunological functions have been related to prostasomes. Under certain aspects, prostasomes resemble synaptosomes. The fusion of prostasomes to spermatozoa enriches spermatozoa with cholesterol and causes bursts of cytoplasmic sperm calcium. The interaction of spermatozoa and prostasomes should be limited to vagina since prostasomes are immobile and do not follow spermatozoa in the superior female genital tract. Calcium bursts would increase spermatozoa motility, where cholesterol would decapacitate spermatozoa, so preventing untimely activation. Since spermatozoa receive many different molecules from prostasomes, additional effects are also possible. Prostasomes makes spermatozoa more apt to be activated by progesterone in the proximity of the ovum. Therefore, the fusion between spermatozoa and prostasomes would influence spermatozoa behaviour under many aspects and might be relevant for fecundation. The richness of molecular species in prostasomes is amazing and these small vesicles are expected to lead to many more discoveries in the field of human reproduction.

Keywords: acrosomal reaction - cytosolic calcium - membrane fusion - prostasomes - semen - spermatozoa

* Correspondence to: Giuseppe ARIENTI,
Dipartimento di Medicina Interna,
Sezione di Biochimica, Università di Perugia,
Via del Giochetto, 06126 Perugia.
Tel./Fax: +39.075.5857417.
E-mail: arienti@unipg.it

Prostasomes

Prostasomes are membranous vesicles secreted by the prostate gland. They were first discovered by Ronquist *et al*., in Sweden [1, 2] who also proposed for them the name of "prostasomes" because of their prostatic origin [1-7]. Prostasome-like particles have been found in the semen of other mammals [8-10]; however, their composition and amounts differ from those of human material. Moreover, different fecundation patterns (intravaginal vs intrauterine) may make these particles functionally dissimilar, depending on the species. The uncertainty of the production site is a major concern, when considering the prostasome-like particles contained in the semen of non-human mammals.

Prostasome morphology

Electron microscopy examination [11, 12] shows round particles surrounded by a membrane and containing electron-dense material. The size of prostasomes has been measured with various methods. Light scattering measurements [12, 13] confirmed microscopy data and revealed a population of particles with an average diameter of 150-200 nm.

Prostasome components

The lipid composition of prostasomes is peculiar; cholesterol is present in high amounts as is sphingomyelin, whereas phosphatidylcholine is less abundant [11, 12]. Therefore, these membranes differ amply from sperm plasma membranes [14, 15] that contain less sphingomyelin and more phosphatidylcholine with a cholesterol:phospholipid ratio of 0.83 [16]. The cholesterol to lipid phosphorus ratio in human prostasomes is about 2 [12]; this may be interesting since cholesterol may have roles in sperm capacitation [17]. Morever, prostasome lipid are rich in saturated fatty acid.

Prostasomes contain many small molecules and ions (Ca^{2+}, Zn^{2+}, GDP, ADP and ATP) and a number of enzymes among which phopholipase A, ATPase and peptidases [18-23], to list only few. Many prostasomal proteins have been recently identified through the analysis of proteome by Utleg *et al*. [24]. These authors found 139 proteins; enzymes account for 33.8% of total, transport structural proteins for 19.4%, chaperone proteins for 5.8%, GTP proteins for 14.4% and signal transduction proteins for 17.3%. Non-identified proteins were 9.4%.

Human semen is rich in prostasomes; ratio prostasome protein/spermatozoa protein is about 2. The presence of neuroendocrine markers such as chromogranin B, neuropeptide Y and vasoactive intestinal polypeptide [7] depicts prostasomes as neuroendocrine-like vesicles.

Physiological roles of prostasomes

Although prostasomes are small particles, they contain many different molecules. Moreover, several prostasomal components are found in other tissues and cell types where they exert known physiological roles. It is, therefore, reasonable to raise the question of their meaning in prostasomes.

The physiological significance of human prostasomes has been long debated and various possibilities have been put forward: among them, the enhancement of sperm motility [25], the liquefaction of semen [26] and immunosuppression [27-31]. The motility of spermatozoa is enhanced by prostasomes [25, 32-37] and antibacterial prop-

Table 1 Lipid content of human prostasomes.

Cholesterol (a)	08 ± 01
Total lipid phosphorus (a)	04 ± 0.1
Cholesterol/Phospholipid ratio	2.0 ± 0.8
Phosphatidylethanolamine (b)	15 ± 4
Phosphatidylcholine (b)	12 ± 3
Sphingomyelin (b)	53 ± 12
Phosphatidylserine	14 ± 3
Phosphatidylinositol	6 ± 3

(a) Results are expressed as $\mu mol.mg^{-1}$ protein ± SE
(b) Results are expressed as percentage of total lipid phosphorus in each phospholipid class ± SE

erties of prostasomes have been described [38, 39], including an antioxidant capacity [see ref. 40].

Prostasome to spermatozoa fusion

It has been reported that prostasomes interact [41] or even fuse [42] with spermatozoa. Interaction and/or fusion between spermatozoa and prostasomes may help throw some light on the role of prostasomes.

A fusion between prostasomes and spermatozoa has first been described by Arienti and coworkers [42] who used a lipophilic fluorescent derivative of rhodamine (octadecyl rhodamine G; R_{18}) to detect the movement of lipid from a membrane population to another [43]. The extent of fusion can be expressed as percentage, taking as 100% the theoretical maximal lipid mixing [44].

The fusion of prostasomes and spermatozoa is a pH and protein-dependent phenomenon (Fig. 2, 3). It increases with decreasing of the pH values of prostasome/spermatozoa mixtures and it is suppressed by the simultaneous destruction of prostasome and spermatozoa proteins. Usually, the fusion is investigated at pH 5.0, but it is still detectable at pH as high as 6 or 7. Yet, it is absent at pH 8. Since the average pH value of semen is around 7.6 [45] small variations of pH may trigger the fusion process. Moreover, in the so-called split ejaculation, spermatozoa appear to be emitted with the first fractions, together with the prostatic secretions, more acidic than the bulk of semen [46].

Fig. 1 Transmission electron micrograph of human semen prostasome vesicles.

The fusion between prostasomes and spermatozoa is never very large (10-20%), even in best conditions, but it is a quick phenomenon with a hyperbolic time course; it is practically finished 10 min after mixing prostasomes and spermato-

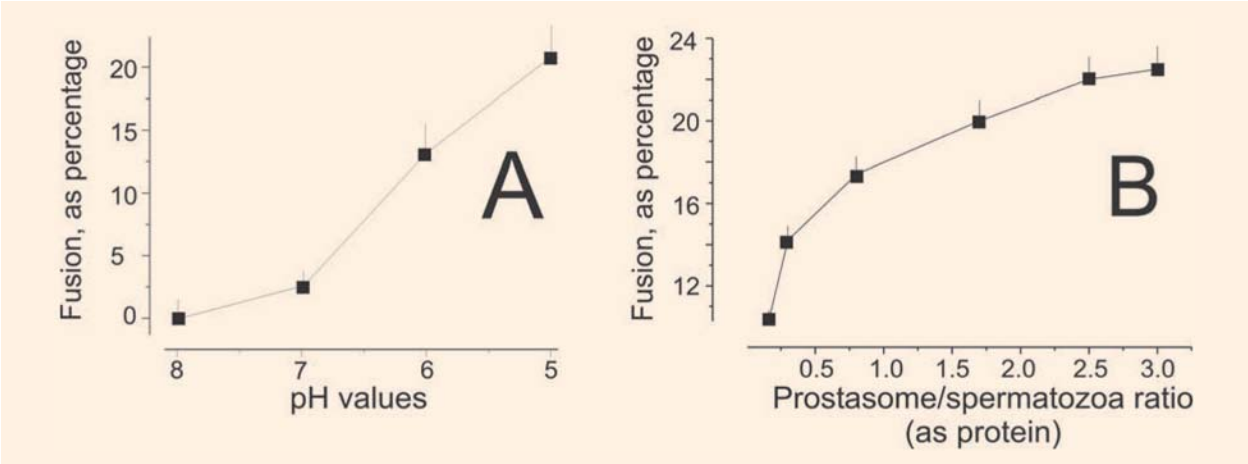

Fig. 2 Fusion of spermatozoa and prostasomes. (A) The effect of pH; (B) Effect of prostasome to spermatozoa ratio.

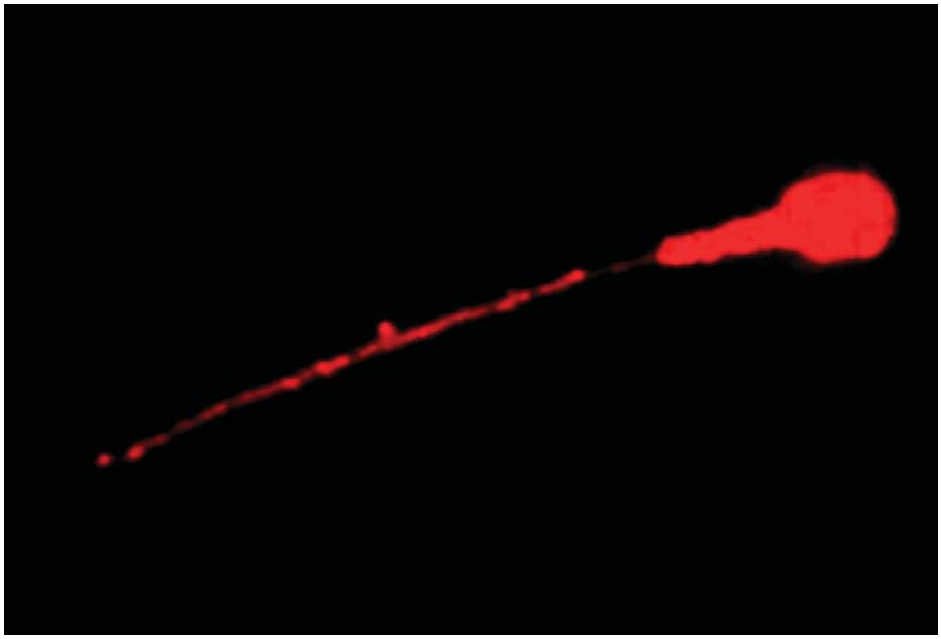

Fig. 3 Appearance of spermatozoa after fusion with R_{18}-loaded prostasomes. Spermatozoa were mixed for 110 min at pH 5.0 with R_{18}-labelled prostasomes. Magnification x 1000.

zoa [42] (Fig. 4). On the other hand, its quickness may have a physiological relevance, considering that the time of contact of prostasomes and spermatozoa at pH values low enough (below 7.0-7.5) to produce fusion is probably rather limited in physiological conditions.

Lipid and membrane-bound protein are both transferred through fusion [47-49] and some characteristics of sperm membranes, such as fluidity, change upon fusion with prostasomes [15]. The fusion appears to be specific for spermatozoa because it does not occur with lymphocytes [50], probably due to the necessary proteins, not yet identified. Other cell types have not been tested.

Spermatozoa cytosolic calcium

The fusion process may transfer all molecule types and ions contained in prostasomes besides lipid and protein. A particular attention has been given to calcium because it is contained in prostasomes [51] and is important for sperm motility and capacitation [52-54]. During the fusion process, calcium is released to spermatozoa as revealed by the increase of sperm $[Ca^{2+}]_i$. The increase of $[Ca^{2+}]_i$ is a transient process since the ion exchange readily and rapidly with extracellular Na^+. Upon omitting Na^+ from the external milieu, the increase of $[Ca^{2+}]_i$, parallels the extent of fusion as measured with R_{18} [55]. The bursts of

Table 2 Possible steps in spermatozoa activation.

Site	Event	Effect	Physiological meaning
Vagina	Spermatozoa/ prostasome fusion	↑ $[Ca^{2+}]_i$ ↑ Cholesterol Spermatozoa acquire a number of lipid and of protein	Increased motility Decreased ability to capacitate Long lasting effects
Uterus and tubae	Migration of spermatozoa	↓ Cholesterol	Spermatozoa acquire ability to capacitate
Near the ovum	Presence of progesterone	↑ $[Ca^{2+}]_i$ Acrosome reaction	Spermatozoa fused with prostasomes react better

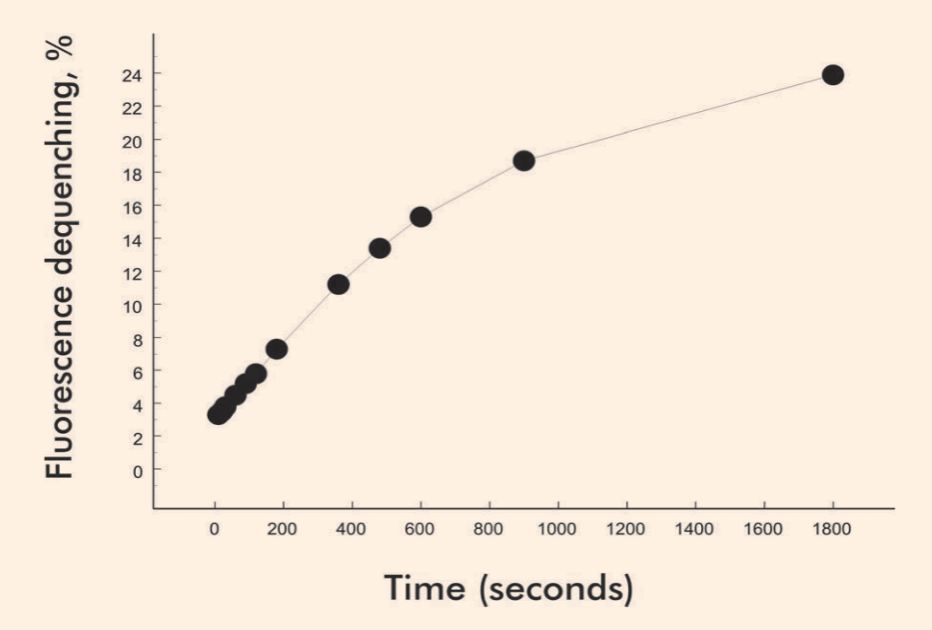

Fig. 4 Time-course of the fusion process. Spermatozoa were mixed with R_{18}-labelled prostasomes at pH 5.

spermatozoa $[Ca^{2+}]_i$ produced by the fusion with prostasomes appear to be physiologically in contrast with the delivery of cholesterol, since this lipid decreases the activation of sperm [17, 56-59], where calcium increases it. The simultaneous delivery of cholesterol and calcium, should activate the spermatozoa, increasing their motility (effect of $[Ca^{2+}]_i$) [37], although not permitting the occurrence of the acrosome reaction (effect of cholesterol). This fact would be interesting because the fusion with prostasomes must occur in vagina, whereas the acrosome reaction is a phenomenon taking place in the proximity of the oocyte.

Spermatozoa capacitation and acrosome reaction

Ejaculated human spermatozoa must be capacitated and undergo the acrosome reaction before fertilizing the oocyte [60]. The molecular mechanisms through which this is accomplished have amply been discussed. Attention has been given to the role of cholesterol to phospholipid ratio in sperm membranes [56, 57, 59, 61-63]. The loss of cholesterol during the migration of spermatozoa toward the ovum may be an activating factor.

The discovery that progesterone may act on spermatozoa through a mechanism that does not require a genomic pathway has thrown some more light on the activation of spermatozoa [64]. The effects of progesterone are mediated essentially by the increase of $[Ca^{2+}]_i$, the stimulation of the activity of phospholipases, the phosphorylation of proteins and efflux of chloride [65-68].

During fusion with prostasomes, lipids, proteins and ions [47-49, 55] can be delivered to spermatozoa. This is likely to further influence the behaviour of spermatozoa and their response to activating or inhibiting agents for some time after the fusion process.

The acrosome reaction is a necessary step in the sperm maturation. Before the passage through the epididymis, spermatozoa are unable to undergo the acrosomal reaction, but they have acquired this capacity when they reach the cauda epidydimis. Yet, spermatozoa are not allowed to acrosome-react, until they arrive in the proximity of the ovum. Therefore the cells maintain a sort of unstable equilibrium that allows them to acrosome react in the upper female genital tract, but not before.

The interaction with zona pellucida signals sperm to undergo the acrosome reaction [60] that can be initiated on capacitated spermatozoa following the activation of extragenomic progesterone receptors [65]. Progesterone also increases spermatozoa $[Ca^{2+}]_i$ and its effects are independent and additive to those produced by the fusion with prostasome [69].

In vagina, spermatozoa may acquire prostasomal components that would, directly or indirectly, modulate the acrosome reaction that takes place in the proximity of the ovum. Progesterone (1 μM at 37°C and in 30 min) does not induce the acrosome reaction [70], although spermatozoa cytosolic calcium increases in these conditions [69]. Usually, progesterone is required in concentrations about 10-15 μM for several hours to induce the acrosome reaction [59, 71, 72].

Previously fused spermatozoa were stimulated with 1 μM progesterone and the increase of sperm cytosolic calcium was larger than that due either to fusion or to progesterone alone [69]. Therefore progesterone and fusion show an independent and additive effect. The acrosome reaction was studied through the FITC-labeled pisum sativum agglutinin [73]. Fused spermatozoa underwent acrosome reaction better than the non-fused cells. This is in agreement with the finding the fused spermatozoa show a larger increase of $[Ca^{2+}]_i$ in the presence of low doses of progesterone [69].

Therefore, the fusion of prostasomes to spermatozoa, occurring in vagina, could alter the properties of spermatozoa producing several effects: (a) $[Ca^{2+}]_i$ bursts (stimulating spermatozoa movement); (b) delivery of cholesterol to spermatozoa (prevents an untimely acrosome reaction and (c) increased response to progesterone (calcium increase and improved acrosome reaction).

Conclusions

Since the time of their discovery, prostasomes, small particles contained in human semen that, since the time of their discovery, have revealed interesting properties. One of the most discussed points is their physiological relevance in human reproductive physiology. Some hypotheses have been put forward, but it is reasonable to think that future research will reveal much on this subject.

It is possible that the contact of spermatozoa with prostasomes may have a large part in spermatozoa maturation. The fusion of prostasomes and spermatozoa may help to keep spermatozoa in the delicate equilibrium involved in capacitation and acrosome reaction processes.

References

1. **Ronquist G., Brody I.,** The prostasome: its secretion and function in man, *Biochim. Biophys. Acta,* **822**: 203-218, 1985
2. **Brody I., Ronquist G., Gottfries A.,** Ultrastructural localization of the prostas, *Ups. J. Med. Sci.,* **88**: 63-80, 1983
3. **Carlsson L., Nilsson O., Larsson A., Stridsberg M., Sahlen G., Ronquist G.,** Characteristics of human prostasomes isolated from three different sources, *Prostate,* **54**: 322-330, 2003
4. **Nilsson B.O., Egevad L., Jin M., Ronquist G., Busch C.,** Distribution of prostasomes in neoplastic epithelial prostate cells, *Prostate,* **39**: 36-40, 1999
5. **Nilsson B.O., Jin MRonquist G.,** Immunolocalization of prostasomes in the human prostate, *Ups. J. Med. Sci.,* **101**: 149-157, 1996
6. **Sahlen G.E., Egevad L., Ahlander A., Norlen B.J., Ronquist G., Nilsson B.O.,** Ultrastructure of the secretion of prostasomes from benign and malignant epithelial cells in the prostate, *Prostate,* **53**: 192-199, 2002
7. **Stridsberg M., Fabiani R., Lukinius A., Ronquist G.,** Prostasomes are neuroendocrine-like vesicles in human semen, *Prostate,* **29**: 287-295, 1996
8. **Arienti G., Carlini E., De Cosmo A.M., Di Profio P., Palmerini C.A.,** Prostasome-like particles in stallion semen, *Biol. Reprod.,* **59**: 309-313, 1998
9. **Minelli A., Moroni M., Martinez E., Mezzasoma I., Ronquist G.,** Occurrence of prostasome-like membrane vesicles in equine seminal plasma, *J. Reprod. Fertil.,* **114**: 237-243, 1998
10. **Frenette G., Lessard C., Sullivan R.,** Selected proteins of "prostasome-like particles" from epididymal cauda fluid are transferred to epididymal caput spermatozoa in bull, *Biol. Reprod.,* **67**: 308-313, 2002
11. **Arvidson G., Ronquist G., Wikander G., Ojteg A.C.,** Human prostasome membranes exhibit very high cholesterol/ phospholipid ratios yielding high molecular ordering, *Biochim. Biophys. Acta,* **984**: 167-173, 1989
12. **Arienti G., Carlini E., Polci A., Cosmi E.V., Palmerini C.A.,** Fatty acid pattern of human prostasome lipid, *Arch. Biochem. Biophys.,* **358**: 391-395, 1998
13. **Bordi F., Cametti C., De Luca F., Carlini E., Palmerini C.A., Arienti G.,** Hydrodynamic radii and lipid transfer in prostasome self-fusion, *Arch. Biochem. Biophys.,* **396**: 10-15, 2001
14. **Mack S.R., Everingham J., Zaneveld L.J.D.,** Isolation and partial characterization of the plasma membrane from human spermatozoa, *J. Expt. Zool.,* **240**: 127-136, 1986
15. **Carlini E., Palmerini C.A., Cosmi E.V., Arienti G.,** Fusion of sperm with prostasomes: effects on membrane fluidity, *Arch. Biochem. Biophys.,* **343**: 6-12, 1997
16. **Poulos A., White I.G.,** The phospholipid composition of human spermatozoa and seminal plasma, *J. Reprod. Fertil.,* **53**: 265-272, 1973
17. **Benoff S.,** Preliminaries to fertilization. The role of cholesterol during capacitation of human spermatozoa, *Hum. Reprod.,* **8**: 2001-2008, 1993

18. **Fabiani R.,** Functional and biochemical characteristics of human prostasomes. Minireview based on a doctoral thesis, *Ups. J. Med. Sci.,* **99**: 73-111, 1994
19. **Ronquist G., Frithz G.G.,** Prostasomes in human semen contain ADP and GDP, *Acta Eur. Fertil.,* **17**: 273-276, 1986
20. **Ek P., Malm J., Lilja H., Carlsson L., Ronquist G.,** Exogenous protein kinases A and C, but not endogenous prostasome-associated protein kinase, phosphorylate semenogelins I and II from human semen, *J. Androl.,* **23**: 806-814, 2002
21. **Fabiani R., Ronquist G.,** Association of some hydrolytic enzymes with the prostasome membrane and their differential responses to detergent and PIPLC treatment, *Prostate,* **27**: 95-101, 1995
22. **Minelli A., Allegrucci C., Mezzasoma I., Ronquist G., Lluis C., Franco R.,** CD26 and adenosine deaminase interaction: its role in the fusion between horse membrane vesicles and spermatozoa, *Biol. Reprod.,* **61**: 802-808, 1999
23. **Olsson I., Ronquist G.,** Isoenzyme pattern of lactate dehydrogenase associated with human prostasomes, *Urol. Int.,* **45**: 346-349, 1990
24. **Utleg A.G., Yi E.C., Xie T., Shannon P., White J.T., Goodlett D.R., Hood L., Lin B.,** Proteomic analysis of human prostasomes, *Prostate,* **56**: 150-161, 2003
25. **Stegmayr B., Ronquist G.,** Promotive effect on human sperm progressive motility by prostasomes, *Urol. Res.,* **10**: 253-257, 1982
26. **Lilja H., Laurel C.B.,** Liquefaction of coagulated human semen, *Scand. J. Clin. Lab. Invest.,* **44**: 447-452, 1984
27. **Kelly R.W., Holland P., Skibinski G., Harrison C., McMillan L., Hargreave T., James K.,** Extracellular organelles (prostasomes) are immunosuppressive components of human semen, *Clin. Exp. Immunol.,* **86**: 550-556, 1991
28. **Skibinski G., Kelly R.W., Harkiss D., James K.,** Immunosuppression by human seminal plasma-extracellular organelles (prostasomes) modulate activity of phagocytic cells, *Am. J. Reprod. Immunol.,* **28**: 97-103, 1992
29. **Kelly R.W.,** Immunosuppressive mechanisms in semen: implications for contraception, *Hum. Reprod.,* **10**: 1686-1693, 1995
30. **Lazarevic M., Skibinski G., Kelly R.W., James K.,** Immunomodulatory effects of extracellular secretory vesicles isolated from bovine semen, *Vet. Immunol. Immunopathol.,* **44**: 237-250, 1995
31. **Babiker A.A., Ronquist G., Nilsson U.R., Nilsson B.,** Transfer of prostasomal CD59 to CD59-deficient red blood cells results in protection against complement-mediated hemolysis, *Am. J. Reprod. Immunol.,* **47**: 183-192, 2002
32. **Carlsson L., Ronquist G., Stridsberg M., Johansson L.,** Motility stimulant effects of prostasome inclusion in swim-up medium on cryopreserved human spermatozoa, *Arch. Androl.,* **38**: 215-221, 1997
33. **Fabiani R., Johansson L., Lundkvist O., Ronquist G.,** Prolongation and improvement of prostasome promotive effect on sperm forward motility, *Eur. J. Obstet. Gynecol. Reprod. Biol.,* **58**: 191-198, 1995
34. **Fabiani R., Johansson L., Lundkvist O., Ulmsten U., Ronquist G.,** Promotive effect by prostasomes on normal human spermatozoa exhibiting no forward motility due to buffer washings, *Eur. J. Obstet. Gynecol. Reprod. Biol.,* **57**: 181-188, 1994
35. **Fabiani R., Johansson L., Lundkvist O., Ronquist G.,** Enhanced recruitment of motile spermatozoa by prostasome inclusion in swim-up medium, *Hum. Reprod.,* **9**: 1485-1489, 1994
36. **Ronquist G., Stegmayr B., Brody I., Gottfries A.,** Prostasomes-a newly discovered organelle that increases sperm motility, *Lakartidningen,* **80**: 810-813, 1983
37. **Arienti G., Carlini E., Nicolucci A., Cosmi E.V., Santi F., Palmerini C.A.,** The motility of human spermatozoa as influenced by prostasomes at various pH levels, *Biol. Cell,* **91**: 51-54, 1999
38. **Carlsson L., Pahlson C., Bergquist M., Ronquist G., Stridsberg M.,** Antibacterial activity of human prostasomes, *Prostate,* **44**: 279-286, 2000
39. **Andersson E., Sorensen O.E., Frohm B., Borregaard N., Egesten A., Malm J.,** Isolation of human cationic antimicrobial protein-18 from seminal plasma and its association with prostasomes, *Hum. Reprod.,* **17**: 2529-2534, 2002
40. **Saez F., Motta C., Boucher D., Grizard G.,** Antioxidant capacity of prostasomes in human semen, *Mol. Hum. Reprod.,* **4**: 667-672, 1998
41. **Ronquist G., Nilsson B.O., Hjerten S.,** Interaction between prostasomes and spermatozoa from human semen, *Arch. Androl.,* **24**: 147-157, 1990
42. **Arienti G., Carlini E., Palmerini C.A.,** Fusion of human sperm to prostasomes at acidic pH, *J. Membr. Biol.,* **155**: 89-94, 1997
43. **Hoekstra D., de Boer T., Klappe K., Wilschut J.,** Fluorescent method for measuring the kinetic of fusion between biological membranes, *Biochemistry,* **23**: 5675-5681, 1984
44. **Corazzi L., Pistolesi R., Arienti G.,** The fusion of liposomes to rat brain microsomal membranes regulates phosphatidylserine synthesis, *J. Neurochem.,* **56**: 207-212, 1991
45. **Raboch J., Skaková J.,** The pH of human ejaculates, *Fertil. Steril.,* **16**: 252-256, 1965
46. **Meinertz H.,** Antisperm antibodies in split ejaculates, *Am. J. Reprod. Immunol.* **26**: 110-113, 1991
47. **Arienti G., Carlini E., Verdacchi R., Cosmi E.V., Palmerini C.A.,** Prostasome to sperm transfer of CD13/aminopeptidase N (EC 3.4.11.2), *Biochim. Biophys. Acta* **1336**: 533-538, 1997
48. **Arienti G., Carlini E., Verdacchi R., Palmerini C.A.,** Transfer of aminopeptidase activity from prostasomes to sperm, *Biochim. Biophys. Acta* **1336**: 269-274, 1997
49. **Arienti G., Polci A., Carlini E., Palmerini C.A.,** Transfer of CD26/dipeptidyl peptidase IV (E.C. 3.5.4.4) from prostasomes to sperm, *FEBS Lett.,* **410**: 343-346, 1997
50. **Arienti G., Carlini E., Saccardi C., Palmerini C.A.,** Interactions between prostasomes and leukocytes, *Biochim. Biophys. Acta,* **1425**: 36-40, 1998
51. **Stegmayr B., Berggren P.O., Ronquist G., Hellman B.,** Calcium, magnesium and zinc contents in organelles of

prostatic origin in human seminal plasma, *Scand. J. Urol. Nephrol.,* **16**: 199-203, 1982
52. **Suarez S.S.,** Hyperactivated motility in sperm, *J. Androl.,* **17**: 331-335, 1996
53. **Ahmad K., Bracho G.E., Wolf D.P., Tash J.S.,** Regulation of human sperm motility and hyperactivation components by calcium, calmodulin and protein phosphatases, *Arch. Androl.,* **35**: 187-208, 1995
54. **Suarez S.S., Dai X.,** Intracellular calcium reaches different levels of elevation in hyperactivated and acrosome-reacted hamster sperm, *Mol. Reprod. Develop.,* **42**: 325-333, 1995
55. **Palmerini C.A., Carlini E., Nicolucci A., Arienti G.,** Increase of human spermatozoa intracellular Ca^{2+} concentration after fusion with prostasomes. *Cell Calcium* **25**: 291-296, 1999
56. **Cross N.L.,** Human seminal plasma prevents sperm from becoming acrosomally reactive to the agonist progesterone: cholesterol is the major inhibitor. *Biol. Reprod.* **54**: 138-145, 1996
57. **Cross N.L., Mahasreshti P.,** Prostasome fraction of human seminal plasma prevents sperm from becoming acrosomally responsive to the agonist progesterone. *Arch. Androl.* **39**: 39-44, 1997
58. **Cross N.L., Razy-Faulkner P.,** Control of human sperm intracellular pH by cholesterol and its relationship to the response of the acrosome to progesterone. *Biol. Reprod.,* **56**: 1169-1174, 1997
59. **Cross N.L.,** Sphingomyelin modulates capacitation of human sperm in vitro. *Biol. Reprod.* **63**: 1129-1134, 2000.
60. **Snell W.J., White J.M.,** The molecules of mammalian fertilization, *Am. J. Reprod. Immunol.,* **28**: 97-103, 1996
61. **Davis B.K.,**. Timing of fertilization in mammals: sperm cholesterol/phospholipid ratio as a determinant of the capacitation interval, *Proc. Natl. Acad. Sci. USA,* **78**: 7560-7564, 1981
62. **Osheroff J.E., Visconti P.E., Valenzuela J.P., Travis A.J., Alvarez J., Kopf G.S.,** Regulation of human sperm capacitation by a cholesterol efflux- stimulated signal transduction pathway leading to protein kinase A- mediated up-regulation of protein tyrosine phosphorylation. *Mol. Hum. Reprod.,* **5**: 1017-1026, 1999
63. **Khorasani A.M., Cheung A.P., Lee C.Y.,** Cholesterol inhibitory effects on human sperm-induced acrosome reaction. *J. Androl.,* **21**: 586-594, 2000
64. **Calogero A.E., Burrello N., Barone N., Palermo I., Grasso U., D'Agata R.,** Effects of progesterone on sperm function: mechanisms of action, *Hum. Reprod.,* **15 Suppl 1**: 28-45, 2000
65. **Baldi E., Luconi M., Bonaccorsi L., Maggi M., Francavilla S., Gabriele A., Properzi G., Forti G.,** Nongenomic progesterone receptor on human spermatozoa: biochemical aspects and clinical implications, *Steroids,* **64**: 143-148, 1999
66. **Patrat C., Serres C., Jouannet P.,** The acrosome reaction in human spermatozoa. *Biol. Cell* **92**: 255-266, 2000.
67. **Kirkman-Brown J.C., Bray C., Stewart P.M., Barratt C.L.R., Publicover S.J.,** Biphasic elevation of $[Ca^{2+}]i$ in individual human spermatozoa exposed to progesterone, *Develop. Biol.,* **222**: 326-335, 2000
68. **Kirkman-Brown J.C., Barratt C.L., Publicover S.J.,** Nifedipine reveals the existence of two discrete components of the progesterone-induced $[Ca^{2+}]_i$ transient in human spermatozoa, *Dev. Biol.*, **259**: 71-82, 2003
69. **Arienti G., Nicolucci A., Santi F., Carlini E., Palmerini C.A.,** Progesterone-induced increase of sperm cytosolic calcium is enhanced by previous fusion of spermatozoa to prostasomes, *Cell Calcium,* **30**: 222-227, 2001
70. **Palmerini C.A., Saccardi C., Carlini E., Fabiani R., Arienti G.,** Fusion of prostasomes to human spermatozoa stimulates the acrosome reaction, *Fertil. Steril.,* **80**: in press, 2003
71. **Sirivaidyapong S., Bevers M.M., Gadella B.M., Colenbrander B.,** Induction of the acrosome reaction in dog sperm cells is dependent on epidymal maturation: the generation of a functional progesterone receptor is involved, *Mol. Reprod. Dev.,* **58**: 451-459, 2001
72. **Kuroda Y., Kaneko S., Yoshimura Y., Nozawa S., Mikoshiba K.,** Influence of progesterone and GABAA receptors on calcium mobilization during human sperm acrosome reaction. *Arch. Androl.,* **42**: 185-191, 1999
73. **Mendoza C., Carreras A., Moos J., Tesarik J.,** Distinction between true acrosomal reaction and degenerative acrosome loss by one-step staining method using pisum sativum agglutinin, *J. Reprod. Fertil.,* **95**: 763, 1992